

**Investigation of trace element geochemistry of moose  
(*Alces alces*) teeth apatite and possible links with  
increased incisoform breakage of Cape Breton Highland  
moose (*Alces alces andersoni*): A Pilot Study**

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## Abstract

Ungulates in the wild are normally free from periodontal disease. Yet, Cape Breton Highland (CBH) moose (*Alces alces andersoni*) of Nova Scotia have displayed an increased incidence of incisiform macro- and micro-fractures, which may have an effect on moose longevity. This condition appears to be rare, as it has only been formally documented in Alaskan (1992) moose (*Alces alces gigas*) and in Manitoban moose (*Alces alces andersoni*), and remains unexplained.

Furthermore, it has also been observed that the CBH moose are chewing their own fallen antlers, a condition known as osteophagia that is common within other ungulates but never before documented within moose populations, and they have also shown a marked increase in bark stripping activity. Both of these types of behaviour are usually associated with a dietary deficiency of some sort within other ungulate populations. Geochemical analyses of chewed fallen antlers from the CBH indicate that they have higher contents of calcium phosphate than those from the control area.

We have selected suites of broken and healthy teeth from the CBH and compared them with moose teeth from Shelburne County, NS, where no incidence of broken incisorform teeth has been documented. Fracture patterns in teeth, and especially tooth enamel (hydroxyapatite) were studied under the petrographic microscope and the electron microprobe. Enamel was carefully isolated from 25 representative samples of teeth from both the CBH and the control area and analyzed chemically by Inductively Coupled Plasma Mass Spectrometry (ICP-MS).

Samples from the problematic CBH area are significantly depleted in Barium, Lead, Strontium, Cobalt and Tin when compared against the control area, and some of these deficiencies have been associated with dental disease in animals elsewhere. Samples from the CBH were divided arbitrarily into two groups: A) those collected north of the CBH National Park, and B) those collected south of the Park. In each case, they were separated with respect of their perceived low, medium and high degree of fracturing, which to some degree correlates with the increasing age of the moose.

Correlation trends based on breakage score and elemental concentration within the CBH are not always consistent. A) North of the Park increased degree of fracturing correlates positively with contents of Cd, Mn, Nb, Rb, Sr, Y, Sn and Bi, and negatively with Al, Cu, Ti, Zn, Mg, As, Th and U. B) South of the Park increased degree of fracturing correlates positively with Ba, Mn, Sr, Mg, Se, As, Y, Th and U, and negatively with Al, Cr, Cu, Ga and Sn. The number of samples is so far insufficient to draw statistically significant conclusions, yet these are the first multielement data available for moose teeth in the region, and raise interesting possibilities. For instance, the much higher concentrations of Sn determined in incisorform teeth of the healthy control group coincides with the relatively high Sn concentrations in rocks and soils in southwestern Nova Scotia, (e.g. East Kemptville tin mine) suggesting that the geochemistry of teeth may be used effectively as a forensic tracer of the source of illegally hunted wildlife.

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## **Chapter 1: Introduction**

### **1.1 General Statement**

For centuries, moose have been an important resource for native North American populations. Recently, within the last 10 years, an increase in incisiform breakage has been documented within the Cape Breton Highland (C.B.H) moose population, leading to concerns for the overall health of the herd.

A previous thesis carried out by Walsh (2003), assembled the background to this problem, and presented descriptive microscopy including that done by Graves and Casey (2002, unpublished). Walsh (2003) also presented some preliminary, yet inconclusive, geochemical data done at the slowpoke 2 reactor in early 2002, reaching the conclusion that the geochemistry on the moose incisor teeth should be a fruitful line of research.

This thesis, which supersedes the above-mentioned study, took over where Walsh (2003) left it, reassesses the nature of breakage, and provides geochemical data for the enamel of unhealthy teeth from C.B.H, and also for a control population in Shelburne County where there is no evidence of breakage. The study suggests that increased incisiform breakage and antler chewing are problems associated with nutritional deficiencies experienced by the CBH moose.

Recent findings also suggest that moose within the C.B.H herd are chewing their own fallen antlers (osteophagia), a phenomenon only ever recorded in other cervids, and the apparent first documentation of such behaviour within moose. There has also been an increased documentation of heavy bark stripping amongst the C.B.H population. Osteophagia is triggered by a phosphorous deficiency, whilst bark stripping is attributed to a depletion of preferred browse.

### **1.2 Introduction**

Until recently, moose (*Alces alces americana*) were the dominant ungulate (Ungulate is Latin for "provided with hoofs") within Nova Scotia (N.S), persisting throughout much of their historical range, despite a number of significant

population declines over the last 300 years (Pulsifer and Nette, 1995). They have been an important resource for the native North Americans for centuries, long before Europeans arrived. Although it was the staple diet for many of the native communities, the hides and bones were just as important. The women worked the hides to provide valuable warmth during winters. The sinews yielded thread and glue, and the bones and antlers provided handles to knives and awls, as well as spoons and various other utensils (Franzmann and Schwartz, 1997).

When Europeans arrived during the early 17<sup>th</sup> century, moose were quickly recognized as being an easily obtainable source of food, tallow and hides, and subsequently they were substantially diminished by overharvesting and habitat loss (Pulsifer and Nette, 1995). Unrestricted moose harvests had a profound effect on moose populations within N.S. Two thirds of the mainland moose population were extirpated by the beginning of the 19<sup>th</sup> century, and the Cape Breton moose population was also in decline (Pulsifer and Nette, 1995).

During the mid 1800's, legislation was introduced that would manage and regulate the harvesting of the moose herds, which included the total ban on hide exports and protection of the cows, in hopes that the populations would recover to sustainable levels (Pulsifer and Nette, 1995). In 1874 the moose season was closed for the first time over concerns for the fluctuating population, and in 1877 it was reopened as mainland populations had increased. Irregular management practices followed that resulted in the moose populations to once again decline and resulted in another closure of the hunting season in 1937. Within 10 years of the closure the mainland population rapidly increased, peaked and crashed between 1949 and 1951 (Pulsifer and Nette, 1995). This resulted in local extirpation, scattered low density moose populations distributed across the mainland, and only two localized regions on the N.S mainland with relatively high density moose populations occurring within the Cobequid-Pictou-Antigonish Highland zones and the Tobeatic wilderness area (Fig 1.1a) (Pulsifer and Nette, 1995).

The moose population on Cape Breton Island did not experience the same success as the mainland populations from early management practices, never

having recovered from a decline at the turn of the 20<sup>th</sup> century (Pulsifer and Nette, 1995). The reason for decline is not clearly understood, and was attributed to unregulated over harvesting of the herd, although this theory has become less popular in recent years because of the inaccessibility of the C.B.H. to the early hunters, and modern day hunters/people for that matter, and it is thought that the current day problems that are occurring within the herd could have been a contributing factor in the past (Nette, personal communication, 2004).

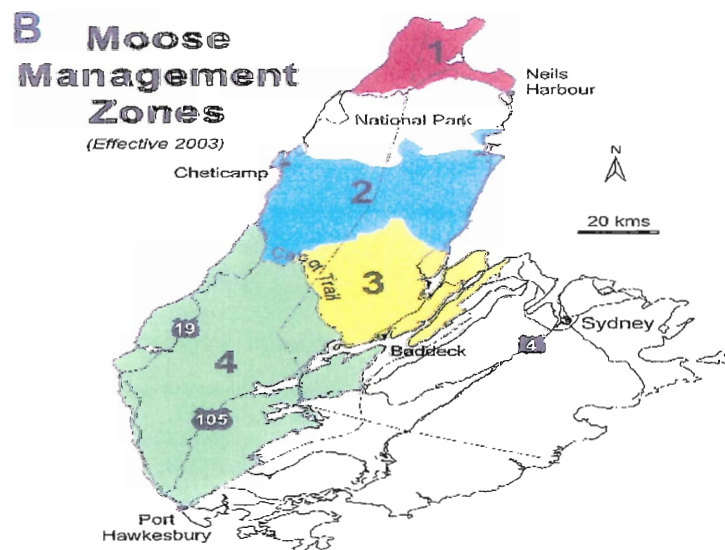
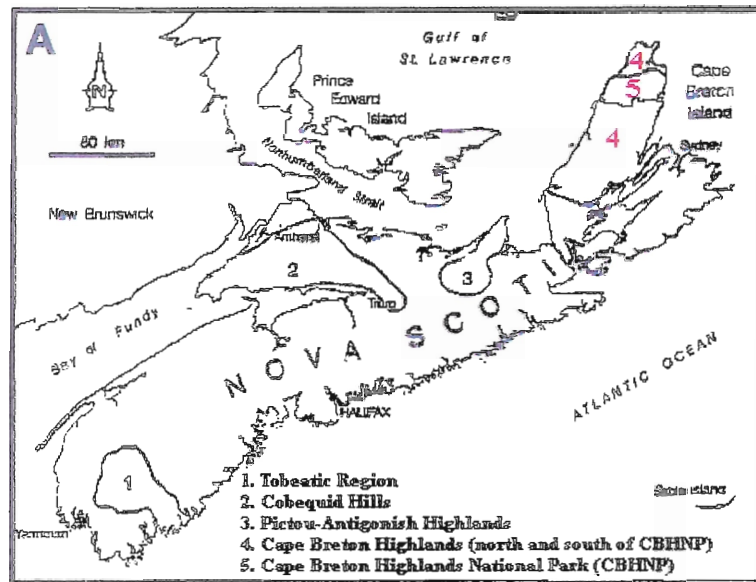


Figure 1.1 (A) Areas where moose populations have declined, are relatively abundant or high. Notice the control population (1) and the study population (4&5) (Pulsifer and Nette, 1995).

(B) Enlarged section of the study area, indicating new zones of management (Nette, personal communication, 2004).



During 1928 – 1929 an attempt at introducing 7 moose from the mainland into the western portion of the island to rebuild the herd proved unsuccessful. A second attempt was made in 1947 and 1948 with 18 moose from Alberta (*Alces alces andersoni*) being introduced within the Cape Breton Highland National Park (CBHNP) and this proved to be successful.

The introduced population, along with the possible remaining survivors of the eastern race, have formed the basis of the present Cape Breton Highland (C.B.H) population, which is now considered the largest and most stable population of moose in Nova Scotia (Figure 1.1a) (Pulsifer and Nette, 1995). The C.B.H herd is now known to be predominantly *Alces alces andersoni* (Broders et al, 1999).

### **1.3 Nature of the problem**

During the past 10 years, hunters and managers of the Nova Scotia Department of Natural Resources (NSDNR), Wildlife Division have observed an increase of incisiform breakage prevalence, that is particularly noticeable within the I1 and I2 incisors of the moose. Observations and documentation have been made on teeth that have been submitted by hunters. The gradual degradation of the teeth has implications for the overall health and vitality of individual moose within the herd, and appears to be playing a role in the poor representation of older aged moose within the population (Nette, personal communication, 2004).

#### **1.3.1 Study Area**

Cape Breton Island was recently (2003) divided into four regions for the purposes of moose management (Figure 1.1b), 1. North of the Park, 2. South of the Park, 3. Baddeck; Margaree, Hunters Mountain to Fraser Mountain Road and 4. Cabot Trail south to Port Hawkesbury. The focus of this study is based on the older management zones of Figure 1.1a, located in northern Cape Breton Island, northern N.S. The highland region is approximately 2400 km<sup>2</sup> (Pulsifer and Nette, 1995).

South of the Park is the largest of the three regions with a land area of 1220 km<sup>2</sup>, CBHNP has an area of 950 km<sup>2</sup>, and the area north of the park is the smallest with a land area of only 230 km<sup>2</sup> (Pulsifer and Nette, 1995).

The CBH's is a characteristic boreal region (Pulsifer and Nette, 1995), with a combination of dense spruce, blanket bogs and barrens that provides habitat for boreal species such as moose, snowshoe hare and lynx (Canadian Geographic, 2004). Black bear and cougar are the large carnivores that survive in generally undisturbed portions of habitat (Canadian Geographic, 2004). It is represented by the highest elevation within the province of Nova Scotia (370m), and also receives the highest annual precipitation for the province, over 1600mm annually (Environment Canada, 2004). During winter, more than 400cm of snow may fall in the region, with the snow pack usually lasting from early or mid November to April-May (Museum of Natural History, 2004). Because no weather stations exist in the regions of high elevation, reliable weather data are not available for the highlands. However, it is known that many days of the year the highlands are covered in a dense fog that results in relative high humidity. The high elevation contributes to a harsh climate for the CBH that results in a short growing season, but tree growth is rapid, except where strong winds stunt growth on exposed ridges (Museum of Natural History, 2004).

The coastal lowlands of Cape Breton are typical of cool, wet, acidic conditions dominated by spruce and fir trees, with the major influences on the regional vegetation resulting from marine climate and extensive disturbances by fire and cutting (Canadian Geographic, 2004).

#### **1.4 Purpose**

This project has been ongoing for five years, with little hard results or conclusions to date (Walsh, 2003). Through the combined efforts of Dalhousie University and the NSDNR, wildlife division, this thesis conducted a preliminary study of the tooth enamel from the C.B.H moose and tooth enamel from the moose of the Tobeatic herd. The Tobeatic herd is used as a control; where there is currently no evidence to suggest that these moose are experiencing the same

problems as the C.B.H herd in regard to incisiform breakage. To accomplish this we analysed for trace elements within the enamel to determine if the moose are deficient/excessive in trace elements, some of which have been previously implicated in both dental studies and physiological studies based on controlled experiments and observations conducted on human and animal populations. Trace elements can be naturally occurring within the environment or also from anthropogenic sources.

## **1.5 Importance**

### **1.5.1 Annual Harvest**

The moose herd in C.B.H is the largest and most stable population within the province of N.S., and is therefore the only area within the province that an annual moose hunt is conducted. Since 1982, other regions of N.S have been closed to hunting (Pulsifer and Nette, 1995) because current herd populations are not stable enough to support sustainable harvesting. It is important for the C.B.H herd to remain at sustainable levels for hunting to continue in the area. Native people of the area still rely on the herd for sustenance and ceremonial purposes (Nette, Personal communication 2004), while the annual hunt also supports numerous big game hunters. The native harvest is unregulated and quite high, with no hard numbers available. From 1986 to 2002 the regulated hunt offered 200 licences a year based on a lottery system with an average of 11,500 to 16,000 applications a year (Nette, personal communication, 2004).

### **1.5.2 Physiology**

Another aspect of importance is moose physiology and browsing habits. Moose, like other ungulates, lack upper incisors. Moose rely on their lower incisors (Figure 1.2a, Figure 4.2) when foraging for food, especially in winter months when vegetation is relatively poor quality woody vegetation (Franzmann and Schwartz, 1997). Woody twigs and branches are cropped and stripped off of their source by placing the vegetation between the incisors and upper maxillary bone (Figure 1.2b).

Increased bark stripping of hardwood species has also been observed within the C.B.H moose, which is strong evidence that suggests moose are close to or have exceeded their carrying capacity (Nette, Personal communication, 2004) as this type of behaviour is usually a response normally associated with food shortage in late winter when preferred browse is unavailable (Franzmann and Schwartz, 1997).

Fracturing is only evident within the incisors, and not the molars. Such fracturing that occurs severely inhibits a moose's foraging ability in winter months, and can lead to severe malnutrition, or in severe cases death of the animal. Moose need to maintain their large body size to meet the demands of changing thermal conditions of their environment (Franzmann and Schwartz, 1997). It is because of the large body size that moose are extremely tolerable of cold conditions, as the large body size acts as insulation (Franzmann and Schwartz, 1997).

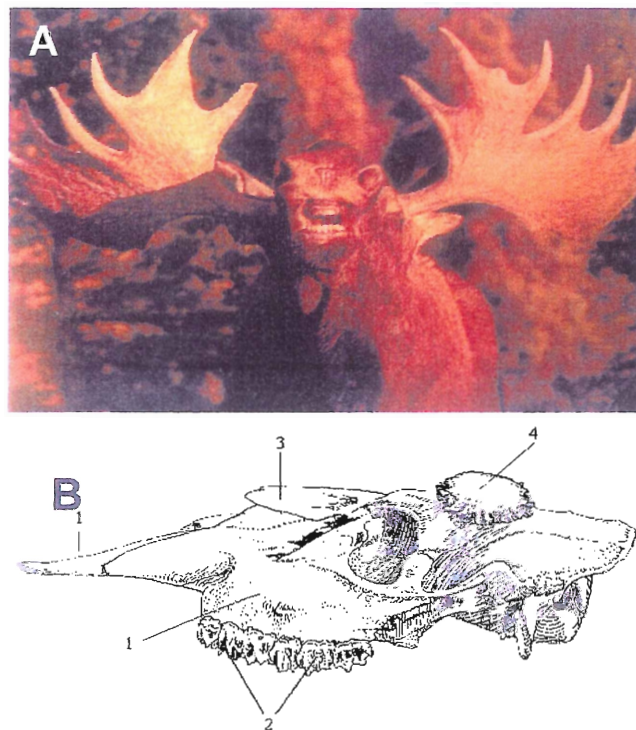


Figure 1.2: (A) Frontal view of Bull Moose displaying lower incisors and upper maxillary (Franzmann and Schwartz, 1997)

(B) Lateral view of a Bull Moose cranium. 1 = maxillary bone, 2 = upper molars, 3 = nasal bone, 4 = antler shaft. Notice that the upper jaw lacks incisors (Modified from Franzmann and Schwartz, 1997)

### **1.5.3 Osteophagia**

There is also evidence to suggest that moose are chewing on their fallen antlers, a phenomenon that is usually only seen to occur within other ungulates, and not moose (Nette, Personal communication, 2004). It is important to document this phenomenon, which is believed to be the first known evidence to suggest that moose eat their own fallen antlers (Nette, Personal communication, 2004).

### **1.5.4 Wildlife Forensics**

This project analysed within moose teeth enamel for trace elements. The project will investigate tooth breakage in the C.B.H moose, and explore the hypothesis that breakage may be a reflection of the trace element content of the enamel, a result of moose chewing their fallen antlers (Osteophagia), the stripping of hardwood bark, or even a combination of two or all three of these factors.

The results of geochemical analysis may be of significance in regard to the developing field of Wildlife Forensics. Analysis of the data may show distinguishable populations based on the trace element composition of the tooth enamel. The Control population on the mainland portion of N.S has been dwindling in numbers, and was recently listed as “endangered” under the Nova Scotia Endangered Species Act. Illegal hunting is an on going problem with the dwindling population (Nette, personal communication, 2004). Under the Nova Scotia Endangered Species Act, any persons that are found guilty of disturbing, killing, injuring, possessing, taking or interfering with or attempting to kill, injure, possess, disturb, take or interfere with an endangered or threatened species can now be charged upwards of \$500,000 and possible imprisonment in accordance with the Endangered Species Act (Endangered Species Act, Chapter 11, sects 13; 22-26).

If successful, this study will demonstrate how it is possible to determine from which area a moose originated (lived). At present, it is difficult if not

impossible to prove in a court of law, where a moose from within Nova Scotia originated. DNA can be used but only if DNA structure of the source population, as well as the alleged source population and other possible source populations are known (Nette, personal communication, 2004).

### **1.6 Previous Work**

Incisoform wear in moose populations of North America appears to be a rare condition, as it has only been formally documented twice before. Previous accounts of tooth wear in moose were illustrated in two separate studies, one conducted on a population of moose in Manitoba and the other in Alaska. Both studies documented different aspects of causation to tooth wear, and a brief discussion of each will follow.

The C.B.H moose were previously studied. A thesis, conducted by Walsh (2003), described the Micro and Macro fracturing of moose teeth, and also attempted to link the geochemistry of C.B.H to the increased incidence of incisoform breakage by utilising microprobe and Neutron Activation Analysis (NAA). Both microprobe and NAA results indicated a correlation between trace element content and fracturing, with fractured teeth indicating a relative depletion of Zn, Sr, Au and Ba. However, the results by Walsh (2003) were not proven to be statistically significant.

Phil Casey, funded through the Natural Sciences and Engineering Research Council of Canada (NSERC), completed a descriptive study on of microscopic breakage within the enamel framework of the C.B.H population. His worked has been continued in the present thesis.

In Alaska, Smith (1992) examined 270 mandibles from a population of moose (*A. a. gigas*) in the Seward Peninsula. Mandibles containing incisors were collected during the 1988-1990 hunting seasons. All moose were aged by cementum annulus analysis. Breakage score was determined by the amount of tooth material that was lost on a scale of 0-5. 0=unbroken, 1= 15% or less missing, 2 = 15-25% lost, 3 = 25-35% lost, 4 = 35-50% lost, and 5 = 50% or more lost.

Smith (1992) did show a positive correlation for moose age and incisoform breakage, although he did not go into great detail describing the fractures. Zn, Cu, K, Co, Fe, Pb, Ca, Mg, Na, Cd, Mn, Cr, Mo, Se, and Al were analysed using Microbeam analysis. Analysis of 40 teeth harvested from the Seward Peninsula, and 20 moose harvested near an area called Galena (areas where moose rarely have broken teeth), did not exhibit any significant differences in trace elements.

Smith (1992) realised that the significance of incisoform breakage to the welfare of the Seward Peninsula moose population is unknown. The concluding factor to the incidence of incisoform breakage within the moose population of the Seward Peninsula was determined by Smith (1992) to be possible early symptoms of density related (overpopulation) problems amongst the moose.

Young and Marty (1986) described 'excessive tooth wear' within a population of Manitoban moose (*A. a. andersoni*). Thirty-five moose from the 1983-1984 annual moose hunt were the subject of their study, 16 displaying tooth wear and 19 controls that did not exhibit tooth wear. Moose were aged by cementum annulus analysis. A detailed description of the incisors was documented.

Crown heights, percent tooth loss, interfacet distance, facet area and microwear were all analysed by Young and Marty (1986). Their methods of analysis for crown height, tooth loss and interfacet distance are very in depth, and the paper should be consulted as it is well beyond the scope of this project. The microwear was studied using a Phillips scanning electron microscope 505 at magnifications of X10 to X700. Teeth were examined for the extent of enamel and dentin loss, the orientation of the wearing forces from striations and also the direction of wear. It is important to mention that the type of 'tooth loss' observed by Young and Marty (1986) is analogous to the fracturing within the CBH moose.

Young and Marty (1986) showed that there were significant ( $P < 0.001$ ) differences between crown heights, percent tooth loss, and interfacet distance between the affected moose and the control. The microwear favoured abrasion by particulate matter of a greater hardness than dentine or enamel. Noting the moose's browsing habits of aquatic vegetation and woody stems and leaf litter,

Young and Marty (1986) noted the possibility of high silica content as being likely source for an abrasive agent, which were inferred to have a minimum size of 1.5 $\mu$ m

No chemical analysis was performed on the teeth. Young and Marty (1986) recommended a field study of the affected moose population by a ruminant biologist, also a direct study of moose feeding habits and local vegetation, and indirect studies of their diet by dung analysis to solve the problem.

### **1.8 Limitations**

The current project is intended as being a pilot study. Based on this approach, the project had limited funding and therefore limited samples were analysed, although those samples analysed are considered as being representative of the populations.

Unfortunately, no comparisons could be made with other moose studies in relation to trace elements present, only against the literature that contained studies on humans and animals. This limits our ability to draw solid conclusions, as biological trace element values from one species to another is always open to question.

### **1.9 Organization of Material**

This thesis is divided into seven chapters. The introductory chapter briefly outlines the focus of the study and why it is of importance. The second chapter provides an overview of moose, with particular attention paid to habitat, nutritional requirements, and affects of antler growth and reproduction. Chapter three describes teeth in detail; tooth mineralization, enamel structure, dental caries and the importance of trace elements in relation to dental caries. Chapter four and five focus on moose teeth, describing Macro and Microfracturing respectively. Chapter five also with the geochemistry of moose teeth, with attention paid to the results of ICP-MS analysis. From the results, comparisons are made in reference to the literature and possible conclusions are drawn.



Chapter five provides full descriptions of microfractures as described by Graves and Casey (2002, unpublished), and introduces new data followed by discussions and conclusions. Chapter six attempts to correlate the geochemistry of moose teeth with the geology/geochemistry of the Cape Breton Highlands, and southwest Nova Scotia. The final chapter draws together all the information and conclusions are drawn based on this study, and provides recommendations for future work.

## **Chapter 2: Nutrition and Antlers**

### **2.1 Introduction**

Ungulates comprise one of the most successful and diverse groups of large mammals alive today, inhabiting every continent except Antarctica and Australia (The Ultimate Ungulate Page, 2003). Ungulates belong to the **Kingdom** Animalia and The **Phylum** Chordata (Subphylum vertebrata). More Specifically, moose are classified as follows: **Order** Artiodactyla (Or 'even toed'), **Suborder** Ruminantia (Four (sometimes 3) stomach chambers), **Infraorder** Pecora, **Family** Cervidae (Deer, with shedding antlers), **Subfamily** Odocoileinae, **Genus** *Alces*, **Species** *Alces alces* (Franzmann and Schwartz, 1997; The Ultimate Ungulate page, 2003).

Ruminants, such as moose, are considered to be the most advanced of the Artiodactyla having four (sometimes three) stomach chambers that allow for the proliferation of micro organisms that allows for the digestion of tough vegetation, which would otherwise be unavailable to the animal (The Ultimate Ungulate Page, 2003).

Moose are found only in the Northern Hemisphere, occurring in Canada, U.S.A, Russia, Finland, Sweden, Denmark, Norway, Lithuania, Estonia, Poland, Czechoslovakia, Manchuria and China occupying the vast, global band of Northern boreal forest dominated by spruce, fir and pine trees (Odum, 1983; Telfer, 1984, in Franzmann and Schwartz, 1997). They belong to the suborder Ruminantia, and Genus *Alces* (*Alces* = Latin for elk) (Franzmann and Schwartz, 1997).

Modern North American moose appear to have originated from Siberia, yet the timing of their North American invasion across Beringia is still disputed. Many people have hypothesised to the exact timing of invasion, ranging from the end of the last glaciation (10,00-14000 years B.P), to the peak of the Wisconsinian glacial period (~18,000 years B.P) (Franzmann and Schwartz, 1997). Although moose have been present on the North American continent for thousands of years, they still continue to exploit new range and territory. It is only

recently that they have invaded eastern portions of North America such as Labrador, where the first sightings occurred in this region in the early 1950's (Chubbs and Schaeffer, 1997).

## **2.2 Habitat**

Moose can thrive in a variety of habitats, including montane forest and riparian communities characteristic of the Northern Rocky Mountains, mixed deciduous hardwood forests of north central and northeastern portions of the United States, and aspen-dominated boreal forests of Canada and boreal forests of Canada and Alaska (Franzmann and Schwartz, 1997). The limiting factors of a moose's geographic distribution are food and cover to the North (Kelsall and Telfer, 1974, in Franzmann and Schwartz, 1997), and climate to the south (Renecker and Hudson, 1986, in Franzmann and Schwartz, 1997).

Abundant woody vegetation is a common link between these characteristic ecoregions and characteristic of most, if not all, moose habitats and is especially an important feature for wintering moose who rely on the availability of these woody plants as a food source (Franzmann and Schwartz, 1997). C.B.H moose inhabit areas ranging from boreal to mixed forest, bogs, rivers lakes and streams.

## **2.3 Extent of Territory**

A home range of a moose represents an area that is familiar to the moose, where the moose can feed, rest, escape predators and meet its other life requirements, and moose tend to return to these areas (or remain in the same home range) for many years (Franzmann and Schwartz, 1997). The C.B.H moose will range approximately 40km<sup>2</sup>, and stay close to an abundant food source in the winter, restricted by snow cover, and during summer months they will range further in search for more succulent food (Nette, Personal Communication, 2004).

## **2.4 Forage / Browse**

The moose is a herbivore that is considered a “generalist” browser, who will ingest moderate amounts of a variety of vegetation such as leaves and bark, shrubs, woody twigs, and aquatic vegetation, as opposed to “specialist” browsers which consume high levels from only a few plant species (McArthur et al. 1991, in Franzmann and Schwartz, 1997). Moose, like all browsers, are equipped with narrow muzzles, prehensile lips and tongues that allow for high quality selection of small plant parts for ingestion. The type of vegetation a moose ingests is dependant on the seasons, and must also meet certain criteria. According to Oldemeyer (1974), forage must be highly palatable, have optimum levels of various nutrient components, have high apparent digestibility of nutrient components, have optimal proportions of volatile fatty acids, have adequate levels of minerals, vitamins and trace elements, and finally be efficiently converted into components necessary for the animals bodily functions.

Moose will consume plant parts from a variety of species, although the majority of moose foods are classified as browse (Woody plants). (Franzmann and Schwartz, 1997).

### **2.4.1 Spring-Summer**

Vegetation growth is at its peak during the warmer months of spring and summer. North American moose will usually rely on deciduous trees and forbs, which are quick growing annual species such as cloudberry (*Rubus chamaemorus*), sundew (*Drosera rotundifolia*), fireweeds (*epilobium angustifolium*) and lupine (*lupinus nootkatensis*) (Peek, 1974). Mushrooms and aquatic plants are considered succulent foods for moose, and moose will also feed on such species when they are present (Peek, 1974)

As seasons change, so does the quality of nutrients available within plant tissues. Actively growing plant tissues represent the highest quality foods available to moose, and they begin their growth phase in early spring, long before actual green-ups occur (Franzmann and Schwartz, 1997). This is due to

the translocation of nutrients from the roots to the twigs and swelling buds that will turn to leaves, and represents a time of rapid increase in the plants nutritive value (Franzmann and Schwartz, 1997). As the plant nears maturity, plant tissues mature and more structural carbohydrate is manufactured, thus decreasing forage quality for moose (Franzmann and Schwartz, 1997).

No detailed information was available on the spring-summer forage preference of Nova Scotia moose populations.

#### **2.4.2 Autumn-Winter**

Fallen leaves are an important food source in early autumn for moose. By consuming freshly fallen leaves in early autumn, a higher digestibility rate is realized than from woody twigs alone, and in addition this food resource can be exploited with minimal amount of travel in areas where high stands of forage biomass occur (Renecker and Hudson, 1985; Franzmann and Schwartz, 1997). However, feeding on this resource declines when snow cover restricts access, and also when more nutritious food becomes available in early spring (Franzmann and Schwartz, 1997).

Moose are inclined to sample new plants they locate (Franzmann and Schwartz, 1997), so therefore browsing habits for moose will vary geographically. A 1996 browsing survey for moose in C.B.H by Basquill and Thompson (1997) revealed moose preferred foraging woody plant species that occurred in the dense undergrowth during winter months. These included deciduous woody plants, such as mountain, red, and striped maple, red buried elder, mountain ash, white birch, and shadbush (Basquill and Thompson, 1997).

Bark stripping is a behaviour amongst ungulates that occurs when areas support poor winter browse, when food is in short supply in late winter when preferred foods are unavailable or when animals are restricted by deep snow cover. Moose perform this task by taking their lower incisor teeth and racking them perpendicular to the tree trunk (Figure 2.1), which is different from the technique used by elk, which normally bark trees by racking the lower incisors vertically along the trunk (Franzmann and Schwartz, 1997). This food source

does not normally comprise a large portion of the moose diet and usually is considered a “starvation” (last resort) food (Franzmann and Schwartz, 1997). Bark stripping is not a common behaviour for moose, and is very rarely observed within populations (Nette, personal communication, 2004).

However, Renecker and Hudson (1985, in Franzmann and Schwartz, 1997) observed moose in Alberta stripping bark mainly from aspen during early spring. This behaviour corresponded with warmer temperatures and the apparent movement of soluble sugars, which is highly palatable and easily absorbed by the moose’s digestive system, up the stem of the plant.

Bark Stripping by C.B.H moose is increasingly evident, especially in areas where moose densities are higher and very heavy browsing of preferred vegetation is evident (Nette, personal Communication, 2004). This could be a sign of moose populations in the area having reached, or even exceeded, their carrying capacity (Nette, personal Communication, 2004).



Figure 2.1: Bark stripping occurs in moose when there is a depletion of preferred browse, or when snow cover restricts the animal. Moose strip bark from trees by racking their teeth perpendicular to the trunk. (Photo by Charles C. Schwartz; Alaska Dept. of fish and game, *In* Franzmann and Schwartz, 1997).

## **2.5 Nutritional Requirements**

Due to their large body size, moose must consume large quantities of food, and because of these large food requirements they must find, consume, and digest food at a rapid rate (Franzmann and Schwartz, 1997). Many factors need to be considered when evaluating forage for moose, such as amounts, availability and nutritional value. For moose, the nutritive value of plants is related to the essential nutrients that they contain, including carbohydrates, fats, water, protein, vitamins and minerals (Franzmann and Schwartz, 1997).

For the purpose of this study, we will be focusing on nutritional requirements in relation to minerals. Minerals are a diverse group of nutrients that play an essential role for moose growth, health, and well being. Calcium, phosphorous, sodium, potassium, magnesium, chlorine and sulphur are minerals that are required in large amounts (mg / g), and are classified as macronutrients (Franzmann and Schwartz, 1997). Minerals required in small amounts are referred to as trace elements, and very little is known about the trace element requirements by moose (Franzmann and Schwartz, 1997).

Minerals will tend to vary in concentration between plant species, and also vary in concentration within a single plant organism. Many species tend to accumulate elements within their leaves or stems, and this differing elemental concentration between different sections of a plant species is important because elemental imbalances can have an impact on moose nutritive health than the absolute amounts present within the plant (Kubota, 1974).

A variety of high quality forage is an important factor in a moose' diet. A study conducted by Ohlson and Staaland (2001) considered the mineral diversity in wild plants, and the benefits to moose. Their study concluded that a diversity of plant species is required for a diversity of minerals for moose, and that feeding on few plant species may result in a deficient, or even toxic mineral nutrition. It is important for large herbivores, such as moose, to feed on a wide spectrum of plants to obtain essential minerals, not only in sufficient amounts, but also in physiologically balanced proportions (Ohlson and Staaland, 2001), because it is possible for plants to hyper accumulate certain minerals, or oppositely plants

may not absorb sufficient quantities of minerals. So therefore, when moose limit themselves to very few forage species, or they are forced to limit their foraging due to external factors, they are put at risk of element deficiency/toxicity.

## **2.6 Antlers**

Antlers are composed of hydroxyapatite, a crystalline calcium phosphate (Franzmann and Schwartz, 1997). Bull moose will begin antler growth in April – March and cease to grow in August (Nette, personal communication, 2004). This has a huge strain on the mineral resources of a moose's body. During antler growth, calcium and phosphorous are resorbed from bones because dietary intake and absorption of these minerals is insufficient to meet mineralization requirements (Banks et. 1968, Hillman et al. 1973, Muir et al. 1987; in Moen and Pastor 1998). This can severely impact the health and well being of the moose because shortly after mineralization of the antlers, moose will enter the rut, and during this period will not eat (Franzmann and Schwartz, 1997), therefore not replenishing any loss of minerals from bones used to support antler growth. This is significant because serious injuries, such as broken bones already weakened by mineral deficiencies, may result due to the characteristic sparring of Bull Moose that occurs during the rut.

Once antlers have been shed, which usually occurs by December for mature bulls (Nette, personal communication, 2004), the mineral constituents of the antlers that are stored are lost to the moose, however, rodents and other animals benefit from chewing antlers to obtain minerals for themselves (Franzmann and Schwartz, 1997).

Bulls do not grow prime antlers until they reach the ages of 5-12 (Franzmann and Schwartz, 1997). Antlers are first apparent in the bull when the calf is 6-7 months old, when tiny thumb-like antlers appear (Franzmann and Schwartz, 1997). When the moose is at one year of age, it will grow a second set of antlers that will be visible, and occasionally palmation of the antlers will occur, with three or more points (Franzmann and Schwartz, 1997). At 2-4 years of age,



sub mature bulls will grow their third to fifth antler sets, which will be characterized by small palms (Franzmann and Schwartz, 1997).

### **2.7 Nutritional Requirements of Antler Growth and Reproduction**

The following discussion has been taken from Moen and Pastor (1998), unless otherwise noted.

There is very little work documenting the nutritional and energetic requirements for moose during the mating season. Moen and Pastor (1998) concluded that antler growth in bulls required up to 25% more energy than the normal basal metabolic, and that a cows requirements for gestation and lactation increased their energy requirements by up to 35%. Protein requirements for the growth of antlers in bulls, and gestation and lactation in cows are most likely met by forage intake and, as mentioned, Ca and P are resorbed from the bones.

### **2.8 Osteophagia in C.B.H moose**

Osteoporosis in moose has been documented from skeletal remains in moose populating the Isle Royale National Park, Michigan (Franzmann and Schwartz, 1997). Observations of antler chewing (Osteophagia) by moose were made in the C.B.H during 2001 (Roger and Nette, 2002). This phenomena has been visually verified (Nette, personal communication, 2004) and also, by preliminary fieldwork by Roger and Nette (2002) inspecting 98 shed antlers, found that over 30% of shed antlers in the C.B.H had been chewed by moose within one month of antlers becoming available after snowmelt (Figure 2.3a). This appears to be the first documented case of osteophagia in moose (Nette, personal communication, 2004).

Evidence suggests that osteophagia is an innate, rather than learned behaviour, which is triggered by a deficiency of phosphate (Denton et al. 1986) that is widespread amongst artiodactyls (Sutcliffe, 1973). Osteophagia occurs in natural conditions and has a distinct geographical distribution that is dependant on the parent rock in which the plants are growing, but can also be influenced by

the occurrence of excessive amounts of calcium, aluminium or iron, which can reduce phosphorous availability to plants (Sutcliffe, 1973).

This behaviour probably presents some inconvenience to the cervidae that exhibit this behaviour, as it reduces time spent grazing, may cause injury to the mouth, and infection by bacteria in the case of bones still covered by rotten flesh, and certainly enhances teeth wear (Sutcliffe, 1977; Denton 1982; both in Barrette, 1985).

Characteristic patterns have been documented in antlers chewed by ruminants such as deer, that produce characteristic fork patterns at then end of the antler (Figure 2.2a), and zigzag margins at the proximal end have been documented (Figure 2.2b) (Sutcliffe, 1973).

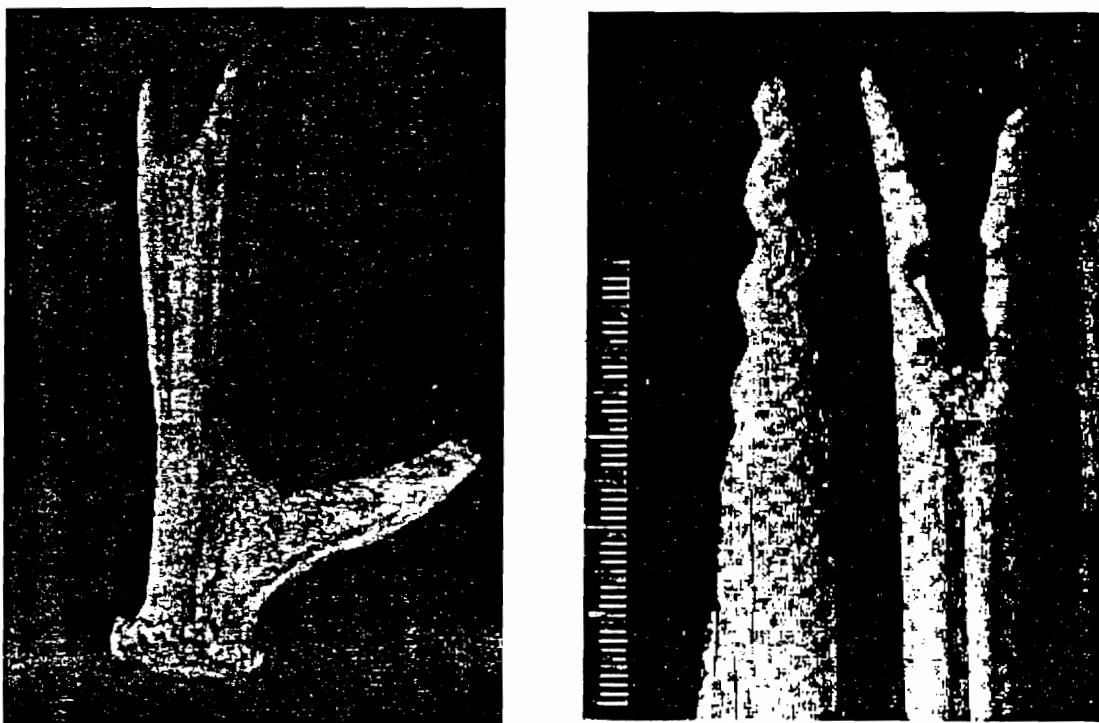


Figure 2.2 (A) Red Deer antler chewed by wild Scottish red deer to create a "fork" pattern.

(B) Norwegian Reindeer metatarsal chewed to create 'fork' with zigzag margins at its proximal end. (Pictures from British Museum (Natural History); In Sutcliffe, 1973)

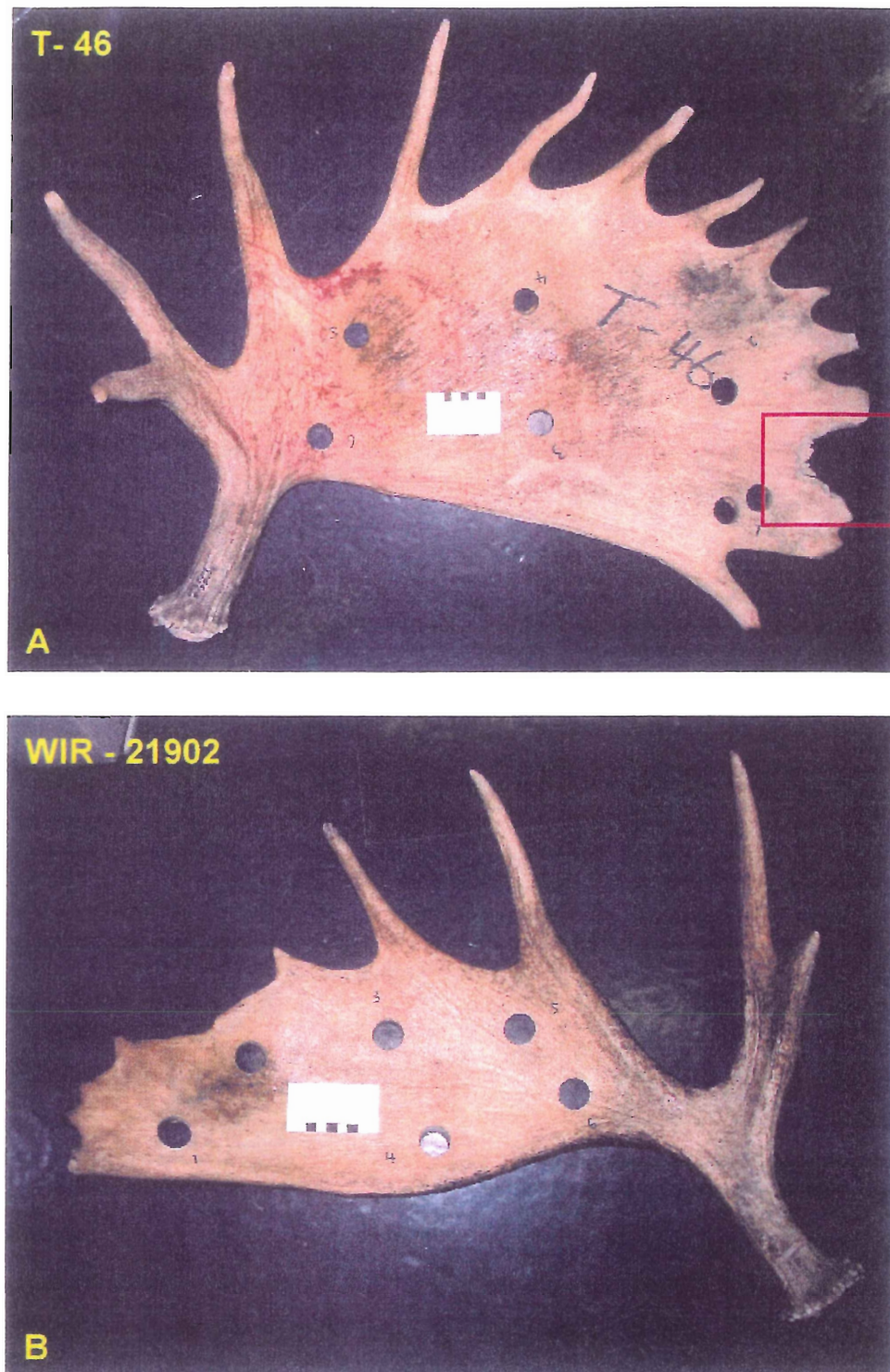


Figure 2.3 (A) Antler from C.B.H, notice the chewed section highlighted in the box. Holes represent core samples taken for geochemical analysis (B) Un-chewed antler from the control area. There is no evidence of chewing having occurred on this antler. Holes represent core samples taken for geochemical analysis.

During a limited (2 day) study within the C.B.H, Roger and Nette (2002) found several antlers that displayed characteristics of being chewed upon by moose within the area (Nette, personal communication, 2004). The nature of these chews and marks did not appear to be the result of rodents, carnivores or black bear as these animals will not leave the characteristic pattern due to different chewing techniques, that usually involve the very tips of the antlers (Nette, personal communication, 2004). White-tailed deer, the only other cervid in the province, seldom inhabit areas of higher elevation such as that of the C.B.H, as snow depth is a restrictive factor (Roger and Nette, 2002). According to Roger and Nette (2002), there was no evidence of deer having been in the area of investigation.

## **2.9 Methods**

Two antlers were received from the NSDNR, Wildlife Division, for geochemical analysis (Figure 2.3). T-46 has been chewed upon by a moose, and comes from the C.B.H (Figure 2.3a). WIR-21902 is an unchewed antler that comes from a herd in the Tobetic Wilderness Area of Shelburne County (Figure 2.3b).

Six drill core samples were taken from each antler, representing various sections of the antler, and were analysed using atomic absorption spectrometry (AAS), Minerals Engineering Center, Dalhousie University.

### **2.9.1 Atomic Absorption Spectrometry (AAS)**

The following has been taken from Hughes et al (1976) unless otherwise noted. Atomic Absorption Spectrometry is analytical technique that measures a wide range of elements within a given sample. The process requires very small sample sizes, typically ~10mg, that are dissolved in strong acids. Therefore, the technique is destructive of the sample, but little damage is caused to the source due to the very small samples needed. Spraying the resulting solution into the flame of the instrument, which is consequently atomised, results in

measurements of elements. Light of a suitable wavelength for a particular element is shone through the flame, and the atoms of the sample absorb some of this light. The amount of light absorbed is proportional to the concentration of the element in the solution, and hence in the original object. Figure 2.4 is a schematic diagram of a typical AAS machine.

To achieve complete analysis of a sample, each element is scanned for separately, thus the spraying technique is repeated several times according to the number of elements of interest. Therefore, the technique is time consuming but it is possible to detect trace elements to parts per million (ppm) level.

### 2.10 Results

AAS analysed for 26 trace elements within both sets of antlers (Table 2.1). The results show that there are generally higher representations of Ca, P, and Mn within the C.B.H antler that has been chewed compared to that of the unchewed antler from Shelburne County. The unchewed antler shows generally higher representation of C, Ba and K compared to the chewed antler of C.B.H. Figure 2.5 shows a plot of Ca versus P between the two antlers, and from this we can see that there are two distinct populations.

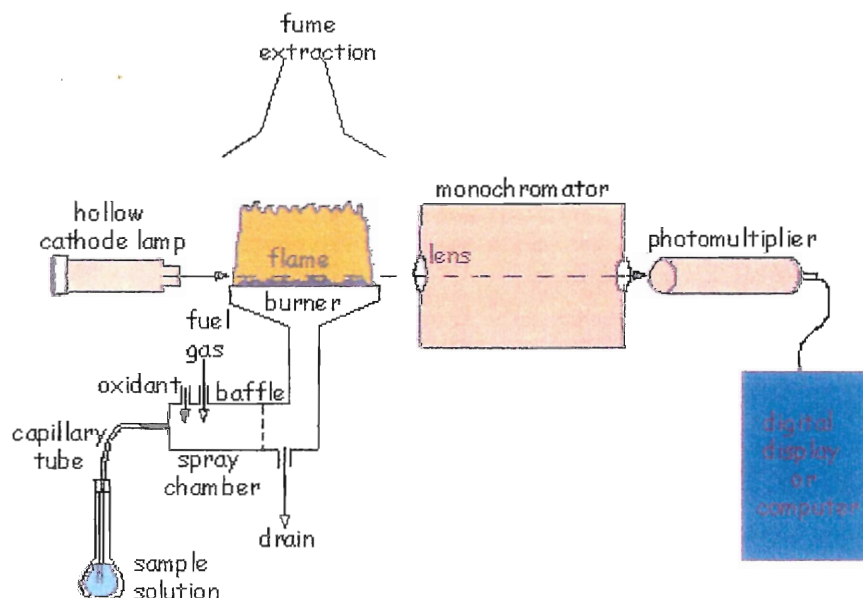


Figure 2.4: A typical Atomic Absorption Spectrophotometer (Hughes et al, 1976).

Table 2.1 : Geochemical Results for Antler Analysis

T-46							WIR 21902						
Si %	0.15	0.84	0.24	0.22	0.21	0.11	Si %	0.12	0.34	0.29	0.28	0.21	0.18
Ti %	<0.009	<0.009	<0.009	<0.009	<0.009	<0.009	Ti %	<0.009	<0.009	<0.009	<0.009	<0.009	<0.009
Al %	0.025	0.046	0.021	0.042	0.017	0.023	Al %	0.035	0.03	0.013	0.011	0.011	0.014
Mg %	0.34	0.27	0.34	0.44	0.46	0.53	Mg %	0.3	0.26	0.26	0.28	0.29	0.31
Ca %	26	24.94	26.95	27.11	27.46	29.1	Ca %	16.7	15.92	15.31	17.45	10.57	17.8
Na %	0.24	0.22	0.25	0.26	0.27	0.36	Na %	0.19	0.16	0.18	0.28	0.3	0.36
P %	9.82	9.43	9.78	9.73	9.95	9.99	P %	6.81	6.37	5.78	6.33	5.94	6.76
C %	18.37	18.56	17.93	17.88	17.61	16.74	C %	27.12	29.74	30.09	28.67	29.8	28.11
As ppm	4	4	2	5	4	4	As ppm	2	3	2	3	8	2
Ba ppm	260	220	230	240	220	240	Ba ppm	380	380	340	340	350	350
Be ppm	<1	<1	<1	<1	<1	<1	Be ppm	<1	<1	<1	<1	<1	<1
Cd ppm	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	Cd ppm	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2
Co ppm	<1	<1	<1	<1	<1	<1	Co ppm	<1	<1	<1	<1	<1	<1
Cr ppm	14	30	19	15	18	22	Cr ppm	12	24	24	29	19	19
Cu ppm	4	5	5	3	5	5	Cu ppm	4	5	6	5	4	3
Fe ppm	844	1950	1200	838	1060	1170	Fe ppm	568	1470	1610	1830	1180	1010
K ppm	95	87	84	67	65	134	K ppm	104	75	158	288	293	309
Li ppm	4	3	5	5	5	5	Li ppm	3	3	3	3	3	3
Mn ppm	37	62	59	88	89	42	Mn ppm	14	17	18	17	17	15
Mo ppm	<1	<1	<1	<1	<1	<1	Mo ppm	<1	<1	<1	<1	<1	<1
Ni ppm	2	4	3	2	1	1	Ni ppm	<1	<1	2	3	5	2
Pb ppm	6	5	<2	<2	6	<2	Pb ppm	4	4	2	3	2	3
Rb ppm	6	8	5	6	3	5	Rb ppm	2	2	2	4	4	4
Sr ppm	285	264	315	318	331	313	Sr ppm	366	358	361	403	381	408
V ppm	8	9	9	8	9	7	V ppm	8	5	6	9	8	7
Zn ppm	113	104	101	100	100	100	Zn ppm	113	109	96	99	100	103

## 2.11 Discussion

The data shows that there are interesting differences in chemical composition between antlers of different geographic regions. It would be premature to make any bold statements on the nature of the differences as we only have two samples and it is impossible to prove the difference has any statistical significance with only two samples. It is interesting to point out the higher representation of Ca and P corresponds with antler T-46, the chewed antler from the CBH (Figure 2.5).

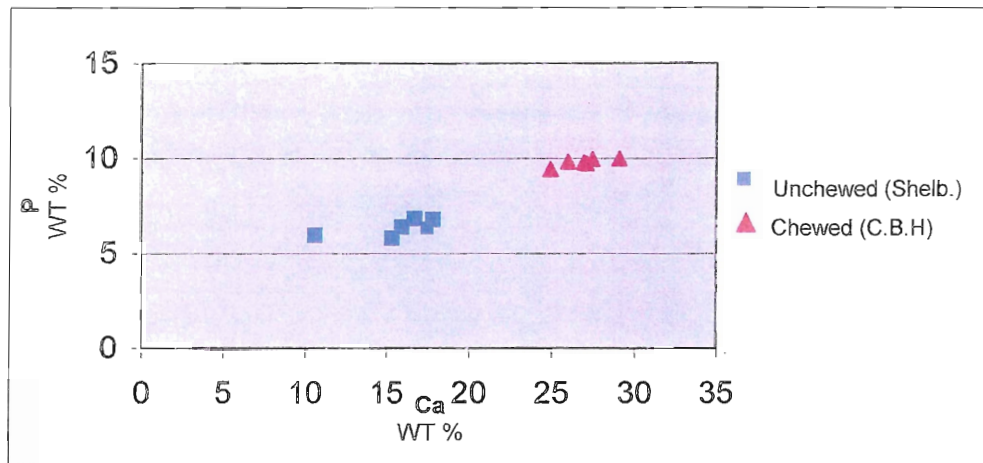


Figure 2.5: Ca/P plot illustrating the differing concentrations between the two antlers. Notice the chewed antler has a higher representation Ca/P.

## 2.12 Conclusion

The concentrations of Ca and P that correspond with chewed antler from CBH (T-46) are significant, and warrants further investigation because we know that osteophagia is triggered by phosphorous deficiency, and the antler T-46 shows high concentrations of this element.

## 2.13 Summary of Chapter

Little is known about trace element benefits/requirements in moose. These minerals are obtained from the forage that moose ingest, and the forage will vary geographically as moose are inclined to sample new plants that they encounter.

It is important for moose to have a variety of high quality forage in their diet. Feeding on few plant species will increase the risk of becoming deficient/toxic of trace elements because certain plant species may hyperaccumulate trace elements or may not readily absorb trace elements into the tissue.

Bark stripping occurs when areas support poor winter browse, when food is in short supply in late winter when preferred foods are unavailable or when deep snow cover restricts animals. Bark Stripping is increasingly evident in areas

of C.B.H that support high-density moose populations where heavy browsing of preferred vegetation is evident.

Osteophagia occurs in natural conditions and has a distinct geographical distribution that is dependant on the parent rock in which the plants are growing which may be deficient in phosphorous. It may also be influenced by excessive concentrations of Ca, Al or Fe, which can reduce phosphorous availability of to plants.

Thus conclusions, if only suggestive, show a higher representation of Ca and P within the antler of the CBH antler, and warrants further investigation.

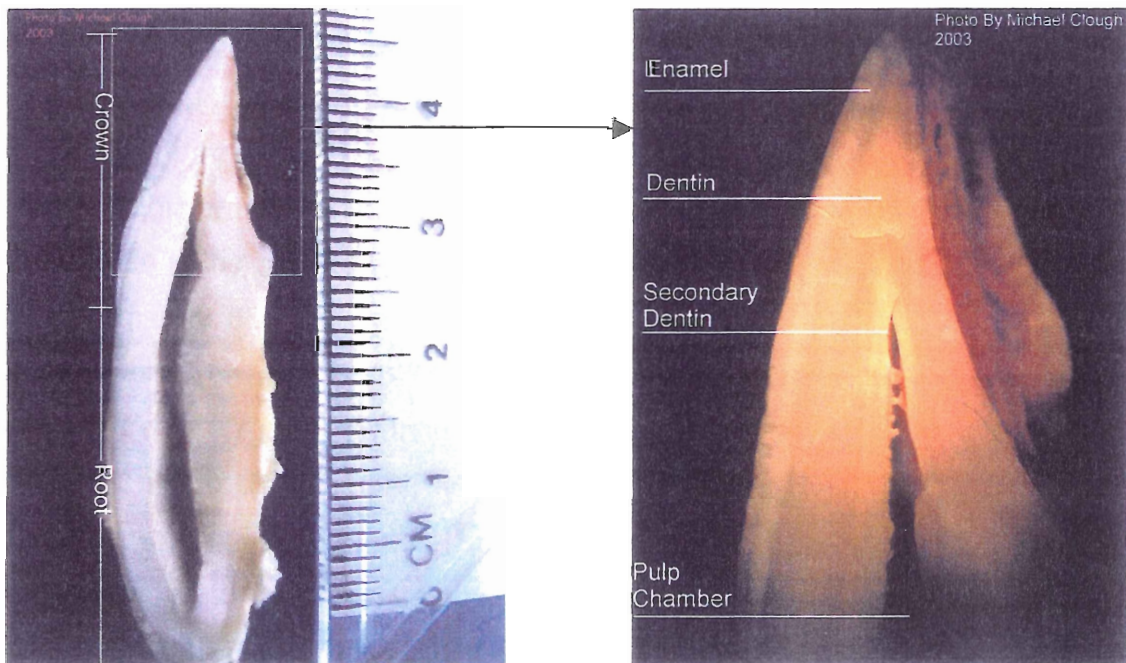


## Chapter 3: Teeth

### 3.1 Introduction: Tooth Physiology

Teeth are composed of both inorganic (mineralised) and organic (non-mineralised) substances. There are four main components of the tooth: Enamel, dentin, pulp and cementum. Enamel, dentin and cementum are all mineralised to varying degrees, whilst the pulp is the only non-mineralised component of the tooth.

Enamel is the outside layering of the tooth and encases the crown, and thins down at the base of the crown giving a knife-edge appearance. It varies in thickness, and is approximately 0.25mm-1.5mm thick in moose teeth (Figure 3.1). It is the hardest substance in the body, very brittle, and is 96% mineralised (Melfi, C. 1988), whilst the other 4% is made up of water and organic carbon. Enamel is similar to apatite (Hardness 4 in the Mohs Scale), and belongs to the apatite family. This has been confirmed by x-ray diffraction (Lazarri, E. 1976). More specifically, hydroxyapatite ( $\text{Ca}_5(\text{PO}_4)_3\text{OH}$ ) is the form present within tooth



**Figure 3.1:** (A) Longitudinal section of moose I2 incisor illustrating enamel, dentin and the pulp chamber. (B) Magnified longitudinal section of I2 incisor crown (6X magnification)

enamel. The function of enamel is to form a resistant covering of the tooth, rendering them suitable for mastication (Baskhar S.N., 1991), bearing the force of cropping food (Marieb and Mallatt, 2001). Although tooth enamel is the hardest calcified tissue in the body it is still subject to attrition: the wearing down of the tooth due to friction of use.

Biologic hydroxyapatite is markedly different to pure hydroxyapatite. Differences include inclusions of small amounts of proteinaceous material and trace elements (Curzon and Cutress, 1983). The trace elements present reflect the composition of the tissue environment during tooth formation and the oral environment after tooth eruption (Curzon and Cutress, 1983). Trace elements are incorporated into the apatite crystal lattice during the mineralization period, and also after mineralization preceding eruption elements can diffuse into the tissue (Curzon and Cutress, 1983).

According to Driessens (1982), tooth enamel, preceding eruption, will remain in physico-chemical equilibrium with its oral environment. This is possible due to enamel being semi-permeable, allowing for small ions and molecules to pass through the enamel of the tooth (Driessens, F.C.M. 1982). All present evidence indicates that once any chemical elements are incorporated within the tooth enamel framework, no mineral substances are withdrawn or decreased by any physiologic process within the tooth (Melfi. C. 1988).

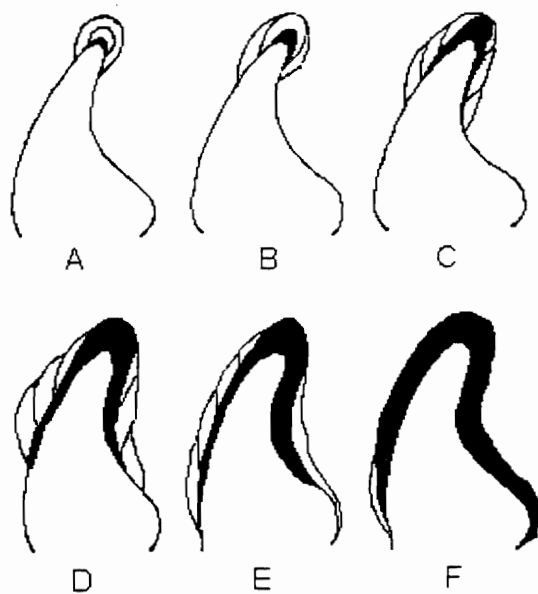
Dentin is the second most mineralised component of the tooth. It consists of approximately 68-79% inorganic material, and the remainder is water and organic carbon (Lazarri, E. 1976). It provides the bulk and general form of the tooth (Figure 3.1 & 3.4). Dentin is very similar to bone both physically and chemically, and unlike enamel, which is hard and brittle, dentin is viscoelastic and subject to slight deformation (Baskhar S.N., 1991). In an intact tooth, dentin is not visible due to the crown being covered with enamel.

Cementum is the mineralised dental tissue covering the anatomic root of the tooth (Figure 3.4). It is the least mineralised component of the tooth being only 45-50% mineralised and 50 –55% organic (Lazarri, E. 1976).

Pulp is the only non-mineralised component of the tooth. It is located within the interior of the tooth and occupies the pulp chamber in the crown and the root canal in the root of the tooth (Figure 3.4).

### 3.2 Mineralization of Enamel

Both deciduous and permanent tooth enamel is mineralised during critical development periods of an animal's lifetime, the second trimester to approximately ten years of age (Goodman and Rose, 1990). There are two stages of mineralization that occur, matrix formation and matrix maturation, although the time interval that takes place between the two appears to be very small (Bhaskar, 1991). Matrix formation involves partial mineralization of the matrix segments, which is in the form of crystalline apatite. While the second stage, maturation, is characterised by the gradual completion of mineralization, which begins at the height of the crown and progresses towards the base of the crown (Figure 3.2) (Bhaskar, 1991). A consequence of this complex calcification



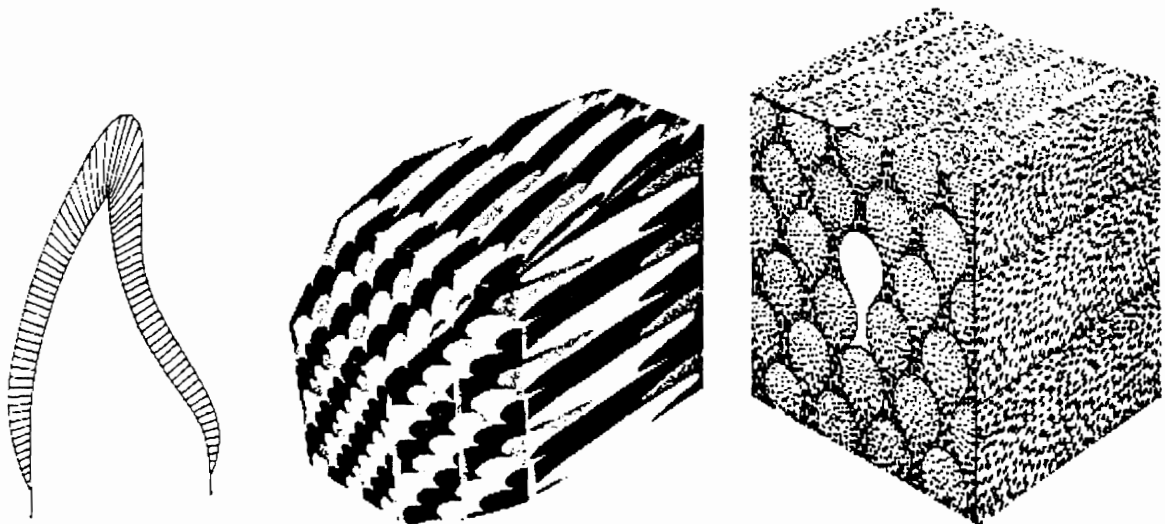
pattern is that it is difficult to ascertain the age of the individual at the time of any calcification defects that may occur (Goodman and Rose, 1990), although the cause of defective enamel formation can be generally classified as being a result of systemic, local or genetic in cause. The most common systemic influences are nutritional deficiencies, febrile disease, and certain chemical intoxications (Provenza, 1988).

**Figure 3.2:** Schematic showing mineralization of an incisor tooth. *Unshaded areas:* Consecutive layers of partially mineralised enamel matrix. *Shaded areas:* Advance of final mineralization during maturation (From Crabb HSM: Proc R Soc Med 52:118, 1959; and Crabb HSM and Darling AI: Arch Oral Biol 2:308, 1960; In Bhaskar, 1991)

### 3.3 Enamel Structure

Enamel is composed of rods or prisms, rod sheaths, and in some regions a cementing interprismatic substance (Bhaskar, 1991). The rods generally form an orientation that is perpendicular to the external surface of the enamel, except near the incisal edge where the rods are at an angle more directly opposed to masticatory surfaces (Figure 3.3a) (Provenza, 1988). Rods rarely, if ever, form straight lines from dentin to the enamel surface but generally follow a wavy course from the dentin to the enamel surface (Bhaskar, 1991).

Various patterns can also be produced when viewing the enamel prisms under a high-powered microscope by changing the plane of sectioning (Figure 3.3a&b) (Bhaskar, 1991).



**Figure 3.3:** (A) Diagrammatic illustration of rod orientation, which is approximately perpendicular to the external surface except near the incisal edge where the rods approach the surface at an angle more directly opposed to masticatory forces (Avery, 1994), 1986)  
(B) Various patterns can be produced by changing plane of sectioning (Meckel AH, Griebstein WJ, and Neal RJ: Arch Oral Biol 10:775, 1965; *In* Bhaskar, 1991)  
(C) (Griebstein WJ: *In* Stack MV and Fearhead RW, editors: Tooth enamel, Bristol, 1965, John Wright and Sons, Ltd, p 190; *In* Bhaskar, 1991)

### 3.4 Dental Caries

The following discussion of dental caries is a summary from Lazarri (1976), unless otherwise noted. Dental Caries, or tooth decay, is a disease of the hard tissue in the teeth whereby the mineral substances of the tooth are dissolved by acid (Melfi, C. 1988), resulting in lesions that penetrate the enamel surface. The process of dental caries progresses to destroy the organic components of the tooth, such as dentin and pulp, which are more susceptible to acid attacks.

Caries will begin as a subsurface demineralisation of the enamel. They will then progress to the dentino-enamel junction (DEJ) (Figure 3.4) and further progress more rapidly through the higher organic containing dentin. Eventually, when left unchecked, caries will involve the pulp and destroy the vitality of the tooth.

The process of dental caries involves salivary bacteria that convert all foods, especially sugars and carbohydrates, into lactic acids. Studies have been done that suggest direct pH measures indicate that carious dissolution takes

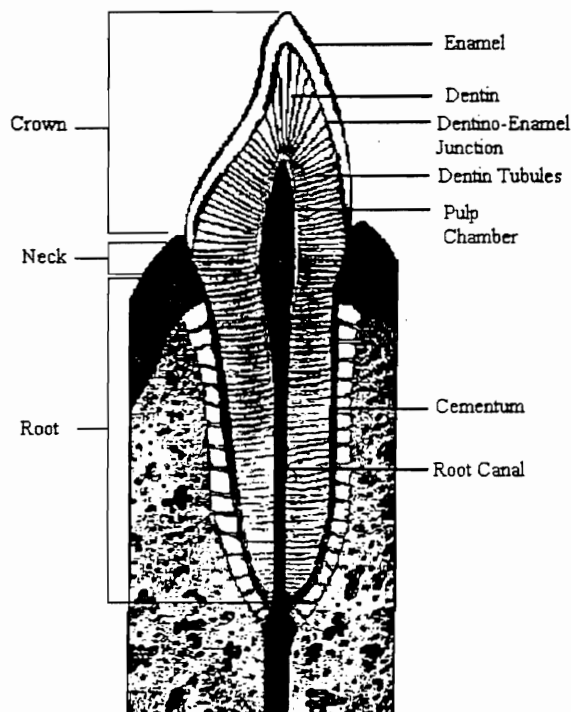


Figure 3.4: Longitudinal Section of canine tooth. (modified from Marieb and Mallat, 2001)

place in an acid environment. Histological studies demonstrate further that breakdown of the organic matrix occurs only after demineralisation of enamel is well established. Once past the protective enamel layer, lysis of the organic component begins, as acidic ions are free to react with and dissolve the more organic components of the tooth structure.

Enamel solubility also plays a major role in susceptibility to caries, and this is a determination of the concentrations of major and trace

elements present within the tooth. Newly erupted teeth are generally more susceptible than mature teeth to carious affects. Teeth undergo both physical and chemical alteration following eruption. Maturation and enhanced resistance is generally associated with exposure to the oral environment of the tooth, where mineral-containing saliva will deposit in areas of the tooth. Carbonates will tend to increase enamel solubility while fluorides tend to decrease enamel solubility. With increasing age, there is an increase in the fluoride and a decrease in the Carbonate concentrations of surface enamel.

Carious enamel and dentin contain more water, more organic matter and less mineral content, when measured on a weight basis, than corresponding sound tissue in the same tooth.

Dental caries is not to be confused with fracturing, that is evident within the moose populations, but is intended to illustrate the relationships that are associated with dental disease and trace elements and will be discussed in further detail in chapter 4.

### **3.5 Trace Elements and Enamel**

Trace elements are selectively taken up by enamel and incorporated within the surface enamel over the lifetime of an individual, which reflect the oral environment (Curzon and Cutress, 1983). Within humans, chemical composition of tooth enamel will change over time as individual's age. According to Curzon and Cutress (1983), various factors will influence the occurrence of changing chemistry within the enamel framework, such as normal wear within an individual's life history, food composition, smoking, geographic location, and cultural habits. Curzon and Cutress (1983) feel substantiating studies are still required before conclusions are drawn upon variation by geographic location. A study done by Cutress, (1972, in Curzon and Cutress, 1983) clearly demonstrated that there were higher concentrations of several elements in tooth enamel from one defined geographic source as opposed to another.

### **3.6 Summary of Chapter**

Enamel is the outside layering of a tooth that forms a protective layer, with the primary function of bearing the force of cropping food and mastication. It is markedly similar to apatite, and belongs to the apatite family. More specifically, enamel takes the form hydroxyapatite.

Trace elements present within the tooth enamel are a reflection of the environment during tooth mineralization, and oral environment after tooth eruption. Due to the semi-permeable nature of the enamel framework, trace elements are able to diffuse across the enamel surface, and will remain in physico-chemical equilibrium with the oral environment. Trace elements are selectively taken up by enamel and incorporated into the enamel framework over the lifetime of an individual. Trace element concentrations within enamel may vary according the geographic location of an individual. Trace element concentrations present within the tooth are a determining factor of enamel solubility.

Caries are lesions within teeth, and are not to be confused with fracturing, which is evident in the C.B.H population, and is intended to illustrate associations between dental disease and trace elements.

Defective enamel formation can be generally classified as being a result of systemic, local or genetic in cause. The most common systemic influences are nutritional deficiencies, febrile disease, and certain chemical intoxications.

## **Chapter 4: Macrofractures and Inductively Coupled Plasma-Mass Spectrometry (ICP-MS)**

### **4.1 Introduction**

Macrofractures are the type of fractures visible with the naked eye. Macrofractures result in a brown staining of the surface fracture, and indicate, within parts of the tooth, where breakage is imminent, and also where breakage has already occurred (Figure 4.3). Macrofractures are distinguishable from possible breakage due to tooth extraction because the surface where the fracturing has occurred is usually rounded out, indicating that fracturing occurred long before the death of the moose (Figure 4.3).

Inductively Coupled Mass Spectrometry (ICP –MS) was the analytical tool utilised for measuring the trace element concentration of the enamel. This technique was favoured due to it being a powerful tool for measuring a variety of trace elements at very low (ppm) detection limits.

### **4.2 Tooth Collection and Storage**

Teeth samples were obtained from the 2001-hunting season. Each year during the annual moose hunt the Nova Scotia Department of Natural Resources (NSDNR), Wildlife Division, require that hunters hand in the lower mandible with teeth intact for age determination by cementum age analysis. Prior to processing, the teeth are kept in a frozen storage locker. It is important to keep the teeth in a cool dry place, preferably in paper envelopes so that the teeth will dry without rotting. In sealed plastic envelopes, there will be decomposition of soft tissues associated with the teeth, creating an undesirable nuisance factor for technicians who handle the teeth later on (Matson's Laboratory, LLC, 2004). Jaws were also categorized for ease of study. Individual moose were assigned with the letter 'M', followed with three digits (eg; M001).

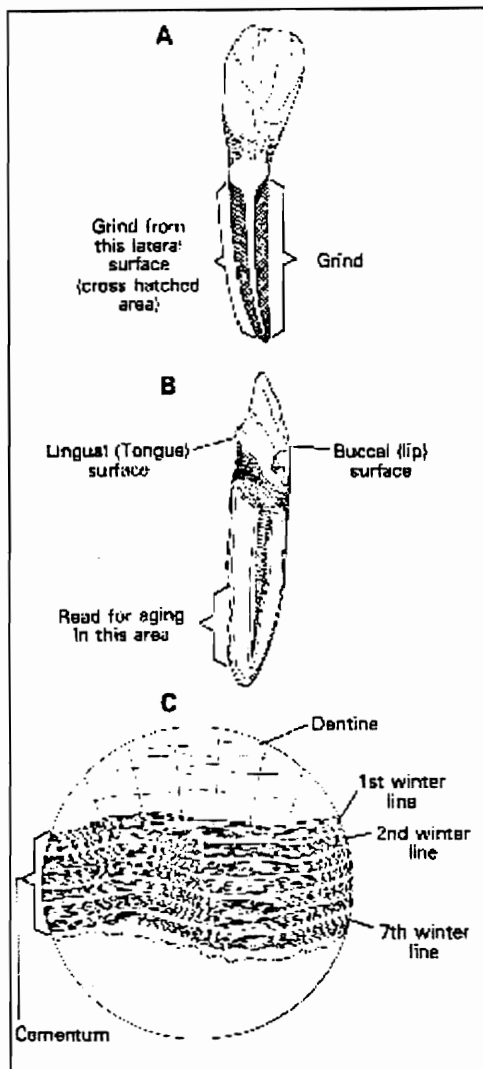


### 4.3 Tooth Extraction

The jaws were soaked in a hot distilled water bath at 70 –90 degrees centigrade for 4-6 hours as stipulated by Matson's Laboratory (2004). The attachment between incisor and mandible of the ungulate is largely soft tissue, so

this was severed by cutting it with a sharp knife on either side of the tooth. After the cut was made, the tooth was loosened by gently twisting and rocking it before pulling it out with an extractor (Matson's Laboratory, LLC, 2004). Care must be taken so as not to inflict damage to the tooth.

For this study, the I1 and I2 incisors were extracted. The I1 incisor was sent for cementum annuli dating to the Matson's Laboratory, LLC (2004). The I2 incisor was used for ICP-MS analysis. Overall, 56 individual I1 incisors were sent away for cementum annuli analysis, whilst the 56 I2 incisors were kept and documented.



**Figure 4.1:** Frontal (A) view of a moose tooth, indicating surface to be ground away for cementum analysis; lateral (B) view of a ground down tooth; and (C) 30-power magnification of a crossed section tooth of a 7-year moose killed in autumn. (Illustrations by Celia Carl Anderson; redrawn from original by Dave Harkness, Alaska Dept. of Fish and Game, in Franzmann and Schwartz, 1997).

### 4.4 Cementum Analysis

The basis for cementum aging is similar to the processes that occur within trees. Annular rings are formed when physical pressure is exerted on the teeth during foraging, which stimulates development of new protective layers of cementum around the roots (Franzmann and Schwartz, 1997). Layers formed in

spring and summer are lightly stained, and are thicker and denser than the darkly stained “annulus”, or ring formed during the winter months (Figure 4.1) (Franzmann and Schwartz, 1997; Matson's Lab, 2004).

#### **4.5 Tooth Categorization and Selection**

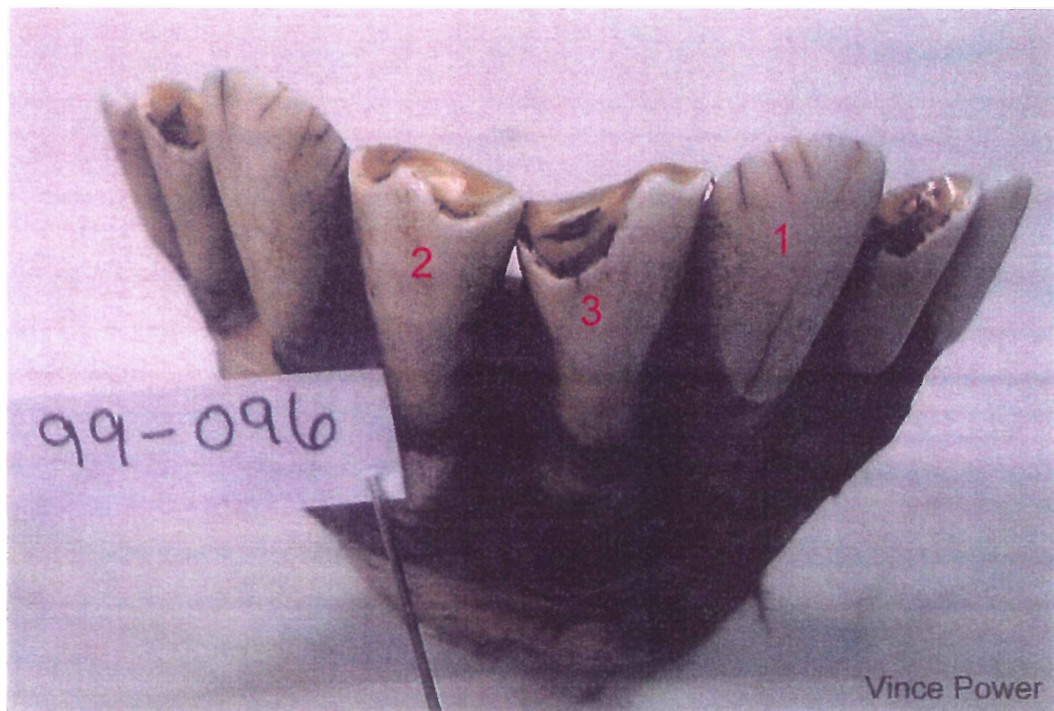
Nette and Power (unpublished, 1999) devised a simple classification system for determining the state of moose teeth in terms of macrofracture present. Based on previous work done by Smith (1992), they categorized the teeth based on three levels of damage, rather than the 5 levels used by Smith (1992). 1) Indicates none/slight damage (<30% of tooth material missing from the cutting surface), 2) Indicates moderate damage (30-60% of tooth material missing from the cutting surface). Finally 3) indicates severe damage (>60% of tooth material missing from the cutting surface).

Figure 4.2 illustrates a healthy set of incisor teeth, where there is no evidence of macrofractures present. Notice there are eight incisors in total, all given a prefix to identify where from the mandible they originate. Figure 4.3 shows a set of incisors with fracturing of various degrees present. Brown stains running perpendicular to the enamel surface are evidence of initial fracturing. The stained fractures appear to weaken the enamel structure, eventually breaking off. After breakage has occurred, the tooth surface will become polished and stained brown again (Figure 4.3). This is an important characteristic of the fracturing because it implies that the moose was living for a period of time after breakage occurred. When fresh breaks occur, the staining and rounded surface will not be present and can imply that breakage occurred just prior to the death of the animal, or even that it occurred after death.

Of the 56 teeth collected and documented, 46 were from the CBH (33 North of the National Park, 13 South of the National park), 5 from Shelburne County (including areas of the Tobeatic Wilderness area), 2 from Baddeck (Victoria County), 1 from coxheath (Cape Breton), 1 from Parrsboro (Cumberland County) and 1 from Sheet Harbour (Halifax County). Table 4.1 shows a list of samples and categorization. From the list, teeth 25 were selected for ICP-MS analysis.



**Figure 4.2:** example of a healthy set of moose teeth, with no evidence of fracturing, from the control area. Notice there are 8 teeth in total (6 incisors, 2 canines), with identifiers I1-I3 and C (Also given a Prefix Right or Left to determine which side of the jaw is being referred to. This is from the perspective of the moose) (Photo by Vince Power, 2002).



**Figure 4.3:** Lower mandible from a C.B.H. moose with varying degrees of fracturing present. Note the brown staining and polished surfaces of the fractured teeth, indicating fracturing occurred some time before the death of the animal (Photo by Vince Power, 2002).

Table 4.1: Categorization of moose teeth from 2001. C.B.H samples obtained from hunting season, others are either live extraction or from road kills

Sample Number	Other Id Number	Location	Age	SEX 1=MALE 2=FEMALE	TOOTH DAMAGE none/slight= 1 moderate= 2 severe= 3	NOTES
m001	7	Cape Breton north	6.5	2	3	
m002	10	Cape Breton north	3.5	1	3	
m003	13	Cape Breton north	3.5	1	1	lots of fine cracks
m004	17	Cape Breton north	3.5	1	1	split vertically-fine cracks
m005	22	Cape Breton South	3.5	1	1	plaque
m006	30	Cape Breton South	3.5	1	1	
m007	37	Cape Breton north	2.5	2	3	mostly missing(breach looks fresh)
m008	38	Cape Breton South	4.5	2	1	Split vertically- lots of cracks
m009	40	Cape Breton north	6.5	2	2-3	
m010	49	Cape Breton north	2.5	2	1	
m011	50	Cape Breton South	2.5	2	1	
m012	51	Cape Breton north	8.5	1	3	
m013	53	Cape Breton north	3.5	2	3	v.black (plaque)
m014	56	Cape Breton north	8.5	2	3	
m015	72	Cape Breton north	3.5	2	1	vertically split
m016	75	Cape Breton South	4.5	2	1	vertically split
m017	77	Cape Breton north	5.5	2	2	
m018	81	Cape Breton South	3.5	2	1	
m019	84	Cape Breton north	unk	1	1	nice-split vertically
m020	87	Cape Breton north	6.5	2	2	
m021	88	Cape Breton north	3.5	1	3	
m022	91	Cape Breton north	4.5	1	2-3	
m023	92	Cape Breton north	2.5	1	3	
m024	96	Cape Breton South	11.5	1	2-3	
m025	97	Cape Breton north	7.5	2	3	some damage(looks fresh)Vert.split
m026	100	Cape Breton South	3.5	1	1-2	old looking damage (Black)
m027	106	Cape Breton South	7.5	1	3	
m028	114	Cape Breton north	6.5	2	2-3	
m029	119	Cape Breton north	3.5	1	1	
m030	124	Cape Breton north	8.5	2	2	
m031	127	Cape Breton north	7.5	1	3	
m032	129	Cape Breton north	4.5	2	1	lots of cracks
m033	134	Cape Breton South	3.5	1	1	lots of cracks
m034	149	Cape Breton north	unk	1	1	
m035	154	Cape Breton north	2.5	1	1-2	
m036	155	Cape Breton north	3.5	1	3	
m037	157	Cape Breton South	7.5	1	3	
m038	164	Cape Breton north	1.5	1	1	
m039	166	Cape Breton north	2.5	1	3	Break looks fresh (vert. Split)
m040	179	Cape Breton north	3.5	1	1	
m041	182	Cape Breton South	3.5	2	2	vertically split
m042	184	Cape Breton South	4.5	2	3	
m043	185	Cape Breton north	3.5	2	1	cracks
m044	186	Cape Breton north	3.5	1	2-3	
m045	197	Cape Breton north	10.5	1	3	
m046	198	Cape Breton north	unk	2	3	
m047	211780	Coxheath Cape Breton	unk	1	3	tiny piece fell off
m048	205758	Baddeck Victoria	adult	2	1-2	brown
m049	205613	Baddeck Victoria	adult	1	1	
m050	201169	Sheet Harbour Halifax	unk	2	1	
m051	52124	Shelburne	2.5	2	1	nice
m052	52124	Parrsboro Cumberland	2.5	2	1	slight v.split
m053	moose8	Tobeatic	live (7)	2	1	nice
m054	moose10	Tobeatic	live (11)	1	1	nice
m055	moose14	Tobeatic	live	2	1	V.nice
m056	moose15	Tobeatic	live	2	1	V.nice

Shelburne County was selected as the control area based on the assumption it is a healthy population, having no documentation of the problems that are plaguing the C.B.H moose. Four teeth from Shelburne County were selected, and for statistical purposes, the single tooth from Parrisborro was included for analysis for reasons owing to the displacement of M054. Ten teeth were selected from North of the Park and 10 from South of the Park. Within both samples of 10, there were 4 teeth with breakage values of 1, and 3 each with breakage values of 2 and 3, giving a total of 10 teeth in each sample (Table 4.2)

#### **4.6 Inductively Coupled Plasma Mass-Spectrometry (ICP-MS)**

ICP-MS analysis is a powerful technique for multi-elemental material. It is particularly useful for analysing those elements that occur in low concentrations, such as the rare earths (La, Ce, Pr, Nd, Sm, Eu, Gd, Tb, Dy, Ho, Er, Tm, Yb, and LU) plus Y, the high field strength elements (Zr, Nb, Hf, Ta, Th, and U), and large ion-lithophile elements (Rb, Sr, Cs, and Ba). (GEOlabs, 2003)

In a typical application, the sample of interest is completely dissolved in acid to a solution suitable for spraying into flowing argon that is passed into a torch, which is inductively heated to approximately 10,000°C (West Coast Analytical Service, 2004). At this temperature, everything is atomized and ionized, forming plasma which provides a rich source of both excited and ionized atoms (West Coast Analytical Service, 2004). By acquiring the mass spectrum of the plasma, data can be obtained for almost the entire periodic table in just minutes (West Coast Analytical Service, 2004), giving detection limits from the low parts-per-trillion (ppt) level, up to hundreds of parts-per-million (ppm) (GEOlabs, 2003).

Table 4.2: Teeth chosen for geochemical analysis

Healthy	AGE	North of the Park	AGE	South of the Park	AGE	Breakage (CBH)
M051	2.5	M003	3.5	M005	3.5	1
M052	2.5	M010	2.5	M006	3.5	1
M053	?	M023	2.5	M016	4.5	1
M055	?	M038	1.5	M018	3.5	1
M056	?	M017	5.5	M008	4.5	2
		M022	4.5	M011	2.5	2
		M030	8.5	M041	3.5	2
		M001	6.5	M027	7.5	3
		M009	6.5	M037	7.5	3
		M012	8.5	M042	4.5	3

#### **4.7 Methods: Sample Preparation**

Great care has to be taken when preparing the samples. Because of its high degree of mineralization, enamel is the preferred substance for analysis as we are analysing for trace elements that potentially occur in extremely low concentrations within animal bones and tissue.

Moose teeth have a layering of enamel surrounding the dentin that is approximately 0.25-1.5mm (Figure 3.1b), so amounts were limiting and care had to be taken so that we could utilize as much of the enamel as possible to make up the required amounts for ICP-MS analysis, which requires a 2.0g sample.

The preparation of the enamel to a fine powder, of 200-mesh size as required by the laboratory for ICP-MS analysis, was preformed in 5 steps:

##### **Step 1: Separation of the root from the crown.**

The crown was separated from the root using a Buhler Isomet Low speed saw with a diamond tip blade in the Fission Tracks Lab at Dalhousie University.

##### **Step 2: Crushing of teeth with Percussion cutter**

Individual crowns were placed into the percussion cutter and carefully pulverized into fragments. At this point we did not want to pulverize to powder. If it was crushed to a complete powder, we could not distinguish the enamel from the other components of the tooth. It was found that by placing the enamel surface

parallel with the crusher and gently crushing the tooth, enamel fragments were more likely to break off as larger fragments, rather than tiny shards.

### **Step 3: Recovery of enamel under binocular microscope**

The fragments were removed from the percussion cutter and placed under the binocular microscope and the enamel pieces were carefully selected and extracted using a fine tipped paintbrush and then placed into an appropriately labelled vial. Enamel was distinguishable under the binocular microscope, owing to its bright white colour compared to dentin and pulp that appeared as a pale yellow colour under the microscope. Some fragments that contained enamel and small fractions of dentin could be separated using a sharp blade.

Step 2 and 3 was repeated several times until sufficient quantities of enamel had been extracted. With practice, it becomes evident which pieces are more likely to yield good enamel fragments. By selecting these fragments and placing them back into the percussion cutter one can increase the enamel concentration and reduce the dentine and pulp waste in the pile when observing under the binocular microscope improving the efficiency of enamel extraction.

### **Step 4: Reducing Enamel to Fine Grain Powder**

The enamel was then reduced to powder using an agate mortar and pestle. An ultrasound bath filled with distilled water was used to clean the mortar and pestle prior to every sample that was prepared to decrease the chances of cross contamination between the samples. Distilled water is important because we are analysing for trace elements, and it is possible for such elements to be found in the drinking/tap water. Ethanol was used to lubricate the enamel and aided in the reduction to a powder, and prevented fragments from scattering. Once there were no large fragments left, the pestle was rinsed with ethanol into the mortar, then the mortar was emptied and the remaining enamel rinsed with ethanol into a drying dish and left to dry at room temperature. Cleaning in warm soapy water, and then rinsing with distilled water prepared the drying dishes. The enamel was left to air dry.

### **Step 5: Recovery of enamel powder**

Recovering the powder from the drying dishes was done with a fine brittle paintbrush. The use of a fine bristle brush over a large bristle brush was preferred as it allowed for more control of recovering the sample as the large bristled brush causes the enamel powder to scatter. The powder was carefully brushed onto a sheet of weighing paper 8cm \* 8cm that was placed on top of a larger sheet of drying paper 15cm \* 15cm in case any material was scattered. New sheets of weighing paper were used for each sample to prevent cross contamination. The enamel powder was then placed into an appropriately labelled vial ready for analysis. The paintbrush was cleaned between each sample by blowing and wiping in paper towel, and more thoroughly cleaned between the sample groups by washing with warm soapy water and rinsing with distilled water and being allowed to air dry. It was discovered that recovery of the enamel powder was easiest immediately after drying, rather than leaving the samples overnight where the enamel was able to stick harder to the drying dish and recovery was much more labour intensive, thus creating more chance of the sample scattering.

For control of analysis, samples M017 and M032 are controls for the North and samples M008 and M020 are controls for the South.

## **4.8 Results**

Appendix A shows the raw data of results for ICP-MS analysis for all 25 samples, and the 52 elements that were analysed. From this data the results were divided into 3 individual populations; those North of the Park; those South of the Park; and the control population. The North and South populations were further divided into three populations within the overall populations in terms of tooth breakage based on the classification scheme devised by Nette and Power (unpublished, 1999) to determine if there were any significant differences in mean values between the various levels of fracturing.

Statistical analysis using one-way ANOVA of variance was used to compare variations between the populations, and also the variations within the population samples. The specified hypothesis for the problems were:



Null: *all means are equal within/between the populations.*

Alternative: *not all means are equal.*

All output was compared with Tables of  $F$  critical values.

Initially, the one sample that originated from the Parrsboro area (M052) was included within the analysis of data, with full statistical analysis using ANOVA (Appendix B). However, following advice by Zentilli and Nette (Personal Communication, 2004) ANOVA was conducted without M052 to see if there were any differences to the outcome of results (Appendix C). There was a clear shift with the mean and Confidence Bands between the two different statistical analyses for numerous elements. Therefore, from this point forward any reference to the control population will include only those from Shelburne County.

By comparing mean values of concentration based on the different breakage rates using ANOVA, it was possible to see varying degrees of a marked increase/decrease in the mean across the different levels of breakage. Appendix D (North of the Park) and Appendix E (South of the Park) illustrate these trends. After performing this analysis, it was evident that there may be correlations between elemental concentration and breakage score. Therefore, a complete analysis of correlating the breakage score against elemental concentration was performed for North of the Park (Appendix F) and South of the Park (Appendix G). Appendix F and Appendix G also show the 2 control samples used within the populations, with associated error (at 95% Confidence of the Standard Error). Error varied between elements, and was quite large for some, yet quite small for others. Refer to the Appendix G and Appendix F to see how the error varied.

Table 4.3: Minor constituents of enamel for North of the Park. Numbers indicate: mean  $\pm$  1 Standard Error with 95% Confidence.

<i>Elements</i>	<i>Symbol</i>	<i>Microgram/ Gram (PPM)</i>	<i>Elements</i>	<i>Symbol</i>	<i>Microgram/ Gram (PPM)</i>
Aluminium	Al	49.1 ~ 28.03	Molybdenum	Mo	0.0503 ~ 0.019
Antimony	Sb	0.0393 ~ 0.0255	Niobium	Nb	0.0226 ~ 0.0179
Arsenic	As	5.080 ~ 0.788	Rubidium	Rb	0.4634 ~ 0.1103
Barium	Ba	125.85 ~ 15.99	Selenium	Se	1.354 ~ 0.098
Beryllium	Be	0.01155 ~ 0.00559	Silicon	Si	185.1 ~ 84.87
Bismuth	Bi	0.01673 ~ 0.00398	Silver	Ag	0.0793 ~ 0.0515
Cadmium	Cd	0.1615 ~ 0.1115	Strontium	Sr	269.8 ~ 22.34
Calcium	Ca	34.81 ~ 1.52	Tantalum	Ta	0.0011 ~ 0.00059
Cesium	Cs	0.00327 ~ 0.00047	Thallium	Tl	0.00711 ~ 0.00167
Chromium	Cr	0.0951 ~ 0.0659	Tin	Sn	0.0633 ~ 0.0259
Cobalt	Co	0	Titanium	Ti	96.55 ~ 15.62
Copper	Cu	22.37 ~ 8.134	Thorium	Th	0.0018 ~ 0.00071
Gallium	Ga	0.2251 ~ 0.015	Tungsten	W	8.67 ~ 4.86
Hafnium	Hf	0.01535 ~ 0.01382	Uranium	U	0.00216 ~ 0.00037
Iron	Fe	107.9 ~ 30.38	Vanadium	V	0.3918 ~ 0.158
Lead	Pb	0.7231 ~ 0.0827	Yttrium	Y	0.0493 ~ 0.0057
Lithium	Li	0.980 ~ 0.423	Zinc	Zn	54.92 ~ 8.62
Magnesium	Mg	3215 ~ 392	Zirconium	Zr	0.674 ~ 0.694
Manganese	Mn	54 ~ 20.97	Rare Earths	REE	0.1663 ~ 0.039

Table 4.4: Minor constituents of enamel for South of the Park. Numbers indicate: mean  $\pm$  1 Standard Error with 95% Confidence.

<i>Elements</i>	<i>Symbol</i>	<i>Microgram/ Gram (PPM)</i>	<i>Elements</i>	<i>Symbol</i>	<i>Microgram/ Gram (PPM)</i>
Aluminium	Al	81.5 ~ 63.11	Molybdenum	Mo	0.06 ~ 0.043
Antimony	Sb	0.0147 ~ 0.0067	Niobium	Nb	0.0176 ~ 0.0042
Arsenic	As	4.603 ~ 0.292	Rubidium	Rb	0.7507 ~ 0.133
Barium	Ba	125.5 ~ 30.97	Selenium	Se	1.3435 ~ 0.0637
Beryllium	Be	0.0176 ~ 0.0052	Silicon	Si	161.4 ~ 59.98
Bismuth	Bi	0.01312 ~ 0.0099	Silver	Ag	0.0275 ~ 0.0112
Cadmium	Cd	0.0617 ~ 0.0198	Strontium	Sr	266.9 ~ 46.26
Calcium	Ca	34.725 ~ 0.76	Tantalum	Ta	0.0019 ~ 0.0012
Cesium	Cs	0.0067 ~ 0.0037	Thallium	Tl	0.0081 ~ 0.0022
Chromium	Cr	0.0687 ~ 0.072	Tin	Sn	0.105 ~ 0.063
Cobalt	Co	0.0348 ~ 0.058	Titanium	Ti	80.57 ~ 7.8
Copper	Cu	12.46 ~ 5.037	Thorium	Th	0.00585 ~ 0.0039
Gallium	Ga	0.1893 ~ 0.0196	Tungsten	W	6.56 ~ 4.61
Hafnium	Hf	0.00599 ~ 0.0048	Uranium	U	0.00464 ~ 0.0029
Iron	Fe	175.9 ~ 162.88	Vanadium	V	0.3818 ~ 0.0894
Lead	Pb	0.888 ~ 0.288	Yttrium	Y	0.0746 ~ 0.033
Lithium	Li	1.076 ~ 0.535	Zinc	Zn	54.51 ~ 9.98
Magnesium	Mg	3644 ~ 442.96	Zirconium	Zr	0.239 ~ 0.25
Manganese	Mn	59.6 ~ 28.8	Rare Earths	REE	0.435 ~ 0.396

Table 4.5 Minor constituents of enamel for the Control Population. Numbers indicate: mean  $\pm$  1 Standard Error with 95% Confidence.

<i>Elements</i>	<i>Symbol</i>	<i>Microgram/ Gram (PPM)</i>	<i>Elements</i>	<i>Symbol</i>	<i>Microgram/ Gram (PPM)</i>
Aluminium	Al	73.88 ~ 13.07	Molybdenum	Mo	0.05318 ~ 0.01824
Antimony	Sb	0.2455 ~ 0.0133	Niobium	Nb	0.02042 ~ 0.00274
Arsenic	As	5.41 ~ 1.033	Rubidium	Rb	0.7816 ~ 0.1513
Barium	Ba	201.4 ~ 42.14	Selenium	Se	1.406 ~ 0.055
Beryllium	Be	0.02726 ~ 0.01589	Silicon	Si	149.4 ~ 14.99
Bismuth	Bi	0.02406 ~ 0.0851	Silver	Ag	0.05818 ~ 0.0195
Cadmium	Cd	0.3762 ~ 0.1834	Strontium	Sr	468.2 ~ 103.88
Calcium	Ca	34.66 ~ 1.55 (wt %)	Tantalum	Ta	0.00069 ~ 0.00041
Cesium	Cs	0.00753 ~ 0.00167	Thallium	Tl	0.00799 ~ 0.00166
Chromium	Cr	0.1651 ~ 0.102	Tin	Sn	0.42 ~ 0.24
Cobalt	Co	0.869 ~ 0.480	Titanium	Ti	87.12 ~ 10.82
Copper	Cu	22.66 ~ 4.66	Thorium	Th	0.00948 ~ 0.00484
Gallium	Ga	0.1676 ~ 0.0133	Tungsten	W	3.69 ~ 2.37
Hafnium	Hf	0.0146 ~ 0.0225	Uranium	U	0.00531 ~ 0.00145
Iron	Fe	318.4 ~ 169.34	Vanadium	V	0.5734 ~ 0.156
Lead	Pb	2.15 ~ 0.856	Yttrium	Y	0.1307 ~ 0.03
Lithium	Li	1.066 ~ 0.429	Zinc	Zn	63.76 ~ 8.68
Magnesium	Mg	2912 ~ 311.64	Zirconium	Zr	0.0567 ~ 0.028
Manganese	Mn	38.4 ~ 18.93	Rare Earths	REE	0.909 ~ 0.629

Two outliers from North of the Park were eliminated from the analysis. M009 was discarded from the Fe analysis, having a value of 1600ppm (averaging 15 times higher in concentration than all other samples for Fe), and M022 was eliminated from the Li analysis, having a value of 17.3ppm (averaging 17 times higher in concentration than all other samples analysis for Li). Three outliers were eliminated from the analysis for South of the Park. M011 was discarded from both Cd and Fe (averaging 110 and 10 times higher in concentration than all other samples within the analysis respectively), and M018 was discarded from Ag (averaging 25 times higher in concentration than all other samples in the analysis).

Basic statistics were calculated for all the sample populations and tabulated for ease of reference. For all 3 populations, the Rare Earth Elements

(REE) was summed, and the mean and Standard Error with 95% confidence was calculated. Table 4.3 shows the mean concentration of elements for the entire population North of the Park with associated Standard Error at the 95% confidence level. Table 4.4 shows the mean concentration of elements for the entire population South of the Park, with associated Standard Error at the 95% Confidence level. And Finally, Table 4.5 shows the mean concentration of elements for the entire population of the control area, with associated Standard Error at the 95% Confidence level.

For ease of discussion, discussion of results will be set out in three sections. The first section will discuss significant differences between elemental concentration for both the population North of the Park and population South of the Park against the control population. The second section will discuss significant differences between the population North of the Park against the control population and significant differences between the population South of the Park against the control population. And finally, the third section will discuss any significant correlations that may be apparent between elemental concentration and breakage score for the population in the C.B.H.

#### **4.9 Healthy Vs C.B.H (North and South of the Park)**

Within the CBH, results show that 5 elements, Barium, Lead, Strontium, Cobalt and Tin differ significantly in the C.B.H moose compared to that of the control population, all showing depleted levels. The following discussions will give individual attention to each of the above-mentioned elements, followed by bringing all the information together and drawing conclusions upon the discussions collectively.

##### **4.9.1 Discussion of Barium (Ba)**

Barium shows significant depletion in both C.B.H populations (Figure 4.4). Barium does occur widely in soils, plants and animal tissues in highly variable concentrations, although little is retained within the tissues of animals because it is poorly absorbed from most diets (Underwood, 1971). According to Underwood (1971) and Frieden (1984), evidence to suggest that Ba performs any essential

biological functions in plants or animals has not been confirmed (or denied), although it has been shown to act as a growth stimulant in plants. Rygh (1949; in Frieden 1984) claimed that the omission of Ba from the diet resulted in depressed growth and reduced calcification of bones and teeth in rats and Guinea Pigs.

The role of Barium in tooth disease is unclear at best. According to Curzon and Cutress (1983), research on Ba and dental disease has been very limited, and is insufficient for any conclusions of any possible role for the element in dental disease. Curzon and Cutress (1983) refer to two studies undertaken to determine the role of Ba in dental disease in humans. One study carried out by Schamschula et al. (1978, in Curzon and Cutress, 1983) identified a positive correlation of Ba with dental caries in surface enamel of human samples taken from Papua-New Guinea.

A similar study carried out by Curzon and Losee (1977, in Curzon and Cutress, 1983) indicated significantly higher Ba concentrations in the enamel of humans from a population sample taken from New England who displayed a high incidence of Caries, compared to a population sample taken from South Carolina who displayed a low incidence of caries.

Shaw and Griffiths (1961, in Curzon and Cutress, 1983) demonstrated Ba as having minor influences on caries prevalence.

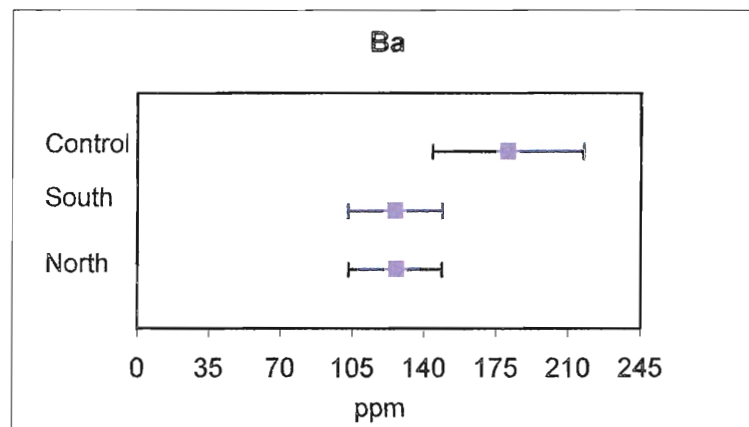


Figure 4.4: ANOVA results for Ba. Individual 95% Confidence Intervals based on pooled standard deviation.

#### 4.9.2 Discussion of Lead (Pb)

Lead shows significant depletion within the C.B.H populations, compared to the control (Figure 4.5). Lead occurs naturally and widely in soils and plants, plants that grow in areas of low lead concentrations rarely contain more than a few ppm (Underwood, 1971). Lead levels within animals, plants and the environment are not equivalent to those that prevailed during the creation and evolution of physiological responses, they represent levels that have been elevated due to anthropogenic activities involving the injection of millions of tons of lead into the environment (Underwood, 1971).

Excessive lead toxicity has been well documented over the years, with symptoms that include anaemia, and neuropathy or encephalopathy (Underwood, 1971). Yet, the possibility that lead in low concentrations performing some essential biochemical function cannot be excluded. Experiments have been conducted that show Pb depletion may result in reduced retention of Fe in animal tissue, and Frieden (1984) gives a full account of the experiments, which provides an in depth discussion. Fe deficiency results in anemia, or reduced haemoglobin.

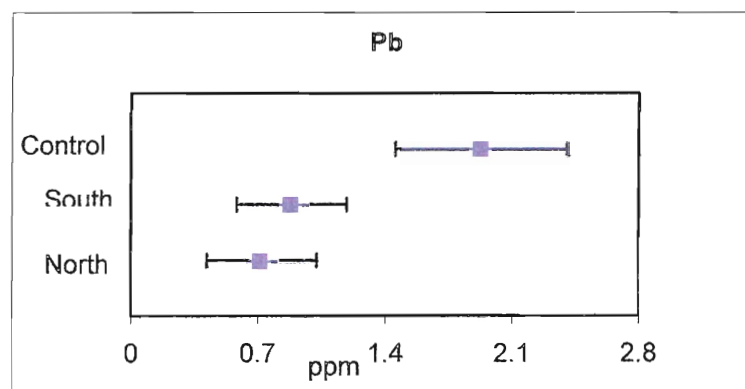


Figure 4.5: ANOVA results for Pb. Individual 95% Confidence Intervals based on pooled standard deviation.

In dental disease, there have been no clear conclusions drawn upon the influence lead may have. An extensive study conducted by Curzon (1977, in Curzon and Cutress, 1983) illustrated that there was a trend toward higher enamel Pb with lower prevalence of caries in human teeth.

Anda (1976, in Curzon and Cutress, 1983) on the other hand found that increased Pb level in teeth was found to be associated with increased industrial exposure and higher caries incidence. Proud (1976, in Curzon and Cutress, 1983) and Moses et al (1976, in Curzon and Cutress, 1983) both illustrated a negative trend.

Studies conducted on animals have also produced mixed results. Wisotzky and Hein (1958, in Curzon and Cutress, 1983) observed Pb to be a caries-promoting agent in male, but not female hamsters. Lazansky (1947, in Curzon and Cutress, 1983) demonstrated, according to a preliminary report, that by brushing the teeth of hamsters with a mascara brush charged with Pb-fluoride solution three times a week that there was a three quarter reduction of caries affected molars.

#### **4.9.3 Discussion of Strontium (Sr)**

Strontium is significantly depleted in the C.B.H moose populations compared to the control area (Figure 4.6). Strontium in animal tissue is a reflection of the concentrations found within drinking water supplies, and there is evidence that suggests it will increase with age, and that it varies with geographic regions (Curzon and Cutress, 1983).

Plants will readily absorb Sr into their tissues and it has been claimed to be a plant growth stimulant (Underwood, 1971), but has not been shown to be essential for plant or animal functions.

Calcium and strontium have a strong interaction, as the distribution of Sr within minerals, rocks, sediments and water is affected by the presence of calcium (Fairbridge, 1972). It is generally found in greatest amounts in calcium rich minerals, and to a lesser extent in potassium rich minerals (Fairbridge, 1972).

Dental studies within humans have been conducted that suggest a negative association between Sr and caries - the prevalence of caries being decreased as environmental Sr increases (Curzon and Cutress, 1983). Studies relating trace elements to dental disease carried out by Curzon and Losee, (1977, 1978, from Curzon and Cutress, 1983) selected geographic areas with high and low caries. Their results showed higher Sr concentrations in enamel from the area of low caries than the high caries area, and also higher enamel Sr in low caries individuals within each area.

Rygh (1949, from Curzon and Cutress, 1983) studied the effects of a number of trace elements on growth and health of rats. He found that a reduction of Sr levels produced poorer calcification and dental decay in the teeth of the rats. Studies done since Rygh's (1949, in Curzon and Cutress, 1983) have generally indicated that increased Sr may lead to a decrease in caries in rodents (Curzon and Cutress, 1983).

Because of its close similarity to Ca with regard to physical and chemical properties, it is theoretically possible that Sr can replace Ca in hydroxyapatite (Curzon and Cutress, 1983). Dedhiya (1974, from Curzon and Cutress, 1983)

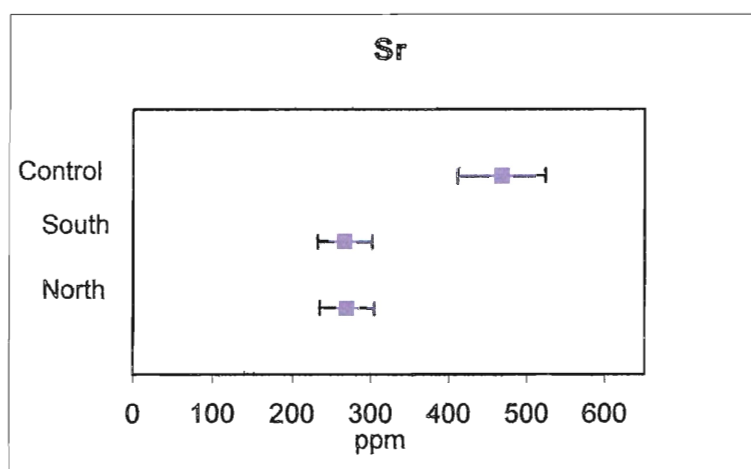


Figure 4.6: ANOVA results for Sr. Individual 95% Confidence Intervals based on pooled standard deviation.



illustrated how the addition of Sr in association with Fluoride affected the dissolution of synthetic apatite. Sr and F both retarded apatite dissolution, although it was much greater for F, and when used in partnership the affect was even greater than for Fluoride alone, and greater than would have been expected if they were added separately, suggesting a synergistic relationship for the two trace elements (Curzon and Cutress, 1983). Herbeson and Handelman (1975, in Curzon and Cutress, 1983), looked at the affects of several trace elements in vitro with effects on acid production and hydroxyapatite dissolution. Their work was of great interest because the study reflected trace element concentrations based on communal water supplies of a previous study conducted by Curzon et al (1970, In Curzon And Cutress, 1983). The study showed that there was a significant effect with Sr and F in combination that reduced the hydroxyapatite dissolution, which was not due to any other factors influencing the results (Curzon and Cutress, 1983).

#### **4.9.4 Discussion of Cobalt (Co)**

Cobalt is significantly depleted within moose populations of CBH (Figure 4.7), not being detected in the population North of the Park. The detection limit for cobalt was 0.1ppm for the lower limit and 120ppm for the upper limit.

Cobalt concentrations generally occur in highest concentrations in the liver, kidneys and bones, although it is widely distributed throughout the bodies of most animals with no excessive accumulation within any particular organ or tissue (Underwood, 1971).

The dietary benefits of cobalt have been known for decades. In 1935 Australian researchers described the effects of "coast disease" and "wasting disease" among sheep and cattle to be the result of insufficient cobalt in dietary intake (Underwood, 1971). Co has a relationship with vitamin B<sub>12</sub> synthesis, and Co deficient animals lack the ability to synthesize vitamin B<sub>12</sub> properly.

Victims of this debilitating disease initially thrive and grow as normal drawing upon B<sub>12</sub> reserves within the liver and other tissues for a period of weeks to several months. Following this is usually a gradual loss of appetite and failure of growth or loss of body weight, and in some animals there is evidence of

hypoplasia within erythrocytic tissues and in bone marrow (Underwood, 1971) due to the restriction of vitamin B<sub>12</sub> synthesis, which consequently increases other metabolic disturbances (Frank et al, in press). Hypoplasia in enamel is well documented, and is a defect that results in a lesser quantity of enamel covering the dentin that would normally be present that occurs during matrix formation, and cannot be predicted with any reliability even in the most severe form of the disease (Bhaskar, 1991).

Frank et al (in press) investigated the Co within moose of Nova Scotia, and concluded that Co and vitamin B<sub>12</sub> was deficient within the herds of the Tobetic and C.B.H regions, but was also observed to some extent within other mainland populations. The problem of 'moose sickness' was the focus of the study, which results in symptoms resembling that of 'coastal' or 'wasting disease'.

There is no clear role between Co concentration of enamel and dental disease. Within humans, only two known studies have been conducted analysing for affects of Co. Navia (1970, in Curzon and Cutress, 1983) concluded that Co was a doubtful as a trace element that promoted dental Caries. Driezen et al (1952, in Curzon and Cutress, 1983) could find no relationship with Co concentration in saliva with dental caries.

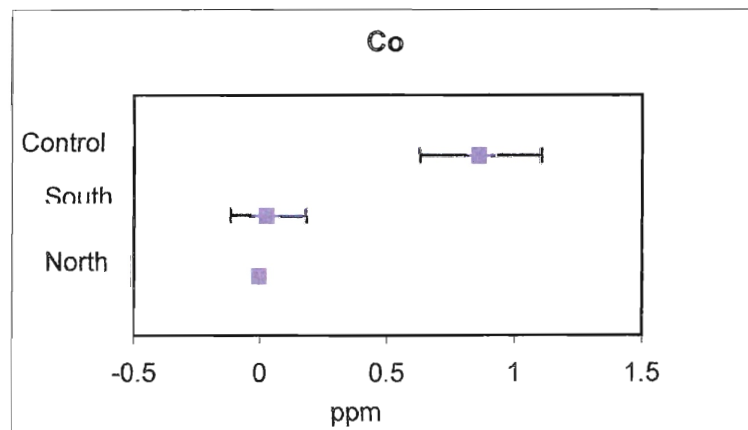


Figure 4.7: ANOVA results for Co. Individual 95% Confidence Intervals based on pooled standard deviation.

In animal experiments, a study conducted by Hendershot and Forsaith (1958, in Curzon and Cutress, 1983) showed there was a higher incidence of caries in rats that were fed Co-EDTA, yet they did not attempt to separate the effect of EDTA alone.

#### 4.9.5 Discussion of Tin (Sn)

Tin is significantly depleted in the C.B.H population compared to the control population (Figure 4.8). Chemically tin is non-toxic and non-reactive, and there is no evidence to support tin as performing any essential functions within plants and animals. It has a low absorption rate and low retention in animal tissues so therefore is generally non-toxic (Underwood, 1971).

The majority of studies conducted with Sn in relation to dental disease have been in the form of SnF<sub>2</sub>, fluoride being a known inhibitor of caries. It is therefore not clear whether Sn has an affect on caries by itself, as a separate entity of F (Curzon and Cutress, 1983). SnF in mouth rinse inhibits acid formation in plaque and also lessens the formation of plaque (Svatun and Attramadal, 1978, in Curzon and Cutress, 1983). Attramadal and Svatun (1980, Curzon and Cutress, 1983) showed that Bacteria in the oral environment show a rapid uptake of Sn greater than other metal cations.

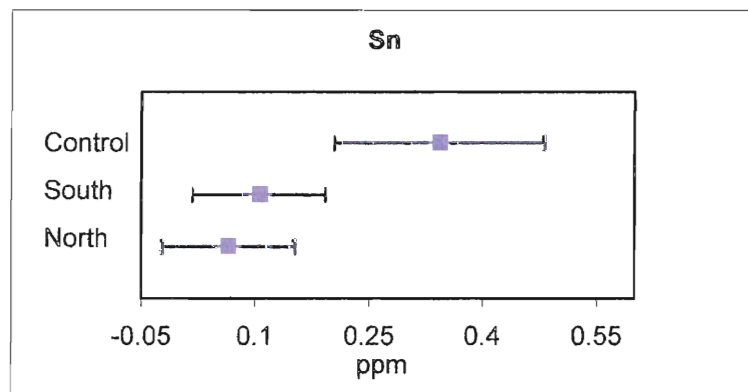


Figure 4.8: ANOVA results for Sn. Individual 95% Confidence Intervals based on pooled standard deviation.

The presence of  $\text{Sn}^{2+}$  may have an inhibitory effect on growth rate and metabolism of bacteria (Aikin and Dean, 1976, Curzon and Cutress, 1983). This is a possible reason for Sn having inhibiting affects on caries (Curzon and Cutress, 1983).

Studies conducted on Sn itself and its relationship with dental disease is very limited, although there has been considerable interest recently according to Curzon and Cutress (1983).

East Kemptville mine is situated in close proximity to the Tobeatic area, where the control population inhabits. This area is considered a major tin province, and when it was operating it was the largest Tin mine in North America (Zentilli, personal communication, 2004). This correlates with our results, which indicate higher levels of Sn within the enamel of the control population. Curzon and Cutress (1983) also describe the marked difference in trace element concentration between different geographic locations. The differences in concentrations between geographical areas would be highly likely as a result of geochemistry in the area.

#### **4.9.6 Conclusions**

Many investigations of trace elements and disease have been conducted for other species and also between species of ruminants, yet there are no data available that adequately describe the recommended dietary intake of trace elements for moose, and biological values from one species to another is always open to question (Frank et al, 2003).

Ba and Sr have both been implicated to result in poorer calcification of bones, and Cobalt is well documented in resulting in wasting disease, loss of appetite and even hypoplasia. The weakening of the teeth in the C.B.H may be a result of poor calcification, which ultimately affects the crystal structure within the enamel framework, and also hypoplasia, which is a defect resulting in the enamel being insufficiently represented in teeth that are affected causing enamel to become weak and vulnerable to caries, and possibly fractures.

Pb should not be excluded, because as Underwood (1971) states, Pb in deficient quantities should not be discounted in relation to performing any biological function. Further investigation should be considered to examine if there are any Fe deficiencies within the moose in response to experiments described by Frieden (1984).

Sn is chemically non-toxic and non-reactive, and has not been proven to have any essential role in the biological functions of plants or animals (Underwood, 1971). The correlation between the East Kemptville tin mine that occurs in close proximity to the control population, and the elevated levels of Tin within the enamel of the control, provides good evidence that trace elements can vary geographically, as a result of the geochemistry within the soils and bedrock. Based on the results for Sn it is possible to determine the origin (where a moose lived) based on tooth geochemistry

There are significant indications based on the results thus far. Further investigation is required, as our results are based on the assumption of only one control population and the populations of the C.B.H. The use of more than one control population with larger sample sizes to reduce any sampling error present would be required before any strong conclusions can be drawn.

#### **4.10 Healthy Vs North of the Park & Healthy Vs South of the Park**

Rubidium, Gallium, Yttrium and Thorium show significantly different values within the population North of the Park compared to the control. Rubidium, Yttrium and Thorium are all depleted, while Gallium shows elevated levels. There are no significant differences of elements South of the Park compared to the control area.

##### **4.10.1 Rubidium (Rb)**

Rubidium is significantly depleted in the population North of the Park compared to both South of the Park and the control population (Figure 4.9). Biological interest has been stimulated because of rubidium's close affiliation with potassium and its presence in living tissues in higher concentrations, relative to those of potassium, than in the terrestrial environment (Underwood, 1971).

Because of its close resemblance to potassium, all plant and animal cells are freely permeable to rubidium, at rates comparable to potassium (Underwood, 1971). According to Frieden (1984) rubidium cannot be excluded from being an essential, even though it has been proven to be essential at accurately determined levels.

Rubidium is not normally taken up by calcified tissues any more than by other parts of the body, and appears to be a normal constituent of enamel, yet due to its close physiochemical relationship with potassium it should be investigated further (Curzon and Cutress, 1983).

#### 4.10.2 Discussion of Yttrium (Y)

Yttrium is significantly depleted within the moose population North of the Park, and although relatively less South of the Park, it is not significantly depleted in this population (Figure 4,10). No information could be found in the literature regarding yttrium's role in biological processes.

The role of Y in relation to dental caries is still unknown, although studies done by Castillo-Mercado and Bibby (1973, in Curzon and Cutress, 1983) produced significant alterations in rat molar morphology by injection of Y during the period of tooth development in rats. Castillo-Mercado and Ludwig (1973, in Curzon and Cutress, 1983) yielded results in an associated study showing that rats being fed a cariogenic diet showed significantly lower caries prevalence when injected with Y. Rats receiving Y through drinking water also displayed lower caries prevalence, but not at a significant level.

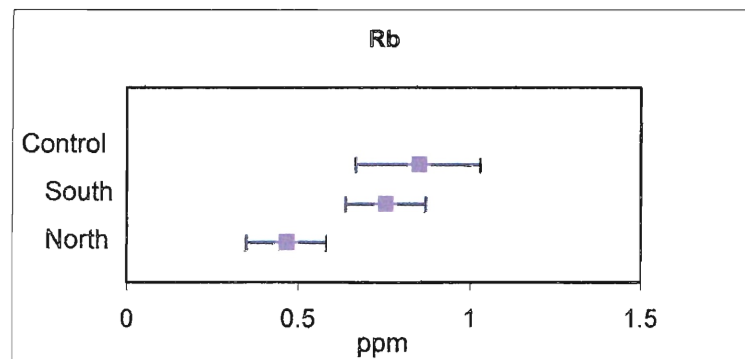


Figure 4.9: ANOVA results for Rb. Individual 95% Confidence Intervals based on pooled standard deviation.

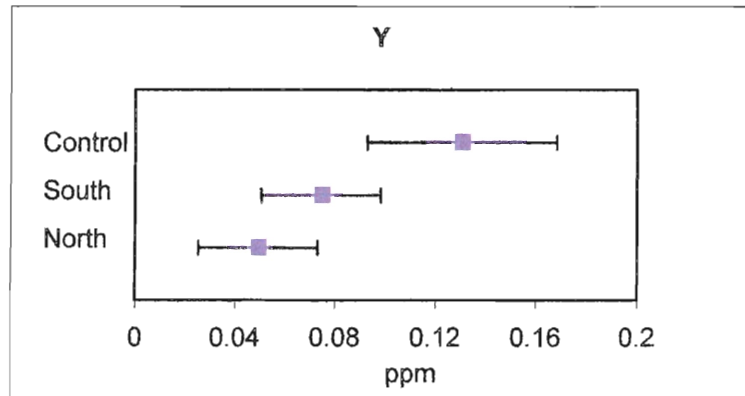


Figure 4.10: ANOVA results for Y. Individual 95% Confidence Intervals based on pooled standard deviation.

#### 4.10.3 Discussion of Thorium (Th)

Thorium is significantly depleted in moose North of the Park compared to the control population (Figure 4.11). No information was found in the literature regarding the role of Thorium in biological functions and dental disease.

#### 4.10.4 Discussion of Gallium (Ga)

Gallium is significantly elevated in the population North of the Park compared to population South of the Park and the control population (Figure 4.12). There is no information in the literature regarding gallium as performing any essential biological function in animals, or in dental disease. Although it has been shown that gallium is an essential micronutrient of the fungus *Aspergillus niger* and the duckweed *Lemna minor* (Fairbridge, 1972).

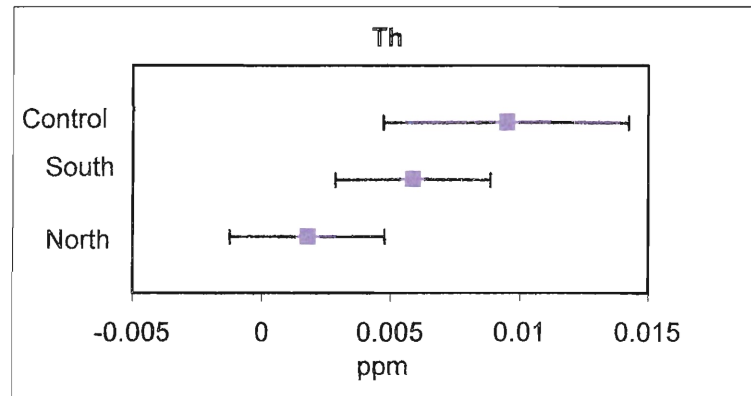


Figure 4.11: ANOVA results for Th. Individual 95% Confidence Intervals based on pooled standard deviation.

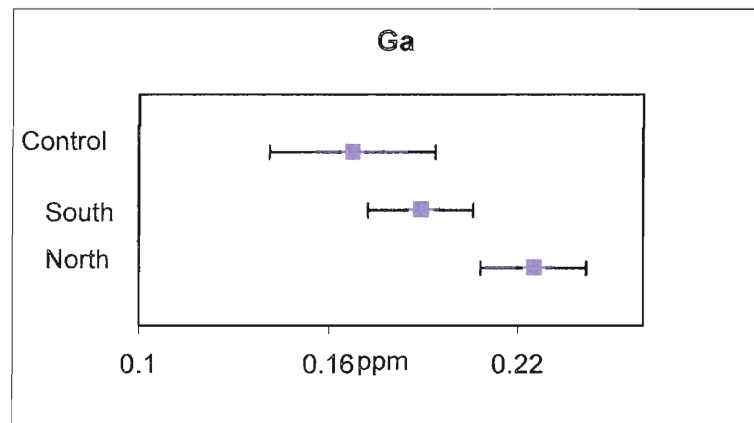


Figure 4.9: ANOVA results for Rb. Individual 95% Confidence Intervals based on pooled standard deviation.

#### 4.10.5 Conclusions

The results for Rubidium, Gallium, Yttrium and Thorium are unclear, particularly for Rubidium and Gallium, which are significantly depleted in the population North of the Park compared to both the control population and the population South of the Park. Although, if the geology differs between North and South of the Park this may account for the differences, as moose very rarely cross over between North and South of the Park, and when they do it is generally to replenish populations (Nette, personal communication, 2004).

Yttrium is the only element discussed that has information in the literature to implicate it as having a role in dental disease, showing a decrease in caries



when in increasing concentrations. This may be of significance as yttrium is depleted in the North, and, although not significant, yttrium does appear to be present in lesser amounts in the south as compared to the control also.

Results for Rb, Ga and Th do not provide any strong conclusions, but may warrant a further investigation with more than one control and increased sample sizes to reduce any sampling error that may be associated with this study.

#### **4.11 Correlations between Element (ppm) and Breakage score**

The correlation data for element concentration and breakage score is very unclear. The correlation trends that are seen in the population North of the Park (Appendix F) are not always consistent with the trends in correlation to the population South of the Park (Appendix G). R-square values are very low in some cases, yet the values can be higher in other cases. The fact that there is disagreement in correlation trends between the two populations, presents a problem for interpretation, as we would expect the data to follow the same trends.

##### **4.11.1 Conclusions**

Correlation data are inconclusive. Undoubtedly the number of samples is so far insufficient to draw useful conclusions, yet these are the first data available for moose teeth in the region, and raise interesting questions. By repeating the experiment with a larger sample size to reduce error associated with the sampling, and reproducing this experiment may further aid in understanding the correlation data. At present, there is no information or studies to compare our correlation data with and therefore, these data must be classified as inconclusive.

#### **4.12 Summary of Chapter**

Macrofractures are the type fractures visible with the naked eye. Based on work done by Smith (1992), Nette and Power (Unpublished, 1999) described and categorized teeth from the C.B.H area to determine the state of their teeth in terms of the amount of fracturing present.

Brown stains running perpendicular to the enamel surface are evidence of initial fracturing. The stained fractures appear to weaken the enamel structure, eventually breaking off. After breakage has occurred, the tooth surface will become polished and stained brown again. This is an important characteristic of the fracturing because it implies that the moose was living for a period of time after breakage occurred. When fresh breaks occur, the staining and rounded surface will not be present and can imply that breakage occurred just prior to the death of the animal, or even that it occurred after death.

Within the CBH, results show that 5 elements, Barium, Lead, Strontium, Cobalt and Tin show significantly depleted levels in the C.B.H moose compared to that of the control population.

Barium and Strontium have both been implicated to result in poorer calcification of bones, and Cobalt is well documented in resulting in wasting disease, loss of appetite and even hypoplasia. The weakening of the teeth in the C.B.H may be a result of poor calcification, which ultimately affects the crystal structure within the enamel framework, and hypoplasia, which is a defect resulting in the enamel being insufficiently represented in teeth that are affected causing enamel to become weak and vulnerable to caries, and possibly fractures.

The correlation between the East Kemptville tin mine that occurs in close proximity to the control population, and the elevated levels of Tin within the enamel of the control, provides good evidence that enamel trace element concentration can vary geographically, as a result of the geochemistry within the soils and bedrock.

Yttrium is depleted in the North, and, although not significant, yttrium does appear to be present in lesser amounts in the south compared to the control. Yttrium has also been implicated in dental disease, showing a decrease in caries when in increasing concentrations.

## **Chapter 5: Microfractures**

### **5.1 Introduction**

Microfractures are fractures that cannot be seen with the naked eye, and require the use of a high-powered microscope to investigate their prevalence within the enamel structure of the moose incisors. This chapter will combine the work of Graves and Casey (2002, unpublished), and new data obtained in 2004. Graves and Casey (2002, unpublished) investigated and categorized characteristic microfracturing, and Microprobe data, within both the control group of Tobeatic, and the unhealthy moose population of CBH. Graves and Casey (2002, unpublished) examined fracturing that was present in both unhealthy and healthy populations, and also unique fractures present only in the unhealthy CBH population. This Chapter will also introduce new data obtained from utilising the Microprobe at Dalhousie University.

### **5.2 Description of Microfractures**

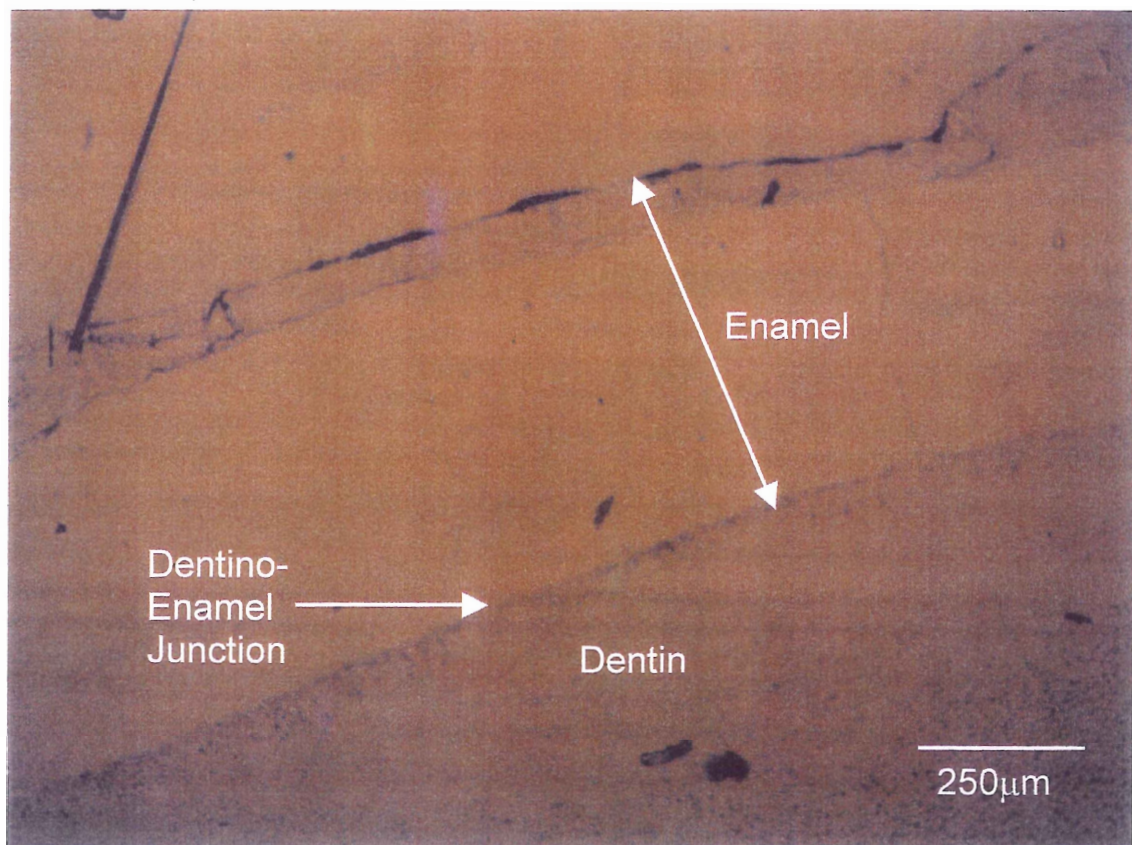
The following discussion and categorization of fracturing was identified analysing cross-sections of incisors under reflective light using the petrographic microscope (Zeiss Axioplan (German)) in the Fission Track Laboratory, Dalhousie University (Graves and Casey, 2002, unpublished). This method was employed following the example set by Young and Marty (1986).

The teeth were selected as being a representative sample of the entire populations, and were chosen on the basis of degree of fracturing (Breakage score) and the area in which the moose was located at the time of harvesting. Because there was no indication that the sex of the animal was a determinant factor when analysing the degree of breakage (Breakage score) (Nette, personal communication, 2004), it was therefore not considered for this analysis also.

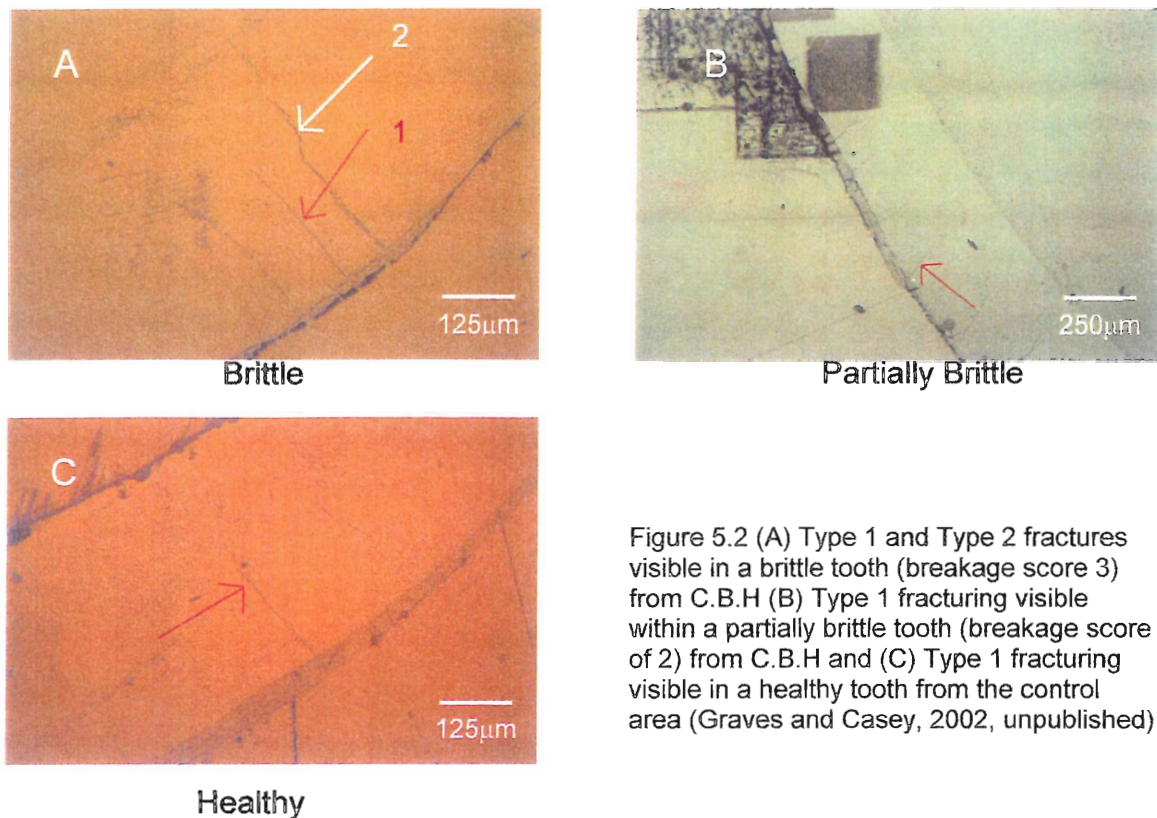
Graves and Casey (2002, unpublished) analysed a representative sample of 4 moose incisors; Healthy, Breakage score 1(CBH), Breakage score 2 (CBH), and Breakage score 3 (CBH), and divided their analysis into three subsections of

degree of breakage; healthy (Tobeatic), unhealthy, partially brittle (C.B.H), and unhealthy, very brittle (C.B.H).

Five different type fractures were common to all teeth analysed, healthy, 1 (none/slight), 2 (moderate) and 3 (severe), and two types of fracturing were unique to the very brittle CBH teeth. Figure 5.1 illustrates a cross-section of a tooth that is visible under the petrographic microscope using reflected light. From Figure 5.1 we can visibly identify the main components of the tooth, dentin as being the dark area on the lower right hand side, the dentino-enamel junction where the enamel forms a protective layer over the dentin, and the outer enamel.



**Figure 5.1:** Cross-section of incisor under reflected light. Enamel, Dentin and Dentino Enamel Junction (DEJ) are clearly visible (Graves and Casey, 2002 unpublished).



Type 1 fractures were described by Graves and Casey (2002, unpublished) as being long, narrow, shallow and following the same orientation of the apatite rods. Figure 5.2 (a, b and c) illustrates the occurrence of this fracture type, which is evident in all teeth types, both healthy and unhealthy.

Type 2 fractures were characterized by Graves and Casey (2002, unpublished) as being similar to the type 1 fractures, only that they were longer and wider than type one fractures. Type two fractures also occur in all teeth

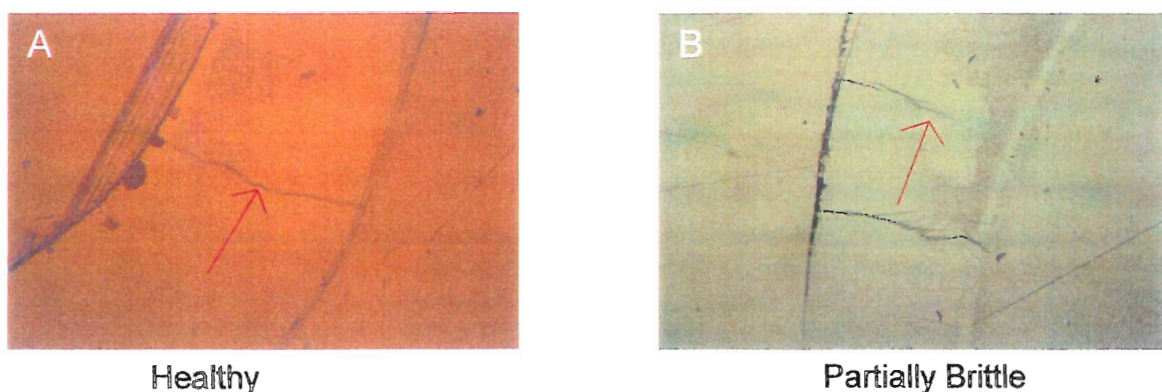


Figure 5.3: Type 2 fracturing (A) evident within a healthy tooth from the control population. A type 2 fracture (B) visible within a partially brittle tooth (Breakage score of 2) from C.B.H (Graves and Casey, 2002, unpublished)

types, healthy and unhealthy, and follow an orientation similar to the apatite crystals. Type two fractures are illustrated in the unhealthy teeth (Figure 5.2a and Figure 5.3b), and in the healthy teeth (Figure 5.3a).

According to Graves and Casey (2002, unpublished), type 3 fracturing is limited to the outside layering of the enamel. It is much smaller than the previous fracture types mentioned, and is characteristically short and shallow fracturing. It does, however, follow the same pattern as the previous two fractures in terms of orientation in that it follows the orientation of the apatite crystal structure. Figure 5.4 (a, b and c) illustrates type three fracturing that is common amongst all teeth.

Type 4 fractures were interpreted by Graves and Casey (2002, unpublished) as being the longest and widest of all fracture types. They are not limited to the enamel layering, but extend through to involve the dentine as well. Type 4 fractures were described also as being the deepest of the cracks and that they were often in filled with an unknown substance (Graves and Casey, 2002 unpublished).

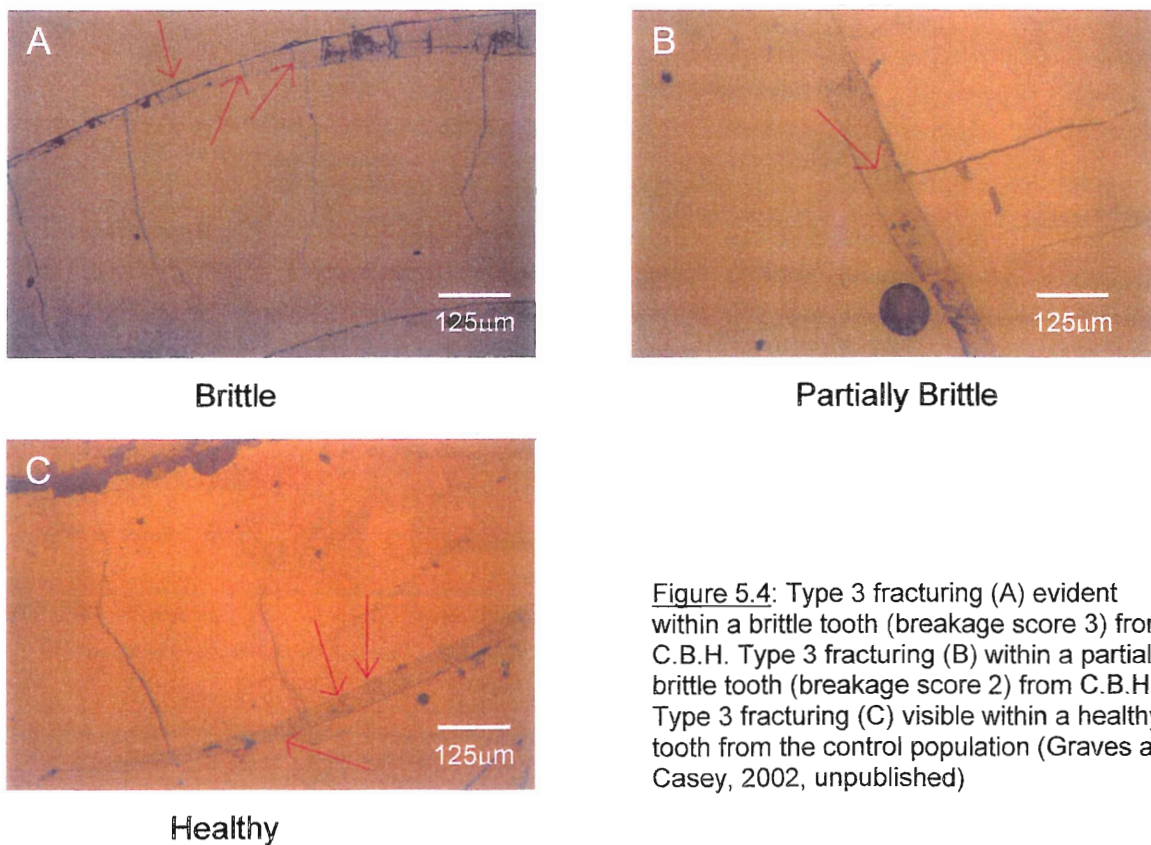
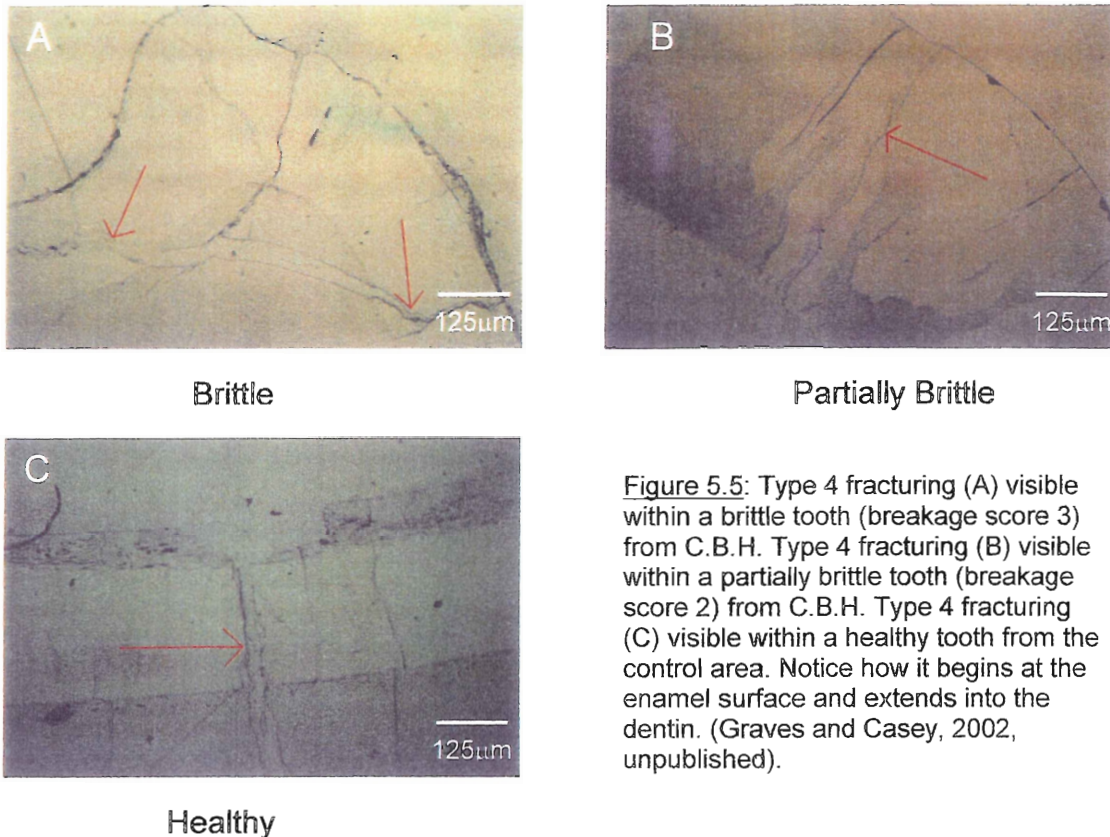


Figure 5.4: Type 3 fracturing (A) evident within a brittle tooth (breakage score 3) from C.B.H. Type 3 fracturing (B) within a partially brittle tooth (breakage score 2) from C.B.H. Type 3 fracturing (C) visible within a healthy tooth from the control population (Graves and Casey, 2002, unpublished)

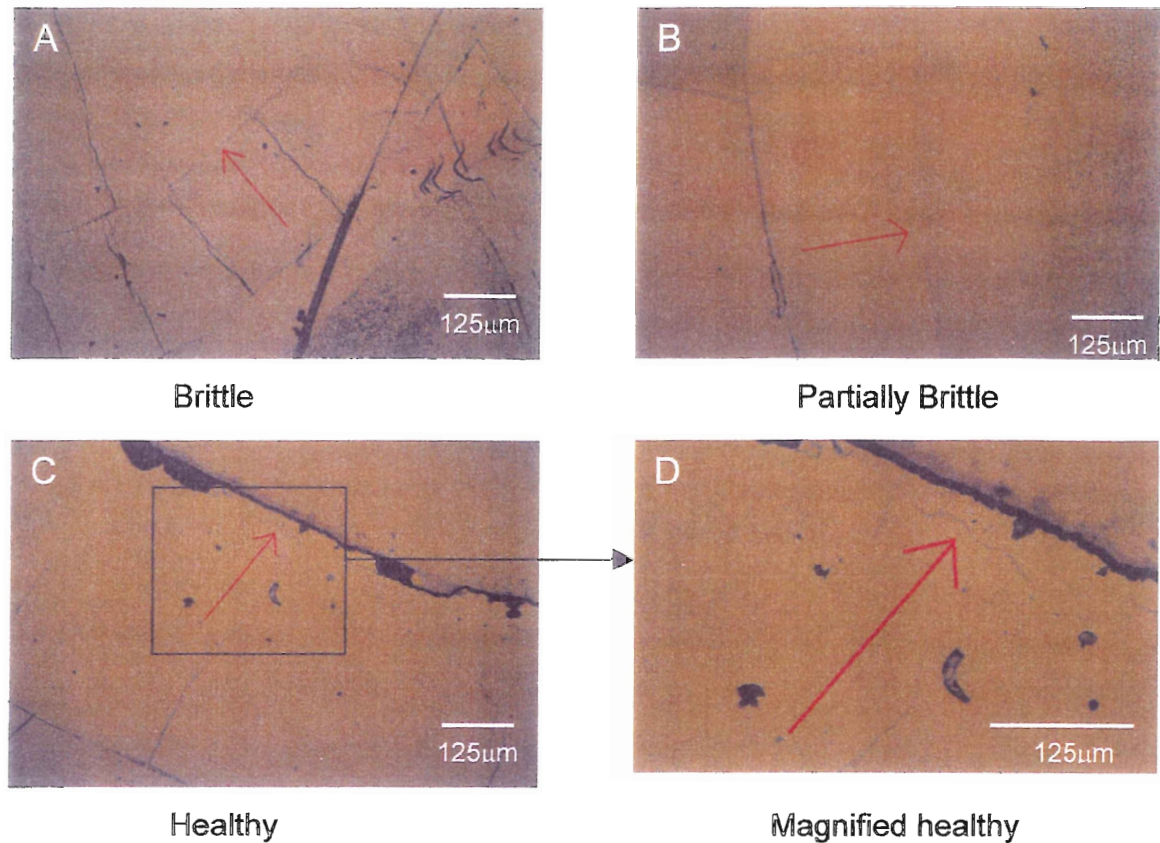


**Figure 5.5:** Type 4 fracturing (A) visible within a brittle tooth (breakage score 3) from C.B.H. Type 4 fracturing (B) visible within a partially brittle tooth (breakage score 2) from C.B.H. Type 4 fracturing (C) visible within a healthy tooth from the control area. Notice how it begins at the enamel surface and extends into the dentin. (Graves and Casey, 2002, unpublished).

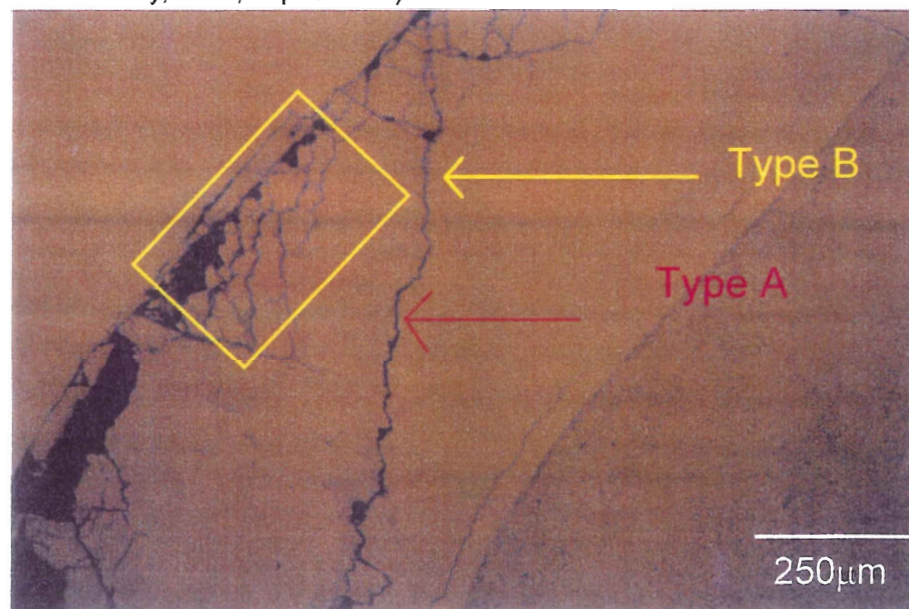
Figure 5.5 (a, b and c) shows the features of a type 4 fracture documented by Graves and Casey (2002, unpublished)

Type 5 fractures are the final fracture type common to all teeth. These fracture types were generally long, narrow, and wavy running parallel and with close proximity to the Dentino-Enamel junction (Graves and Casey, 2002 unpublished). Figure 5.6 (a, b, c and d) illustrates the nature of these fracture types, with figure 5.6d showing a magnified view from a healthy tooth.

The final type of fracturing was unique to the CBH populations limited to animals that exhibited a breakage score of 3. Two fracture types (A & B) were identified as occurring within the incisor studied by Graves and Casey (2002, unpublished). Type A was a long jagged line cutting 45 degrees across the enamel layer, from the Dentino-Enamel Junction to the outer edge of the enamel (Figure 5.7). Type B fractures were very randomized fracturing in terms of directions, they are very jagged, and are restricted to the outer  $\frac{1}{3}$  of the enamel (Figure 5.7).



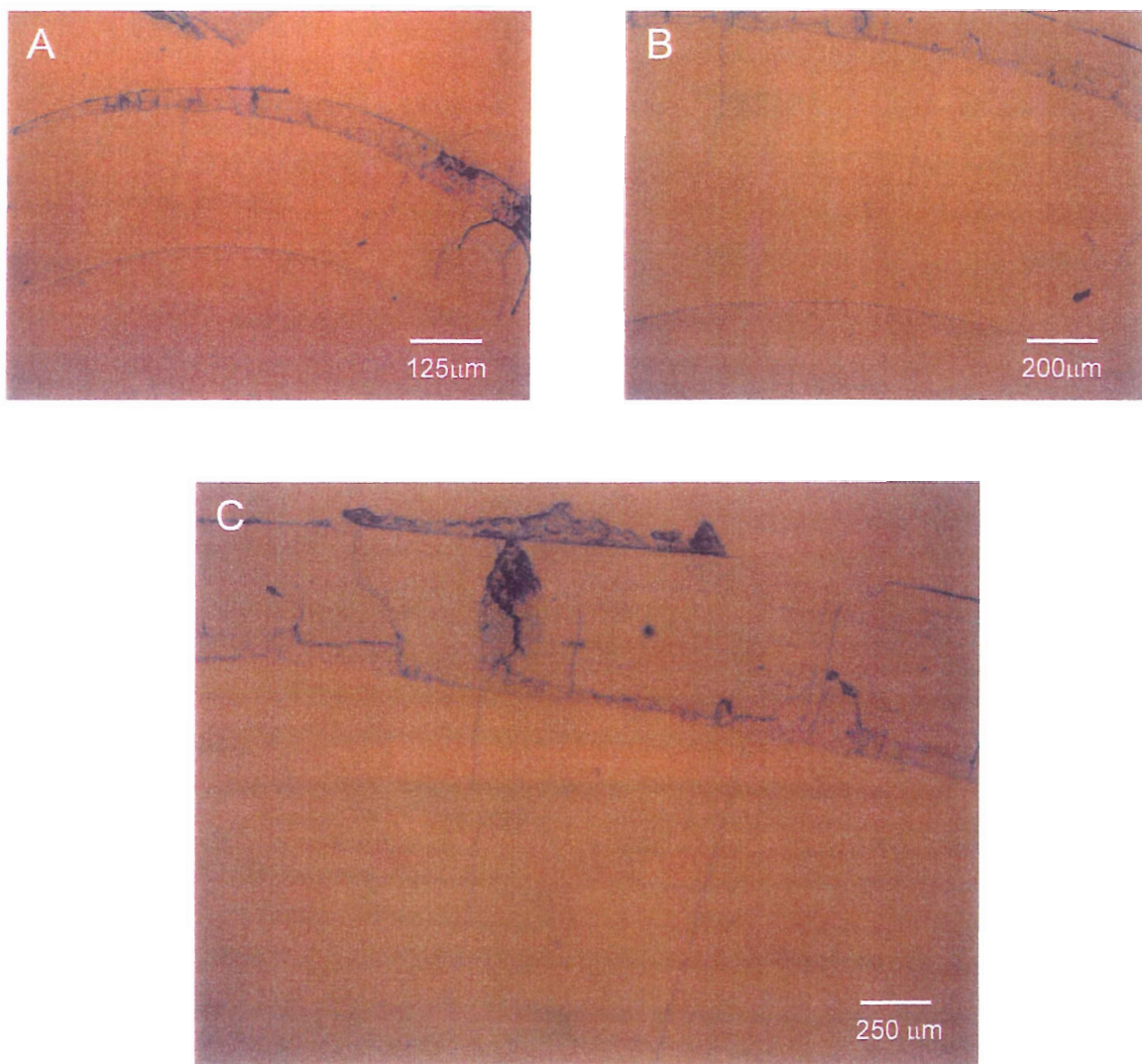
**Figure 5.6:** Type 5 fracturing (A) visible within a brittle tooth (breakage score 3) from C.B.H. Type 5 fracturing (B) visible within a partially brittle tooth (breakage score 2) from C.B.H. Type 5 fracturing visible within a healthy tooth from the control population. Magnified view (D) of a type 5 fracture. Notice it runs parallel, and not perpendicular like the other fracture types. (Graves and Casey, 2002, unpublished)



**Figure 5.7:** Type A fracture (red arrow) extending through the entire enamel framework at a  $45^{\circ}$  angle. Type B fracture (yellow arrow) occurs in the outer enamel and exhibits a dendritic pattern (Graves and Casey, 2002, unpublished)



An interesting feature that was not documented by Graves and Casey (2002, unpublished) is the fracturing that appears to cease at the outer most enamel layer, where another layer of enamel has appeared to have been calcified. Figure 5.8 (a, b and c) illustrates this phenomenon, as we clearly see that type 1 and 2 fractures will cease at the outer layer of enamel and also type 3 fractures will not involve the rest of the enamel, only the outside layering.



**Figure 5.8:** Section of enamel from M051 (control population). A, B, and C are different locations within the enamel at varying magnifications. Notice that the fracturing ceases to include the outer enamel (Graves and Casey, 2002, unpublished)

### 5.2.1 Scanning Electron Micrograph (SEM)

Figures 5.9 to 5.13 are sections of enamel that were photographed using the Scanning Electron Micrograph (SEM) in the microprobe lab at Dalhousie University. The enamel samples were mounted onto one individual slide and prepared at the same, with no difference in preparation between individual samples. This is important to note because we see varying degrees of fracturing amongst these four grains.

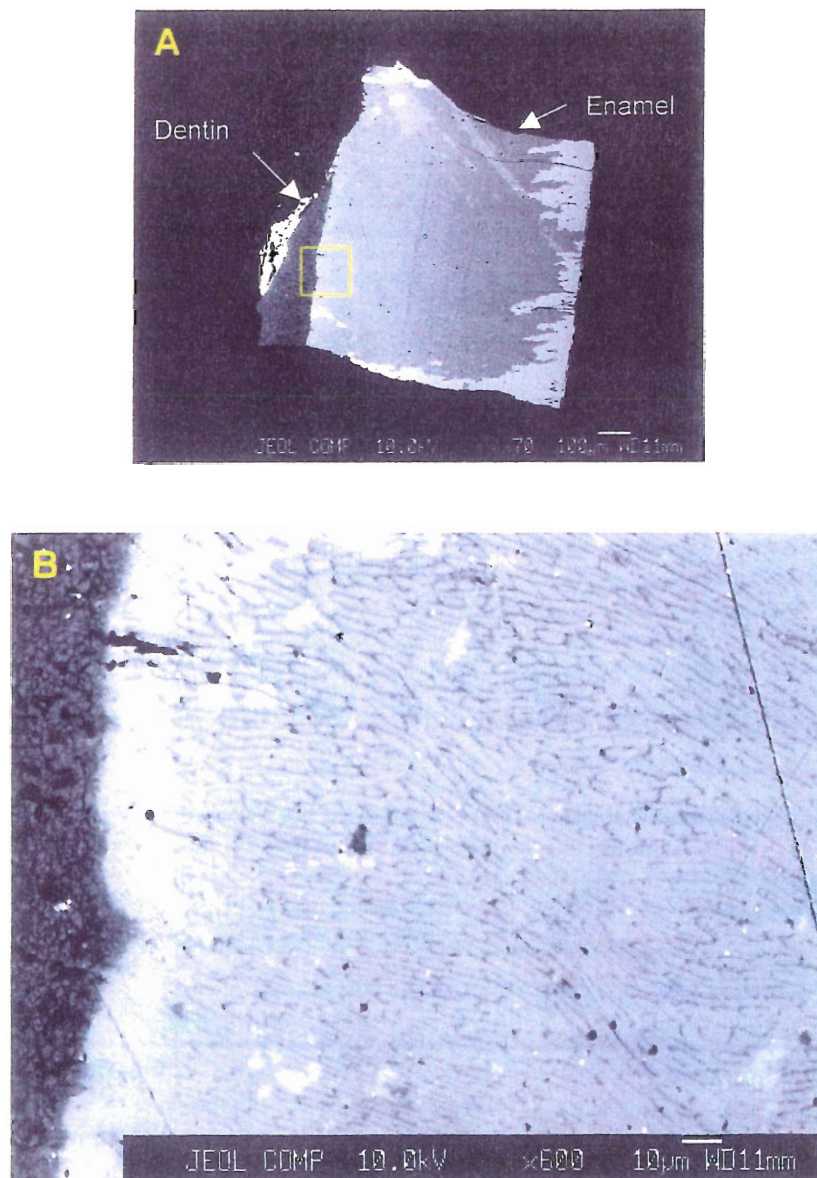


Figure 5.9: (A) Scanning Electron Micrograph (SEM) image of sample M056. (B) Magnified section of M056. Notice the wavy crystal structure.

The fracturing present within these samples appears to be similar in nature and orientation to the microfractures described by Graves and Casey (2002, unpublished). Also visible in the magnified sections of Figure 5.9b, Figure 5.10b and Figure 5.11b are the various prism structure patterns that are produced by cutting different planes of the enamel (Figure 3.3a).

Figure 5.9 is a sample taken from M052, a healthy moose from the control population. This sample shows very little fracturing.

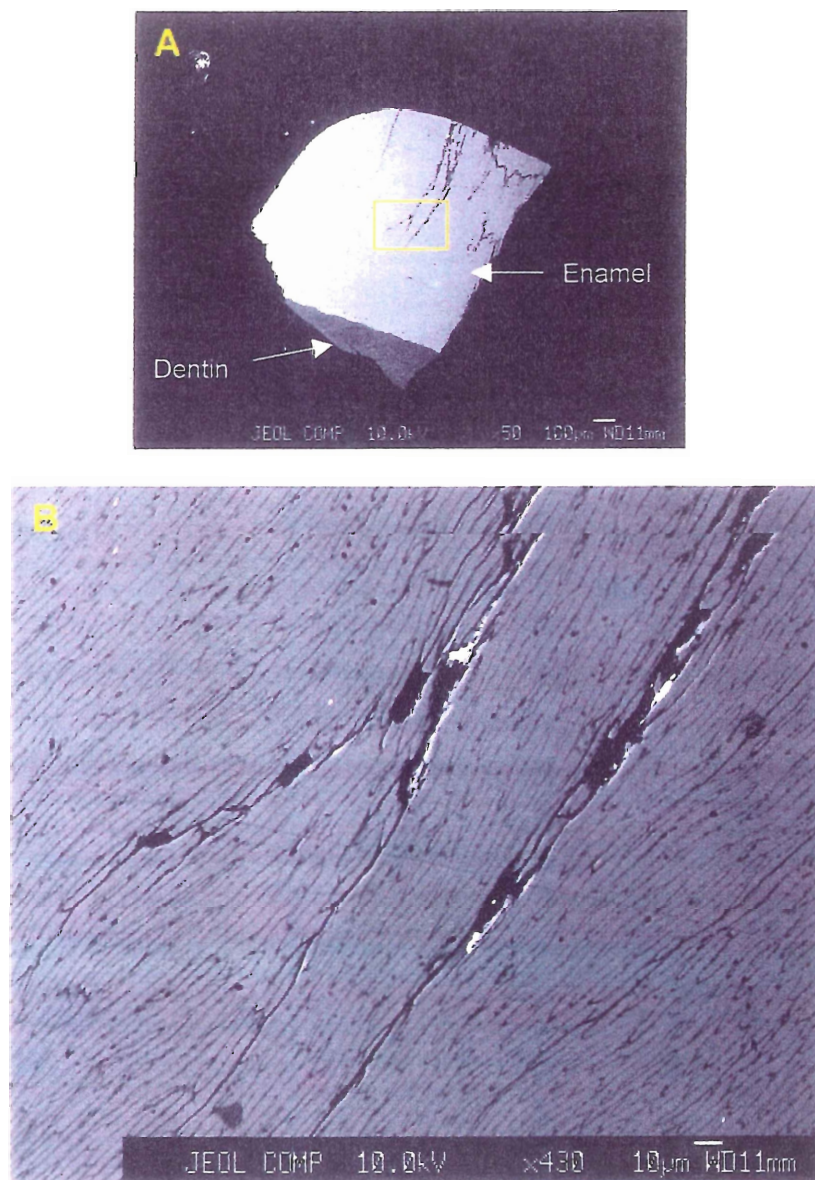


Figure 5.10: (A) Scanning Electron Micrograph (SEM) image of sample M027 (South of Park with breakage score 3) (B) Magnified section of M027. Notice the wavy crystal structure and orientation of fracturing.

Figure 5.10 is an enamel sample taken from M027, which exhibited a breakage score of 3 and was from the population South of the Park. Higher rates of fracturing are visible within this tooth sample compared to Figure 5.9.

Figure 5.11 is an enamel sample taken from M012, which exhibited a breakage score of 3 and was from the population North of the Park. Also visible are a higher frequency of fracturing compared to Figure 5.9.

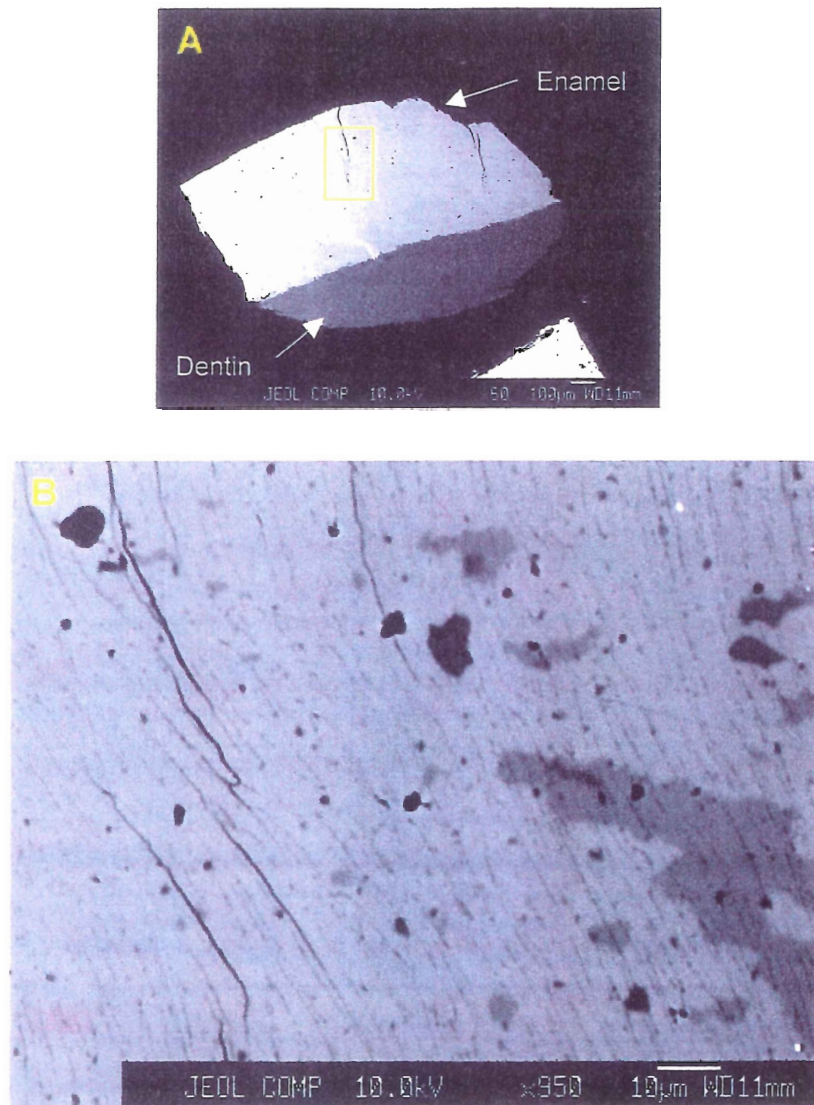
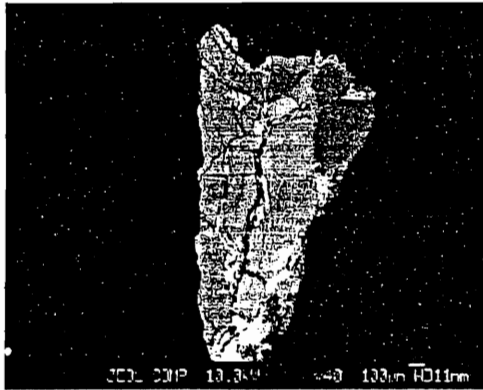
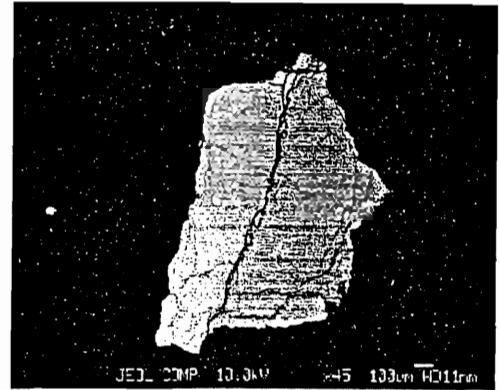


Figure 5.11: (A) Scanning Electron Micrograph (SEM) image of sample M012 (North of Park with breakage score 3) (B) Magnified section of M012. Notice the wavy crystal structure and orientation of fracturing.



**Figure 5.12:** Scanning Electron Micrograph (SEM) image of sample M030 (North of the Park with breakage score 3). Notice the degree of fracturing.



**Figure 5.13:** Scanning Electron Micrograph (SEM) image of sample M042 (South of the Park with breakage score 3). Notice the degree of fracturing.

Figure 5.12 and 5.13 are both from C.B.H (North and South of the Park respectively) with breakage score of 3. These samples are of interest due to the nature of fracturing. Unlike the previous samples, the fracturing has no preferred orientation and seems to be random and bearing close resemblance to Type B fracturing described by Graves and Casey (2002, unpublished). Prior to the microprobe analysis the samples did not exhibit this degree of fracturing, it was only when they were inserted into the microprobe that they began to fracture. This type of fracturing was not limited to the unhealthy teeth, and did occur in the one control sample, although it did occur in a higher frequency within the unhealthy teeth ( $1/4$  in Control,  $3/4$  South of the Park, and  $3/4$  North of the Park)

### 5.2.2 Discussion of Microfractures

There is no knowledge of microfractures occurring in the literature, and therefore comparisons with other studies cannot be made. Although, lamellae in teeth were initially misinterpreted as being 'cracks'. The following discussion on enamel lamellae and Stripes of Retzius was taken from Bhaskar (1991), unless otherwise noted.

In dentistry, the term 'cracks' was originally used to describe narrow, fissurelike structures that are seen in almost all enamel surfaces. It has since been demonstrated that these 'cracks' are in fact dental lamellae, thin, leaflike structures that extend from the enamel surface toward the dentino-enamel

junction (DEJ). They occur in the enamel fairly even spaced, and longer lamellae appear thicker than short ones, and occasionally they will extend into the dentin. In cross sectional views, enamel lamellae may be confused with cracks. Lamellae consist of organic material with little mineral content. It has been suggested that enamel lamellae may be a site of weakness in a tooth.

During cavity preparation, dentists are aware of the tendency of enamel to fracture along cleavage planes paralleling the course of the enamel rods (Avery, 1994).

Stripes of Retzius are lines or stripes (striae) that represent disturbed periods of growth. Accentuated stria of retzius (or Wilson band) is an observed in teeth of individuals as a result of some kind of 'trauma' (Goodman and Rose, 1990). A Wilson band will appear as a "marked incremental line", and has been associated with experimentally induced hypoplasias in sheep (Suckling and Thurley, 1984; in Goodman and Rose, 1990).

### **5.2.3 Conclusions for microfractures**

Type 1 – 5 microfractures described by Graves and Casey (2002, unpublished) are most likely to have been misinterpreted as being enamel lamellae. Based on descriptions by Bhaskar (1991), the microfractures show very close resemblance to enamel lamellae. They occur in all the teeth analysed (Healthy and unhealthy), and the fracturing begins at the enamel surface, and extends towards the DEJ, with type 4 involving the dentin (Figure 5.5). The fractures are fairly evenly spaced, and this is particularly noticeable in Figure 5.4a. The longer fractures, such as type 4, are noticeably wider than smaller fractures, such as type 1 fractures.

Figure 5.8 could possibly be a Wilson band, although further investigation would need to be conducted to draw any hard conclusions because Graves and Casey (unpublished, 2002) were interested primarily in microfractures. It is not known whether this characteristic was present amongst all the teeth from the C.B.H population, and also the control population, or restricted to the unhealthy population.

Type A and Type B fracturing are of significance. These fracture types are only present within the C.B.H populations. Due to the nature of the fracturing, it appears that these samples are more brittle, allowing for fracturing to occur in any direction.

The SEM images provide high resolution, high magnification images of the enamel structure. The fracturing present cannot be ruled out as being a result of the preparation of the slide, which required a polished surface for the microprobe, as it is established that enamel does have a tendency to fracture along the cleavage planes that parallel the enamel rods (Avery, 1994). It is interesting to note, however, that there appears to be a higher incidence of fracturing having occurred within the unhealthy teeth, than with the healthy teeth.

The fracturing that has occurred in Figure 5.12 and 5.13 is very interesting. Although it occurred in both the C.B.H and control populations, the frequency was higher in the unhealthy teeth. These fractures occurred after insertion into the microprobe, and the cause of this problem was most likely a strain put on the samples due to the environment within the probe, which creates a vacuum affect (Stoffyn, personal communication, 2004).

Although the possibility of fracturing occurring as a result of sample preparation, or the microprobe environment cannot be ruled out, it is important to note that the frequency of fracturing was higher in the unhealthy teeth, which may indicate that the C.B.H samples are brittle and weak compared to the control population.

### **5.3 Microprobe analysis**

Before the enamel was crushed to a fine powder, enamel grains were taken from twelve sample vials and were mounted onto a single slide. The slide was prepared by Gordon Brown, a thin section technologist at Dalhousie University, ready for analysis in the microprobe at Dalhousie University.

The following discussion on the Electron Microprobe is a summary of conversations with the Electron Microprobe technician, P. Stoffyn (2004). The Electron Microprobe works by focusing a beam on the sample; in this study it was a beam with a diameter of 5 $\mu$ m exciting an area on the sample that was

10 $\mu$ m diameter. The beam bombards the surface with electrons, exciting the atoms within the sample, and as these samples return to their non-excited state they emit X-rays of characteristic wavelength to particular elements. Using a wavelength and/or dispersive detector, the electron microprobe collects and records the X-rays for each element within the sample. The sampled X-rays are compared to the X-rays of a standard control sample, which the machine is calibrated before a sample is analysed. X-ray counts are converted into element concentration to provide a complete chemical analysis of the elements of interest within the sample.

### **5.3.1 Results for Electron Microprobe**

Nine elements were analysed in twelve samples, with a total of four samples from each population. Six points across the enamel surface were analysed, beginning from the inner enamel (Near the Dentino-enamel junction (DEJ)) towards the outer enamel. The nine elements that were analysed for were Calcium (Ca), Sodium (Na), Magnesium (Mg), Chlorine (Cl), Nitrogen (N), Fluorine (F), Oxygen (O), Sulphur (S) and Potassium (K). Of these nine elements, F and O were either not present or were present in levels below the detection limit of for the electron microprobe. The raw data output for the microprobe analysis is set out in appendix H.

Due to the nature of fracturing that occurred when the samples were placed into the electron microprobe (Figure 5.12 & 5.13), the orientation of the enamel surface was not always clear, and thus proving difficult in determining the location of the inner and outer enamel layers when performing any analysis across the enamel surface of these samples. Owing to this, the samples that will be included in the results are those that did not exhibit the fracturing such as Figure 5.12 & 5.13, only those samples where the outer and inner enamel could be distinguished (For example Figure 5.9). These include 3 samples from the control area (M051, M052 & M056), one from North of the Park (M012) and one from South of the Park (M027).

There is little variation in elemental concentration between the populations. Nitrogen shows an increase in concentration towards the outer



enamel, magnesium and sulphur show decreasing trends towards the outer enamel, sodium, and chloride remain constant throughout the enamel whilst potassium varies within and between the samples and shows no trends.

Appendix I illustrates these trends.

### **5.3.2 Discussion and Conclusion for Electron Microprobe**

The microprobe data was slightly useful. There was no indication for the elements that were analysed being depleted/elevated within the study area compared to the control population.

Elemental concentration varying across the enamel framework was well illustrated between all samples, all following similar trends.

Overall, incisors of the different population showed similar trends and concentration.

### **5.4 Summary of Chapter**

Five different type fractures were common to all teeth analysed, healthy, 1 (none/slight), 2 (moderate) and 3 (severe). Two fracture types were unique to the CBH populations limited to animals that exhibited a breakage score of 3.

With the use of the Scanning Electron Micrograph (SEM) within the Electron Microprobe at Dalhousie University, fracturing was evident within cross-sections of enamel at high magnification. The fracturing present within these samples appears to be similar in nature and orientation to the microfractures described by Graves and Casey (2002, unpublished).

Lamellae in teeth were initially misinterpreted by dentists as being 'cracks', and in cross-sectional views of enamel, lamellae may be misinterpreted as being 'cracks. Lamellae are, thin, leaflike structures that extend from the enamel surface toward the dentino-enamel junction (DEJ). They occur in the enamel fairly even spaced, and longer lamellae appear thicker than short ones, and occasionally they will extend into the dentin.

Graves and Casey (2002, unpublished) most likely misinterpreted lamellae as being 'cracks' within the tooth, as it is a common feature in all populations analysed.

Fractures that were present in the Scanning Electron Micrograph (SEM) images are most likely due to preparation of the samples and the process of microprobe analysis. It is important to note that the frequency of fracturing was higher in the unhealthy teeth, which may indicate that the C.B.H samples are brittle and weak compared to the control population.

During cavity preparation, dentists are aware of the tendency of enamel to fracture along cleavage planes paralleling the course of the enamel rods (Avery, 1994).

## **CHAPTER 6: Correlation between the Geochemistry of the Moose Teeth and Geochemistry of the surficial geology**

### **6.1 Introduction**

This chapter discusses the five elements that show significant differences between the study area and the control population, Barium, Lead, Strontium, Cobalt and Tin. Each element is discussed individually, and compared/correlated with the geology and geochemistry of the Cape Breton Highlands and areas within Shelburne County as described in previous studies in the literature. The information is drawn together and conclusions made based on the correlations with the teeth geochemistry and the geochemistry of the geology of the areas in question.

### **6.2 Cape Breton Highlands regional geology**

The geology of the Cape Breton Highlands (C.B.H's) is composed of a complex assemblage of units. The area has been subdivided into three pre-Carboniferous tectonostratigraphic zones and/or terranes by Barr and Raeside (1989) (after Fyffe and Fricker (1987) and Williams et al. (1988)). The three units are referred to as the Blair River Complex, occurring in the northernmost region of the Cape Breton Highlands, the Aspy Terrane, occurring in the North-western region, and finally the Bras d'Or Terrane, occurring in the southern regions of the Cape Breton Highlands (Figure 6.1) (Barr and Jamieson, 1991; Barr et al 1987; Raeside and Barr, 1990).

#### **6.2.1 Blair River Complex**

The Blair River complex is composed of a distinctive assemblage of basement rocks including felsic and mafic gneisses, monzodiorite, anorthosite, and syentite (Neale 1963a, 1963b; Raeside et al. 1986; *In* Barr and Raeside, 1987). It includes the metamorphosed plutonic and stratified lithologies north of the Red River and west of the Wilkie Brook fault systems (Barr et al. 1987).

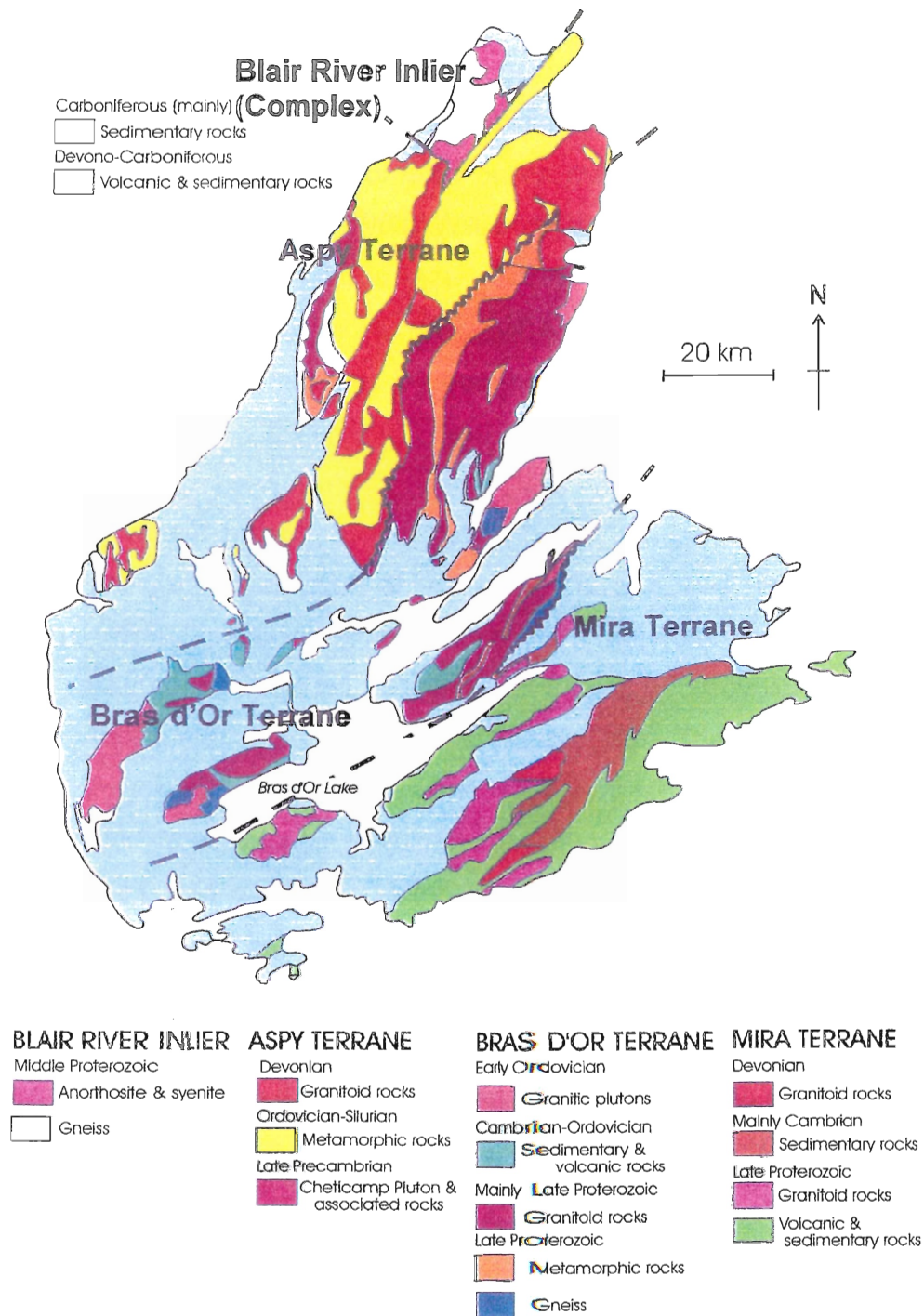


Figure 6.1: Simplified version of the complex geology of the C.B.H. Notice the dominant unit of Aspy Terrane (Barr, 2002, in Walsh, 200, unpublished)

related dioritic, tonalitic, granodioritic, and granitic plutons are also present. Also particularly abundant in the plutons situated within the Highlands region are plutons containing magmatic epidote and high-aluminium hornblende.

The three proterozoic components of the Bras d'Or Terrane have been recognized in the Brookville Terrane of southern New Brunswick, and late Proterozoic gneiss, late proterozoic-early Cambrian calc-alkalic plutons and Ordovician granitic plutons have been reported in parts of the Hermitage Flexure of Southern Newfoundland (Raeside and Barr, 1990).

#### **6.2.4 Geochemistry of the Cape Breton Highlands (CBH's)**

Sangster et al (1990) examined marble-hosted zinc occurrences at Lime Hill and Meat Cove within the highlands region on Cape Breton Island. It was concluded that the area of study was geologically, geochemically, and isotopically (S and Pb) similar to a distinctive group of zinc occurrences hosted by Grenville Supergroup marble in Ontario, Quebec and New York. It was also concluded that minor elements commonly associated with base metal sulphide deposits, including Ag, Au, Sb, Co, Ni, and Mo are anomalously low compared to the similar Grenville Supergroup (Sangster et al. 1990).

### **6.3 Southwest Nova Scotia regional Geology**

The control area situated in Southwestern Nova Scotia occurs within an area known geologically as the Meguma Supergroup (Figure 6.2). The Meguma Supergroup consists of thick metamorphosed siliclastic sequences, and is subdivided into two groups: Goldenville Formation and the overlying Halifax Formation, also referred to as the Goldenville-Halifax Transition (GHT) zone (Figure 6.3) (Graves and Zentilli, 1988). The Goldenville Group is composed of Cambrian greywacke with minor slates and the Halifax Group is composed of the Early Ordovician slates with minor greywacke (Schenk, 1995<sup>a,b</sup>).

The area is also known for the East Kemptville cassiterite deposit, North America's only producing primary tin mine (Richardson et al., 1988).

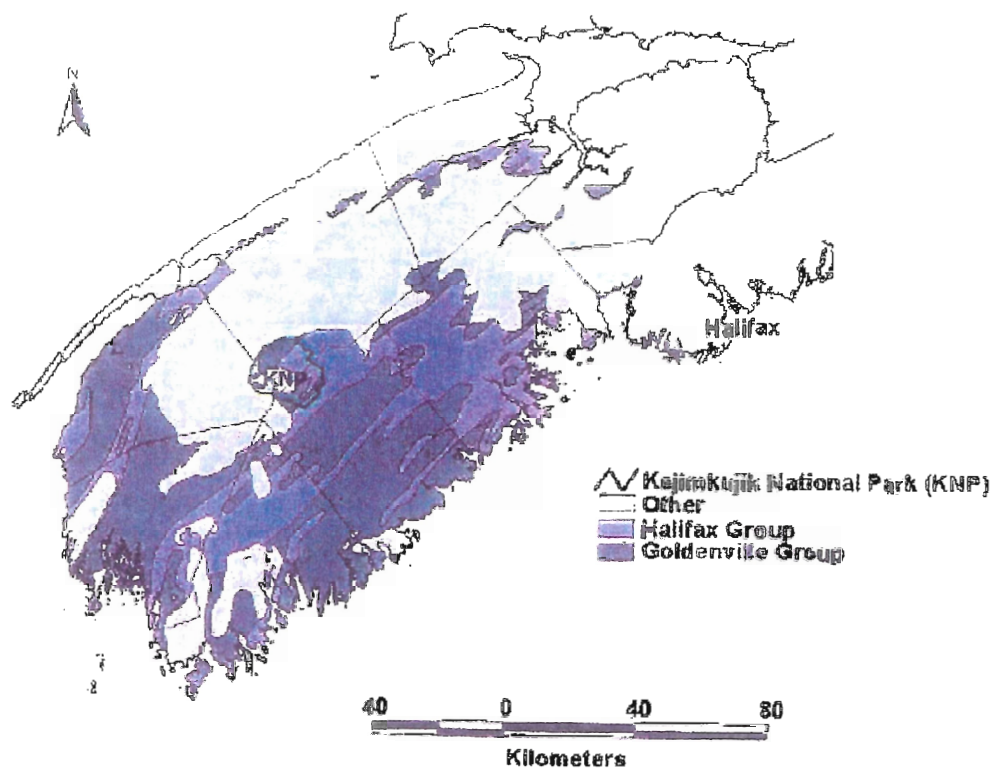


Figure 6.2: The Meguma Supergroup of southwestern mainland Nova Scotia, including the Goldenville Group and the Halifax Group. (Keppie, 2000, in Culgin, 2001, in Walsh, 2003, unpublished).

### 6.3.1 Geochemistry of Southwest Nova Scotia

East Kemptonville, situated in close proximity to the control area, was for a while North America's only producing primary tin mine, with a cassiterite (Tin Oxide) deposit of 58 million tonnes (0.165% tin) (Richardson et al., 1988). Manganiferous calcareous argillite and black slate at the base of the Halifax Formation are preferentially enriched in Mn, total C, Ba, Pb, Co, Zn, Cu, Mo, W and Au over average crustal values and over lithologies of the Goldenville Halifax transition zone (GHT) (Graves and Zentilli, 1988).



#### **6.4 Discussion of trace elements of C.B.H moose teeth**

The following discussions on the trace elements that show depleted levels, with respect to the control area, in the moose teeth enamel of the C.B.H moose, Barium, Lead, Strontium, Cobalt and Tin, is taken from Fairbridge (1972), unless otherwise noted.

##### **6.4.1 Barium (Ba)**

Barium has an atomic weight of 137.56 with atomic number 56, and belongs to the series of elements known as the alkaline earth elements. Metallic Barium does not occur naturally in nature. It occurs in nature as: barite  $\text{BaSO}_4$ ; Witherite (The most common barium-bearing carbonate)  $\text{BaCO}_3$ ; The less common barium minerals are: Barytoanglesite  $(\text{Ba, Pb})\text{SO}_4$ ; Bromlite  $\text{CaBa}(\text{CO})_3$ . Barium silicates are uncommon and include the feldspar of Celsian  $\text{BaAl}_2\text{Si}_2\text{O}_8$ . In terms of the overall geochemical cycle, Barium most closely resembles Strontium, another important alkaline earth trace element of slightly smaller size.

In igneous and metamorphic rocks the most important occurrence of Barium is as a trace element in the tectosilicate mineral, orthoclase feldspar, yet orthoclase from the late stage pegmatitic rocks are typically low in Barium. Generally, felsic igneous rocks contain more barium than basic rocks, and Alkalic rocks such as syenite contain the highest levels of Barium (upwards of 1600ppm).

The Meguma Supergroup is locally enriched in Ba (Graves and Zentilli, 1988), and the occurrence of alkalic rocks within the C.B.H (Raeside and Barr, 1990; Barr and Jamieson, 1991) also suggests that this region should also have an abundance of Ba, however this is not reflected in the geochemistry of the moose teeth, as the control area shows significantly higher concentrations against the study area.



### **6.4.2 Lead (Pb)**

Lead has an atomic weight of 207.19 and atomic number 82. It is the most abundant of the heavy metals within the earth's crust, about 15ppm by weight, and galena, PbS, is the major lead mineral, but there are a large number of sulphates (about twenty) that are also abundant.

Shale's deposits are where the bulk of sedimentary lead is found at an average concentration of 20 ppm, and concentrations in metamorphic deposits are wholly dependant on the original rock concentration but are generally found in highest concentrations within potassium-rich metamorphic rocks such as mica schists and gneiss.

The Meguma Supergroup is locally enriched with Pb (Graves and Zentilli, 1988), and there are also known sphalerite-bearing marbles that contain minor Pb within the C.B.H (Sangster et al. 1990). The moose teeth from the control area have significantly higher Pb concentrations within their tooth enamel.

### **6.4.3 Strontium (Sr)**

Strontium has an atomic weight of 87.63 and atomic number 38, and is the least abundant of the alkaline earth metals. Strontianite,  $\text{SrCO}_3$ , and celestite,  $\text{SrCO}_4$ , are the main ores for strontium. Calcium and strontium have a strong interaction, as the distribution of Sr within minerals, rocks, sediments and water is affected by the presence of calcium. It is generally found in greatest amounts in calcium rich minerals, and to a lesser extent in potassium rich minerals. Basaltic rocks show a large range of values for strontium, as very little coherence of strontium and calcium is seen. In contrast, granitic rocks show a definite coherence between the two elements, with increasing calcium content implying increasing strontium content. There are regional variations in the Sr content of granitic rocks, as samples from Africa show 160ppm, while the Canadian Shield show values of 305ppm.

Strontium behaviour in sedimentary rocks is almost unpredictable because of the many influences on Sr content in low temperature deposition. Generally, shales, limestones and metamorphic schists do not seem to show strontium

coherence with calcium. Strontium contents for various minerals have been determined, and the highest concentrations occur in plagioclases and feldspars, such as monzonites and syenites, with values as high as 1600ppm.

Based on the geology, it is expected that Sr would be in higher concentrations in C.B.H moose teeth as the area is locally abundant with sub-alkalic to alkalic rocks (Raeside and Barr, 1990; Barr and Jamieson, 1991), this is not the case however. Moose teeth enamel from the control area show significantly higher levels of Sr.

However, the behaviour of Sr in sedimentary stages is almost unpredictable according to Fairbridge (1972) and many factors, which are beyond the scope of this thesis, may affect the Sr content of rocks.

#### **6.4.4 Cobalt (Co)**

Cobalt has an atomic weight of 58.9332 and atomic number 27. It is a hard bluish white metal and  $\text{Co}^{59}$  is the only known stable nuclide to exist in nature. The bulk of Cobalt is found within pyroxenes and olivines, with greatest concentrations occurring in sulphide bodies.

Weathering and sedimentation of cobalt displays no marked differentiation, as the simple trivalent state is unstable in water yet redzate sediments concentrate cobalt due to precipitation of insoluble sulphides. Shales exhibit a mean concentration of cobalt that is similar to the crust of the earth, 25ppm.

One would expect Co to be in higher concentrations in the moose teeth enamel from the C.B.H due to the presence of sulphide deposits (Sangster et al. 1990) where one would expect locally enriched Co occurrences. This is not the case, as the control area shows significantly higher concentrations of Co compared to the study area.

#### **6.4.5 Tin (Sn)**

Tin has an atomic weight of 118.69 and atomic number 50. Tin has 10 stable isotopes in nature, the largest of any element, yet is relatively rare in the earth's crust. It is generally associated in granites, but is rather enigmatic as it is

quite abundant in some places, but extremely rare in others. The most common mineral by far is the oxide, cassiterite. Cassiterite is the main tin-forming mineral and because of durability it is generally found in weathered areas as saprolites and placer accumulations.

Tin does not readily associate with other elements, so the list of possible minerals is short, however it can replace iron, scandium and titanium.

One would expect tin to be higher in concentration within the teeth of the control area compared to the C.B.H due to the cassiterite deposits, and this is case

### **6.5 Conclusions**

Based on the literature, interpretations are somewhat confusing. Barium and Strontium are usually to be found in highest concentrations within Alkalic rocks such as monzonites and syenites, in concentrations upwards of 1600ppm. The Blair river Complex and Aspy Terrane are both alkalic-subalkalic in composition (Barr et al. 1987; Barr and Jamieson, 1991), and these two units form the majority of land area in the C.B.H (Figure 6.1).

Areas in southwest Nova Scotia are considered to be enriched in Barium, amongst others (Graves and Zentilli, 1988), and due to a close geochemical relationship with Sr, it would be expected that Sr would also be enriched.

However, due to the unpredictable nature of Sr and the high variations associated with this element, further investigation is required.

Cobalt would be expected to be in higher concentrations in the C.B.H due to the parent rock present, but as Sangster et al. (1990) concluded, minor elements commonly associated with base metal sulphide deposits, including Ag, Au, Sb, Co, Ni, and Mo are low compared to similar rock types of the Grenville Supergroup.

Due to the locality of the East Kemptville mine that occurs in close proximity with the control area, and also knowing that tin is relatively rare other than being abundant in cassiterite, there is a strong correlation between the tin concentrations within moose teeth enamel and the regional geology of the

geographic locality of moose origin. Based on these results for tin, Ba, Sr, and Co it is possible to determine the geographic origin of a moose.

### **6.6 Summary of Chapter**

Sangster et al (1990) concluded that marble-hosted zinc occurrences at Lime Hill and Meat Cove within the C.B.H were anomalously low in Ag, Au, Sb, Co, Ni, and Mo compared the zinc occurrences hosted by Grenville supergroup marble in Ontario, Quebec and New York, which is considered geologically, geochemically, and isotopically (S and Pb) similar to occurrences within the C.B.H.

Southwest N.S is known for the East Kemptville cassiterite deposit, North Americas only producing primary tin mine East Kemptville, situated in close proximity to the control area, was for a while North Americas only producing primary tin mine, with a cassiterite (Tin Oxide) deposit of 58 million tonnes (0.165% tin) (Richardson et al., 1988).

Manganiferous calcareous argillite and black slate at the base of the Halifax Formation are preferentially enriched in Mn, total C, Ba, Pb, Co, Zn, Cu, Mo, W and Au over average crustal values and over lithologies of the Goldenville Halifax transition zone (GHT) (Graves and Zentilli, 1988).

Generally, felsic igneous rocks contain more barium than basic rocks, and Alkalic rocks such as syenite contain the highest levels of Barium (upwards of 1600ppm).

Sr is generally found in greatest amounts in calcium rich minerals, and to a lesser extent in potassium rich minerals. Strontium contents for various minerals have been determined, and the highest concentrations occur in plagioclases and feldspars, such as monzonites and syenites, with values as high as 1600ppm.

Based on these results for tin, Ba, Sr, and Co it is possible to determine the geographic origin of a moose.

## **Chapter 7: Conclusions and Recommendations for Future Work.**

### **7.1 Conclusions**

- Moose require trace elements in small amounts, although little is known about trace element benefits/requirements in moose. These minerals are obtained from the forage that moose ingest, and the forage will vary geographically as moose are inclined to sample new plants that they encounter.
- Moose require a variety of plant species in order to obtain their nutritional requirements, and are therefore considered a 'generalist' browser. This forage must be highly palatable for the moose, and must contain optimum levels of various nutrient and mineral components. Actively growing plant tissues represent the highest quality foods available to moose.
- Increased bark stripping amongst C.B.H moose has been observed, and this behaviour usually occurs when areas support poor winter browse, when food is in short supply in late winter when preferred foods are unavailable or when animals are restricted by deep snow cover. It is increasingly evident in areas of C.B.H that support high-density moose populations where heavy browsing of preferred vegetation is evident.
- Bark stripping is not a common behaviour for moose, and is very rarely observed within populations. It is referred to as being a 'starvation' (last resort) food and does not normally comprise a large portion of a moose's diet.
- Bark stripping has implications on moose vitality, as it is important for moose to have a variety of high quality forage in their diet. Bark stripping is suggestive of moose feeding on few plant species, and this will increase the risk of becoming deficient/toxic of trace elements because certain plant species may hyperaccumulate trace elements or may not readily absorb trace elements into the tissue. It is important to

feed on a wide spectrum of plants to obtain essential nutrients not only in sufficient amounts but also in physiologically balanced proportions.

- Osteophagia has also been observed within the C.B.H population, which is triggered by a deficiency of phosphorous. It appears to be the first documented case of osteophagia in moose. This phenomenon occurs in natural conditions and has a distinct geographical distribution that is dependant on the parent rock in which the plants are growing which may be deficient in phosphorous. It may also be influenced by excessive concentrations of Ca, Al or Fe, which can reduce phosphorous availability of to plants.
- Geochemical analysis of the antlers generally shows a higher concentration of Ca/P ratio and Mn within the chewed C.B.H antler. The unchewed, control, antler generally shows higher concentrations of C Ba and K
- Barium, Lead, Strontium, Cobalt and Tin are significantly depleted within the tooth enamel of C.B.H moose compared to that of the control population.
- It has been demonstrated that the omission of Ba and Sr from the diet may result in depressed growth and reduced calcification of bones and teeth in rats and Guinea Pigs.
- The dietary benefits of cobalt have been known for decades. Co has a relationship with vitamin B<sub>12</sub> synthesis, and Co deficient animals lack the ability to synthesize vitamin B<sub>12</sub> properly. Symptoms usually include a gradual loss of appetite and failure of growth or loss of body weight, and in some animals there is evidence of hypoplasia within erythrocytic tissues and in bone marrow (Underwood, 1971)
- Hypoplasia in enamel is well documented, and is a defect that results in a lesser quantity of enamel covering the dentin that would normally be present that occurs during matrix formation
- Excessive lead toxicity has been well documented over the years, with symptoms that include anaemia, and neuropathy or encephalopathy

(Underwood, 1971). Yet, the possibility that lead in low concentrations performing some essential biochemical function cannot be excluded

- The weakening of the teeth in the C.B.H may be a result of poor calcification, which ultimately affects the crystal structure within the enamel framework, and also hypoplasia, which is a defect resulting in the enamel being insufficiently represented in teeth that are affected causing enamel to become weak and vulnerable to caries, and possibly fractures.
- In dentistry, the term 'cracks' was originally used to describe narrow, fissurelike structures that are seen in almost all enamel surfaces. It has since been demonstrated that these 'cracks' are in fact dental lamellae, thin, leaflike structures that extend from the enamel surface toward the dentino-enamel junction (DEJ).
- Lamellae consist of organic material with little mineral content. It has been suggested that enamel lamellae may be a site of weakness in a tooth.
- During cavity preparation, dentists are aware of the tendency of enamel to fracture along cleavage planes paralleling the course of the enamel rods (Avery, 1994).
- Type 1 – 5 microfractures described by Graves and Casey (2002, unpublished) are most likely to have been misinterpreted as being enamel lamellae. Based on descriptions by Bhaskar (1991), the microfractures show very close resemblance to enamel lamellae. They occur in all the teeth analysed (Healthy and unhealthy), and the fracturing begins at the enamel surface, and extends towards the DEJ, with type 4 involving the dentin. The fractures are fairly evenly spaced. The longer fractures, such as type 4, are noticeably wider than smaller fractures, such as type 1 fractures.

- Type A and Type B fracturing are of significance. These fracture types are only present within the C.B.H populations. Due to the nature of the fracturing, it appears that these samples are more brittle, allowing for fracturing to occur in any direction.
- The SEM images provide high resolution, high magnification images of the enamel structure. The fracturing present cannot be ruled out as being a result of the preparation of the slide, which required a polished surface for the microprobe, as it is established that enamel does have a tendency to fracture along the cleavage planes that parallel the enamel rods (Avery, 1994). It is interesting to note, however, that there appears to be a higher incidence of fracturing having occurred within the unhealthy teeth, than with the healthy teeth. Although, the frequency of the fracturing seen in the SEM images was higher within the C.B.H teeth, suggesting C.B.H teeth are more brittle compared to the control.
- The microprobe data was slightly useful. There was no indication for the elements that were analysed being depleted/elevated within the study area compared to the control population, although there were interesting trends observed across the enamel structure.
- The geology of the Cape Breton Highlands (C.B.H's) is composed of a complex assemblage of units
- The Blair River complex is composed of a distinctive assemblage of basement rocks including felsic and mafic gneisses, monzodiorite, anorthosite, and syentite.
- The Aspy Terrane include interlayered mafic and felsic metavolcanic rocks and metasedimentary rock s of Ordovician to Silurian age. Both mafic, felsic and intermediate rocks within the Aspy Terrane are all subalkalic.



- The Bras d'Or Terrane includes low pressure, amphibolite-facies, with locally migmatic, and gneissic 'basement. Large areas of mainly greenschists-facies quartzite, marble, and meta-greywacke, and a large volume of Late Precambrian subduction zone-related dioritic, tonalitic, granodioritic, and granitic plutons are also present.
- The Meguma Supergroup consists of thick metamorphosed siliclastic sequences, and is subdivided into two groups: Goldenville Formation and the overlying Halifax Formation, also referred to as the Goldenville-Halifax Transition (GHT) zone (Graves and Zentilli, 1988)
- The Halifax Formation are preferentially enriched in Mn, total C, Ba, Pb, Zn, Cu, Mo, W and Au (Graves and Zentilli, 1988)
- The area is also known for the East Kemptville cassiterite deposit, which was once North America's only producing primary tin mine
- Barium and Strontium are usually to be found in highest concentrations within Alkalic rocks such as monzonites and syenites, in concentrations upwards of 1600ppm. The Blair river Complex and Aspy Terrane are both alkalic-subalkalic in composition (Barr et al. 1987; Barr and Jamieson, 1991), and these two units form the majority of land area in the C.B.H
- Areas in southwest Nova Scotia are considered to be enriched in Barium, amongst others (Graves and Zentilli, 1988), and due to a close geochemical relationship with Sr, it would be expected that Sr would also be enriched. However, due to the unpredictable nature of Sr and the high variations associated with this element, further investigation is required.
- Due to the locality of the East Kemptville mine that occurs in close proximity with the control area, and also knowing that tin is relatively rare other than being abundant in cassiterite, there is a strong correlation between the tin concentrations within moose teeth enamel and the regional geology of the geographic locality of moose origin.

- Based on these results for tin, it is possible to determine the geographic origin of a moose.

## **7.2 Recommendations for Future Work**

- This thesis is based on the assumption of the control area being a 'healthy' population. Due to there being little/no data in the literature regarding biological values for trace elements for moose, further investigation is required, with larger sampling criteria and also the use of more control populations to verify results. It is recommended that moose teeth samples from Newfoundland be included in any further work, due to the similarities of the regional geology between the C.B.H and areas of Newfoundland, where there is no documentation of similar problems with incisiform breakage within Newfoundland moose.
- Complete studies on moose browse specific to C.B.H. would be required. Moose will sample new plant species when they encounter them, so diets will vary geographically. Biogeochemical analysis of moose forage may further enhance understanding of the problem.
- Preliminary results of moose antler geochemistry suggest higher Ca/P ratios within C.B.H antlers. Further investigation is required, with a larger sample size.
- Results for Ba and Sr are unclear, and require further investigation. Sr in animal tissue is a reflection of the concentrations found within drinking water supplies, therefore investigation of weathering rates of elements may be required, as this may be a contributing factor to the depleted levels of certain elements within the moose of the C.B.H
- Enamel lamellae consist of organic material with little mineral content. It has been suggested that enamel lamellae may be a site of weakness in a tooth. It should be investigated whether lamellae occur in higher frequency within the C.B.H moose teeth than control areas.

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**Appendix A: ICP-MS Results for tooth enamel**



**CERTIFICATE OF ANALYSIS**

GL JOB#: 03-0375  
 CLIENT: Zentilli  
 DATE:  
 Method: Custom Analysis

**Certified Reference Materials**

Lab ID	Element Client ID	Sample Mass mg	Li ppb	Be ppb	Mg ppb	Al ppb	Si ppb	Ca ppb	Sc ppb	Ti ppb	V ppb	Cr ppb	Mn ppb	Fe ppb	Co ppb	Ni ppb	Cu ppb
<b>Matrix Matched NRC Water</b>																	
SLRS-4	Measured Values		0.602	0.0648	1,770	63.1	2,120	71,400	1.00	1.87	0.458	0.313	3.54	102	0.0270	N.D.	1.82
	Reference Values		0.582	0.0520	1,740	60.6	2,050	68,900	1.04	1.92	0.400	0.257	3.48	105	0.0174	N.D.	1.75
(+80ppm Ca & 0.2% HCl)			0.54 ± 0.14	0.007 ± 0.002	1600 ± 100	54 ± 4	1,864 ± 96	68,200		1.48 ± 0.16	0.32 ± 0.03	0.33 ± 0.02	3.37 ± 0.18	103 ± 5	0.033 ± 0.006	0.67 ± 0.08	1.81 ± 0.08
Lab ID	Element Client ID	Sample Mass mg	Li ppm	Be ppm	Mg ppm	Al ppm	Si ppm	Ca wt%	Sc ppm	Ti ppm	V ppm	Cr ppm	Mn ppm	Fe ppm	Co ppm	Ni ppm	Cu ppm
<b>Apatite</b>																	
CTA-AC-1	Measured Values	98	1.40	0.301	502	3,110	10,700	34.5	4.92	910	97.7	1.67	355	4,960	1.93	N.D.	40.8
	Reference Values				425	4100	5700 ± 1300	32.7 ± 3.1	0.244 ± 0.035	2927 ± 554	104 ± 10	1.7	317 ± 50	5000	2.72 ± 0.28	9	54 ± 4.5
<b>Phosphate Rock</b>																	
BCR-32	Measured Values	101	1.67	1.97	2,260	1,780	5,530	34.2	11.1	187	128	217	15.1	1,460	0.369	26.5	29.3
	Reference Values				2412 ± 60	2911 ± 32	9770 ± 560	36.9 ± 0.2		171 ± 10	153 ± 7	257 ± 16	18.8 ± 1.3	1608 ± 70	0.59 ± 0.06	34.6 ± 1.9	33.7 ± 1.4

Bold: Recommended values  
 Italic: Information values

**Total Procedural Blanks**

Samples Lab ID	Element Client ID	Sample Mass mg	Li ppm	Be ppm	Mg ppm	Al ppm	Si ppm	Ca wt%	Sc ppm	Ti ppm	V ppm	Cr ppm	Mn ppm	Fe ppm	Co ppm	Ni ppm	Cu ppm
RBblank-1		100	0.0205	0.00400	10.4	4.94	272	0.0348	0.262	0.188	0.224	N.D.	0.0320	1.46	0.0120	0.0325	0.651
RBblank-2		100	0.0115	0.0400	4.87	7.61	277	0.0277	0.277	0.314	0.474	0.146	0.0860	5.07	0.0125	0.0460	4.90
RBblank-3		100	0.00550	0.0200	2.16	6.69	263	0.00601	0.266	0.191	0.641	0.0585	N.D.	2.45	0.00650	N.D.	0.424
Samples Lab ID	Element Client ID	Sample Mass mg	Li ppm	Be ppm	Mg ppm	Al ppm	Si ppm	Ca wt%	Sc ppm	Ti ppm	V ppm	Cr ppm	Mn ppm	Fe ppm	Co ppm	Ni ppm	Cu ppm
03-0375-01	M003	48	0.533	N.D.	4,440	22.7	552	35.8	1.74	153	0.631	0.129	35.1	121	N.D.	N.D.	48.5
03-0375-02	M010	98	0.653	0.00765	3,740	144	250	35.1	1.33	126	0.429	0.314	47.9	96.0	N.D.	N.D.	41.0
03-0375-03	M023	103	0.389	0.0257	2,770	19.3	144	31.4	1.24	101	0.347	0.159	31.7	126	N.D.	N.D.	21.3
03-0375-04	M038	95	0.401	0.0200	2,790	57.9	194	36.1	1.34	98.5	0.478	0.175	26.1	213	N.D.	N.D.	12.1
03-0375-05	M017	116	1.77	0.00819	2,780	20.1	93.0	30.4	1.12	76.0	0.268	N.D.	113	78.5	N.D.	N.D.	7.97
03-0375-06	M032	103	1.71	0.00243	3,020	13.0	108	29.6	1.11	72.7	0.167	N.D.	46.2	133	N.D.	N.D.	28.3
03-0375-07	M022	88	17.3	0.0210	2,830	31.6	135	36.3	1.36	87.7	0.236	N.D.	30.3	99.5	N.D.	N.D.	28.1
03-0375-08	M030	110	1.49	0.00318	3,960	119	119	35.4	1.34	88.8	0.243	0.0341	103	82.8	N.D.	N.D.	12.4
	Rep. Soln. Analysis	110	1.39	0.0123	3,970	122	72.2	35.2	1.38	70.0	0.209	0.00818	101	81.9	N.D.	N.D.	12.1
	M001	120	2.12	0.0142	3,400	32.3	130	35.1	1.30	77.5	0.187	N.D.	118	85.1	N.D.	N.D.	13.3
03-0375-10	M009	58	1.01	0.0161	2,640	24.8	128	34.4	1.39	74.0	0.982	1.40	39.3	1,600	N.D.	N.D.	19.8
03-0375-11	M012	129	0.488	0.00233	2,680	22.8	98.5	38.5	1.38	84.7	0.168	N.D.	29.3	41.7	N.D.	N.D.	9.09
03-0375-12	M005	122	1.59	0.0135	2,790	329	195	33.8	1.28	78.0	0.316	0.323	52.4	97.2	N.D.	N.D.	31.5
03-0375-13	M008	117	0.676	0.0145	3,200	18.3	113	35.9	1.32	76.6	0.230	N.D.	30.5	118	N.D.	N.D.	9.93
03-0375-14	M016	116	0.524	0.0142	3,290	17.7	62.2	35.2	1.21	84.0	0.309	N.D.	31.5	59.9	0.0513	N.D.	7.80
03-0375-15	M018	116	0.611	0.0147	2,650	52.5	141	36.8	1.41	79.1	0.432	0.185	52.6	152	N.D.	N.D.	19.9
03-0375-16	M008	139	0.306	0.0176	3,160	22.3	86.8	34.7	1.23	75.3	0.183	N.D.	39.6	54.8	N.D.	N.D.	4.92
03-0375-17	M020	95	0.234	0.0168	3,210	16.6	104	33.6	1.24	71.7	0.348	0.00737	12.3	50.2	N.D.	N.D.	15.1
03-0375-18	M011	83	1.66	0.0343	4,720	162	369	32.8	1.52	114	0.729	0.175	51.5	914	0.297	N.D.	16.8
03-0375-19	M041	139	0.485	0.0115	4,220	12.9	92.9	33.9	1.26	80.0	0.354	N.D.	14.4	44.9	N.D.	N.D.	5.42
03-0375-20	M027	82	0.930	0.0317	3,900	139	287	35.7	1.46	80.4	0.497	N.D.	111	162	N.D.	N.D.	8.44
03-0375-21	M037	106	3.16	0.0104	4,460	34.2	111	33.8	1.33	70.0	0.365	N.D.	58.4	47.3	N.D.	N.D.	5.51
03-0375-22	M042	105	0.858	0.0143	4,020	30.1	147	35.2	1.41	70.1	0.320	N.D.	168	111	N.D.	N.D.	9.31
03-0375-23	M051	44	1.81	N.D.	2,860	60.6	137	33.9	1.48	77.2	0.824	0.150	65.0	211	0.903	N.D.	26.0
03-0375-24	M052	91	0.466	0.0220	3,490	80.0	139	32.4	1.33	97.4	0.481	0.107	54.0	361	1.05	N.D.	16.2
03-0375-25	M053	87	1.17	0.0448	2,660	59.9	144	34.5	1.39	70.9	0.555	0.103	38.7	205	N.D.	N.D.	24.3
03-0375-26	M055	81	0.902	0.0426	2,960	95.8	179	35.3	1.37	92.2	0.654	0.370	19.6	639	0.882	N.D.	28.7
03-0375-27	M056	117	0.981	0.0269	2,590	73.1	148	37.2	1.46	97.9	0.353	0.0957	14.7	176	1.51	N.D.	18.1

Duplicate Analyses Samples Lab ID	Element Client ID	Sample Mass mg	Li ppm	Be ppm	Mg ppm	Al ppm	Si ppm	Ca wt%	Sc ppm	Ti ppm	V ppm	Cr ppm	Mn ppm	Fe ppm	Co ppm	Ni ppm	Cu ppm
03-0375-20	M017	135	1.96	0.0133	3,000	23.6	81.2	31.1	1.20	61.7	0.151	0.261	117	105	N.D.	N.D.	2.90
	Absolute Difference		0.19	0.0051	220	3.5	-11.8	0.7	0.08	-14.3	-0.117	0.261	4	27			-5.07
	% Difference		11%	62%	8%	17%	-13%	2%	7%	-19%	-44%	100%	4%	34%			-64%
03-0375-210	M037	87	2.82	0.0299	4,130	41.9	105	31.9	1.36	56.7	0.310	1.04	58.6	178	N.D.	N.D.	7.14
	Absolute Difference		-0.34	0.0195	-330	7.7	-6	-1.9	0.03	-13.3	-0.055	1.04	0.2	131			1.63
	% Difference		-11%	188%	-7%	23%	-5%	-6%	2%	-19%	-15%	100%	0%	277%			30%

A-1



## CERTIFICATE OF ANALYSIS

GL JOB#: 03-0375  
 CLIENT: Zentilli  
 DATE:  
 Method: Custom Analysis

## Certified Reference Materials

Lab ID	Element Client ID	Zn ppb	Ga ppb	As ppb	Rb ppb	Sr ppb	Y ppb	Zr ppb	Nb ppb	Mo ppb	Ag ppb	Cd ppb	Sn ppb	Sb ppb	Cs ppb	Ba ppb	La ppb	Ce ppb
Matrix Matched NRC Water																		
SLRS-4	Measured Values	1.25	0.0219	2.95	1.62	31.5	0.151	0.110	0.00730	0.277	0.00520	0.0475	0.00870	0.276	0.00700	12.9	0.298	0.397
(+80ppm Ca & 0.2% HCl)	Reference Values	0.93 ± 0.1	0.0119 ± 0.0008	0.68 ± 0.06	1.53 ± 0.1	26.3 ± 3.2	0.146 ± 0.016	0.12 ± 0.03	0.21 ± 0.02	0.035 ± 0.01	0.012 ± 0.002	0.23 ± 0.04	0.009 ± 0.002	12.2 ± 0.6	0.287 ± 0.016	0.36 ± 0.024		
	Measured Values	20.8	23.5	9.32	10.9	16.500	295	10.5	7.28	1.30	0.0939	0.237	0.405	0.246	0.261	795	1.550	2.410
CTA-AC-1	Reference Values	38 ± 7.6				2000	272 ± 53	57								767 ± 79	2176 ± 94	3226 ± 175
Phosphate Rock																		
BCR-32	Measured Values	209	1.75	10.3	3.11	904	245	22.3	0.439	2.80	0.979	19.6	0.353	2.75	0.240	98.9	93.9	29.8
	Reference Values	253 ± 6		9.5 ± 0.5						24	2	20.8 ± 0.7		3				

Bold: Recommended values

#alic: Information values

## Total Procedural Blanks

Samples Lab ID	Element Client ID	Zn ppm	Ga ppm	As ppm	Rb ppm	Sr ppm	Y ppm	Zr ppm	Nb ppm	Mo ppm	Ag ppm	Cd ppm	Sn ppm	Sb ppm	Cs ppm	Ba ppm	La ppm	Ce ppm
RBlank-1		1.44	0.0355	3.28	0.0180	0.381	0.0105	0.00650	0.00800	0.0125	0.0100	0.0110	0.00650	0.0115	0.00700	0.147	0.00950	0.0110
RBlank-2		9.30	0.0320	3.09	0.0155	2.08	0.0555	0.00350	0.00350	0.0275	0.0100	0.0260	0.00900	0.0100	0.00300	0.450	0.168	0.247
RBlank-3		1.47	0.0385	3.38	0.0100	0.140	0.00550	0.113	0.00250	0.0220	0.00550	0.0120	N.D.	0.00700	0.00350	0.0555	0.00300	0.00450
Rep. Soln. Analysis																		
03-0375-01	M003	70.3	0.267	8.06	0.335	243	0.0438	0.0427	0.0135	0.0448	0.294	0.0665	0.0438	0.141	0.00417	122	0.0781	0.0438
03-0375-02	M010	74.4	0.229	5.08	0.403	268	0.0474	0.0755	0.0163	0.0500	0.0730	0.107	0.0974	0.0245	0.00408	121	0.0901	0.0296
03-0375-03	M023	43.7	0.220	3.87	0.287	239	0.0417	0.0364	0.0102	0.0306	0.114	0.165	0.0286	0.0126	0.00194	128	0.0689	0.0199
03-0375-04	M038	47.8	0.226	5.01	0.554	226	0.0495	3.35	0.0137	0.106	0.0426	0.0284	0.0374	0.0353	0.00316	82.1	0.114	0.120
03-0375-05	M017	48.5	0.166	3.83	0.688	255	0.0435	0.0897	0.00905	0.0237	0.0142	0.0746	0.0573	0.0134	0.00216	161	0.0810	0.0246
03-0375-06	M032	47.2	0.199	4.22	0.917	249	0.0354	0.140	0.00971	0.0209	0.0257	0.243	0.0223	0.0117	0.00340	159	0.0738	0.0126
03-0375-07	M022	45.5	0.242	5.11	0.326	280	0.0483	0.0358	0.0222	0.0335	0.0722	0.240	0.0494	0.0273	0.00398	147	0.0756	0.0239
03-0375-08	M030	79.4	0.192	4.58	0.390	293	0.0541	1.31	0.0100	0.0245	0.0227	0.0714	0.0491	0.0227	0.00273	164	0.0905	0.0300
		77.9	0.160	4.42	0.420	285	0.0636	1.40	0.0127	0.0236	0.0295	0.0695	0.0500	0.0177	0.00318	163	0.0964	0.0314
03-0375-09	M001	49.2	0.220	4.14	0.725	246	0.0504	0.0363	0.0133	0.0613	0.0342	0.0683	0.0454	0.0813	0.00292	96.4	0.0638	0.0392
03-0375-10	M009	45.3	0.238	6.37	0.331	313	0.0455	1.68	0.104	0.102	0.104	0.0491	0.168	0.0241	0.00357	118	0.0661	0.0107
03-0375-11	M012	45.3	0.234	4.56	0.480	338	0.0725	0.0570	0.0132	0.0279	0.0159	0.640	0.0736	0.0120	0.00310	120	0.0705	0.0446
03-0375-12	M005	68.0	0.204	3.97	0.682	193	0.0516	0.0730	0.0193	0.0459	0.0265	0.100	0.144	0.0123	0.00574	118	0.0881	0.0549
03-0375-13	M006	49.9	0.204	4.55	0.608	360	0.0654	0.0346	0.0141	0.0154	0.0261	0.0538	0.171	0.00655	0.00256	176	0.128	0.0765
03-0375-14	M016	38.5	0.200	4.67	0.757	199	0.0302	1.28	0.0108	0.0276	0.0595	0.0388	0.0284	0.00398	0.00259	81.5	0.0397	0.0112
03-0375-15	M018	48.8	0.228	4.70	0.533	192	0.0414	0.0513	0.0177	0.242	0.0703	0.0612	0.199	0.0147	0.00474	72.6	0.0543	0.0384
03-0375-16	M008	36.0	0.181	4.12	0.738	285	0.0432	0.0327	0.0122	0.0284	0.0191	0.0396	0.0504	0.00935	0.00360	84.4	0.0486	0.0194
03-0375-17	M020	42.7	0.185	4.91	0.872	278	0.0442	0.0289	0.0147	0.0300	0.0200	0.0579	0.109	0.00684	0.00263	79.8	0.0432	0.0179
03-0375-18	M011	55.1	0.196	5.18	1.25	224	0.190	0.195	0.0349	0.0554	0.0524	6.90	0.322	0.0422	0.0187	110	0.139	0.143
03-0375-19	M041	37.9	0.144	4.02	0.585	278	0.0471	0.640	0.0140	0.0270	0.0144	0.0342	0.0295	0.0151	0.00396	115	0.0680	0.0212
03-0375-20	M027	85.9	0.201	5.53	0.908	290	0.0774	0.0274	0.0201	0.0287	0.0238	0.0329	0.0207	0.0213	0.0171	151	0.153	0.160
03-0375-21	M037	49.2	0.145	4.47	0.570	236	0.0462	0.0269	0.0127	0.0208	0.0108	0.0646	0.0108	0.0104	0.00425	114	0.101	0.0769
03-0375-22	M042	72.5	0.158	4.43	0.809	415	0.153	0.0295	0.0190	0.108	0.0143	0.121	0.0452	0.0100	0.00381	235	0.687	0.941
03-0375-23	M051	80.3	0.172	7.37	0.689	490	0.169	0.0273	0.0216	0.0659	0.0739	0.201	0.151	0.0477	0.00568	171	0.660	0.916
03-0375-24	M052	60.6	0.162	4.31	0.521	267	0.123	0.0374	0.0214	0.0214	0.0407	0.430	0.336	0.0192	0.00604	295	0.345	0.326
03-0375-25	M053	62.1	0.144	5.29	0.926	474	0.0966	3.27	0.0149	0.0557	0.0494	0.292	0.0287	0.00747	0.00920	172	0.176	0.180
03-0375-26	M055	53.6	0.177	5.37	0.891	546	0.164	0.0778	0.0228	0.0759	0.0889	0.717	0.449	0.0185	0.00988	200	0.236	0.256
03-0375-27	M056	62.2	0.173	4.71	0.891	564	0.101	0.0842	0.0214	0.0470	0.0380	0.241	0.744	0.0299	0.00684	179	0.121	0.0816

## Duplicate Analyses

Samples Lab ID	Element Client ID	Zn ppm	Ga ppm	As ppm	Rb ppm	Sr ppm	Y ppm	Zr ppm	Nb ppm	Mo ppm	Ag ppm	Cd ppm	Sn ppm	Sb ppm	Cs ppm	Ba ppm	La ppm	Ce ppm
03-0375-5D	M017	44.1	0.122	3.81	0.721	268	0.0452	2.21	0.00826	0.0993	0.0170	0.0778	0.0437	0.00815	0.00333	167	0.0915	0.0285
	Absolute Difference	-4.4	-0.044	-0.02	0.033	13	0.0017	2.12	0.00021	0.0756	0.0028	0.0032	-0.0136	-0.00525	0.00117	6	0.0105	0.0039
	% Difference	-9%	-27%	-1%	5%	5%	4%	2363%	2%	31%	20%	4%	-24%	-3%	54%	4%	13%	16%
03-0375-21D	M037	63.3	0.176	5.88	0.593	228	0.0440	0.0769	0.0157	0.0634	0.0224	0.149	0.0806	0.0478	0.00448	109	0.0716	0.0358
	Absolute Difference	14.1	0.031	1.41	0.023	-8	-0.0022	0.0500	0.0030	0.0426	0.0114	0.084	0.0698	0.0374	0.00023	-5	-0.0294	-0.0411
	% Difference	23%	21%	32%	4%	-3%	-5%	186%	24%	205%	107%	130%	646%	360%	5%	-4%	-29%	-53%



CERTIFICATE OF ANALYSIS

GL JOB#: 03-0375
CLIENT: Zentilli
DATE:
Method: Custom Analysis

Certified Reference Materials

Table with 16 columns (Element Client ID, Pr, Nd, Sm, Eu, Tb, Gd, Dy, Ho, Er, Tm, Yb, Lu, Hf, Ta) and 3 rows for Matrix Matched NRC Water (BLRS-4) showing Measured and Reference values.

Table with 16 columns (Element Client ID, Pr, Nd, Sm, Eu, Tb, Gd, Dy, Ho, Er, Tm, Yb, Lu, Hf, Ta) and 2 rows for Apatite (CTA-AC-1) showing Measured and Reference values.

Table with 16 columns (Element Client ID, Pr, Nd, Sm, Eu, Tb, Gd, Dy, Ho, Er, Tm, Yb, Lu, Hf, Ta) and 2 rows for Phosphate Rock (BCR-32) showing Measured and Reference values.

bold: Recommended values
italic: Information values

Total Procedural Blanks

Table with 16 columns (Element Client ID, Pr, Nd, Sm, Eu, Tb, Gd, Dy, Ho, Er, Tm, Yb, Lu, Hf, Ta) and 3 rows for RBlank-1, RBlank-2, RBlank-3.

Large table with 16 columns (Element Client ID, Pr, Nd, Sm, Eu, Tb, Gd, Dy, Ho, Er, Tm, Yb, Lu, Hf, Ta) and 27 rows for Rep. Soln. Analysis (MO01-MO56).

Duplicate Analyses

Table with 16 columns (Element Client ID, Pr, Nd, Sm, Eu, Tb, Gd, Dy, Ho, Er, Tm, Yb, Lu, Hf, Ta) and 5 rows for duplicate analyses (MO17, MO37).



## CERTIFICATE OF ANALYSIS

GL JOB#: 03-0375

CLIENT: Zentilli

DATE:

Method: Custom Analysis

## Certified Reference Materials

Lab ID	Element Client ID	W ppb	Tl ppb	Pb ppb	Bi ppb	Th ppb	U ppb
<b>Matrix Matched NRC Water</b>							
SLRS-4	Measured Values	0.0148	0.00690	0.103	0.00600	0.0158	0.0482
(+60ppm Ca & 0.2% HCl)	Reference Values	0.0150	0.00830	0.107	0.00590	0.0163	0.0489
	Reference Values	0.0133 ± 0.0020 0.0076 ± 0.0012		0.088 ± 0.007	0.018 ± 0.008		0.050 ± 0.003

Lab ID	Element Client ID	W ppm	Tl ppm	Pb ppm	Bi ppm	Th ppm	U ppm
<b>Apatite</b>							
CTA-AC-1	Measured Values	0.470	0.00867	3.66	0.00765	21.1	3.86
	Reference Values					21.8 ± 2.1	4.4 ± 0.9

Lab ID	Element Client ID	W ppm	Tl ppm	Pb ppm	Bi ppm	Th ppm	U ppm
<b>Phosphate Rock</b>							
BCR-32	Measured Values	0.300	0.0960	4.05	0.0173	2.15	112
	Reference Values			5		2	125

Bold: Recommended values

Italic: Information values

## Total Procedural Blanks

Samples Lab ID	Element Client ID	W ppm	Tl ppm	Pb ppm	Bi ppm	Th ppm	U ppm
RBlank-1		0.0170	0.0105	0.0780	0.0150	0.00500	0.00900
RBlank-2		0.0115	0.00300	2.23	0.00650	0.00450	0.0200
RBlank-3		0.00650	0.00400	0.447	0.00650	0.00150	0.00350

Samples Lab ID	Element Client ID	W ppm	Tl ppm	Pb ppm	Bi ppm	Th ppm	U ppm
03-0375-01	M003	16.9	0.00729	0.958	0.0188	0.00417	0.00208
03-0375-02	M010	20.8	0.00510	0.824	0.0173	0.00255	0.00306
03-0375-03	M023	4.43	0.00534	0.479	0.0150	0.000485	0.00243
03-0375-04	M038	2.47	0.00526	0.665	0.0132	0.00211	0.00263
03-0375-05	M017	2.76	0.0138	0.789	0.0181	0.000862	0.00345
03-0375-06	M032	23.3	0.0136	0.411	0.0316	0.000971	0.000971
03-0375-07	M022	1.77	0.00968	0.677	0.0142	0.00284	0.00227
03-0375-08	M030	2.39	0.00636	0.775	0.0123	0.000909	0.00182
	Rep. Soln. Analysis	2.27	0.00545	0.737	0.0109	0.00136	0.00682
03-0375-09	M001	2.81	0.00542	0.673	0.0150	0.00167	0.00250
03-0375-10	M009	19.1	0.00714	0.774	0.00714	0.000893	0.00179
03-0375-11	M012	3.03	0.00581	0.806	0.0295	0.00116	0.000775
03-0375-12	M005	1.53	0.00533	1.15	0.0115	0.00656	0.00533
03-0375-13	M006	3.04	0.00470	0.608	0.00684	0.00128	0.00171
03-0375-14	M016	0.988	0.00819	0.534	0.00517	0.000862	0.000862
03-0375-15	M018	2.18	0.00603	1.33	0.00819	0.00216	0.00259
03-0375-16	M008	1.74	0.00504	0.519	0.00324	0.000719	0.00108
03-0375-17	M020	39.3	0.00526	0.509	0.0126	0.00105	0.00158
03-0375-18	M011	2.92	0.0151	1.88	0.0584	0.0151	0.0151
03-0375-19	M041	18.0	0.00647	0.330	0.00504	0.00180	0.00324
03-0375-20	M027	2.61	0.0116	0.980	0.00915	0.0104	0.0110
03-0375-21	M037	1.61	0.00708	0.849	0.00849	0.00189	0.00189
03-0375-22	M042	12.2	0.0114	0.702	0.0105	0.0176	0.00333
03-0375-23	M051	8.39	0.00682	3.30	0.0205	0.0182	0.00568
03-0375-24	M052	1.80	0.00879	3.01	0.0242	0.00989	0.00440
03-0375-25	M053	1.88	0.0109	1.06	0.0178	0.00517	0.00460
03-0375-26	M055	3.37	0.00617	1.96	0.0407	0.00998	0.00802
03-0375-27	M056	3.01	0.00726	1.42	0.0171	0.00427	0.00385

## Duplicate Analyses

Samples Lab ID	Element Client ID	W ppm	Tl ppm	Pb ppm	Bi ppm	Th ppm	U ppm
03-0375-5D	M017	2.64	0.0122	0.591	0.0137	0.00111	0.00333
	Absolute Difference	-0.12	-0.0016	-0.198	-0.0044	0.00025	-0.00012
	% Difference	-4%	-12%	-26%	-24%	23%	-3%
03-0375-21D	M037	1.56	0.00821	2.07	0.0149	0.00239	0.00224
	Absolute Difference	-0.05	0.00113	1.22	0.0064	0.00110	0.00035
	% Difference	-3%	16%	144%	75%	58%	19%

Appendix B: one-way ANOVA of variance results: Control  
(Including Parrsboro), North of the Park,  
South of the Park



### One-way Analysis of Variance

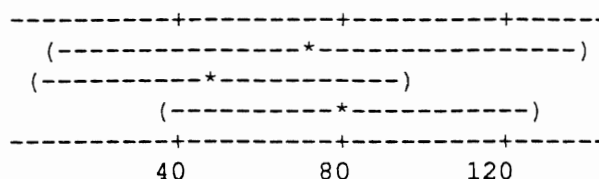
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	5549	2775	0.54	0.590
Error	22	112759	5125		
Total	24	118308			

Level	N	Mean	StDev
Al (H)	5	73.88	14.92
Al (N)	10	49.10	45.32
Al (S)	10	81.52	101.86

Pooled StDev = 71.59

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

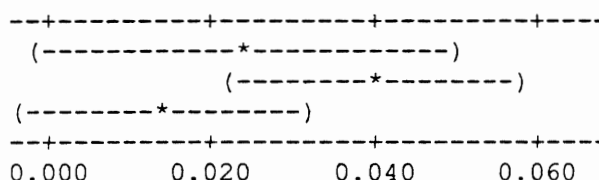
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.003070	0.001535	1.97	0.163
Error	22	0.017132	0.000779		
Total	24	0.020202			

Level	N	Mean	StDev
Sb (H)	5	0.02455	0.01518
Sb (N)	10	0.03933	0.04105
Sb (S)	10	0.01465	0.01077

Pooled StDev = 0.02791

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

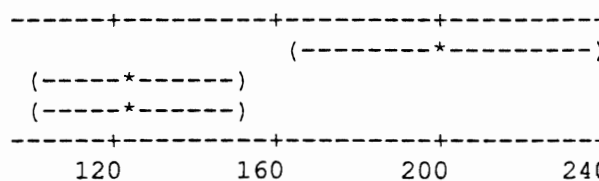
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	22932	11466	6.71	0.005
Error	22	37618	1710		
Total	24	60550			

Level	N	Mean	StDev
Ba (H)	5	201.40	48.17
Ba (N)	10	125.85	25.80
Ba (S)	10	125.52	49.83

Pooled StDev = 41.35

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1.73	0.86	0.45	0.645
Error	22	42.56	1.93		
Total	24	44.29			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Cd (H)	5	0.376	0.209
Cd (N)	10	0.161	0.180
Cd (S)	10	0.746	2.163

Pooled StDev = 1.391

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0312	0.0156	1.24	0.310
Error	22	0.2777	0.0126		
Total	24	0.3090			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Cr (H)	5	0.1651	0.1165
Cr (N)	10	0.0951	0.1062
Cr (S)	10	0.0687	0.1164

Pooled StDev = 0.1124

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	601	300	2.92	0.075
Error	22	2260	103		
Total	24	2861			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Cu (H)	5	22.66	5.31
Cu (N)	10	22.37	13.13
Cu (S)	10	12.46	8.13

Pooled StDev = 10.13

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	74518	37259	0.29	0.748
Error	22	2792078	126913		
Total	24	2866596			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Fe (H)	5	318.4	193.2
Fe (N)	10	257.1	473.9
Fe (S)	10	175.9	262.8

Pooled StDev = 356.2

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	7.368	3.684	13.67	0.000
Error	22	5.928	0.269		
Total	24	13.296			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Pb (H)	5	2.1500	0.9771
Pb (N)	10	0.7231	0.1334
Pb (S)	10	0.8876	0.4653

Pooled StDev = 0.5191

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	14.2	7.1	0.62	0.545
Error	22	250.7	11.4		
Total	24	264.9			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Li (H)	5	1.066	0.490
Li (N)	10	2.612	5.197
Li (S)	10	1.076	0.863

Pooled StDev = 3.376

## One-way Analysis of Variance

B-4

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1515	758	0.53	0.596
Error	22	31445	1429		
Total	24	32961			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Mn (H)	5	38.40	21.60
Mn (N)	10	54.03	33.68
Mn (S)	10	59.63	46.39

Pooled StDev = 37.81

## One-way Analysis of Variance

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.00049	0.00024	0.10	0.905
Error	22	0.05352	0.00243		
Total	24	0.05401			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Mo (H)	5	0.05318	0.02081
Mo (N)	10	0.05029	0.03082
Mo (S)	10	0.06000	0.06931

Pooled StDev = 0.04932

## One-way Analysis of Variance

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000124	0.000062	0.17	0.843
Error	22	0.007952	0.000361		
Total	24	0.008077			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Nb (H)	5	0.02042	0.00314
Nb (N)	10	0.02258	0.02885
Nb (S)	10	0.01761	0.00686

Pooled StDev = 0.01901

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.5346	0.2673	7.19	0.004
Error	22	0.8175	0.0372		
Total	24	1.3521			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Rb (H)	5	0.7816	0.1726
Rb (N)	10	0.4634	0.1780
Rb (S)	10	0.7507	0.2142

Pooled StDev = 0.1928

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0046	0.0023	0.11	0.900
Error	22	0.4771	0.0217		
Total	24	0.4817			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ag (H)	5	0.0582	0.0223
Ag (N)	10	0.0793	0.0831
Ag (S)	10	0.0950	0.2142

Pooled StDev = 0.1473

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	159844	79922	14.93	0.000
Error	22	117800	5355		
Total	24	277644			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Sr (H)	5	468.20	118.55
Sr (N)	10	269.80	35.95
Sr (S)	10	266.85	74.50

Pooled StDev = 73.17

## One-way Analysis of Variance

B-6

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1286	643	1.82	0.185
Error	22	7755	353		
Total	24	9041			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ti (H)	5	87.12	12.34
Ti (N)	10	96.55	25.21
Ti (S)	10	80.57	12.59

Pooled StDev = 18.78

## One-way Analysis of Variance

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.1398	0.0699	1.71	0.204
Error	22	0.8994	0.0409		
Total	24	1.0391			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
V (H)	5	0.5734	0.1780
V (N)	10	0.3918	0.2550
V (S)	10	0.3818	0.1443

Pooled StDev = 0.2022

## One-way Analysis of Variance

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	328	164	0.81	0.459
Error	22	4466	203		
Total	24	4794			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Zn (H)	5	63.76	9.90
Zn (N)	10	54.92	13.90
Zn (S)	10	54.51	16.11

Pooled StDev = 14.25

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000827	0.000414	3.41	0.051
Error	22	0.002669	0.000121		
Total	24	0.003497			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Be (H))	5	0.02726	0.01812
Be (N)	10	0.01155	0.00901
Be (S)	10	0.01763	0.00833

Pooled StDev = 0.01102

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1988252	994126	2.51	0.105
Error	22	8727532	396706		
Total	24	10715784			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Mg (H)	5	2912.0	355.8
Mg (N)	10	3215.0	633.2
Mg (S)	10	3643.5	715.9

Pooled StDev = 629.8

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	5091	2545	0.22	0.804
Error	22	253995	11545		
Total	24	259086			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Si (H)	5	149.4	17.1
Si (N)	10	185.1	136.8
Si (S)	10	161.4	96.8

Pooled StDev = 107.4

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.08	0.04	0.01	0.989
Error	22	80.02	3.64		
Total	24	80.10			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ca (H)	5	34.660	1.773
Ca (N)	10	34.810	2.448
Ca (S)	10	34.725	1.225

Pooled StDev = 1.907

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0137	0.0069	0.45	0.643
Error	22	0.3356	0.0153		
Total	24	0.3494			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Se (H)	5	1.4060	0.0627
Se (N)	10	1.3535	0.1581
Se (S)	10	1.3435	0.1027

Pooled StDev = 0.1235

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	2.9069	1.4534	25.00	0.000
Error	22	1.2788	0.0581		
Total	24	4.1857			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Co (H)	5	0.8690	0.5478
Co (N)	10	0.0000	0.0000
Co (S)	10	0.0348	0.0935

Pooled StDev = 0.2411

\* NOTE \* All values in column are identical.



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.012655	0.006328	9.15	0.001
Error	22	0.015221	0.000692		
Total	24	0.027876			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ga (H)	5	0.16760	0.01511
Ga (N)	10	0.22505	0.02419
Ga (S)	10	0.18930	0.03170

Pooled StDev = 0.02630

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	2.43	1.21	1.21	0.318
Error	22	22.09	1.00		
Total	24	24.52			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
As (H)	5	5.410	1.179
As (N)	10	5.080	1.271
As (S)	10	4.604	0.472

Pooled StDev = 1.002

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.02214	0.01107	7.82	0.003
Error	22	0.03112	0.00141		
Total	24	0.05326			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Y (H)	5	0.13072	0.03421
Y (N)	10	0.04927	0.00923
Y (S)	10	0.07460	0.05341

Pooled StDev = 0.03761

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1.182	0.591	0.62	0.548
Error	22	21.039	0.956		
Total	24	22.221			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Zr (H)	5	0.6993	1.4373
Zr (N)	10	0.6739	1.1179
Zr (S)	10	0.2389	0.4120

Pooled StDev = 0.9779

0.00      0.60      1.20

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.2742	0.1371	7.23	0.004
Error	22	0.4170	0.0190		
Total	24	0.6912			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Sn (H)	5	0.3417	0.2774
Sn (N)	10	0.0633	0.0418
Sn (S)	10	0.1050	0.1020

Pooled StDev = 0.1377

0.00      0.15      0.30      0.45

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000835	0.0000418	2.66	0.092
Error	22	0.0003452	0.0000157		
Total	24	0.0004287			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Cs (H)	5	0.007528	0.001899
Cs (N)	10	0.003273	0.000753
Cs (S)	10	0.006656	0.006016

Pooled StDev = 0.003961

0.0030      0.0060      0.0090

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.1751	0.0875	3.71	0.041
Error	22	0.5189	0.0236		
Total	24	0.6940			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
La (H)	5	0.3076	0.2138
La (N)	10	0.0785	0.0147
La (S)	10	0.1504	0.1927

Pooled StDev = 0.1536

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.3294	0.1647	3.15	0.063
Error	22	1.1503	0.0523		
Total	24	1.4797			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ce (H)	5	0.3519	0.3281
Ce (N)	10	0.0379	0.0309
Ce (S)	10	0.1542	0.2811

Pooled StDev = 0.2287

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.002997	0.001498	3.14	0.063
Error	22	0.010505	0.000477		
Total	24	0.013501			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Pr (H)	5	0.03452	0.03049
Pr (N)	10	0.00465	0.00311
Pr (S)	10	0.01655	0.02728

Pooled StDev = 0.02185

### One-way Analysis of Variance

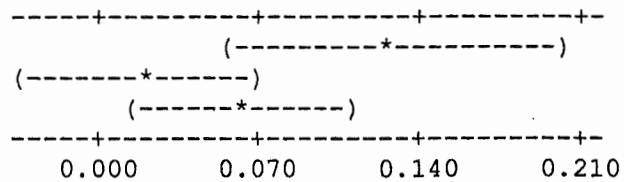
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.04084	0.02042	3.32	0.055
Error	22	0.13541	0.00616		
Total	24	0.17625			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Nd (H)	5	0.12904	0.10710
Nd (N)	10	0.01870	0.01383
Nd (S)	10	0.06200	0.09877

Pooled StDev = 0.07845



### One-way Analysis of Variance

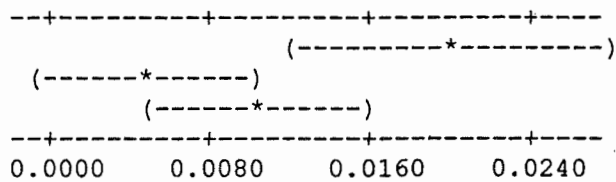
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0007887	0.0003944	5.26	0.014
Error	22	0.0016498	0.0000750		
Total	24	0.0024385			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Sm (H)	5	0.020052	0.009042
Sm (N)	10	0.004690	0.002503
Sm (S)	10	0.010404	0.011862

Pooled StDev = 0.008660



### One-way Analysis of Variance

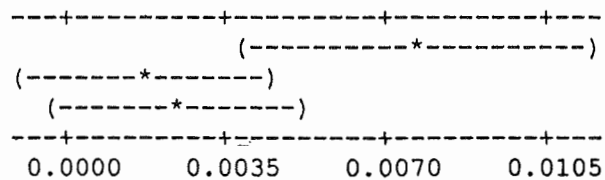
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0001215	0.0000607	3.46	0.049
Error	22	0.0003860	0.0000175		
Total	24	0.0005075			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Eu (H)	5	0.007594	0.008028
Eu (N)	10	0.001854	0.002248
Eu (S)	10	0.002374	0.003033

Pooled StDev = 0.004189



### One-way Analysis of Variance

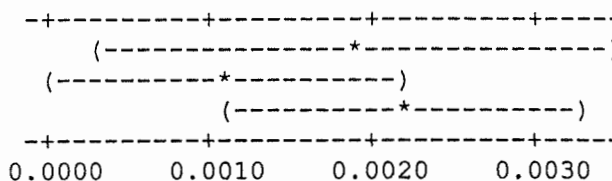
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000063	0.0000031	1.04	0.369
Error	22	0.0000660	0.0000030		
Total	24	0.0000723			

Level	N	Mean	StDev
Tb (H)	5	0.001893	0.001003
Tb (N)	10	0.001100	0.000691
Tb (S)	10	0.002198	0.002532

Pooled StDev = 0.001732

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

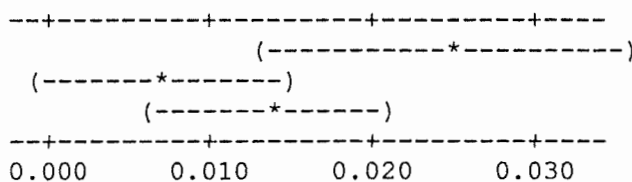
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.001074	0.000537	3.71	0.041
Error	22	0.003181	0.000145		
Total	24	0.004255			

Level	N	Mean	StDev
Gd (H)	5	0.02461	0.01246
Gd (N)	10	0.00670	0.00334
Gd (S)	10	0.01353	0.01653

Pooled StDev = 0.01202

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

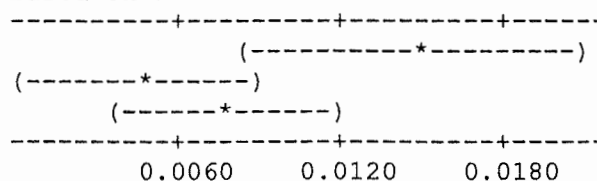
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0003446	0.0001723	3.78	0.039
Error	22	0.0010024	0.0000456		
Total	24	0.0013471			

Level	N	Mean	StDev
Dy (H)	5	0.014796	0.008002
Dy (N)	10	0.004633	0.002983
Dy (S)	10	0.007777	0.008604

Pooled StDev = 0.006750

Individual 95% CIs For Mean  
Based on Pooled StDev



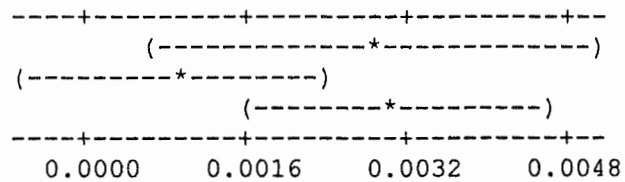
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000271	0.0000136	2.44	0.110
Error	22	0.0001222	0.0000056		
Total	24	0.0001494			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ho (H)	5	0.002904	0.001718
Ho (N)	10	0.000925	0.000620
Ho (S)	10	0.003116	0.003448



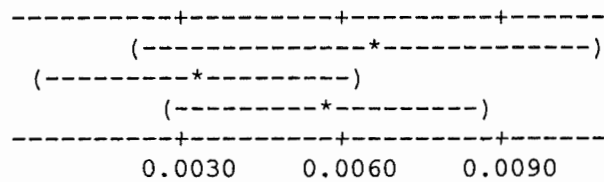
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000480	0.0000240	1.12	0.343
Error	22	0.0004701	0.0000214		
Total	24	0.0005181			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Er (H)	5	0.006454	0.003824
Er (N)	10	0.003228	0.001276
Er (S)	10	0.005795	0.006641



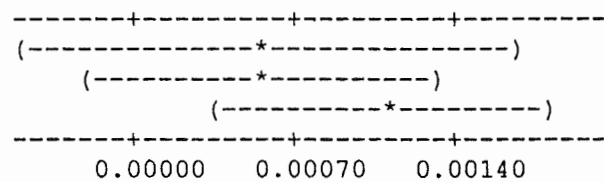
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000016	0.0000008	0.62	0.550
Error	22	0.0000289	0.0000013		
Total	24	0.0000305			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Tm (H)	5	0.000584	0.000594
Tm (N)	10	0.000564	0.000945
Tm (S)	10	0.001090	0.001471



### One-way Analysis of Variance

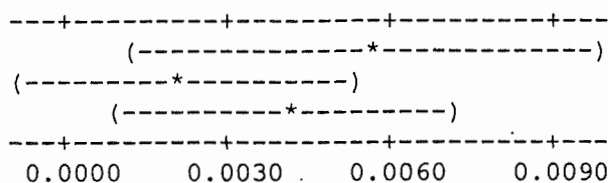
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000425	0.0000212	0.98	0.392
Error	22	0.0004777	0.0000217		
Total	24	0.0005202			

Level	N	Mean	StDev
Yb (H)	5	0.005616	0.002658
Yb (N)	10	0.002196	0.001596
Yb (S)	10	0.004102	0.006884

Pooled StDev = 0.004660

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

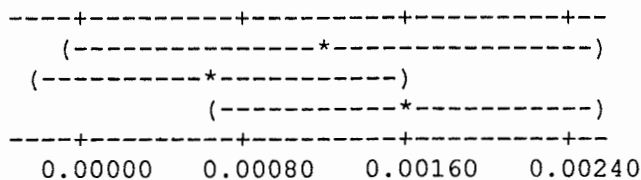
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000044	0.0000022	1.08	0.356
Error	22	0.0000443	0.0000020		
Total	24	0.0000487			

Level	N	Mean	StDev
Lu (H)	5	0.001206	0.000645
Lu (N)	10	0.000667	0.000595
Lu (S)	10	0.001599	0.002094

Pooled StDev = 0.001419

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

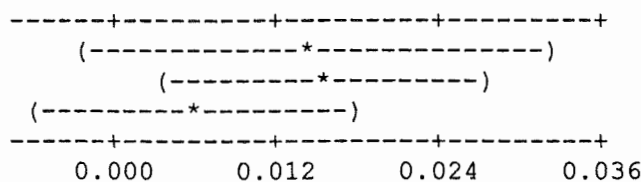
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000500	0.000250	0.72	0.498
Error	22	0.007643	0.000347		
Total	24	0.008143			

Level	N	Mean	StDev
Hf (H)	5	0.01459	0.02566
Hf (N)	10	0.01535	0.02230
Hf (S)	10	0.00599	0.00769

Pooled StDev = 0.01864

Individual 95% CIs For Mean  
Based on Pooled StDev



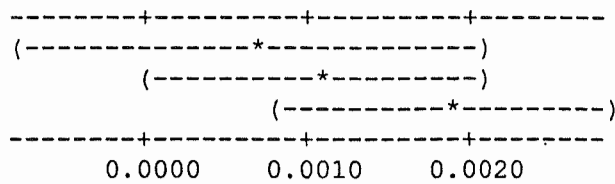
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000055	0.0000028	1.11	0.346
Error	22	0.0000544	0.0000025		
Total	24	0.0000599			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ta (H)	5	0.000691	0.000478
Ta (N)	10	0.001080	0.000959
Ta (S)	10	0.001863	0.002242



Pooled StDev = 0.001573

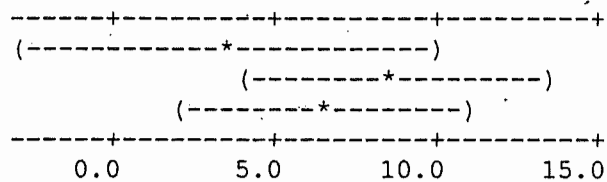
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	83.9	41.9	0.85	0.439
Error	22	1079.8	49.1		
Total	24	1163.7			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
W (H)	5	3.690	2.716
W (N)	10	8.670	7.832
W (S)	10	6.558	7.441



Pooled StDev = 7.006

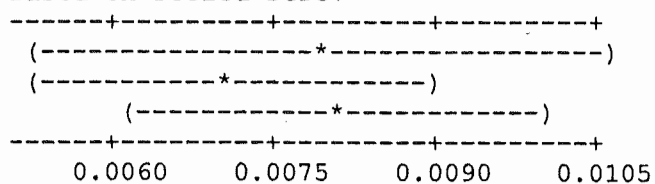
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000056	0.0000028	0.32	0.726
Error	22	0.0001881	0.0000085		
Total	24	0.0001936			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Tl (H)	5	0.007988	0.001892
Tl (N)	10	0.007108	0.002701
Tl (S)	10	0.008105	0.003465



Pooled StDev = 0.002924



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000399	0.000199	1.43	0.260
Error	22	0.003064	0.000139		
Total	24	0.003462			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Bi (H)	5	0.02406	0.00971
Bi (N)	10	0.01673	0.00641
Bi (S)	10	0.01312	0.01604

Pooled StDev = 0.01180

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0002119	0.0001060	4.71	0.020
Error	22	0.0004950	0.0000225		
Total	24	0.0007069			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Th (H)	5	0.009482	0.005525
Th (N)	10	0.001771	0.001150
Th (S)	10	0.005853	0.006333

Pooled StDev = 0.004743

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000454	0.0000227	2.34	0.120
Error	22	0.0002139	0.0000097		
Total	24	0.0002594			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
U (H)	5	0.005310	0.001654
U (N)	10	0.002157	0.000616
U (S)	10	0.004638	0.004709

Pooled StDev = 0.003118

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1.840	0.920	3.51	0.047
Error	22	5.760	0.262		
Total	24	7.600			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
REE Heal	5	0.9088	0.7179
REE Sout	10	0.4351	0.6379
REE Nort	10	0.1663	0.0632

Pooled StDev = 0.5117

Appendix C: one-way ANOVA of variance results: Control  
(Excluding Parrsboro), North of the Park,  
South of the Park

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	5421	2710	0.50	0.611
Error	21	112712	5367		
Total	23	118133			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Al (H)	4	72.35	16.77
Al (N)	10	49.10	45.32
Al (S)	10	81.52	101.86

Pooled StDev = 73.26

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.003050	0.001525	1.87	0.178
Error	21	0.017097	0.000814		
Total	23	0.020147			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Sb (H)	4	0.02589	0.01718
Sb (N)	10	0.03933	0.04105
Sb (S)	10	0.01465	0.01077

Pooled StDev = 0.02853

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	10016	5008	3.64	0.044
Error	21	28882	1375		
Total	23	38898			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ba (H)	4	180.50	13.48
Ba (N)	10	125.85	25.80
Ba (S)	10	125.52	49.83

Pooled StDev = 37.09

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.2513	0.1256	5.35	0.014
Error	20	0.4697	0.0235		
Total	22	0.7210			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Cd (H)	4	0.3628	0.2391
Cd (N)	10	0.1615	0.1798
Cd (S)	9	0.0617	0.0303

Pooled StDev = 0.1532

0.00 0.16 0.32 0.48

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0354	0.0177	1.36	0.279
Error	21	0.2735	0.0130		
Total	23	0.3089			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Cr (H)	4	0.1797	0.1291
Cr (N)	10	0.0951	0.1062
Cr (S)	10	0.0687	0.1164

Pooled StDev = 0.1141

0.00 0.10 0.20 0.30

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	648	324	3.08	0.067
Error	21	2208	105		
Total	23	2855			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Cu (H)	4	24.27	4.50
Cu (N)	10	22.37	13.13
Cu (S)	10	12.46	8.13

Pooled StDev = 10.25

8.0 16.0 24.0 32.0

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	110880	55440	1.41	0.267
Error	20	785994	39300		
Total	22	896874			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Fe (H)	4	307.8	221.4
Fe (N)	9	107.9	46.5
Fe (S)	10	175.9	262.8

Pooled StDev = 198.2

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	4.389	2.195	9.21	0.001
Error	21	5.004	0.238		
Total	23	9.393			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Pb (H)	4	1.9350	0.9823
Pb (N)	10	0.7231	0.1334
Pb (S)	10	0.8876	0.4653

Pooled StDev = 0.4881

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.156	0.078	0.15	0.863
Error	20	10.577	0.529		
Total	22	10.734			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Li (H)	4	1.2158	0.4118
Li (N)	9	0.9804	0.6492
Li (S)	10	1.0764	0.8626

Pooled StDev = 0.7272

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1819	909	0.61	0.551
Error	21	31141	1483		
Total	23	32960			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Mn (H)	4	34.50	22.82
Mn (N)	10	54.03	33.68
Mn (S)	10	59.63	46.39

Pooled StDev = 38.51

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.00059	0.00030	0.12	0.889
Error	21	0.05226	0.00249		
Total	23	0.05285			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Mo (H)	4	0.06112	0.01252
Mo (N)	10	0.05029	0.03082
Mo (S)	10	0.06000	0.06931

Pooled StDev = 0.04989

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000124	0.000062	0.16	0.850
Error	21	0.007951	0.000379		
Total	23	0.008075			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Nb (H)	4	0.02018	0.00357
Nb (N)	10	0.02258	0.02885
Nb (S)	10	0.01761	0.00686

Pooled StDev = 0.01946

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.6042	0.3021	8.66	0.002
Error	21	0.7326	0.0349		
Total	23	1.3368			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Rb (H)	4	0.8468	0.1069
Rb (N)	10	0.4634	0.1780
Rb (S)	10	0.7507	0.2142

Pooled StDev = 0.1868

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.01289	0.00645	1.95	0.168
Error	20	0.06607	0.00330		
Total	22	0.07897			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ag (H)	4	0.06255	0.02309
Ag (N)	10	0.07926	0.08308
Ag (S)	9	0.02749	0.01713

Pooled StDev = 0.05748

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	208669	104334	32.61	0.000
Error	21	67199	3200		
Total	23	275867			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Sr (H)	4	518.50	43.28
Sr (N)	10	269.80	35.95
Sr (S)	10	266.85	74.50

Pooled StDev = 56.57



**One-way Analysis of Variance**

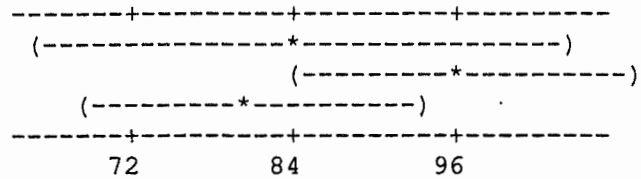
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1331	666	1.83	0.185
Error	21	7623	363		
Total	23	8954			

Level	N	Mean	StDev
Ti (H)	4	84.55	12.61
Ti (N)	10	96.55	25.21
Ti (S)	10	80.57	12.59

Pooled StDev = 19.05

Individual 95% CIs For Mean  
Based on Pooled StDev



**One-way Analysis of Variance**

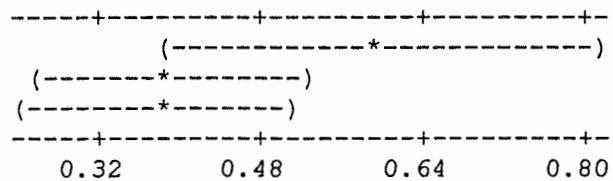
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.1471	0.0735	1.74	0.200
Error	21	0.8887	0.0423		
Total	23	1.0358			

Level	N	Mean	StDev
V (H)	4	0.5965	0.1967
V (N)	10	0.3918	0.2550
V (S)	10	0.3818	0.1443

Pooled StDev = 0.2057

Individual 95% CIs For Mean  
Based on Pooled StDev



**One-way Analysis of Variance**

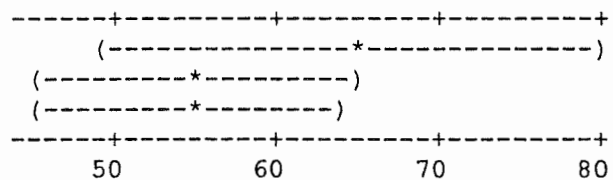
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	323	161	0.76	0.480
Error	21	4454	212		
Total	23	4777			

Level	N	Mean	StDev
Zn (H)	4	64.55	11.25
Zn (N)	10	54.92	13.90
Zn (S)	10	54.51	16.11

Pooled StDev = 14.56

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000837	0.000419	3.34	0.055
Error	21	0.002635	0.000125		
Total	23	0.003472			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Be (H)	4	0.02858	0.02065
Be (N)	10	0.01155	0.00901
Be (S)	10	0.01763	0.00833

Pooled StDev = 0.01120

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	2377771	1188886	3.00	0.071
Error	21	8309927	395711		
Total	23	10687699			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Mg (H)	4	2767.5	171.9
Mg (N)	10	3215.0	633.2
Mg (S)	10	3643.5	715.9

Pooled StDev = 629.1

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	4322	2161	0.18	0.838
Error	21	253860	12089		
Total	23	258182			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Si (H)	4	152.0	18.6
Si (N)	10	185.1	136.8
Si (S)	10	161.4	96.8

Pooled StDev = 109.9

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.73	0.37	0.10	0.901
Error	21	73.63	3.51		
Total	23	74.37			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	CI Lower	CI Upper
Ca (H)	4	35.225	1.436	(---*---)	(---*---)
Ca (N)	10	34.810	2.448	(---*---)	(---*---)
Ca (S)	10	34.725	1.225	(---*---)	(---*---)

Pooled StDev = 1.873

33.6      34.8      36.0      37.2

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0200	0.0100	0.64	0.537
Error	21	0.3284	0.0156		
Total	23	0.3484			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	CI Lower	CI Upper
Se (H)	4	1.4250	0.0532	(---*---)	(---*---)
Se (N)	10	1.3535	0.1581	(---*---)	(---*---)
Se (S)	10	1.3435	0.1027	(---*---)	(---*---)

Pooled StDev = 0.1251

1.280      1.360      1.440      1.520

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	2.1733	1.0867	18.43	0.000
Error	21	1.2379	0.0589		
Total	23	3.4112			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	CI Lower	CI Upper
Co (H)	4	0.8238	0.6216	(---*---)	(---*---)
Co (N)	10	0.0000	0.0000	(---*---)	(---*---)
Co (S)	10	0.0348	0.0935	(---*---)	(---*---)

Pooled StDev = 0.2428

0.00      0.35      0.70      1.05

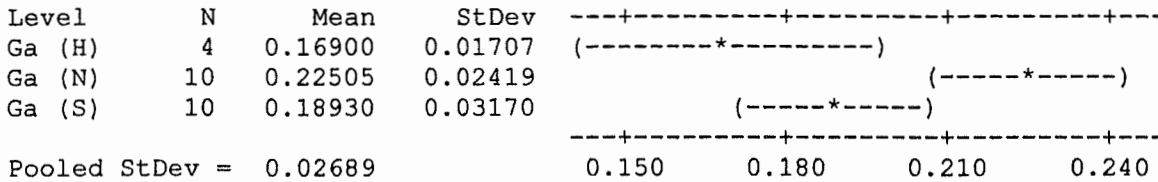
\* NOTE \* All values in column are identical.

**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.011248	0.005624	7.78	0.003
Error	21	0.015181	0.000723		
Total	23	0.026429			

Individual 95% CIs For Mean  
Based on Pooled StDev

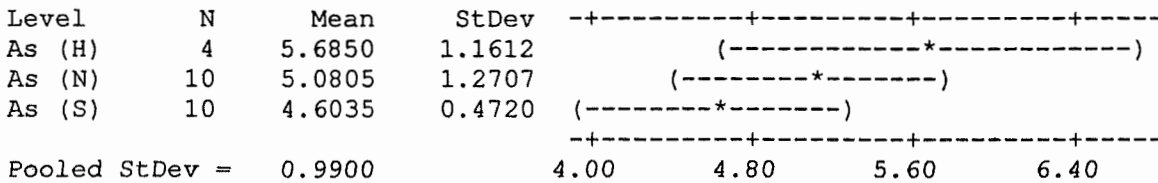


**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	3.506	1.753	1.79	0.192
Error	21	20.581	0.980		
Total	23	24.088			

Individual 95% CIs For Mean  
Based on Pooled StDev

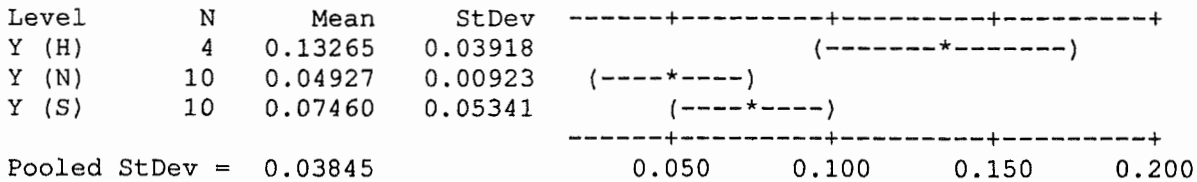


**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.01988	0.00994	6.72	0.006
Error	21	0.03105	0.00148		
Total	23	0.05093			

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

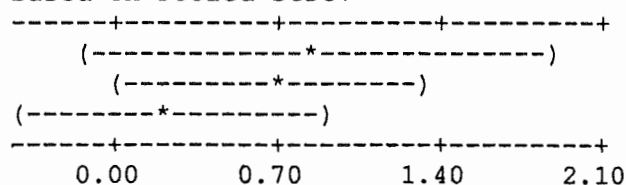
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1.502	0.751	0.77	0.476
Error	21	20.491	0.976		
Total	23	21.993			

Level	N	Mean	StDev
Zr (H)	4	0.8648	1.6037
Zr (N)	10	0.6739	1.1179
Zr (S)	10	0.2389	0.4120

Pooled StDev = 0.9878

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

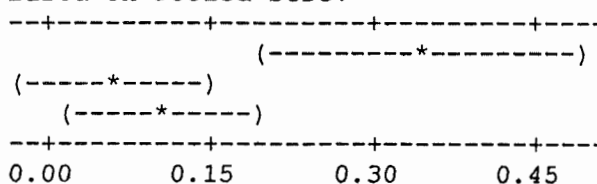
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.2324	0.1162	5.85	0.010
Error	21	0.4170	0.0199		
Total	23	0.6494			

Level	N	Mean	StDev
Sn (H)	4	0.3432	0.3203
Sn (N)	10	0.0633	0.0418
Sn (S)	10	0.1050	0.1020

Pooled StDev = 0.1409

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

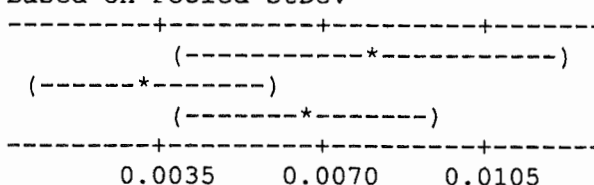
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000859	0.0000430	2.64	0.095
Error	21	0.0003424	0.0000163		
Total	23	0.0004284			

Level	N	Mean	StDev
Cs (H)	4	0.007900	0.001972
Cs (N)	10	0.003273	0.000753
Cs (S)	10	0.006656	0.006016

Pooled StDev = 0.004038

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.1385	0.0692	2.81	0.083
Error	21	0.5171	0.0246		
Total	23	0.6556			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
La (H)	4	0.2983	0.2457
La (N)	10	0.0785	0.0147
La (S)	10	0.1504	0.1927

Pooled StDev = 0.1569

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.2970	0.1485	2.71	0.090
Error	21	1.1495	0.0547		
Total	23	1.4464			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ce (H)	4	0.3584	0.3785
Ce (N)	10	0.0379	0.0309
Ce (S)	10	0.1542	0.2811

Pooled StDev = 0.2340

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.002676	0.001338	2.68	0.092
Error	21	0.010502	0.000500		
Total	23	0.013178			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Pr (H)	4	0.03490	0.03519
Pr (N)	10	0.00465	0.00311
Pr (S)	10	0.01655	0.02728

Pooled StDev = 0.02236

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.03530	0.01765	2.74	0.088
Error	21	0.13541	0.00645		
Total	23	0.17071			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Nd (H)	4	0.12855	0.12367
Nd (N)	10	0.01870	0.01383
Nd (S)	10	0.06200	0.09877

Pooled StDev = 0.08030

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0006016	0.0003008	3.88	0.037
Error	21	0.0016283	0.0000775		
Total	23	0.0022299			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Sm (H)	4	0.019015	0.010092
Sm (N)	10	0.004690	0.002503
Sm (S)	10	0.010404	0.011862

Pooled StDev = 0.008806

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000465	0.0000232	1.52	0.242
Error	21	0.0003211	0.0000153		
Total	23	0.0003675			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Eu (H)	4	0.005793	0.008018
Eu (N)	10	0.001854	0.002248
Eu (S)	10	0.002374	0.003033

Pooled StDev = 0.003910

**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000061	0.0000031	0.97	0.394
Error	21	0.0000659	0.0000031		
Total	23	0.0000720			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Tb (H)	4	0.001816	0.001142
Tb (N)	10	0.001100	0.000691
Tb (S)	10	0.002198	0.002532

Pooled StDev = 0.001772

**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000692	0.000346	2.40	0.115
Error	21	0.003027	0.000144		
Total	23	0.003719			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Gd (H)	4	0.02184	0.01249
Gd (N)	10	0.00670	0.00334
Gd (S)	10	0.01353	0.01653

Pooled StDev = 0.01201

**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0002716	0.0001358	2.86	0.080
Error	21	0.0009988	0.0000476		
Total	23	0.0012704			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Dy (H)	4	0.014370	0.009174
Dy (N)	10	0.004633	0.002983
Dy (S)	10	0.007777	0.008604

Pooled StDev = 0.006897



### One-way Analysis of Variance

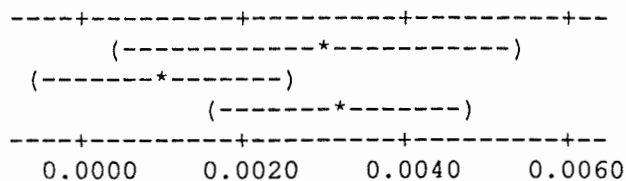
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000268	0.0000134	2.31	0.124
Error	21	0.0001222	0.0000058		
Total	23	0.0001490			

Level	N	Mean	StDev
Ho (H)	4	0.002942	0.001981
Ho (N)	10	0.000925	0.000620
Ho (S)	10	0.003116	0.003448

Pooled StDev = 0.002412

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

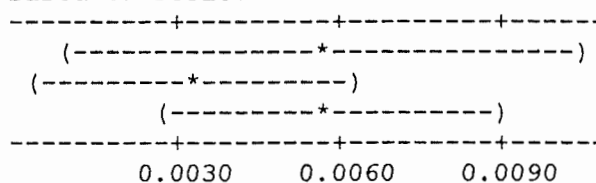
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000379	0.0000190	0.87	0.435
Error	21	0.0004597	0.0000219		
Total	23	0.0004976			

Level	N	Mean	StDev
Er (H)	4	0.005733	0.004004
Er (N)	10	0.003228	0.001276
Er (S)	10	0.005795	0.006641

Pooled StDev = 0.004679

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

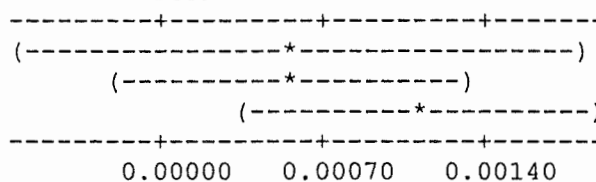
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000016	0.0000008	0.57	0.575
Error	21	0.0000289	0.0000014		
Total	23	0.0000305			

Level	N	Mean	StDev
Tm (H)	4	0.000593	0.000685
Tm (N)	10	0.000564	0.000945
Tm (S)	10	0.001090	0.001471

Pooled StDev = 0.001173

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000367	0.0000184	0.81	0.459
Error	21	0.0004775	0.0000227		
Total	23	0.0005142			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Yb (H)	4	0.005510	0.003057
Yb (N)	10	0.002196	0.001596
Yb (S)	10	0.004102	0.006884

Pooled StDev = 0.004768

0.0000      0.0035      0.0070      0.0105

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000044	0.0000022	1.04	0.372
Error	21	0.0000443	0.0000021		
Total	23	0.0000487			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Lu (H)	4	0.001233	0.000742
Lu (N)	10	0.000667	0.000595
Lu (S)	10	0.001599	0.002094

Pooled StDev = 0.001452

0.0000      0.0010      0.0020      0.0030

### One-way Analysis of Variance

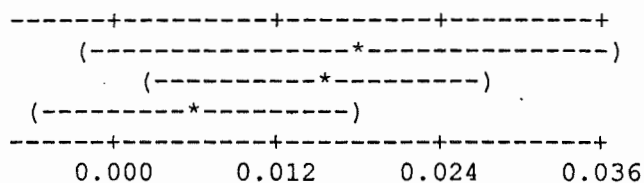
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000590	0.000295	0.83	0.451
Error	21	0.007484	0.000356		
Total	23	0.008074			

Level	N	Mean	StDev
Hf (H)	4	0.01741	0.02872
Hf (N)	10	0.01535	0.02230
Hf (S)	10	0.00599	0.00769

Pooled StDev = 0.01888

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

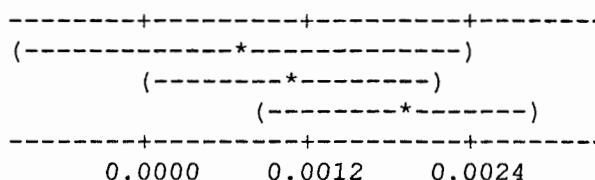
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000049	0.0000025	0.95	0.403
Error	21	0.0000544	0.0000026		
Total	23	0.0000593			

Level	N	Mean	StDev
Ta (H)	4	0.000727	0.000545
Ta (N)	10	0.001080	0.000959
Ta (S)	10	0.001863	0.002242

Pooled StDev = 0.001609

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

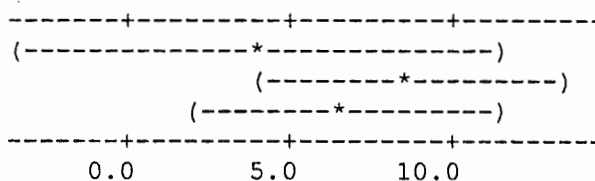
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	62.0	31.0	0.61	0.555
Error	21	1075.3	51.2		
Total	23	1137.4			

Level	N	Mean	StDev
W (H)	4	4.162	2.889
W (N)	10	8.670	7.832
W (S)	10	6.558	7.441

Pooled StDev = 7.156

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000051	0.0000025	0.28	0.755
Error	21	0.0001873	0.0000089		
Total	23	0.0001923			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Tl (H)	4	0.007788	0.002123
Tl (N)	10	0.007108	0.002701
Tl (S)	10	0.008105	0.003465

Pooled StDev = 0.002986

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.000341	0.000171	1.17	0.330
Error	21	0.003063	0.000146		
Total	23	0.003405			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Bi (H)	4	0.02403	0.01121
Bi (N)	10	0.01673	0.00641
Bi (S)	10	0.01312	0.01604

Pooled StDev = 0.01208

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0001867	0.0000933	3.96	0.035
Error	21	0.0004948	0.0000236		
Total	23	0.0006814			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Th (H)	4	0.009380	0.006374
Th (N)	10	0.001771	0.001150
Th (S)	10	0.005853	0.006333

Pooled StDev = 0.004854

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	0.0000461	0.0000230	2.27	0.128
Error	21	0.0002129	0.0000101		
Total	23	0.0002590			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
U (H)	4	0.005537	0.001817
U (N)	10	0.002157	0.000616
U (S)	10	0.004638	0.004709

Pooled StDev = 0.003184

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	2	1.554	0.777	2.83	0.081
Error	21	5.758	0.274		
Total	23	7.312			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
REE Heal	4	0.8989	0.8285
North	10	0.1663	0.0632
South	10	0.4351	0.6379

Pooled StDev = 0.5236

Appendix D: one-way ANOVA of variance results: Control, North of the Park breakage score 1. North of the Park breakage score 2; North of the Park breakage score 3

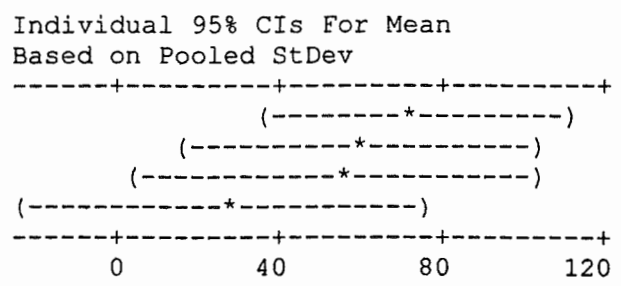
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	4257	1419	0.91	0.468
Error	11	17165	1560		
Total	14	21423			

Level	N	Mean	StDev
Al (H)	5	73.88	14.92
NTH Al 1	4	60.97	58.04
NTH Al 2	3	55.72	55.32
NTH Al 3	3	26.63	5.01

Pooled StDev = 39.50



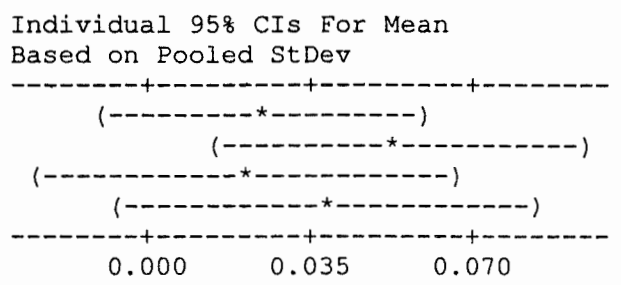
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.00254	0.00085	0.65	0.598
Error	11	0.01428	0.00130		
Total	14	0.01682			

Level	N	Mean	StDev
Sb (H)	5	0.02455	0.01518
NTH Sb 1	4	0.05335	0.05916
NTH Sb 2	3	0.02085	0.00755
NTH Sb 3	3	0.03913	0.03702

Pooled StDev = 0.03603



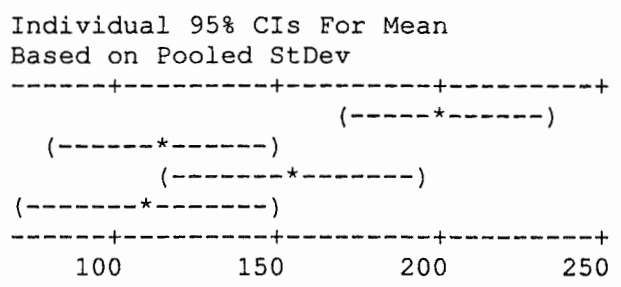
### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	23190	7730	7.66	0.005
Error	11	11106	1010		
Total	14	34296			

Level	N	Mean	StDev
Ba (H)	5	201.40	48.17
NTH Ba 1	4	113.28	21.01
NTH Ba 2	3	157.00	8.89
NTH Ba 3	3	111.47	13.09

Pooled StDev = 31.78



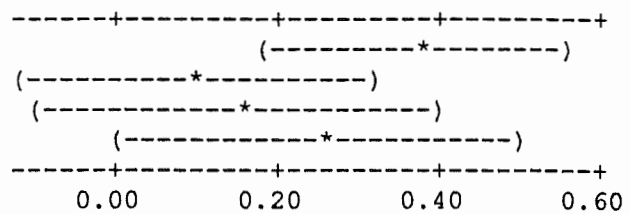
**One-way Analysis of Variance**

## Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.1954	0.0651	1.69	0.227
Error	11	0.4243	0.0386		
Total	14	0.6197			

Level	N	Mean	StDev
Cd (H)	5	0.3762	0.2092
NTH Cd 1	4	0.0967	0.0564
NTH Cd 2	3	0.1567	0.0843
NTH Cd 3	3	0.2525	0.3358

Pooled StDev = 0.1964

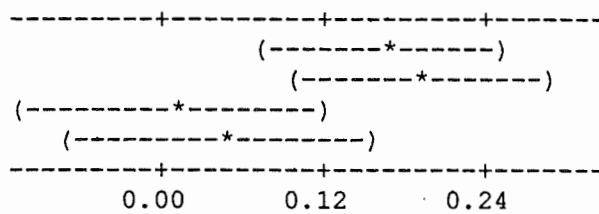
Individual 95% CIs For Mean  
Based on Pooled StDev**One-way Analysis of Variance**

## Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.08374	0.02791	3.48	0.054
Error	11	0.08831	0.00803		
Total	14	0.17205			

Level	N	Mean	StDev
Cr (H)	5	0.16514	0.11647
NTH Cr 1	4	0.19425	0.08208
NTH Cr 2	3	0.01137	0.01969
NTH Cr 3	3	0.04667	0.08083

Pooled StDev = 0.08960

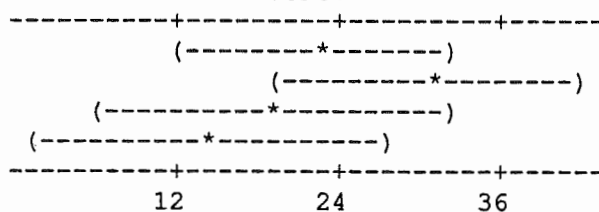
Individual 95% CIs For Mean  
Based on Pooled StDev**One-way Analysis of Variance**

## Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	511	170	1.62	0.241
Error	11	1155	105		
Total	14	1665			

Level	N	Mean	StDev
Cu (H)	5	22.66	5.31
NTH Cu 1	4	30.72	16.90
NTH Cu 2	3	19.53	7.95
NTH Cu 3	3	14.06	5.40

Pooled StDev = 10.25

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	450511	150170	0.95	0.448
Error	11	1732405	157491		
Total	14	2182915			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Fe (H)	5	318.4	193.2
NTH Fe 1	4	139.0	51.0
NTH Fe 2	3	96.0	11.8
NTH Fe 3	3	575.6	887.4

Pooled StDev = 396.9

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	6.794	2.265	6.27	0.010
Error	11	3.972	0.361		
Total	14	10.766			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Pb (H)	5	2.1500	0.9771
NTH Pb 1	4	0.7315	0.2066
NTH Pb 2	3	0.6840	0.0877
NTH Pb 3	3	0.7510	0.0694

Pooled StDev = 0.6009

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	85.6	28.5	1.88	0.191
Error	11	166.4	15.1		
Total	14	252.0			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Li (H)	5	1.066	0.490
NTH Li 1	4	0.494	0.124
NTH Li 2	3	6.843	9.057
NTH Li 3	3	1.206	0.833

Pooled StDev = 3.890

### One-way Analysis of Variance

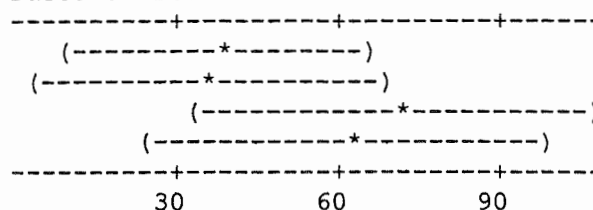
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	3293	1098	1.26	0.336
Error	11	9597	872		
Total	14	12891			

Level	N	Mean	StDev
Mn (H)	5	38.40	21.60
NTH Mn 1	4	35.20	9.24
NTH Mn 2	3	70.97	37.11
NTH Mn 3	3	62.20	48.58

Pooled StDev = 29.54

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

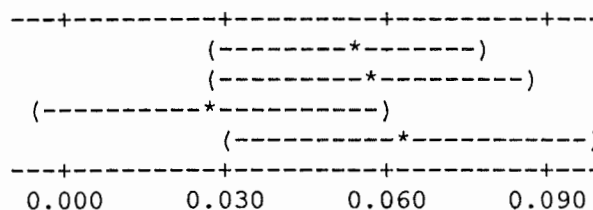
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.002459	0.000820	1.15	0.373
Error	11	0.007850	0.000714		
Total	14	0.010309			

Level	N	Mean	StDev
Mo (H)	5	0.05318	0.02081
NTH Mo 1	4	0.05785	0.03313
NTH Mo 2	3	0.02677	0.00593
NTH Mo 3	3	0.06373	0.03711

Pooled StDev = 0.02671

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

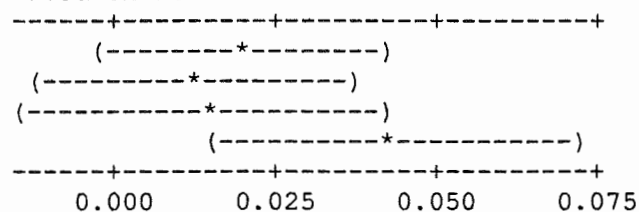
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.001892	0.000631	1.23	0.346
Error	11	0.005653	0.000514		
Total	14	0.007545			

Level	N	Mean	StDev
Nb (H)	5	0.02042	0.00314
NTH Nb 1	4	0.01343	0.00250
NTH Nb 2	3	0.01386	0.00723
NTH Nb 3	3	0.04350	0.05239

Pooled StDev = 0.02267

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.3690	0.1230	3.63	0.049
Error	11	0.3730	0.0339		
Total	14	0.7420			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Rb (H)	5	0.7816	0.1726
NTH Rb 1	4	0.3948	0.1163
NTH Rb 2	3	0.5063	0.2589
NTH Rb 3	3	0.5120	0.1989

Pooled StDev = 0.1841

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.01952	0.00651	1.55	0.256
Error	11	0.04607	0.00419		
Total	14	0.06559			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ag (H)	5	0.05818	0.02225
NTH Ag 1	4	0.13090	0.11260
NTH Ag 2	3	0.03828	0.02940
NTH Ag 3	3	0.05137	0.04649

Pooled StDev = 0.06472

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	136510	45503	8.00	0.004
Error	11	62551	5686		
Total	14	199061			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Sr (H)	5	468.20	118.55
NTH Sr 1	4	244.00	17.57
NTH Sr 2	3	275.00	20.95
NTH Sr 3	3	299.00	47.57

Pooled StDev = 75.41

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	3881	1294	5.18	0.018
Error	11	2746	250		
Total	14	6626			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ti (H)	5	87.12	12.34
NTH Ti 1	4	119.62	25.48
NTH Ti 2	3	83.62	8.04
NTH Ti 3	3	78.73	5.46

Pooled StDev = 15.80

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.2203	0.0734	1.34	0.311
Error	11	0.6015	0.0547		
Total	14	0.8218			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
V (H)	5	0.5734	0.1780
NTH V 1	4	0.4713	0.1194
NTH V 2	3	0.2322	0.0132
NTH V 3	3	0.4457	0.4646

Pooled StDev = 0.2339

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	549	183	1.09	0.393
Error	11	1842	167		
Total	14	2391			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Zn (H)	5	63.76	9.90
NTH Zn 1	4	59.05	15.54
NTH Zn 2	3	57.58	18.93
NTH Zn 3	3	46.77	2.12

Pooled StDev = 12.94

### One-way Analysis of Variance

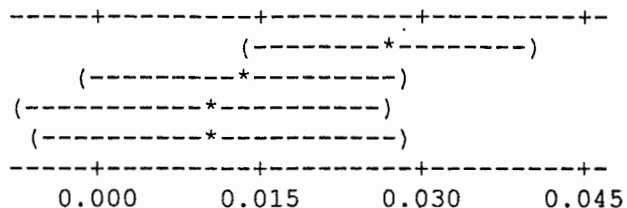
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000846	0.000282	1.53	0.261
Error	11	0.002022	0.000184		
Total	14	0.002868			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Be (H)	5	0.02726	0.01812
NTH Be 1	4	0.01334	0.01165
NTH Be 2	3	0.00983	0.00973
NTH Be 3	3	0.01088	0.00746

Pooled StDev = 0.01356



### One-way Analysis of Variance

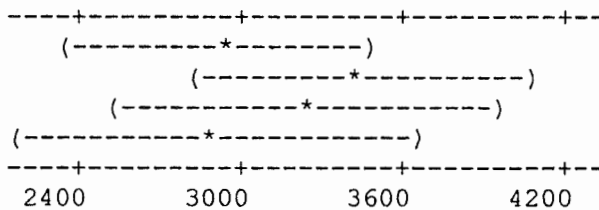
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	785513	261838	0.79	0.523
Error	11	3635247	330477		
Total	14	4420760			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Mg (H)	5	2912.0	355.8
NTH Mg 1	4	3435.0	808.6
NTH Mg 2	3	3230.0	633.2
NTH Mg 3	3	2906.7	427.7

Pooled StDev = 574.9



### One-way Analysis of Variance

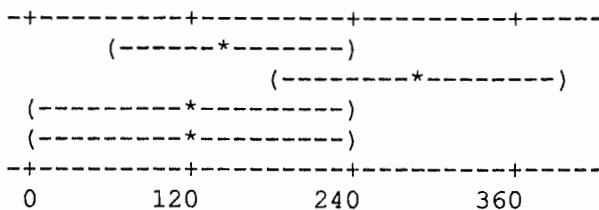
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	70782	23594	2.52	0.112
Error	11	103064	9369		
Total	14	173846			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Si (H)	5	149.40	17.10
NTH Si 1	4	285.00	183.19
NTH Si 2	3	118.17	17.27
NTH Si 3	3	118.83	17.64

Pooled StDev = 96.80



### One-way Analysis of Variance

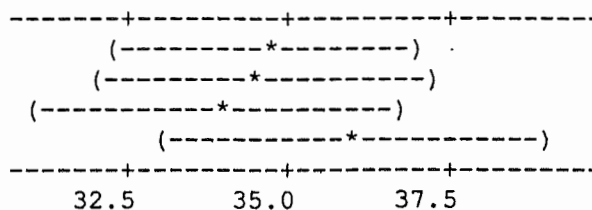
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	6.98	2.33	0.43	0.736
Error	11	59.59	5.42		
Total	14	66.58			

Level	N	Mean	StDev
Ca (H)	5	34.660	1.773
NTH Ca 1	4	34.600	2.174
NTH Ca 2	3	33.900	3.407
NTH Ca 3	3	36.000	2.193

Pooled StDev = 2.328

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

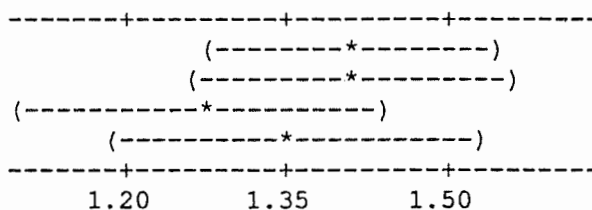
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0432	0.0144	0.77	0.536
Error	11	0.2067	0.0188		
Total	14	0.2499			

Level	N	Mean	StDev
Se (H)	5	1.4060	0.0627
NTH Se 1	4	1.4125	0.2229
NTH Se 2	3	1.2717	0.1360
NTH Se 3	3	1.3567	0.0493

Pooled StDev = 0.1371

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

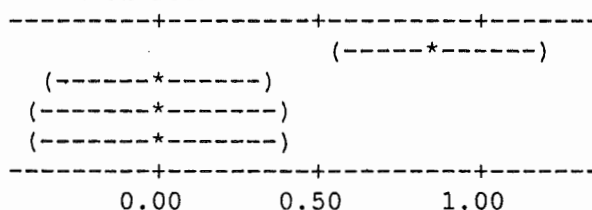
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	2.517	0.839	7.69	0.005
Error	11	1.200	0.109		
Total	14	3.717			

Level	N	Mean	StDev
Co (H)	5	0.8690	0.5478
NTH Co 1	4	0.0000	0.0000
NTH Co 2	3	0.0000	0.0000
NTH Co 3	3	0.0000	0.0000

Pooled StDev = 0.3303

#### Individual 95% CIs For Mean Based on Pooled StDev



\* NOTE \* All values in column are identical.

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.012680	0.004227	10.33	0.002
Error	11	0.004500	0.000409		
Total	14	0.017180			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ga (H)	5	0.16760	0.01511
NTH Ga 1	4	0.23550	0.02133
NTH Ga 2	3	0.20550	0.03196
NTH Ga 3	3	0.23067	0.00945

Pooled StDev = 0.02023

0.150      0.180      0.210      0.240

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	1.87	0.62	0.37	0.777
Error	11	18.58	1.69		
Total	14	20.45			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
As (H)	5	5.410	1.179
NTH As 1	4	5.505	1.791
NTH As 2	3	4.572	0.543
NTH As 3	3	5.023	1.185

Pooled StDev = 1.300

3.6      4.8      6.0      7.2

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.022323	0.007441	15.62	0.000
Error	11	0.005240	0.000476		
Total	14	0.027563			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Y (H)	5	0.13072	0.03421
NTH Y 1	4	0.04560	0.00351
NTH Y 2	3	0.04728	0.00738
NTH Y 3	3	0.05613	0.01438

Pooled StDev = 0.02182

0.040      0.080      0.120      0.160

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	4	1.53	0.38	0.28	0.887
Error	14	19.22	1.37		
Total	18	20.75			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Zr (H)	5	0.699	1.437
Zr (H1)	4	0.057	0.029
NTH Zr 1	4	0.876	1.649
NTH Zr 2	3	0.487	0.714
NTH Zr 3	3	0.591	0.943

Pooled StDev = 1.172

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	4	0.4830	0.1208	3.35	0.040
Error	14	0.5042	0.0360		
Total	18	0.9872			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Sn (H)	5	0.3417	0.2774
Sn (H1)	4	0.4200	0.2485
NTH Sn 1	4	0.0518	0.0310
NTH Sn 2	3	0.0461	0.0055
NTH Sn 3	3	0.0957	0.0642

Pooled StDev = 0.1898

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000605	0.0000202	11.43	0.001
Error	11	0.0000194	0.0000018		
Total	14	0.0000799			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Cs (H)	5	0.007528	0.001899
NTH Cs 1	4	0.003412	0.001117
NTH Cs 2	3	0.003163	0.000708
NTH Cs 3	3	0.003197	0.000336

Pooled StDev = 0.001328



### One-way Analysis of Variance

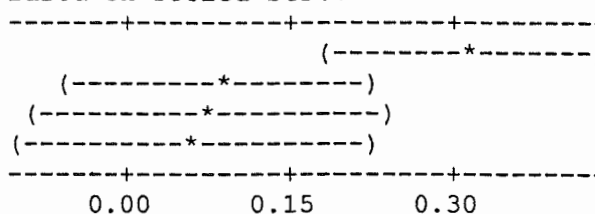
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.1756	0.0585	3.50	0.053
Error	11	0.1842	0.0167		
Total	14	0.3597			

Level	N	Mean	StDev
La (H)	5	0.3076	0.2138
NTH La 1	4	0.0853	0.0198
NTH La 2	3	0.0812	0.0081
NTH La 3	3	0.0668	0.0034

Pooled StDev = 0.1294

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

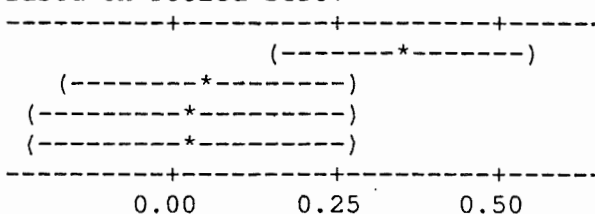
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.3302	0.1101	2.77	0.092
Error	11	0.4377	0.0398		
Total	14	0.7679			

Level	N	Mean	StDev
Ce (H)	5	0.3519	0.3281
NTH Ce 1	4	0.0531	0.0457
NTH Ce 2	3	0.0242	0.0057
NTH Ce 3	3	0.0315	0.0182

Pooled StDev = 0.1995

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

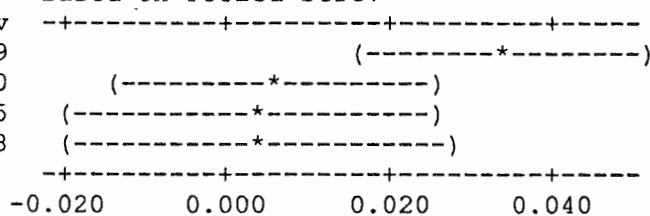
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.002988	0.000996	2.89	0.084
Error	11	0.003792	0.000345		
Total	14	0.006780			

Level	N	Mean	StDev
Pr (H)	5	0.03452	0.03049
NTH Pr 1	4	0.00593	0.00420
NTH Pr 2	3	0.00314	0.00085
NTH Pr 3	3	0.00445	0.00308

Pooled StDev = 0.01857

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.04075	0.01358	3.15	0.069
Error	11	0.04743	0.00431		
Total	14	0.08819			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Nd (H)	5	0.12904	0.10710
NTH Nd 1	4	0.02348	0.02003
NTH Nd 2	3	0.01365	0.00791
NTH Nd 3	3	0.01738	0.01050

Pooled StDev = 0.06567

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0007870	0.0002623	7.53	0.005
Error	11	0.0003831	0.0000348		
Total	14	0.0011701			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Sm (H)	5	0.020052	0.009042
NTH Sm 1	4	0.004627	0.001695
NTH Sm 2	3	0.004497	0.002234
NTH Sm 3	3	0.004967	0.004327

Pooled StDev = 0.005901

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0001137	0.0000379	1.39	0.297
Error	11	0.0002994	0.0000272		
Total	14	0.0004131			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Eu (H)	5	0.007594	0.008028
NTH Eu 1	4	0.002573	0.003075
NTH Eu 2	3	0.001640	0.001709
NTH Eu 3	3	0.001110	0.001923

Pooled StDev = 0.005217

### One-way Analysis of Variance

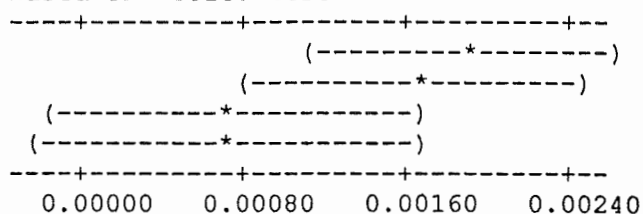
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000042	0.0000014	2.49	0.115
Error	11	0.0000062	0.0000006		
Total	14	0.0000104			

Level	N	Mean	StDev
Tb (H)	5	1.89E-03	1.00E-03
NTH Tb 1	4	1.66E-03	7.78E-04
NTH Tb 2	3	7.55E-04	3.50E-04
NTH Tb 3	3	6.95E-04	2.48E-04

Pooled StDev = 7.51E-04

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

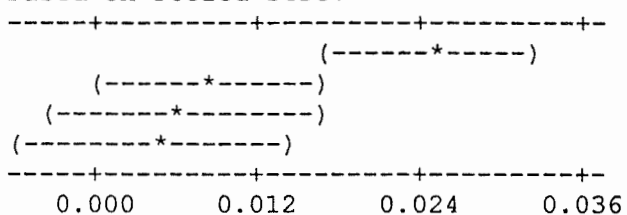
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0010988	0.0003663	5.81	0.012
Error	11	0.0006930	0.0000630		
Total	14	0.0017918			

Level	N	Mean	StDev
Gd (H)	5	0.024612	0.012465
NTH Gd 1	4	0.008602	0.004110
NTH Gd 2	3	0.006347	0.002696
NTH Gd 3	3	0.004513	0.001771

Pooled StDev = 0.007937

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

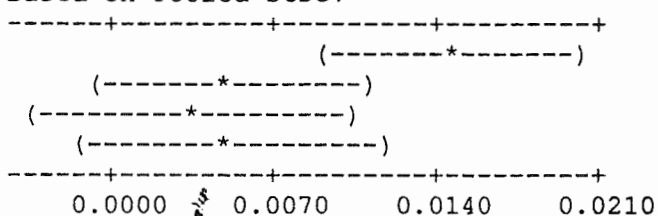
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0003511	0.0001170	3.91	0.040
Error	11	0.0003295	0.0000300		
Total	14	0.0006805			

Level	N	Mean	StDev
Dy (H)	5	0.014796	0.008002
NTH Dy 1	4	0.005145	0.002915
NTH Dy 2	3	0.003377	0.001520
NTH Dy 3	3	0.005207	0.004648

Pooled StDev = 0.005473

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

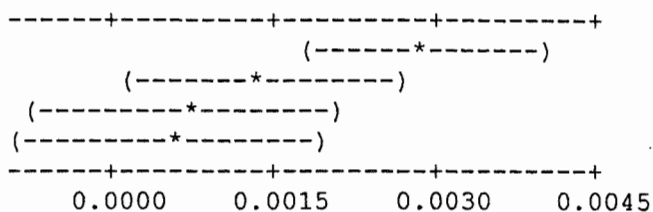
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000147	0.0000049	3.94	0.039
Error	11	0.0000136	0.0000012		
Total	14	0.0000283			

Level	N	Mean	StDev
Ho (H)	5	0.002904	0.001718
NTH Ho 1	4	0.001413	0.000524
NTH Ho 2	3	0.000675	0.000403
NTH Ho 3	3	0.000526	0.000588

Pooled StDev = 0.001114

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

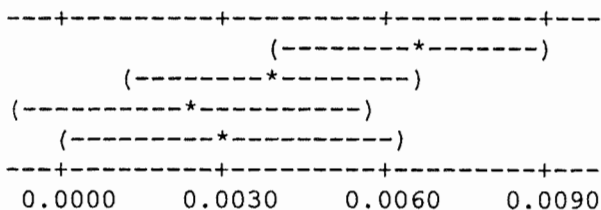
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000392	0.0000131	2.09	0.160
Error	11	0.0000687	0.0000062		
Total	14	0.0001078			

Level	N	Mean	StDev
Er (H)	5	0.006454	0.003824
NTH Er 1	4	0.003987	0.001679
NTH Er 2	3	0.002397	0.000396
NTH Er 3	3	0.003047	0.000838

Pooled StDev = 0.002499

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

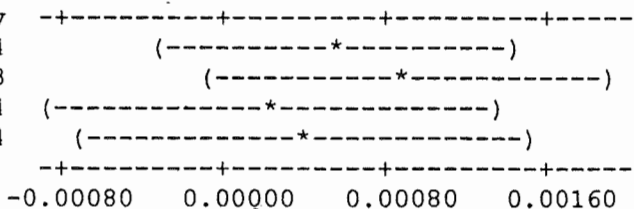
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000008	0.0000003	0.35	0.790
Error	11	0.0000086	0.0000008		
Total	14	0.0000094			

Level	N	Mean	StDev
Tm (H)	5	5.84E-04	5.94E-04
NTH Tm 1	4	9.10E-04	1.50E-03
NTH Tm 2	3	2.69E-04	2.85E-04
NTH Tm 3	3	3.97E-04	3.88E-04

Pooled StDev = 8.85E-04

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000028	0.0000009	1.46	0.278
Error	11	0.0000069	0.0000006		
Total	14	0.0000097			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ta (H)	5	6.91E-04	4.78E-04
NTH Ta 1	4	1.66E-03	1.17E-03
NTH Ta 2	3	6.58E-04	2.59E-04
NTH Ta 3	3	7.26E-04	9.42E-04

Pooled StDev = 7.94E-04

0.00000    0.00080    0.00160    0.00240

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	133.8	44.6	0.92	0.461
Error	11	530.4	48.2		
Total	14	664.2			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
W (H)	5	3.690	2.716
NTH W 1	4	11.150	9.068
NTH W 2	3	5.720	6.312
NTH W 3	3	8.313	9.342

Pooled StDev = 6.944

0.0    6.0    12.0    18.0

### One-way Analysis of Variance

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000364	0.0000121	2.89	0.084
Error	11	0.0000462	0.0000042		
Total	14	0.0000826			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
T1 (H)	5	0.007988	0.001892
NTH T1 1	4	0.005747	0.001033
NTH T1 2	3	0.009907	0.003676
NTH T1 3	3	0.006123	0.000902

Pooled StDev = 0.002049

0.0050    0.0075    0.0100

### One-way Analysis of Variance

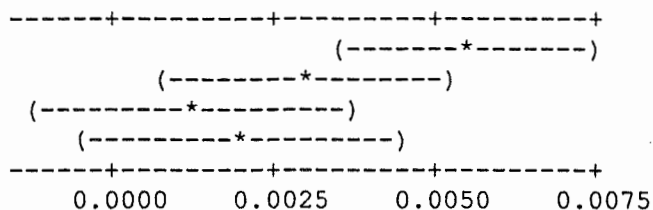
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000447	0.0000149	3.61	0.049
Error	11	0.0000455	0.0000041		
Total	14	0.0000902			

Level	N	Mean	StDev
Yb (H)	5	0.005616	0.002658
NTH Yb 1	4	0.003058	0.001701
NTH Yb 2	3	0.001258	0.000697
NTH Yb 3	3	0.001987	0.001942

Pooled StDev = 0.002033

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

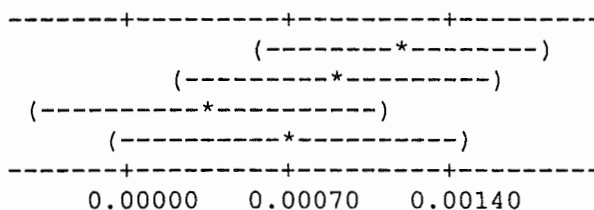
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000015	0.0000005	1.26	0.337
Error	11	0.0000043	0.0000004		
Total	14	0.0000058			

Level	N	Mean	StDev
Lu (H)	5	1.21E-03	6.45E-04
NTH Lu 1	4	8.90E-04	8.87E-04
NTH Lu 2	3	3.43E-04	3.02E-04
NTH Lu 3	3	6.95E-04	2.48E-04

Pooled StDev = 6.27E-04

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

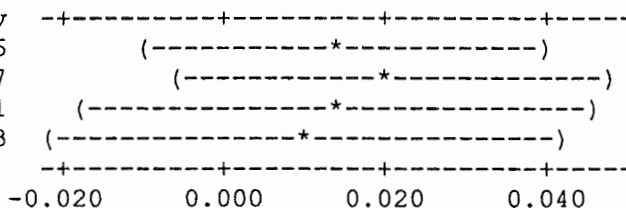
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000211	0.000070	0.11	0.951
Error	11	0.006901	0.000627		
Total	14	0.007112			

Level	N	Mean	StDev
Hf (H)	5	0.01459	0.02566
NTH Hf 1	4	0.02074	0.03437
NTH Hf 2	3	0.01337	0.01241
NTH Hf 3	3	0.01014	0.01443

Pooled StDev = 0.02505

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0001820	0.0000607	0.90	0.474
Error	11	0.0007444	0.0000677		
Total	14	0.0009264			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Bi (H)	5	0.024060	0.009711
NTH Bi 1	4	0.016075	0.002473
NTH Bi 2	3	0.017117	0.006764
NTH Bi 3	3	0.017213	0.011343

Pooled StDev = 0.008226

0.0070      0.0140      0.0210      0.0280

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0002004	0.0000668	5.58	0.014
Error	11	0.0001318	0.0000120		
Total	14	0.0003322			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Th (H)	5	0.009482	0.005525
NTH Th 1	4	0.002329	0.001515
NTH Th 2	3	0.001556	0.001112
NTH Th 3	3	0.001241	0.000395

Pooled StDev = 0.003461

0.0000      0.0050      0.0100

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000344	0.0000115	9.66	0.002
Error	11	0.0000131	0.0000012		
Total	14	0.0000475			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
U (H)	5	0.005310	0.001654
NTH U 1	4	0.002550	0.000409
NTH U 2	3	0.002100	0.000244
NTH U 3	3	0.001688	0.000867

Pooled StDev = 0.001090

0.0020      0.0040      0.0060

## One-way Analysis of Variance

### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	1.845	0.615	3.24	0.064
Error	11	2.089	0.190		
Total	14	3.935			

### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	
REE Health	5	0.9088	0.7179	(-----*-----)
REE N 1	4	0.2006	0.0884	(-----*-----)
REE N 2	3	0.1437	0.0235	(-----*-----)
REE N 3	3	0.1433	0.0419	(-----*-----)

Pooled StDev = 0.4358

0.00      0.50      1.00



Appendix E: one-way ANOVA of variance results: Control, South of the Park breakage score 1. South of the Park breakage score 2, South of the Park breakage score 3

### One-way Analysis of Variance

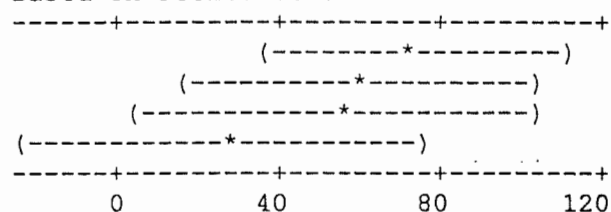
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	4257	1419	0.91	0.468
Error	11	17165	1560		
Total	14	21423			

Level	N	Mean	StDev
Al (H)	5	73.88	14.92
STH Al 1	4	60.97	58.04
STH Al 2	3	55.72	55.32
STH Al 3	3	26.63	5.01

Pooled StDev = 39.50

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

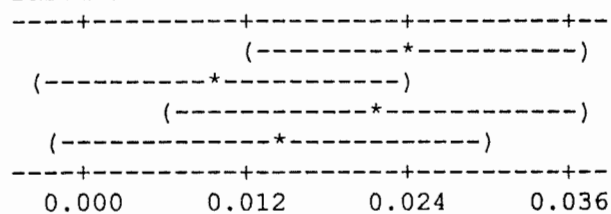
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000574	0.000191	1.22	0.347
Error	11	0.001719	0.000156		
Total	14	0.002293			

Level	N	Mean	StDev
Sb (H)	5	0.02455	0.01518
STH Sb 1	4	0.00986	0.00472
STH Sb 2	3	0.02180	0.01801
STH Sb 3	3	0.01390	0.00641

Pooled StDev = 0.01250

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

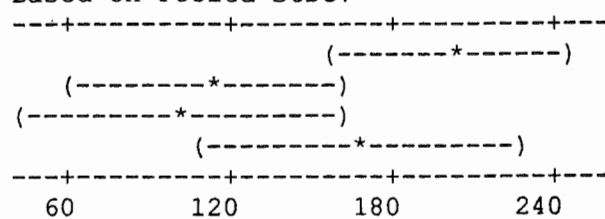
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	26608	8869	4.03	0.037
Error	11	24213	2201		
Total	14	50821			

Level	N	Mean	StDev
Ba (H)	5	201.40	48.17
STH Ba 1	4	112.02	46.96
STH Ba 2	3	102.37	17.73
STH Ba 3	3	166.67	62.00

Pooled StDev = 46.92

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	11.18	3.73	1.30	0.323
Error	11	31.54	2.87		
Total	14	42.72			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	
Cd (H)	5	0.376	0.209	(-----*-----)
STH Cd 1	4	0.063	0.026	(-----*-----)
STH Cd 2	3	2.328	3.960	(-----*-----)
STH Cd 3	3	0.073	0.045	(-----*-----)

Pooled StDev = 1.693

-2.0                      0.0                      2.0                      4.0

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0590	0.0197	1.46	0.279
Error	11	0.1483	0.0135		
Total	14	0.2073			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	
Cr (H)	5	0.1651	0.1165	(-----*-----)
STH Cr 1	4	0.1270	0.1571	(-----*-----)
STH Cr 2	3	0.0596	0.1000	(-----*-----)
STH Cr 3	3	0.0000	0.0000	(-----*-----)

Pooled StDev = 0.1161

-0.12                      0.00                      0.12                      0.24

\* NOTE \* All values in column are identical.

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	515.0	171.7	3.50	0.053
Error	11	539.3	49.0		
Total	14	1054.3			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev	
Cu (H)	5	22.660	5.312	(-----*-----)
STH Cu 1	4	17.282	10.847	(-----*-----)
STH Cu 2	3	10.743	5.725	(-----*-----)
STH Cu 3	3	7.753	1.991	(-----*-----)

Pooled StDev = 7.002

0                      10                      20                      30

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	179146	59715	1.00	0.431
Error	11	659528	59957		
Total	14	838674			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Fe (H)	5	318.4	193.2
STH Fe 1	4	106.8	38.6
STH Fe 2	3	337.1	499.6
STH Fe 3	3	106.8	57.5

Pooled StDev = 244.9

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	5.321	1.774	3.39	0.058
Error	11	5.760	0.524		
Total	14	11.081			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Pb (H)	5	2.1500	0.9771
STH Pb 1	4	0.9055	0.3943
STH Pb 2	3	0.9075	0.8471
STH Pb 3	3	0.8437	0.1391

Pooled StDev = 0.7236

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	1.411	0.470	0.83	0.506
Error	11	6.245	0.568		
Total	14	7.656			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Li (H)	5	1.0658	0.4895
STH Li 1	4	0.8503	0.4971
STH Li 2	3	0.8050	0.7482
STH Li 3	3	1.6493	1.3088

Pooled StDev = 0.7535

### One-way Analysis of Variance

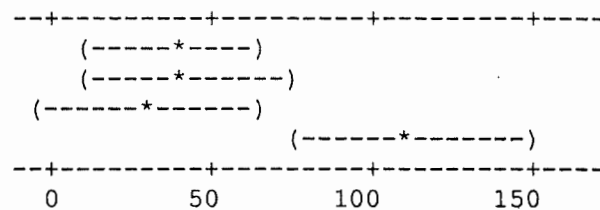
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	13679	4560	5.54	0.015
Error	11	9059	824		
Total	14	22737			

Level	N	Mean	StDev
Mn (H)	5	38.40	21.60
STH Mn 1	4	41.75	12.42
STH Mn 2	3	30.63	18.98
STH Mn 3	3	112.47	54.81

Pooled StDev = 28.70

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

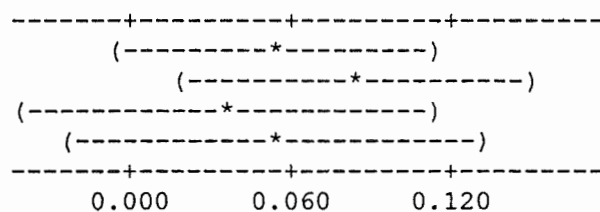
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.00395	0.00132	0.35	0.789
Error	11	0.04118	0.00374		
Total	14	0.04513			

Level	N	Mean	StDev
Mo (H)	5	0.05318	0.02081
STH Mo 1	4	0.08273	0.10692
STH Mo 2	3	0.03720	0.01580
STH Mo 3	3	0.05250	0.04823

Pooled StDev = 0.06118

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

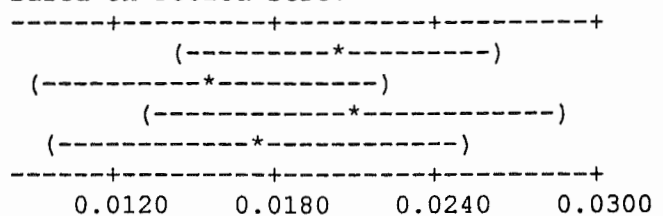
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000752	0.0000251	0.67	0.590
Error	11	0.0004137	0.0000376		
Total	14	0.0004889			

Level	N	Mean	StDev
Nb (H)	5	0.020420	0.003140
STH Nb 1	4	0.015475	0.003800
STH Nb 2	3	0.020783	0.012228
STH Nb 3	3	0.017267	0.003993

Pooled StDev = 0.006133

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

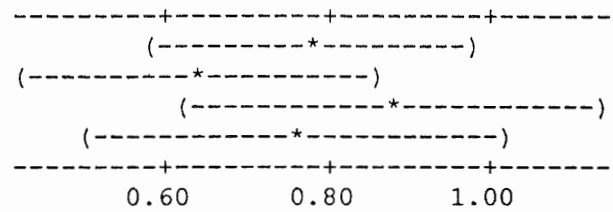
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0984	0.0328	0.83	0.507
Error	11	0.4370	0.0397		
Total	14	0.5354			

Level	N	Mean	StDev
Rb (H)	5	0.7816	0.1726
STH Rb 1	4	0.6450	0.0963
STH Rb 2	3	0.8800	0.3388
STH Rb 3	3	0.7623	0.1738

Pooled StDev = 0.1993

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

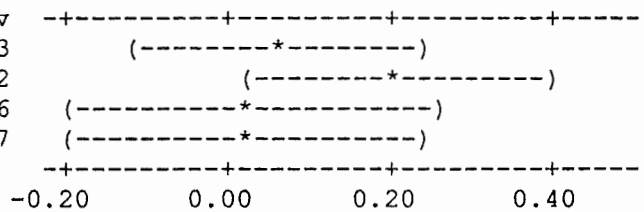
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0836	0.0279	0.91	0.466
Error	11	0.3359	0.0305		
Total	14	0.4195			

Level	N	Mean	StDev
Ag (H)	5	0.0582	0.0223
STH Ag 1	4	0.2038	0.3332
STH Ag 2	3	0.0288	0.0206
STH Ag 3	3	0.0163	0.0067

Pooled StDev = 0.1748

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

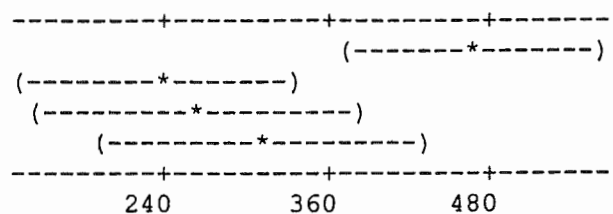
Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	145619	48540	5.58	0.014
Error	11	95690	8699		
Total	14	241308			

Level	N	Mean	StDev
Sr (H)	5	468.20	118.55
STH Sr 1	4	236.00	82.72
STH Sr 2	3	261.17	32.23
STH Sr 3	3	313.67	91.82

Pooled StDev = 93.27

Individual 95% CIs For Mean  
Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	520	173	1.15	0.372
Error	11	1658	151		
Total	14	2178			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ti (H)	5	87.12	12.34
STH Ti 1	4	79.42	3.22
STH Ti 2	3	89.17	21.75
STH Ti 3	3	73.50	5.98

Pooled StDev = 12.28

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.1511	0.0504	1.94	0.182
Error	11	0.2855	0.0260		
Total	14	0.4366			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
V (H)	5	0.5734	0.1780
STH V 1	4	0.3217	0.0832
STH V 2	3	0.4495	0.2461
STH V 3	3	0.3940	0.0920

Pooled StDev = 0.1611

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	1298	433	2.77	0.091
Error	11	1715	156		
Total	14	3012			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Zn (H)	5	63.76	9.90
STH Zn 1	4	51.30	12.26
STH Zn 2	3	44.12	9.54
STH Zn 3	3	69.20	18.57

Pooled StDev = 12.49

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000394	0.000131	0.78	0.530
Error	11	0.001853	0.000168		
Total	14	0.002247			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Be (H)	5	0.02726	0.01812
STH Be 1	4	0.01422	0.00053
STH Be 2	3	0.02100	0.01187
STH Be 3	3	0.01880	0.01134

Pooled StDev = 0.01298

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	4707285	1569095	7.86	0.004
Error	11	2195438	199585		
Total	14	6902723			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Mg (H)	5	2912.0	355.8
STH Mg 1	4	2982.5	310.6
STH Mg 2	3	4041.7	782.9
STH Mg 3	3	4126.7	294.8

Pooled StDev = 446.7

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	8005	2668	0.38	0.772
Error	11	78037	7094		
Total	14	86042			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Si (H)	5	149.40	17.10
STH Si 1	4	127.80	55.42
STH Si 2	3	185.77	158.69
STH Si 3	3	181.67	92.98

Pooled StDev = 84.23



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	5.75	1.92	1.04	0.415
Error	11	20.35	1.85		
Total	14	26.10			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ca (H)	5	34.660	1.773
STH Ca 1	4	35.425	1.266
STH Ca 2	3	33.617	0.718
STH Ca 3	3	34.900	0.985

Pooled StDev = 1.360

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.02861	0.00954	1.10	0.389
Error	11	0.09504	0.00864		
Total	14	0.12364			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Se (H)	5	1.4060	0.0627
STH Se 1	4	1.3050	0.0835
STH Se 2	3	1.3383	0.1578
STH Se 3	3	1.4000	0.0656

Pooled StDev = 0.0929

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	2.337	0.779	6.80	0.007
Error	11	1.261	0.115		
Total	14	3.598			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Co (H)	5	0.8690	0.5478
STH Co 1	4	0.0128	0.0257
STH Co 2	3	0.0990	0.1715
STH Co 3	3	0.0000	0.0000

Pooled StDev = 0.3386

\* NOTE \* All values in column are identical.

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.006562	0.002187	4.85	0.022
Error	11	0.004963	0.000451		
Total	14	0.011525			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ga (H)	5	0.16760	0.01511
STH Ga 1	4	0.21650	0.01700
STH Ga 2	3	0.17433	0.02706
STH Ga 3	3	0.16800	0.02931

Pooled StDev = 0.02124

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	2.368	0.789	1.18	0.362
Error	11	7.363	0.669		
Total	14	9.731			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
As (H)	5	5.4100	1.1787
STH As 1	4	4.4725	0.3412
STH As 2	3	4.5717	0.5821
STH As 3	3	4.8100	0.6239

Pooled StDev = 0.8181

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.01552	0.00517	2.25	0.140
Error	11	0.02533	0.00230		
Total	14	0.04085			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Y (H)	5	0.13072	0.03421
STH Y 1	4	0.04715	0.01498
STH Y 2	3	0.09360	0.08350
STH Y 3	3	0.09220	0.05492

Pooled StDev = 0.04799

### One-way Analysis of Variance

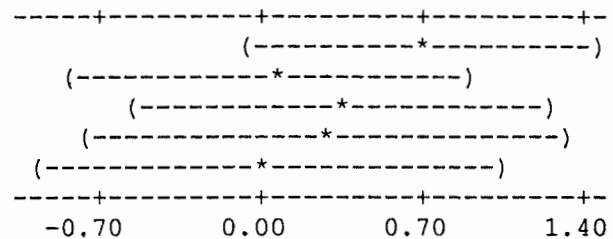
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	4	1.262	0.315	0.46	0.764
Error	14	9.594	0.685		
Total	18	10.856			

Level	N	Mean	StDev
Zr (H)	5	0.6993	1.4373
Zr (H1)	4	0.0567	0.0285
STH Zr 1	4	0.3597	0.6137
STH Zr 2	3	0.2886	0.3152
STH Zr 3	3	0.0279	0.0014

Pooled StDev = 0.8278

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

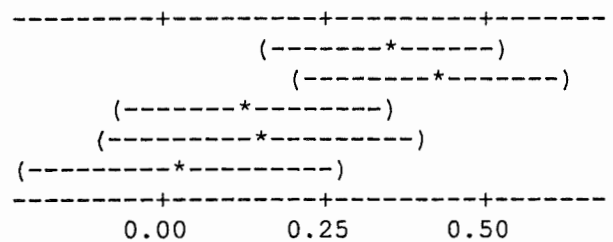
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	4	0.3899	0.0975	2.44	0.096
Error	14	0.5594	0.0400		
Total	18	0.9493			

Level	N	Mean	StDev
Sn (H)	5	0.3417	0.2774
Sn (H1)	4	0.4200	0.2485
STH Sn 1	4	0.1356	0.0749
STH Sn 2	3	0.1437	0.1564
STH Sn 3	3	0.0256	0.0177

Pooled StDev = 0.1999

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

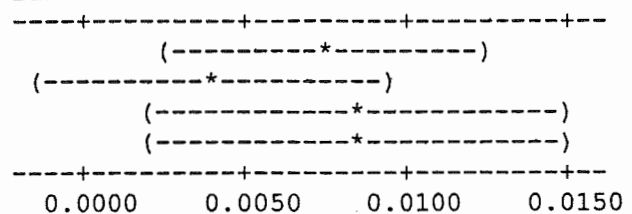
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000530	0.0000177	0.67	0.588
Error	11	0.0002897	0.0000263		
Total	14	0.0003427			

Level	N	Mean	StDev
Cs (H)	5	0.007528	0.001899
STH Cs 1	4	0.003908	0.001592
STH Cs 2	3	0.008590	0.008766
STH Cs 3	3	0.008387	0.007549

Pooled StDev = 0.005132

#### Individual 95% CIs For Mean Based on Pooled StDev



**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.1704	0.0568	3.17	0.068
Error	11	0.1969	0.0179		
Total	14	0.3673			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
La (H)	5	0.3076	0.2138
STH La 1	4	0.0775	0.0393
STH La 2	3	0.0843	0.0486
STH La 3	3	0.0843	0.0486

Pooled StDev = 0.1338

**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.3745	0.1248	1.53	0.262
Error	11	0.8976	0.0816		
Total	14	1.2720			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Ce (H)	5	0.3519	0.3281
STH Ce 1	4	0.0453	0.0275
STH Ce 2	3	0.0610	0.0711
STH Ce 3	3	0.3926	0.4767

Pooled StDev = 0.2857

**One-way Analysis of Variance**

Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.003379	0.001126	1.53	0.262
Error	11	0.008115	0.000738		
Total	14	0.011495			

Individual 95% CIs For Mean  
Based on Pooled StDev

Level	N	Mean	StDev
Pr (H)	5	0.03452	0.03049
STH Pr 1	4	0.00508	0.00307
STH Pr 2	3	0.00878	0.01018
STH Pr 3	3	0.03961	0.04561

Pooled StDev = 0.02716

### One-way Analysis of Variance

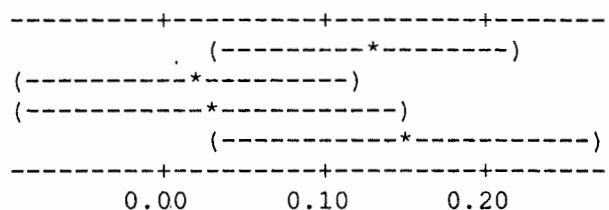
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.04797	0.01599	1.75	0.215
Error	11	0.10070	0.00915		
Total	14	0.14867			

Level	N	Mean	StDev
Nd (H)	5	0.12904	0.10710
STH Nd 1	4	0.01820	0.00554
STH Nd 2	3	0.03318	0.04168
STH Nd 3	3	0.14923	0.16007

Pooled StDev = 0.09568

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

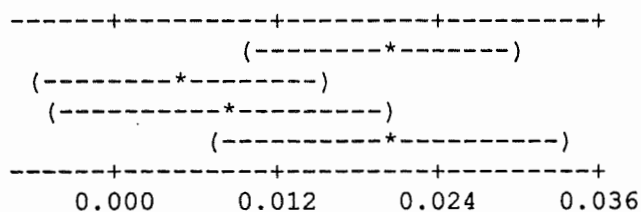
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000767	0.000256	2.47	0.116
Error	11	0.001137	0.000103		
Total	14	0.001904			

Level	N	Mean	StDev
Sm (H)	5	0.02005	0.00904
STH Sm 1	4	0.00466	0.00116
STH Sm 2	3	0.00796	0.00776
STH Sm 3	3	0.02051	0.01851

Pooled StDev = 0.01017

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

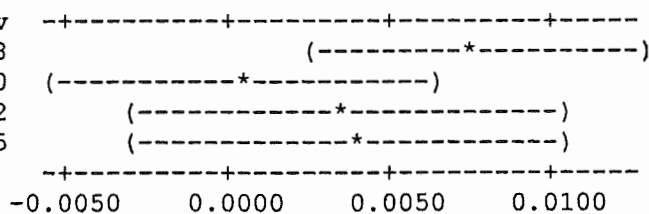
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0001166	0.0000389	1.36	0.306
Error	11	0.0003148	0.0000286		
Total	14	0.0004314			

Level	N	Mean	StDev
Eu (H)	5	0.007594	0.008028
STH Eu 1	4	0.000410	0.000820
STH Eu 2	3	0.003590	0.004352
STH Eu 3	3	0.003777	0.002925

Pooled StDev = 0.005349

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

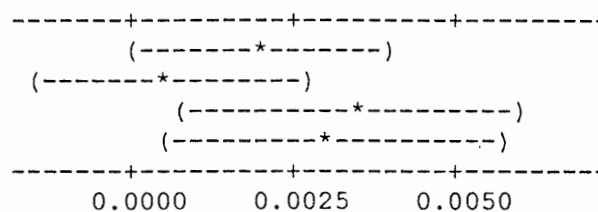
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000170	0.0000057	1.38	0.299
Error	11	0.0000450	0.0000041		
Total	14	0.0000620			

Level	N	Mean	StDev
Tb (H)	5	0.001893	0.001003
STH Tb 1	4	0.000624	0.000788
STH Tb 2	3	0.003417	0.003867
STH Tb 3	3	0.003078	0.002149

Pooled StDev = 0.002023

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

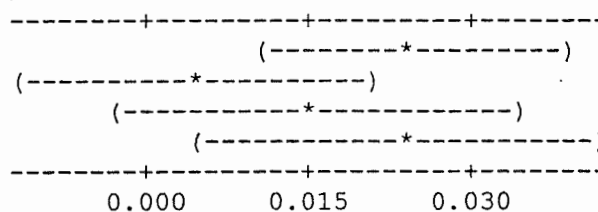
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.001048	0.000349	1.57	0.251
Error	11	0.002441	0.000222		
Total	14	0.003489			

Level	N	Mean	StDev
Gd (H)	5	0.02461	0.01246
STH Gd 1	4	0.00456	0.00106
STH Gd 2	3	0.01540	0.01585
STH Gd 3	3	0.02363	0.02563

Pooled StDev = 0.01490

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

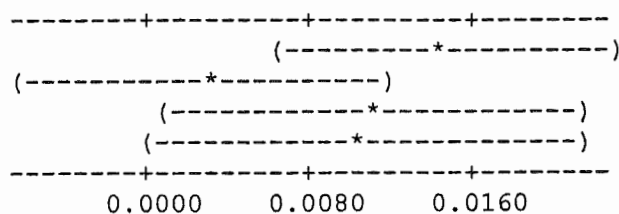
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0003185	0.0001062	1.52	0.264
Error	11	0.0007680	0.0000698		
Total	14	0.0010865			

Level	N	Mean	StDev
Dy (H)	5	0.014796	0.008002
STH Dy 1	4	0.002975	0.000706
STH Dy 2	3	0.011290	0.014212
STH Dy 3	3	0.010667	0.007294

Pooled StDev = 0.008356

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000052	0.0000017	0.17	0.915
Error	11	0.0001137	0.0000103		
Total	14	0.0001189			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ho (H)	5	0.002904	0.001718
STH Ho 1	4	0.002569	0.004066
STH Ho 2	3	0.004200	0.004825
STH Ho 3	3	0.002761	0.001692

Pooled StDev = 0.003215

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000852	0.0000284	0.84	0.500
Error	11	0.0003717	0.0000338		
Total	14	0.0004569			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Er (H)	5	0.006454	0.003824
STH Er 1	4	0.002850	0.001126
STH Er 2	3	0.009837	0.011906
STH Er 3	3	0.005680	0.003598

Pooled StDev = 0.005813

### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000081	0.0000027	2.18	0.148
Error	11	0.0000136	0.0000012		
Total	14	0.0000217			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Tm (H)	5	0.000584	0.000594
STH Tm 1	4	0.000210	0.000243
STH Tm 2	3	0.002267	0.002355
STH Tm 3	3	0.001085	0.000689

Pooled StDev = 0.001113

### One-way Analysis of Variance

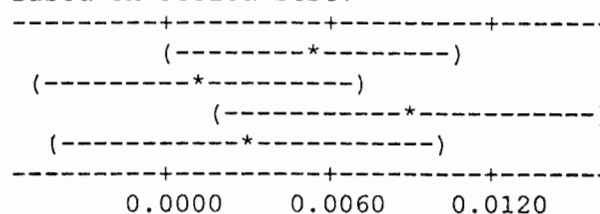
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0001224	0.0000408	1.32	0.318
Error	11	0.0003400	0.0000309		
Total	14	0.0004624			

Level	N	Mean	StDev
Yb (H)	5	0.005616	0.002658
STH Yb 1	4	0.001171	0.000424
STH Yb 2	3	0.009150	0.012439
STH Yb 3	3	0.002960	0.000941

Pooled StDev = 0.005560

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

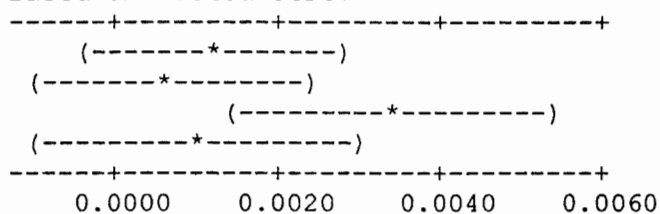
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000147	0.0000049	1.99	0.174
Error	11	0.0000270	0.0000025		
Total	14	0.0000417			

Level	N	Mean	StDev
Lu (H)	5	0.001206	0.000645
STH Lu 1	4	0.000640	0.000253
STH Lu 2	3	0.003393	0.003478
STH Lu 3	3	0.001085	0.000689

Pooled StDev = 0.001567

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

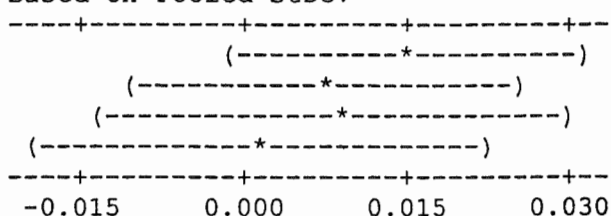
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000348	0.000116	0.42	0.744
Error	11	0.003064	0.000279		
Total	14	0.003412			

Level	N	Mean	StDev
Hf (H)	5	0.01459	0.02566
STH Hf 1	4	0.00783	0.01060
STH Hf 2	3	0.00838	0.00675
STH Hf 3	3	0.00113	0.00123

Pooled StDev = 0.01669

#### Individual 95% CIs For Mean Based on Pooled StDev





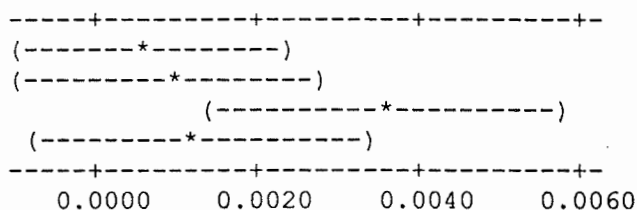
### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000186	0.0000062	2.12	0.156
Error	11	0.0000322	0.0000029		
Total	14	0.0000507			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
Ta (H)	5	0.000691	0.000478
STH Ta 1	4	0.000950	0.000634
STH Ta 2	3	0.003657	0.003737
STH Ta 3	3	0.001288	0.001026



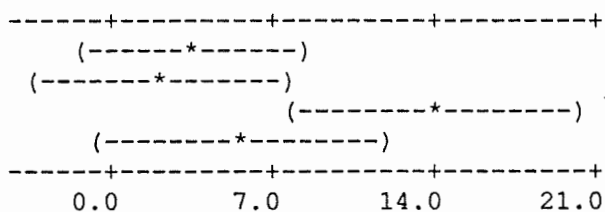
### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	274.1	91.4	3.57	0.050
Error	11	281.1	25.6		
Total	14	555.2			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
W (H)	5	3.690	2.716
STH W 1	4	1.935	0.884
STH W 2	3	13.807	9.511
STH W 3	3	5.473	5.847



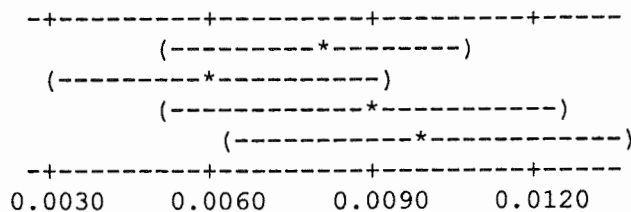
### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000297	0.0000099	1.18	0.363
Error	11	0.0000927	0.0000084		
Total	14	0.0001224			

#### Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev
T1 (H)	5	0.007988	0.001892
STH T1 1	4	0.006062	0.001519
STH T1 2	3	0.008907	0.005404
STH T1 3	3	0.010027	0.002554



### One-way Analysis of Variance

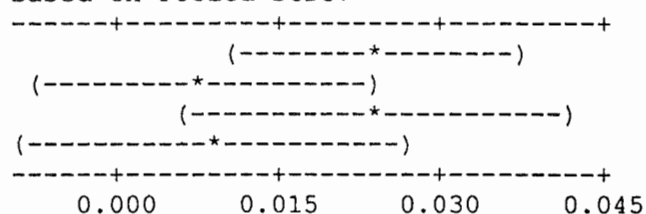
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.000890	0.000297	1.48	0.273
Error	11	0.002202	0.000200		
Total	14	0.003092			

Level	N	Mean	StDev
Bi (H)	5	0.02406	0.00971
STH Bi 1	4	0.00793	0.00268
STH Bi 2	3	0.02379	0.03001
STH Bi 3	3	0.00938	0.00102

Pooled StDev = 0.01415

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

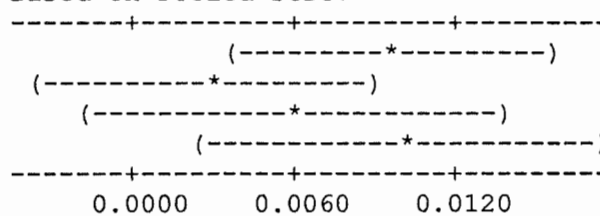
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0001340	0.0000447	1.25	0.339
Error	11	0.0003930	0.0000357		
Total	14	0.0005270			

Level	N	Mean	StDev
Th (H)	5	0.009482	0.005525
STH Th 1	4	0.002716	0.002619
STH Th 2	3	0.005927	0.007958
STH Th 3	3	0.009963	0.007864

Pooled StDev = 0.005977

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

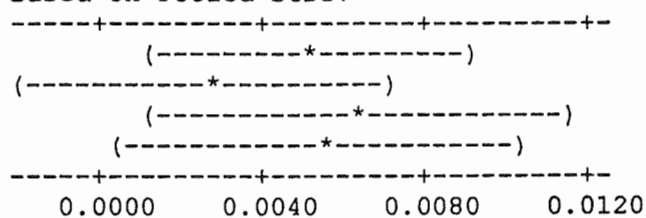
#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	0.0000306	0.0000102	0.62	0.618
Error	11	0.0001815	0.0000165		
Total	14	0.0002120			

Level	N	Mean	StDev
U (H)	5	0.005310	0.001654
STH U 1	4	0.002623	0.001938
STH U 2	3	0.006557	0.007460
STH U 3	3	0.005407	0.004897

Pooled StDev = 0.004062

#### Individual 95% CIs For Mean Based on Pooled StDev



### One-way Analysis of Variance

#### Analysis of Variance

Source	DF	SS	MS	F	P
Factor	3	1.990	0.663	1.63	0.239
Error	11	4.482	0.407		
Total	14	6.472			

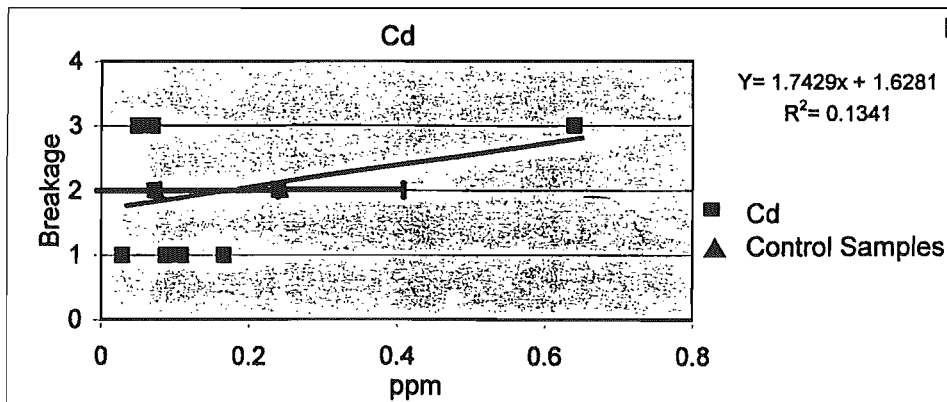
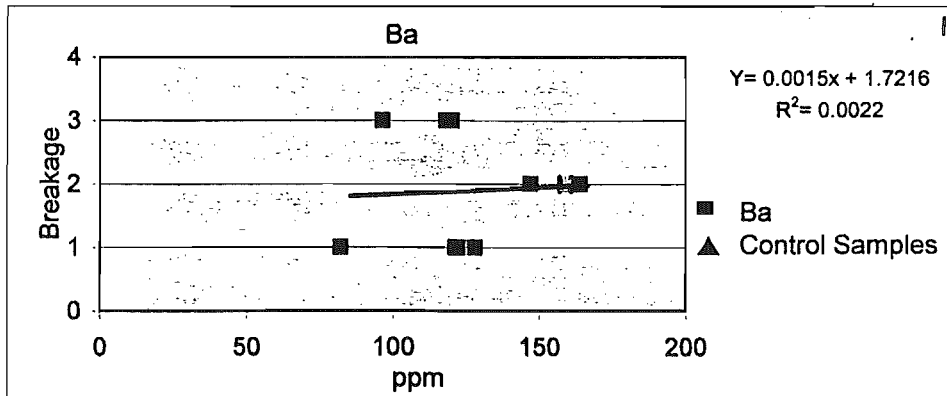
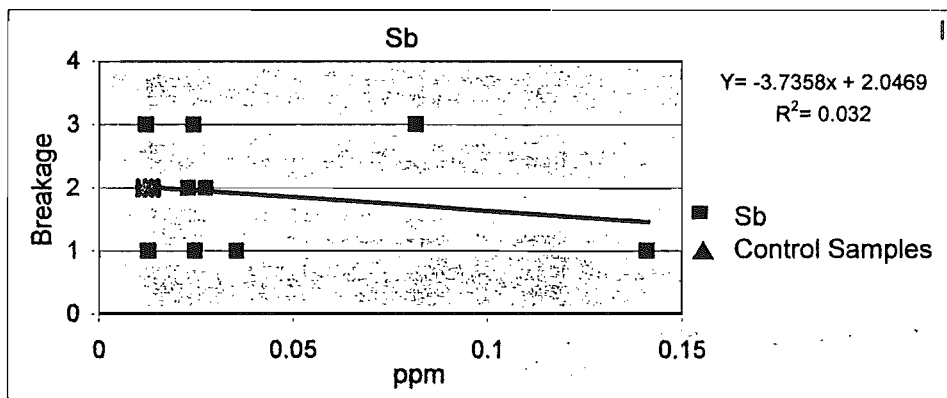
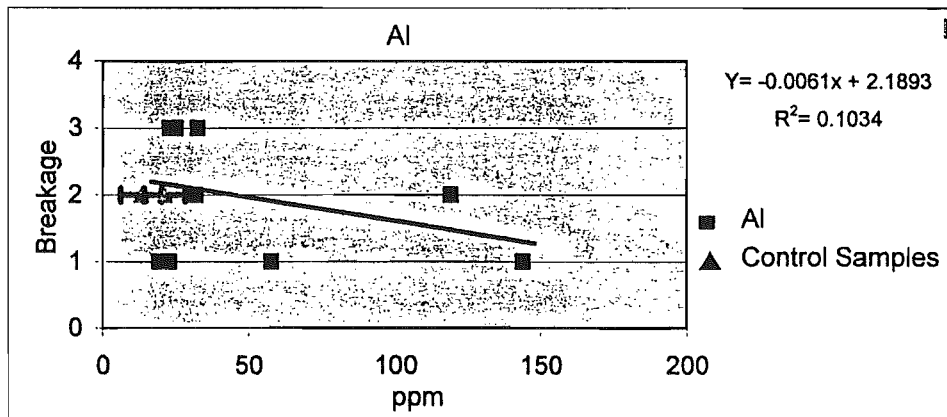
#### Individual 95% CIs For Mean Based on Pooled StDev

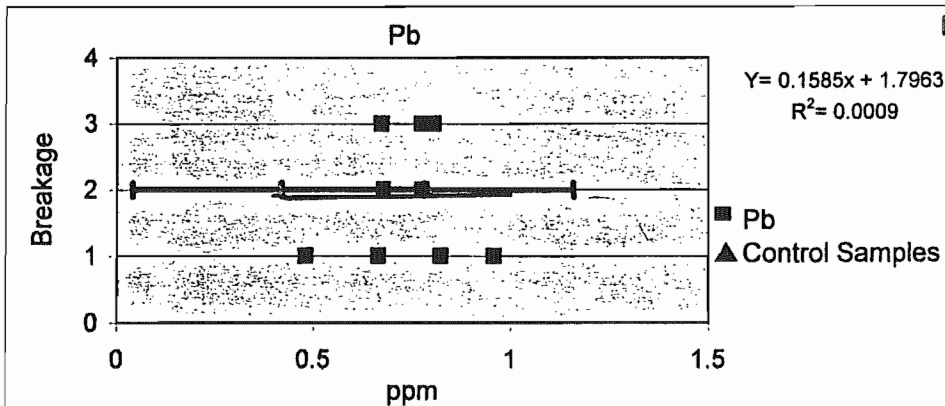
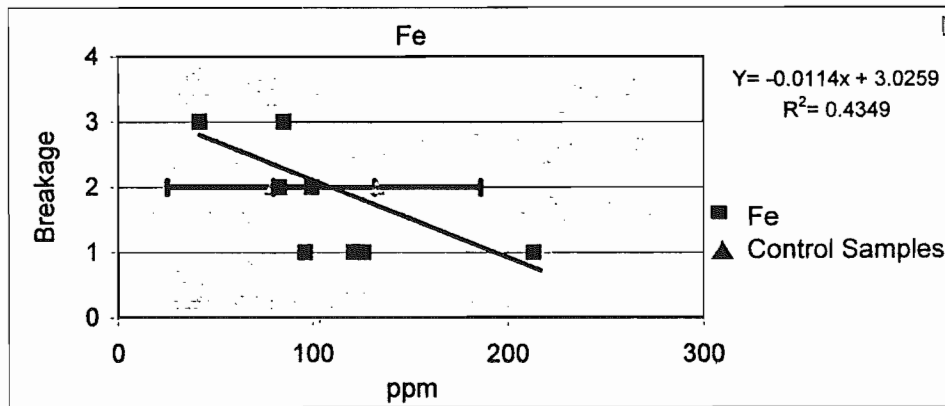
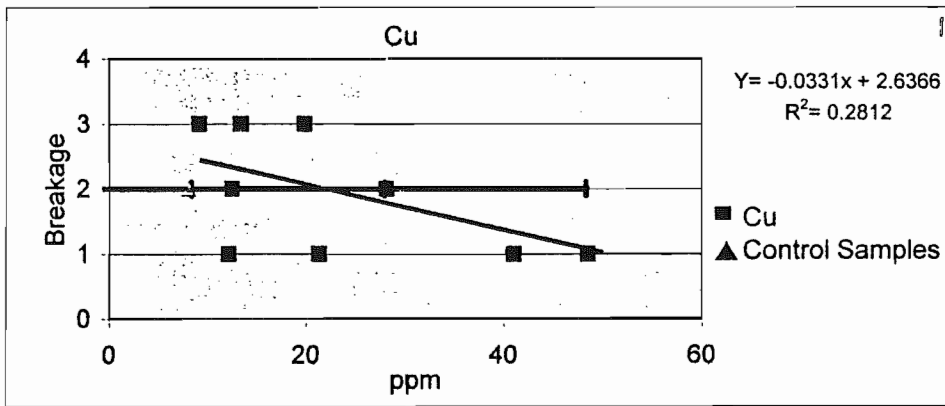
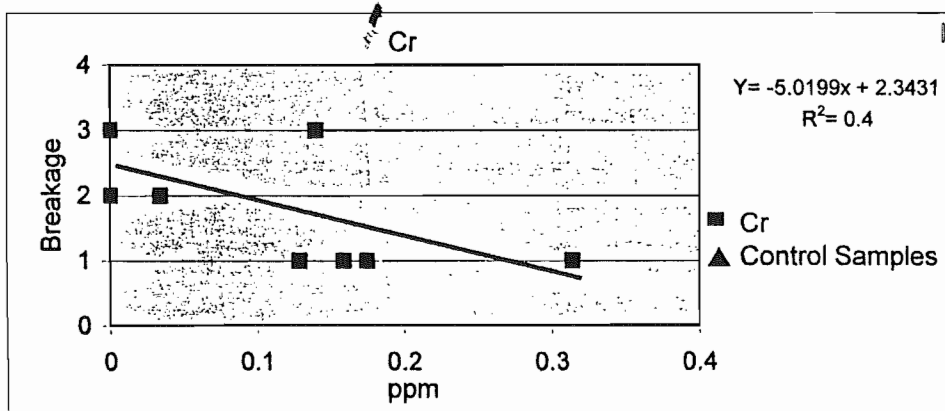
Level	N	Mean	StDev	
REE Health	5	0.9088	0.7179	(-----*-----)
REE S 1	4	0.1667	0.0742	(-----*-----)
REE S 2	3	0.2577	0.2512	(-----*-----)
REE S 3	3	0.9704	1.0672	(-----*-----)

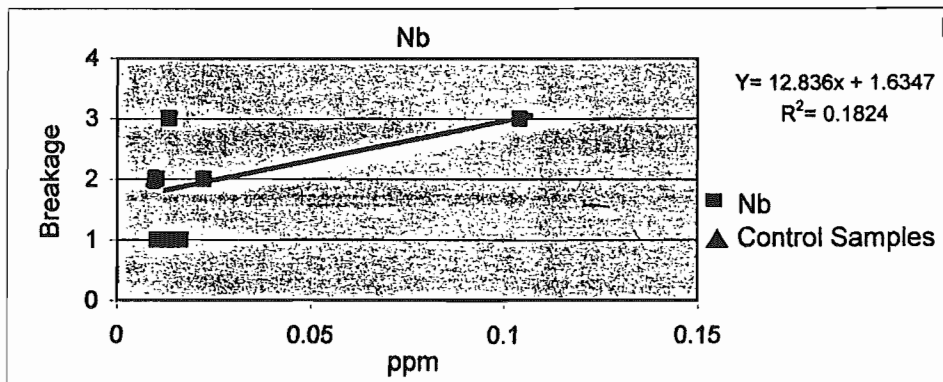
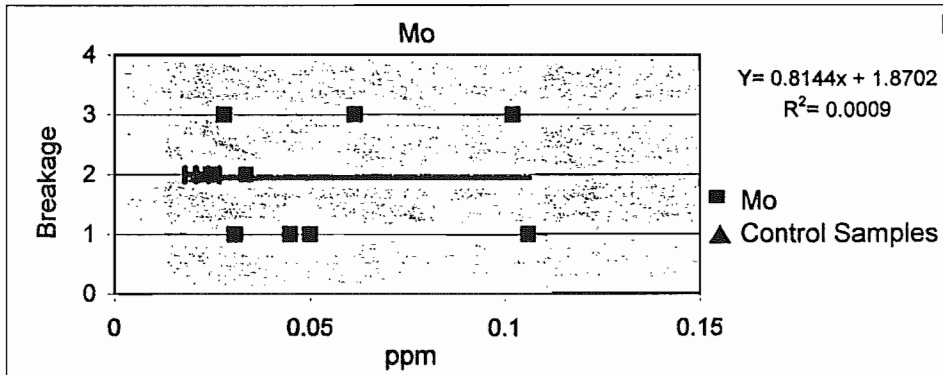
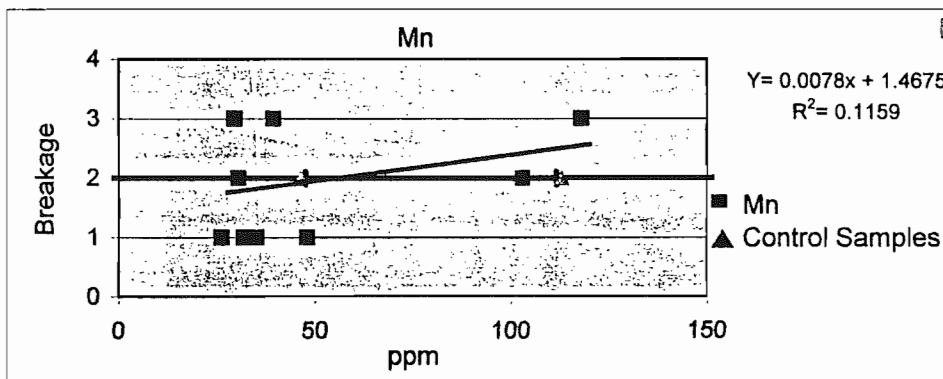
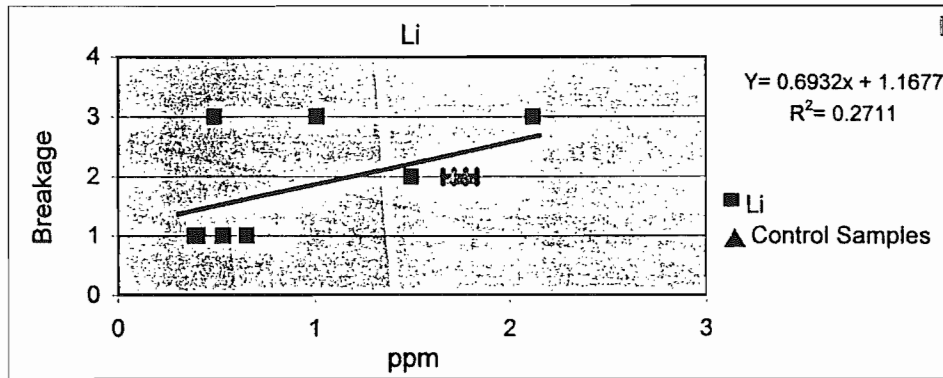
Pooled StDev = 0.6383

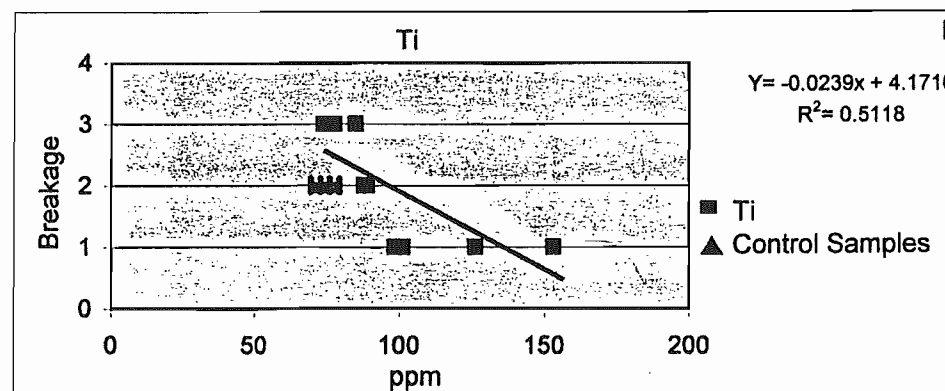
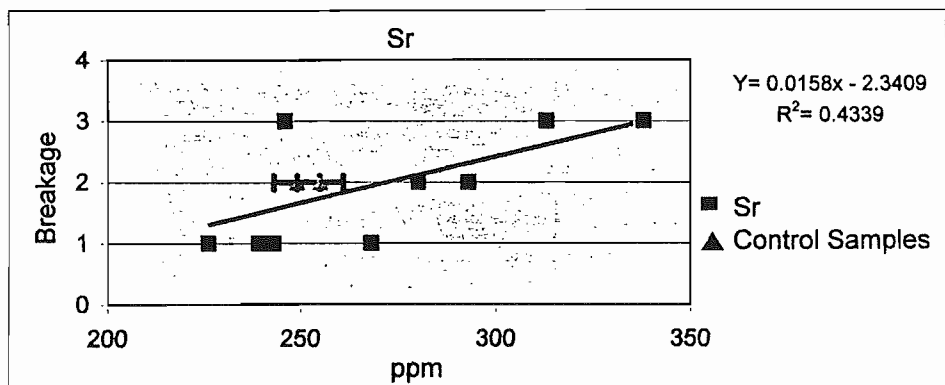
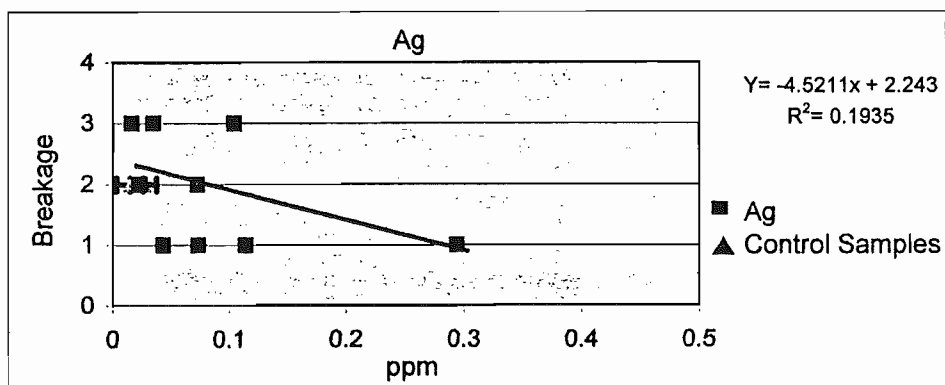
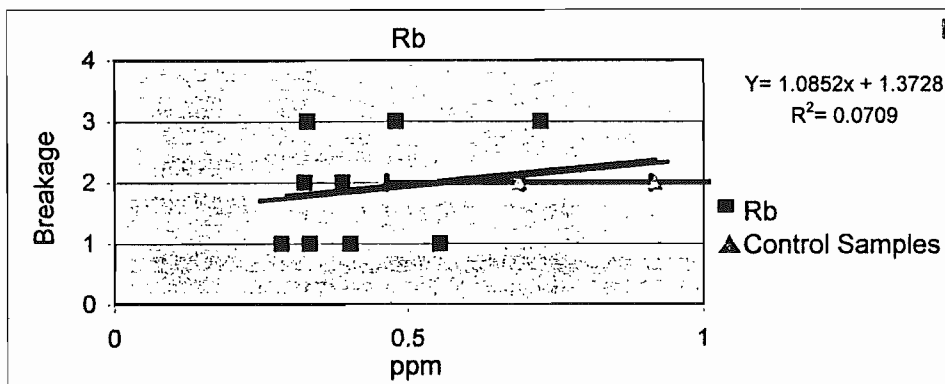
0.00      0.70      1.40

Appendix F: Correlation results for North of the Park:  
Breakage score Vs Elemental concentration  
(Control samples with 95% confidence  
included)

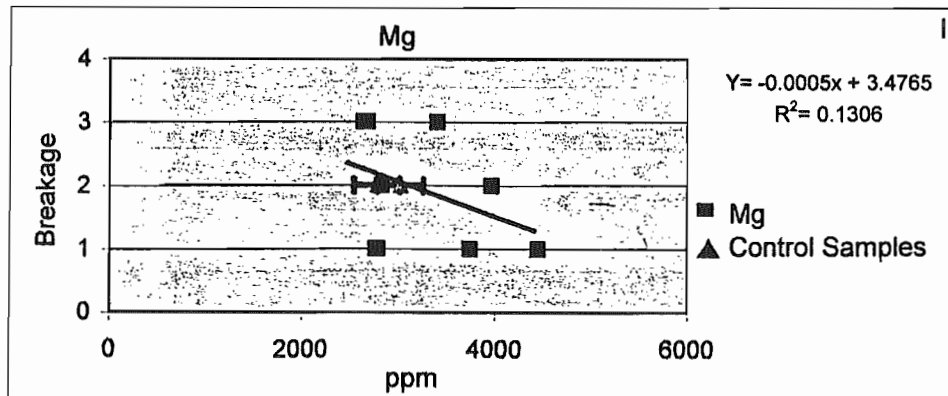
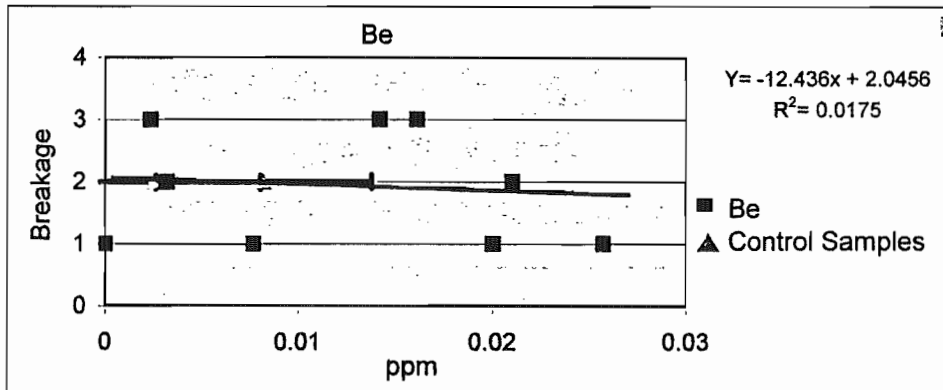
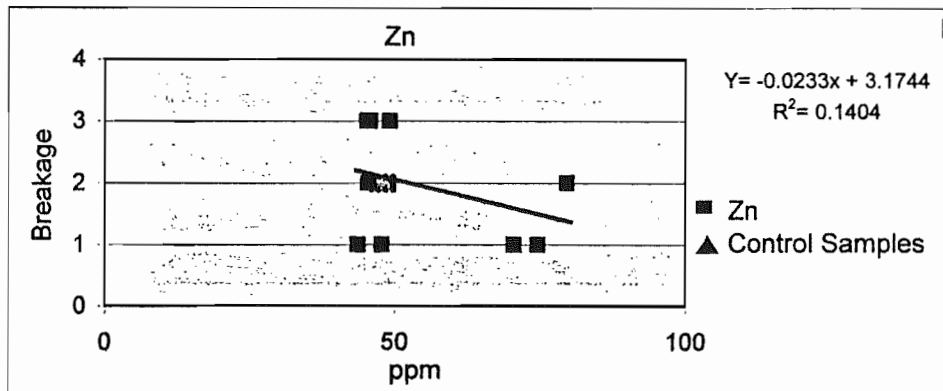
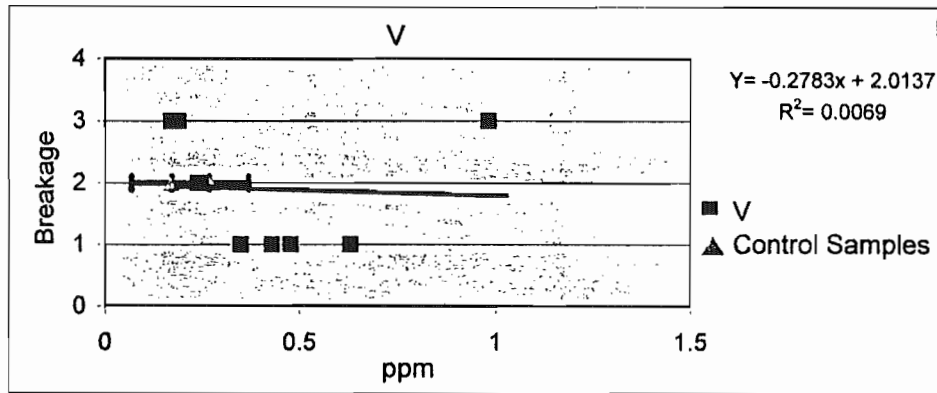


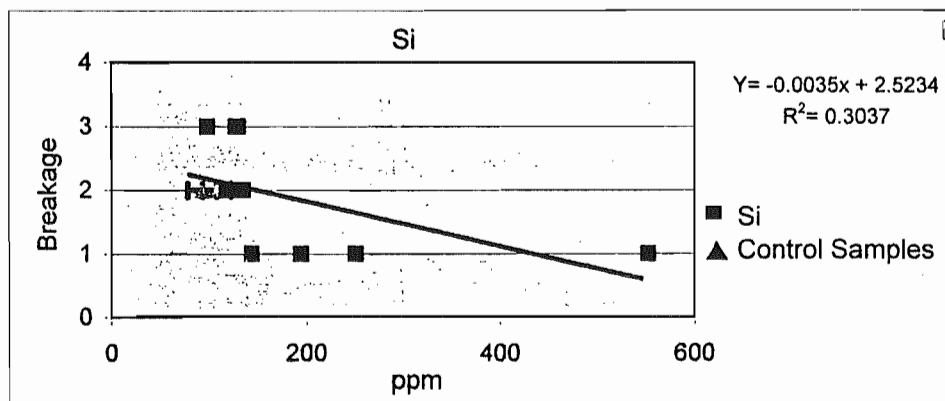


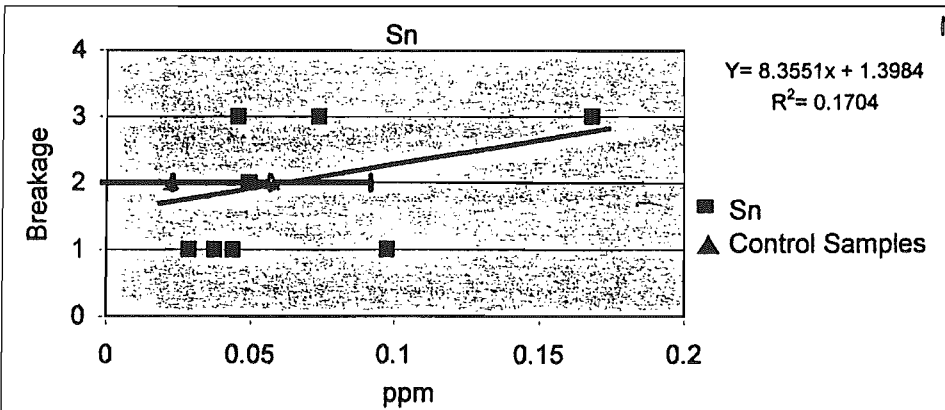
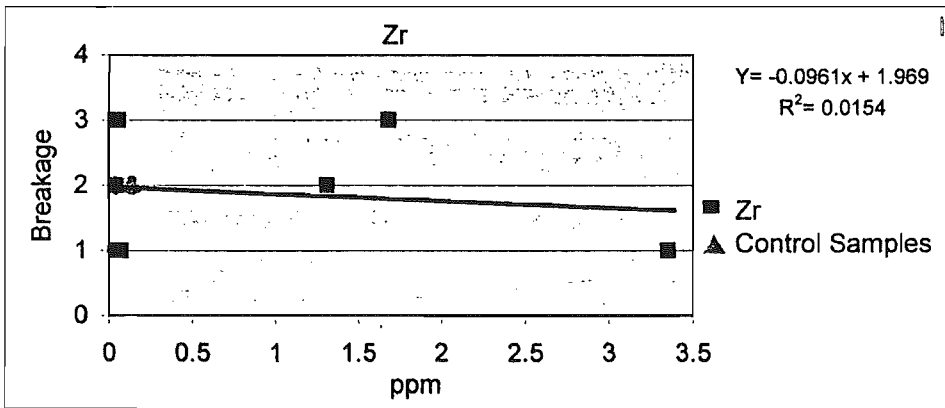
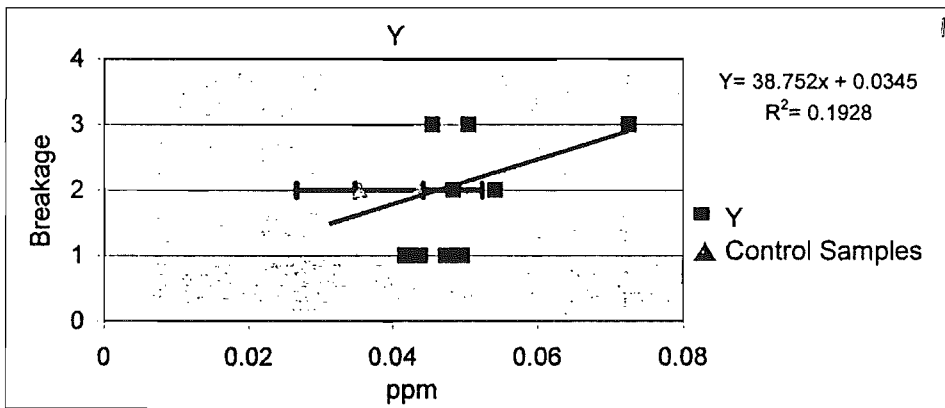
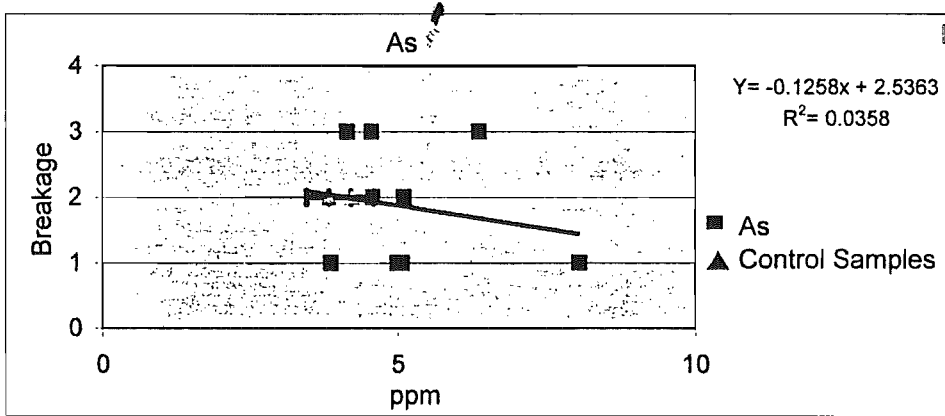


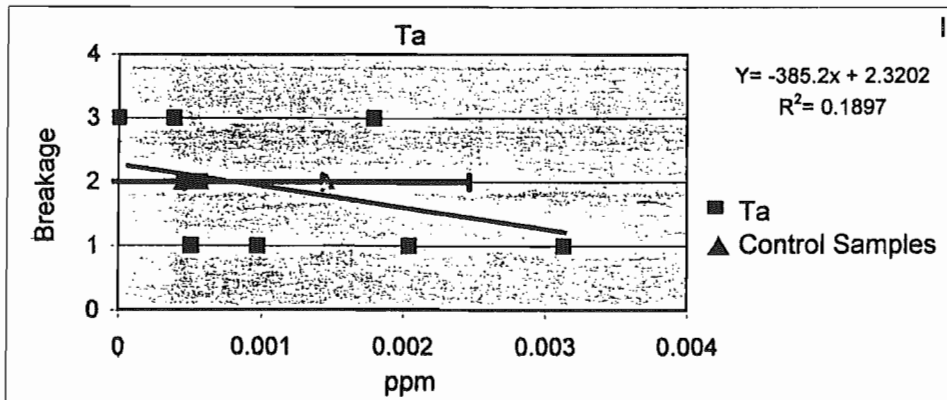
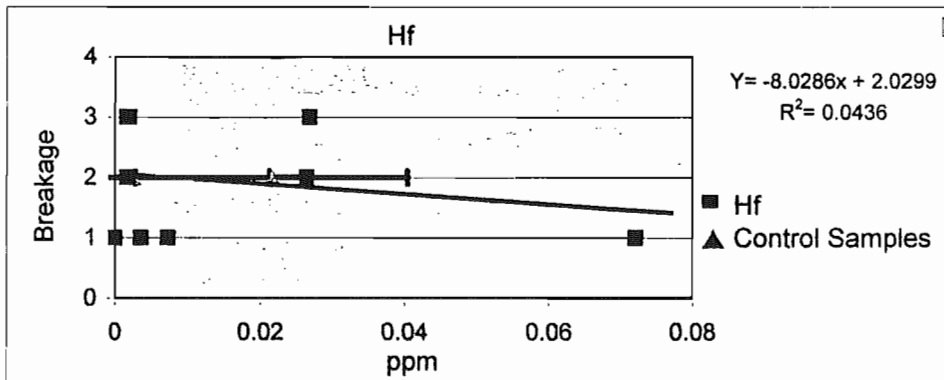
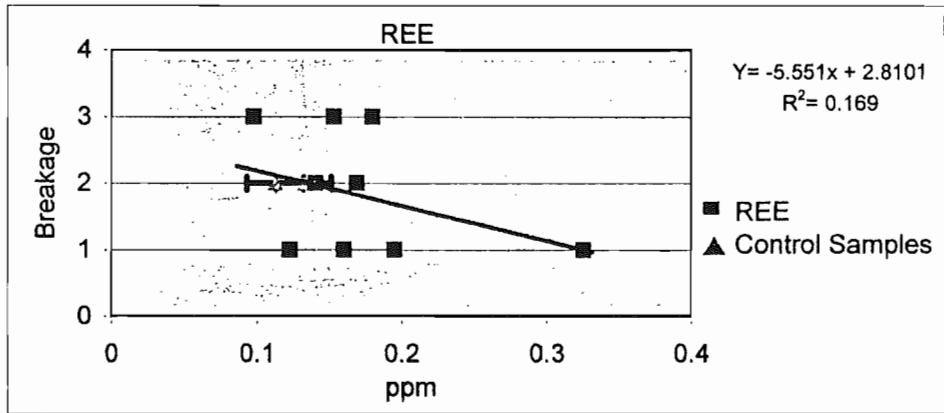
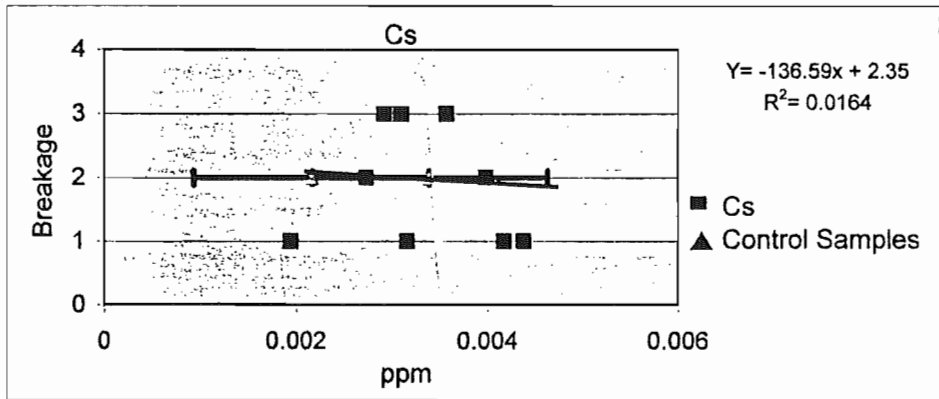


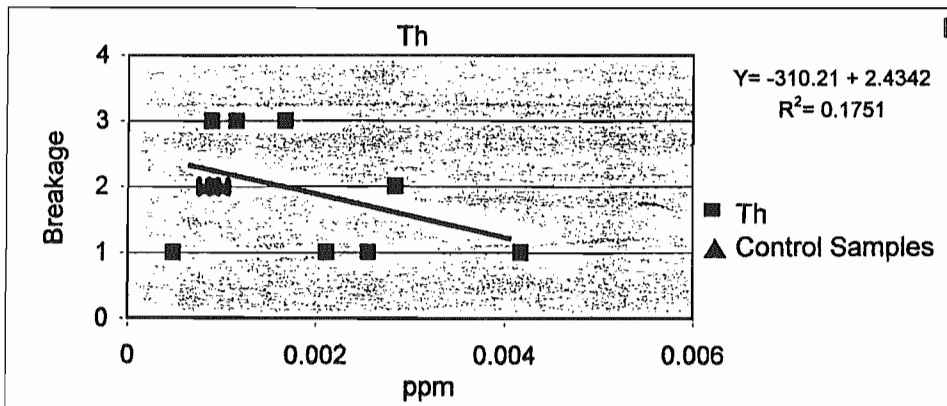
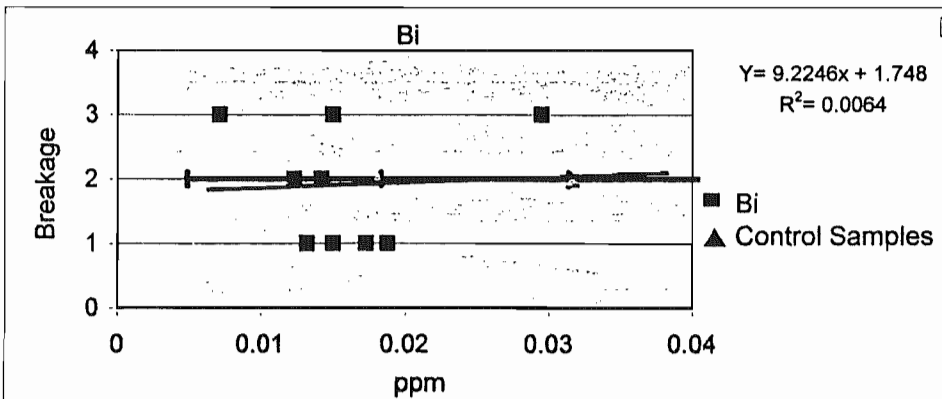
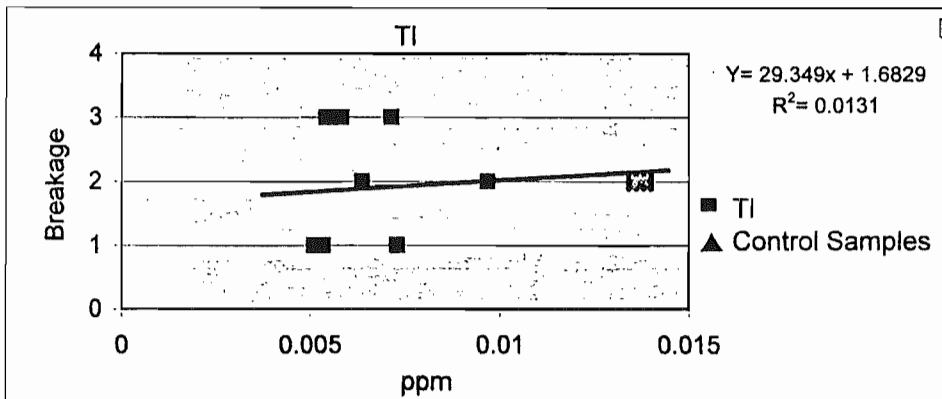
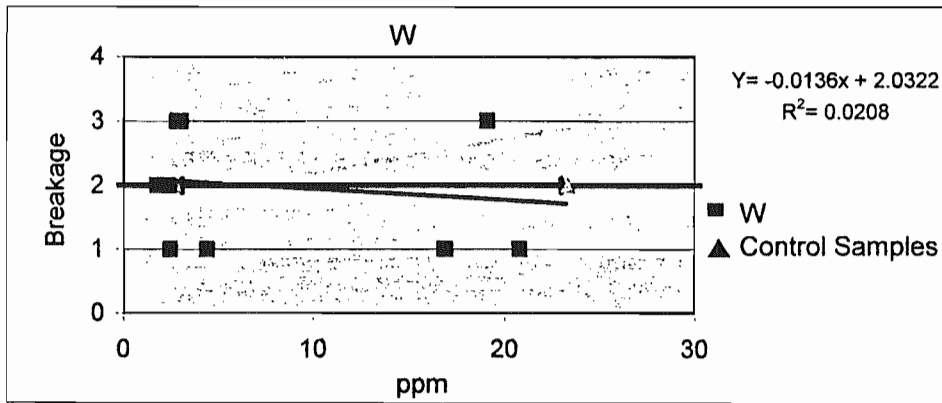


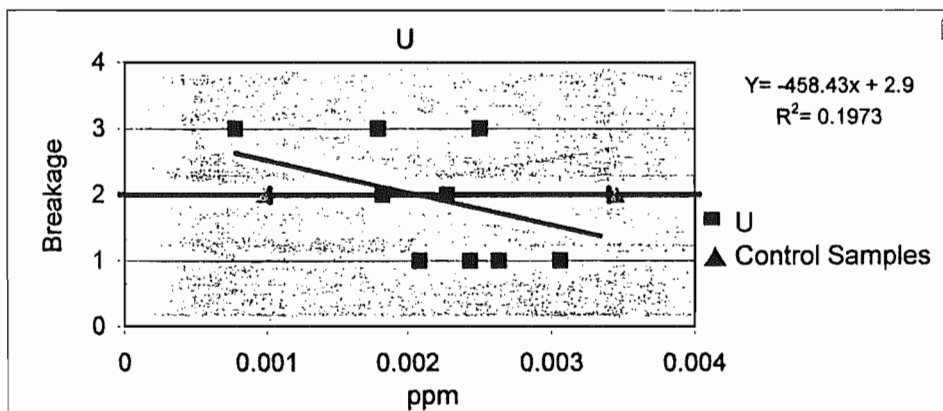




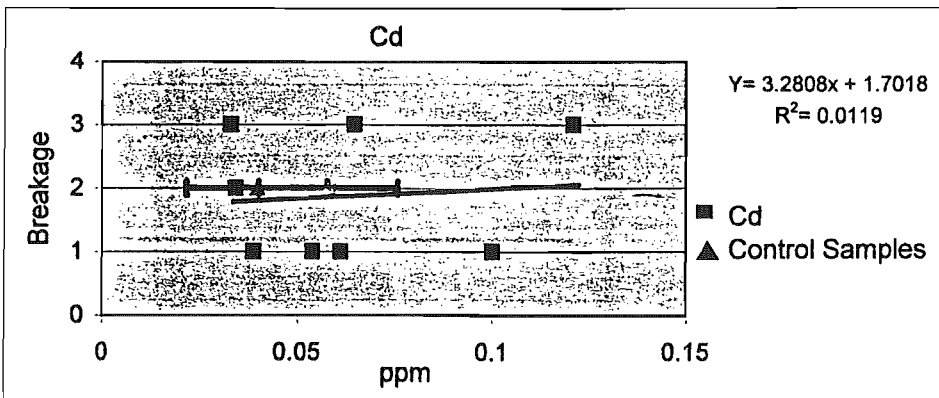
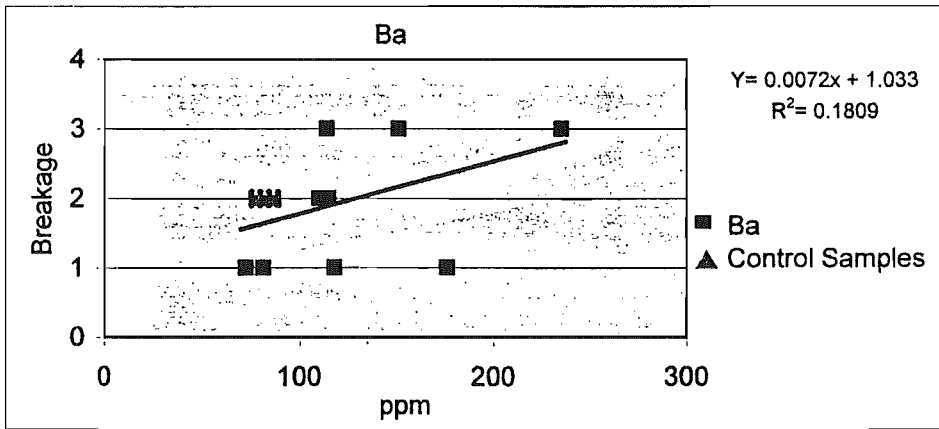
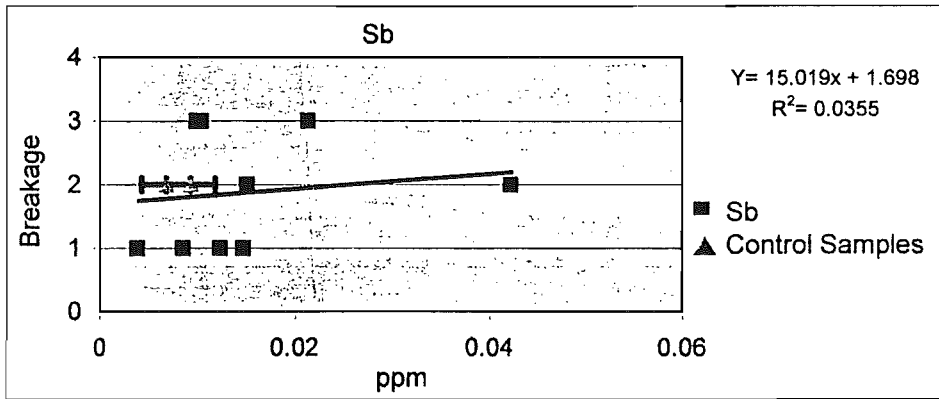
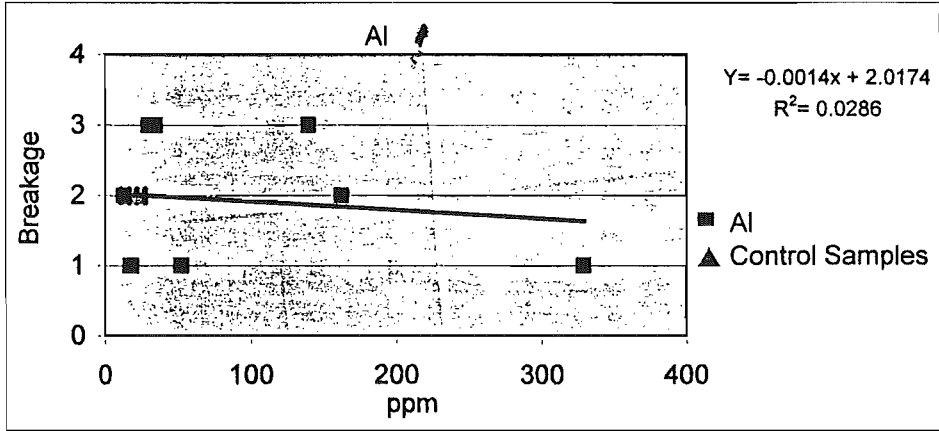






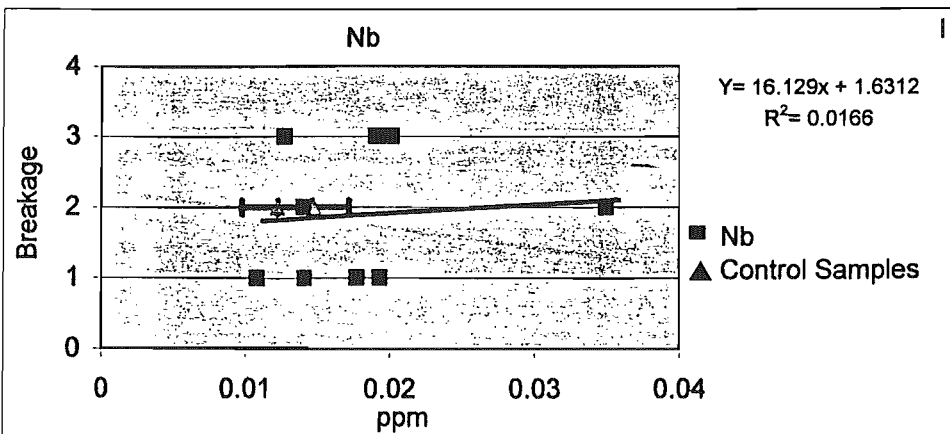
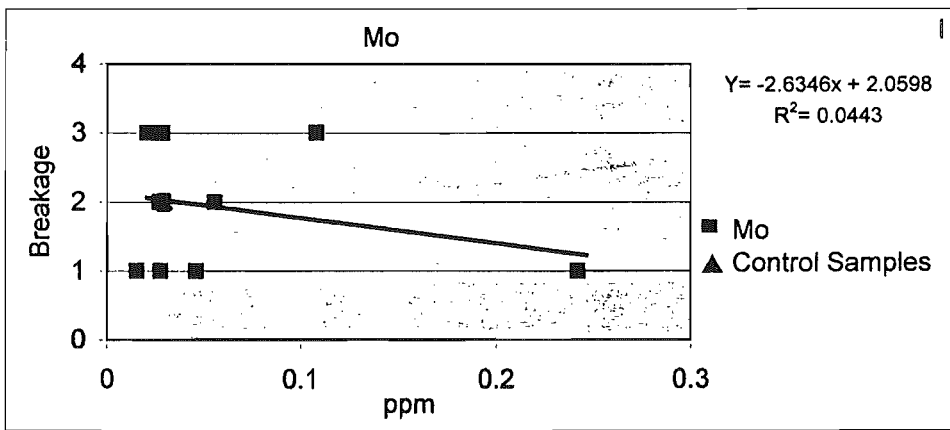
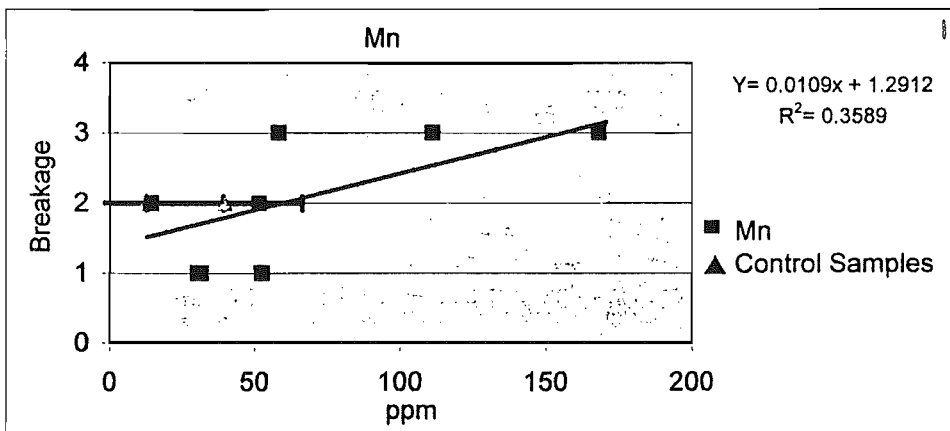
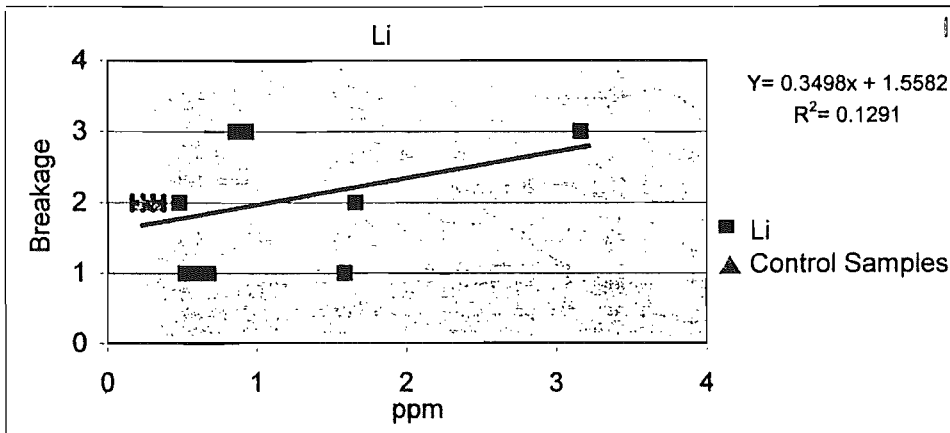


Appendix G: Correlation results for South of the Park:  
Breakage score Vs Elemental concentration  
(Control samples with 95% confidence  
included)

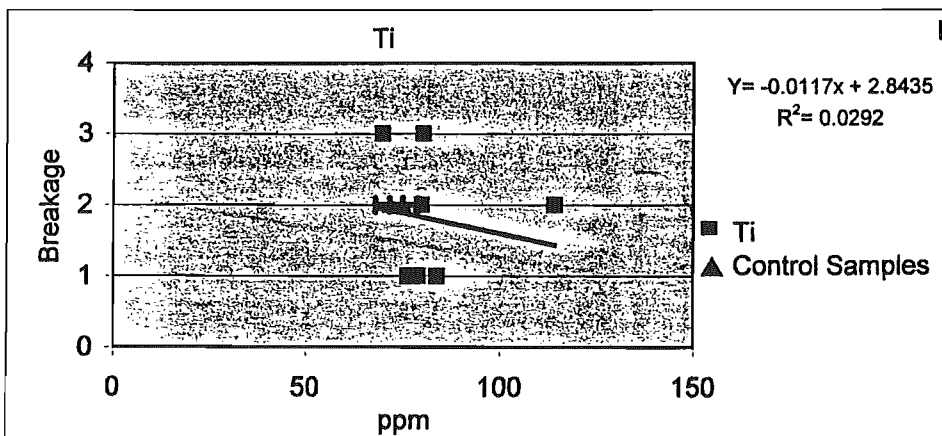
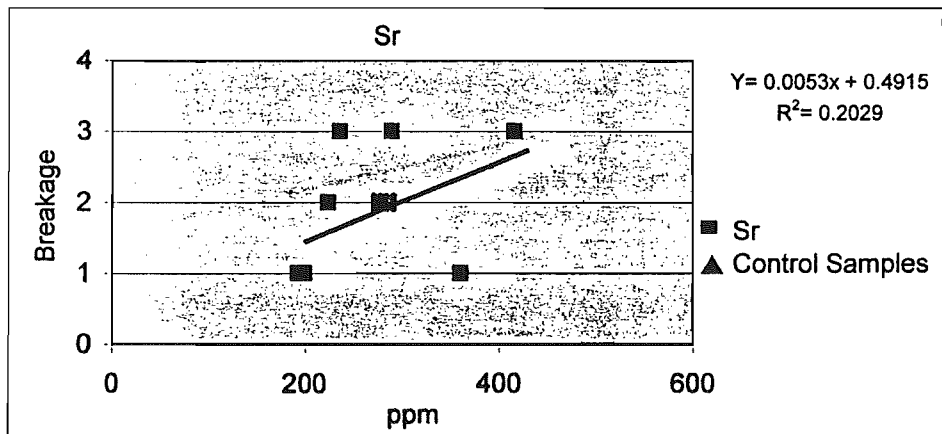
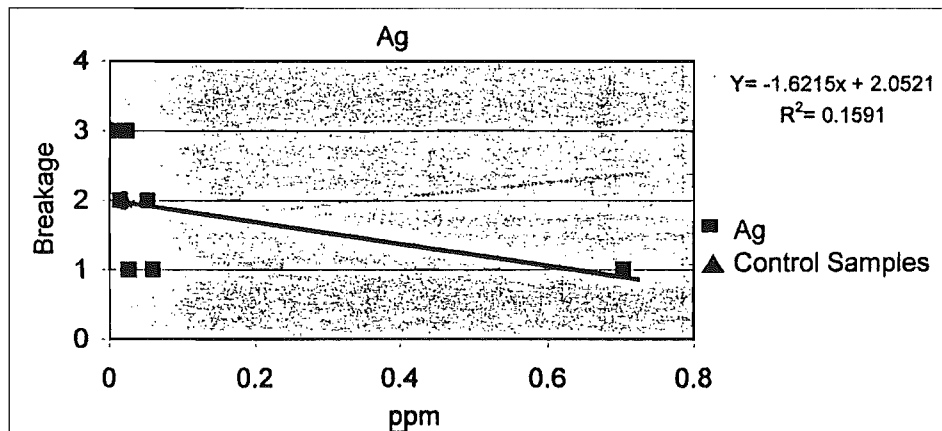
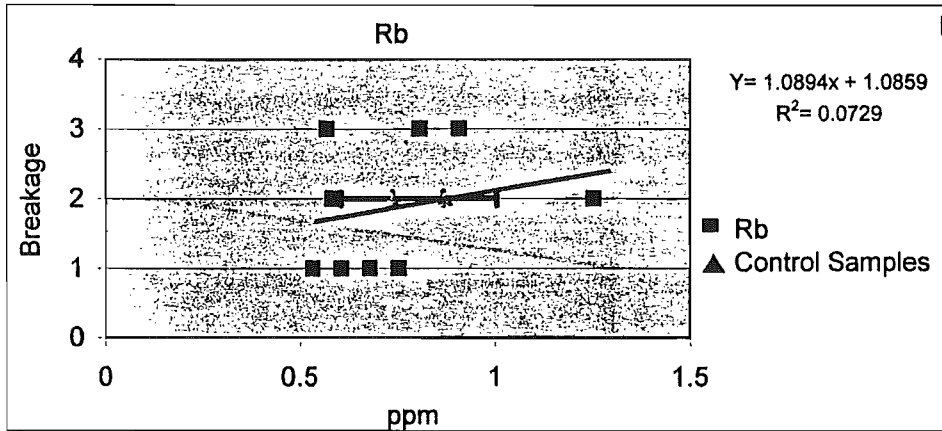




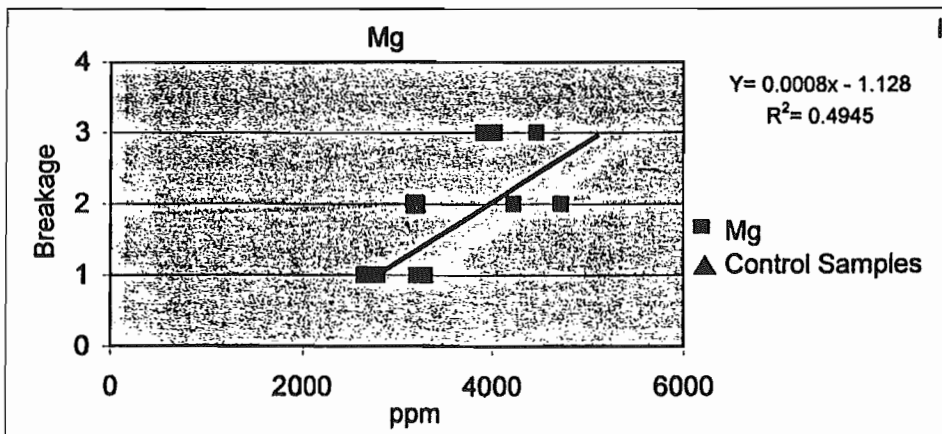
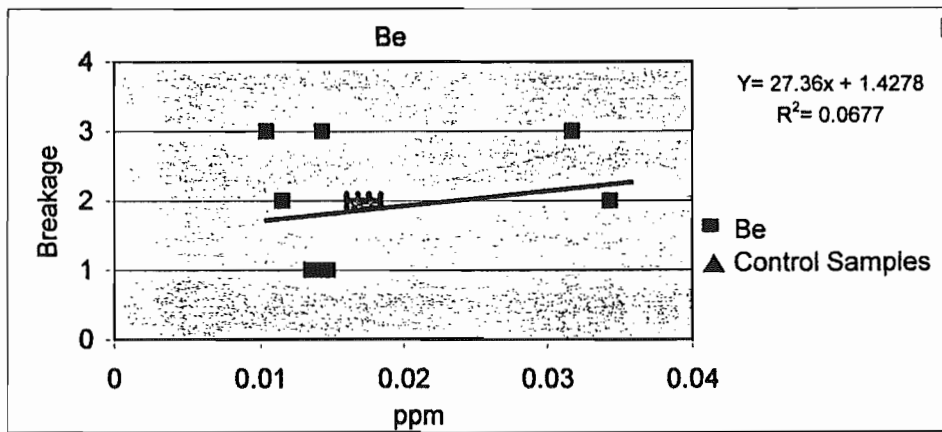
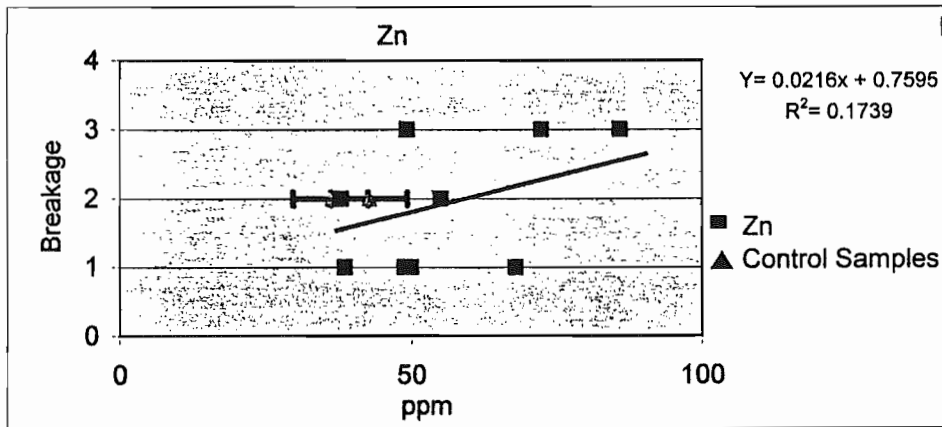
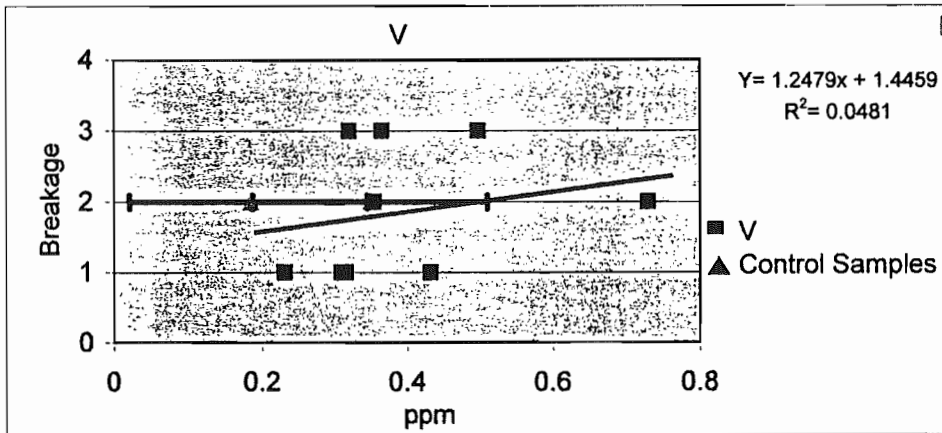




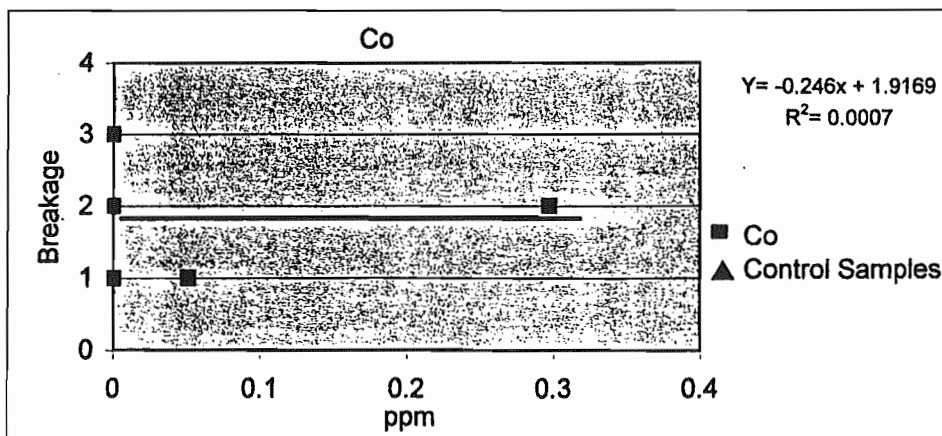
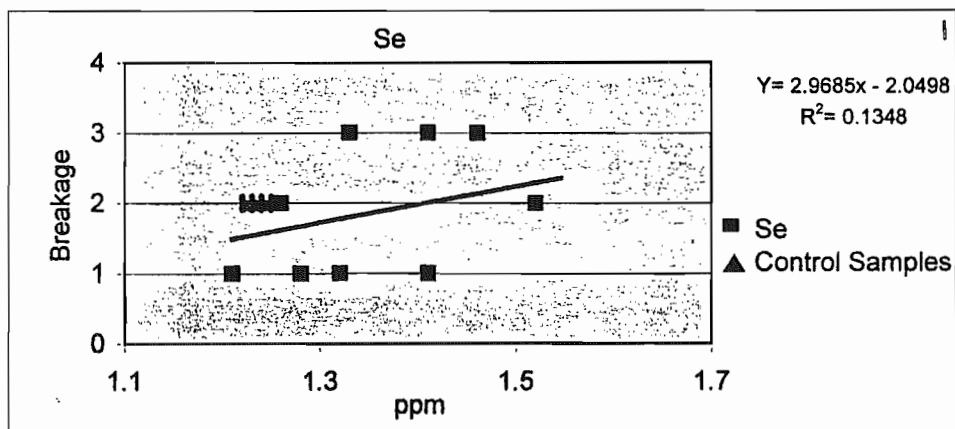
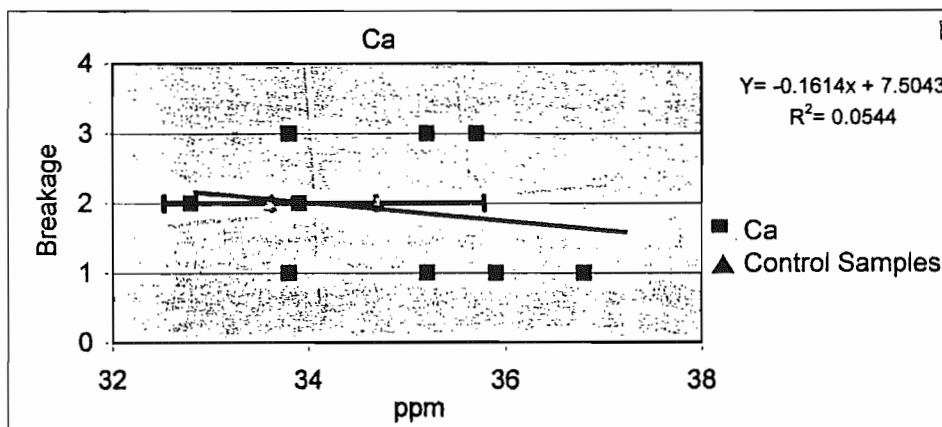
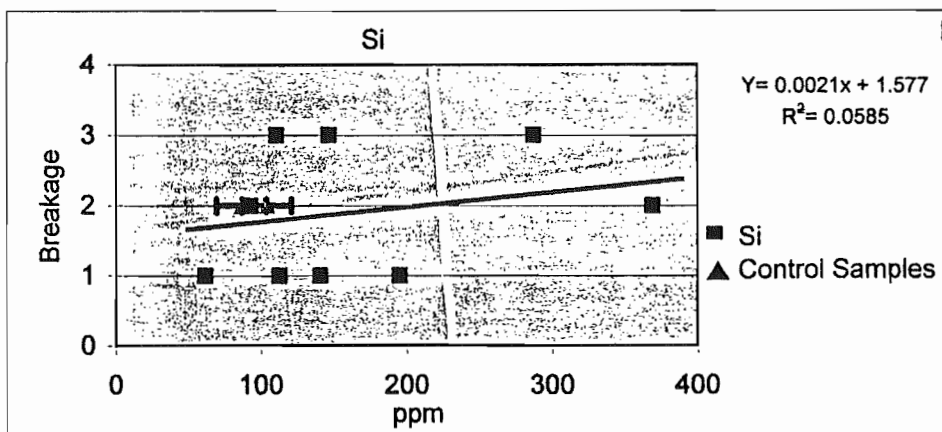
11



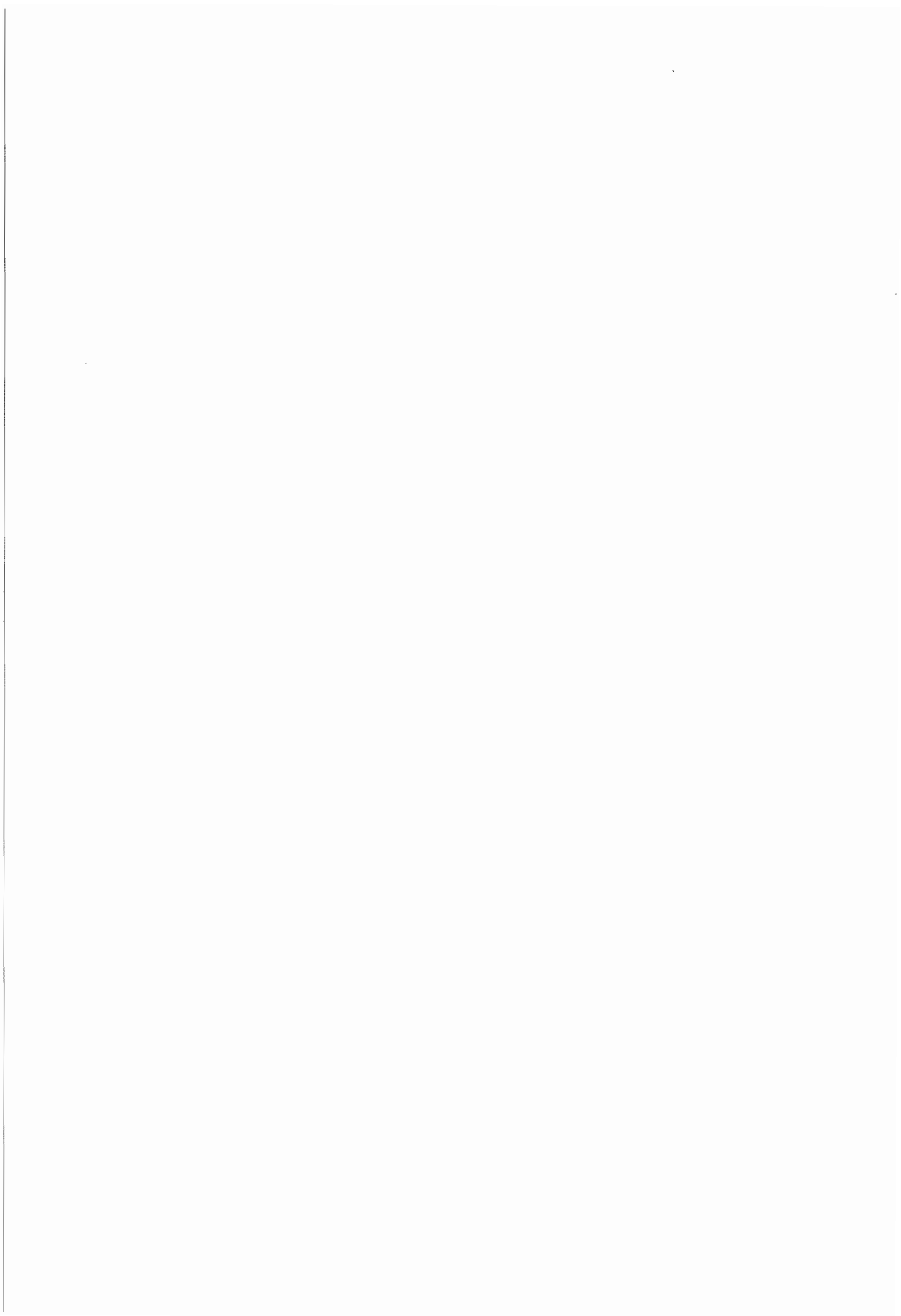


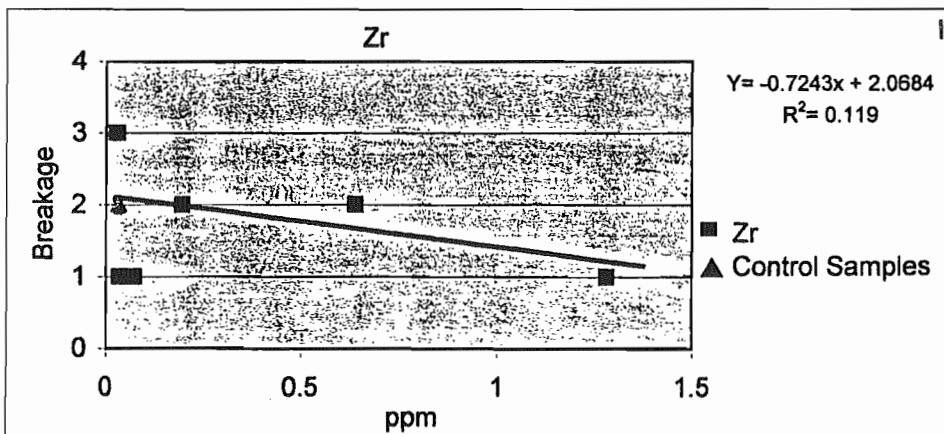
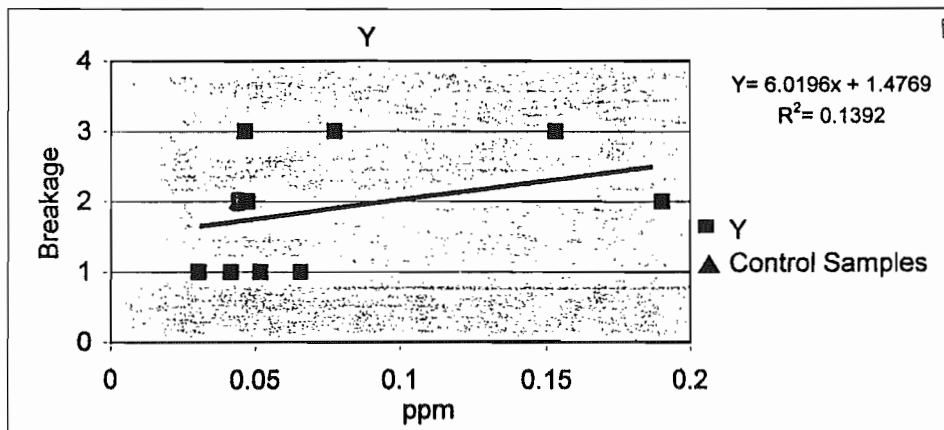
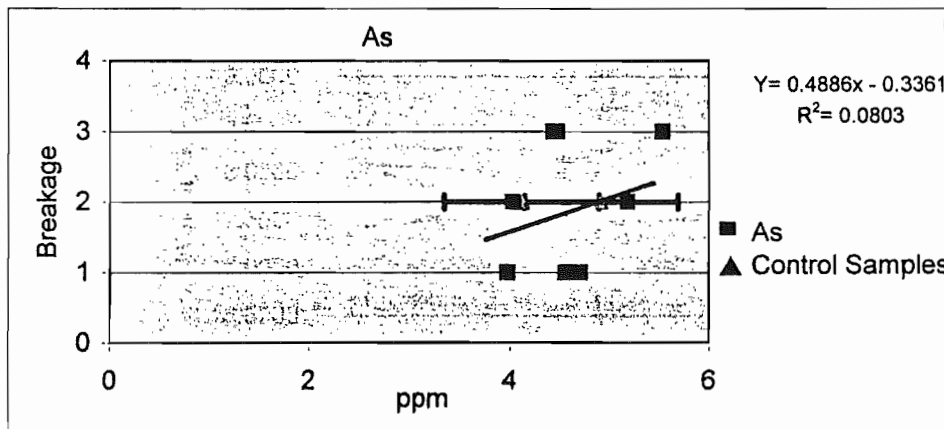
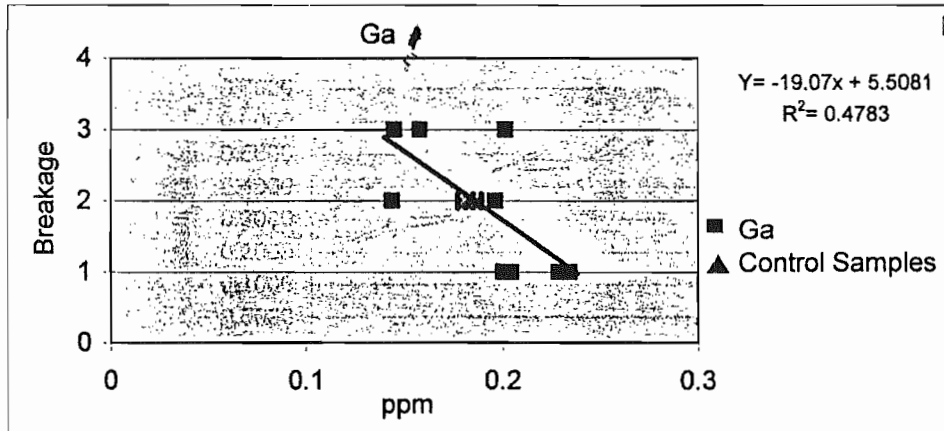




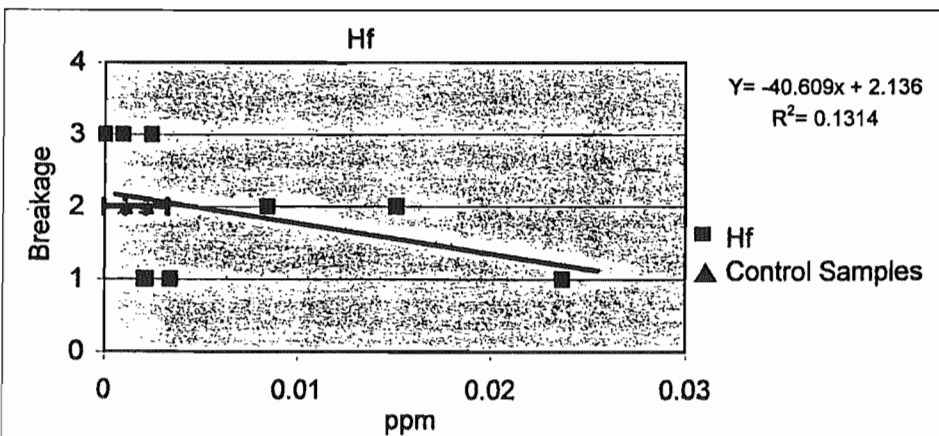
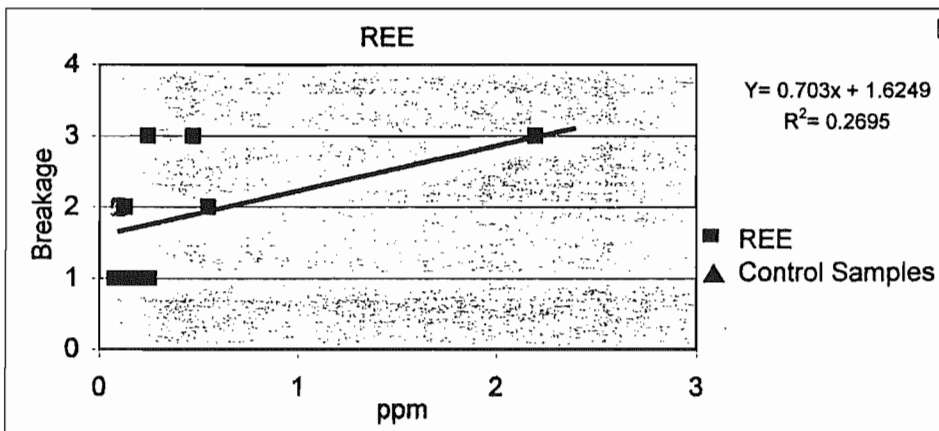
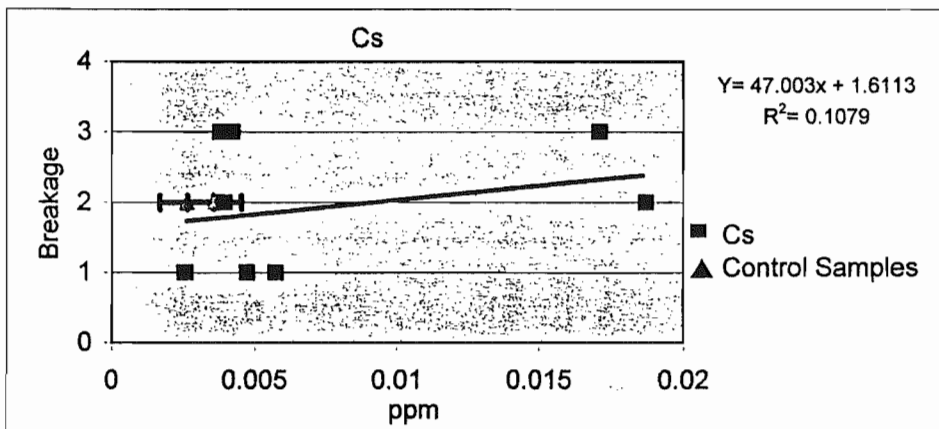
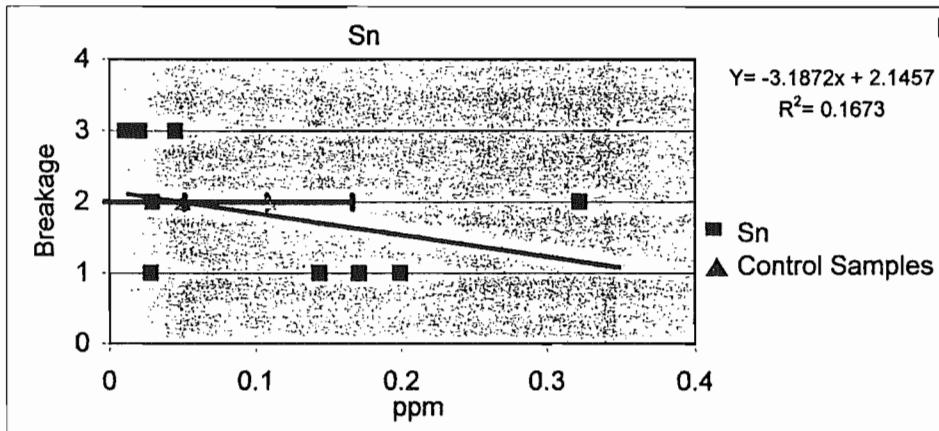


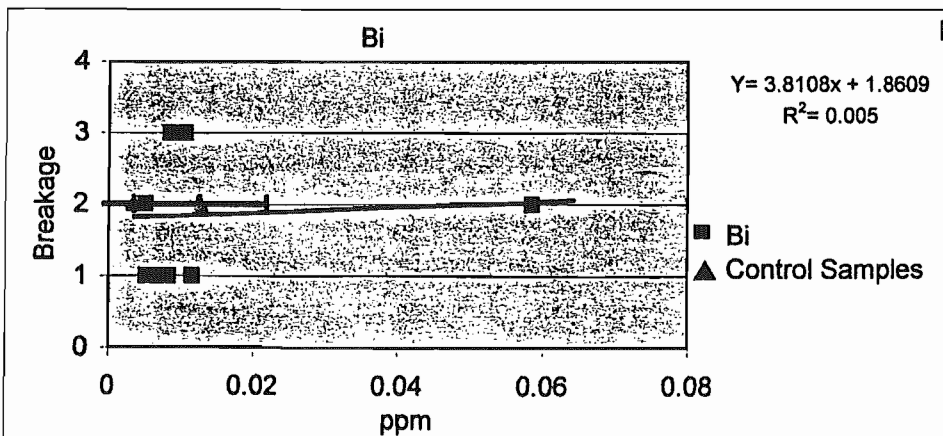
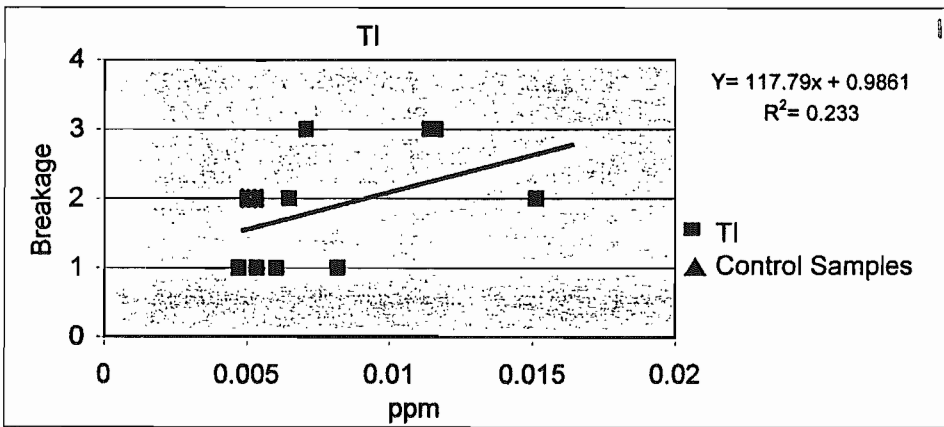
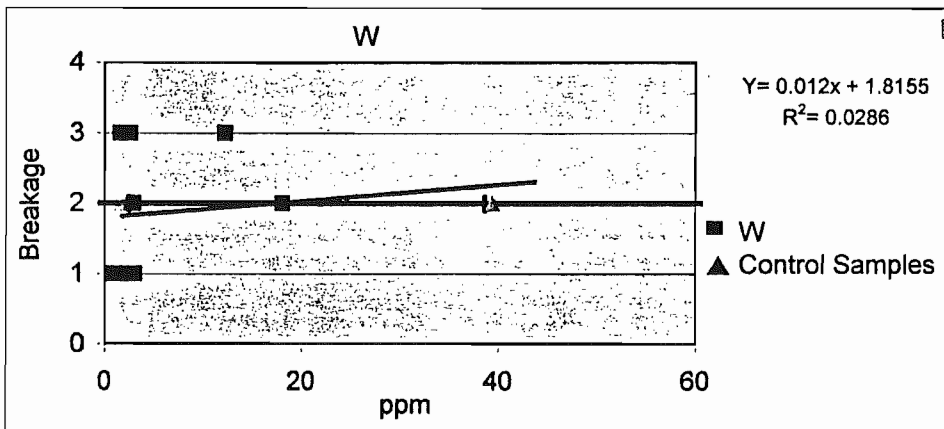
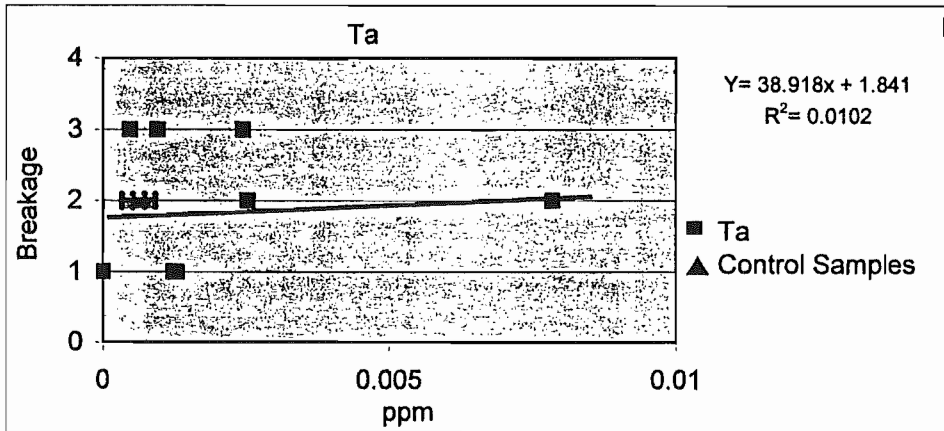














Appendix H: Microprobe data for moose teeth samples

Sample: M056

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.0939	39.7997	0.7486	0.1984	0.2233	0.0000	0.0000	48.2798	0.0187	0.0289	107.3913
18.0526	38.8891	0.6890	0.1676	0.3016	0.0000	0.0000	47.9445	0.0000	0.0447	106.0891
18.0075	38.3668	0.7109	0.1725	0.3169	0.0000	0.0000	45.6269	0.0111	0.0307	103.2432
18.4495	38.1659	0.6276	0.1450	0.4485	0.0000	0.0000	45.3068	0.0026	0.0400	103.1859
18.2682	39.7790	0.6116	0.1561	0.4627	0.0000	0.0000	46.6130	0.0161	0.0229	105.9295

Sample: M053

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.1230	38.4656	0.6136	0.2102	0.3869	0.0000	0.0000	46.1806	0.0212	0.0230	104.0241
18.3346	38.9433	0.6496	0.2207	0.3772	0.0000	0.0000	47.8889	0.0085	0.0285	106.4513
18.4903	38.8505	0.7051	0.2300	0.3781	0.0000	0.0000	47.9501	0.0255	0.0406	106.6701
18.1579	37.6285	0.6492	0.2093	0.3572	0.0000	0.0000	45.7623	0.0281	0.0295	102.8220
18.1689	38.9375	0.6525	0.2149	0.3573	0.0000	0.0000	47.7674	0.0127	0.0258	106.1369
18.2758	38.1379	0.6795	0.2217	0.3581	0.0000	0.0000	45.8613	0.0060	0.0480	103.5882

Sample: M052

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.3896	37.5155	0.7362	0.1674	0.2437	0.0000	0.0000	47.3534	0.0145	0.0174	104.4376
18.2155	38.0838	0.6927	0.1701	0.3114	0.0000	0.0000	46.2920	0.0068	0.0378	103.8101
18.3133	38.8030	0.6613	0.1819	0.3375	0.0000	0.0000	47.3295	0.0102	0.0323	105.6690
18.1650	38.5022	0.6369	0.1712	0.4075	0.0000	0.0000	46.6680	0.0127	0.0137	104.5772
18.0183	38.6541	0.6060	0.1494	0.4713	0.0000	0.0000	46.1876	0.0076	0.0341	104.1283
18.4279	38.1244	0.6042	0.1216	0.5509	0.0000	0.0000	45.7362	0.0145	0.0239	103.6036

Sample: M051

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
17.3373	34.2998	0.9727	0.2020	0.1265	0.0000	0.0000	50.5963	0.0338	0.0307	103.5990
17.7384	39.3477	0.7455	0.1513	0.3091	0.0000	0.0000	48.8035	0.0177	0.0230	107.1361
17.5904	39.1714	0.7871	0.1593	0.3815	0.0000	0.0000	48.3569	0.0025	0.0157	106.4648
17.5511	39.4119	0.6899	0.1709	0.4230	0.0000	0.0000	48.1471	0.0059	0.0515	106.4512
17.7509	39.2512	0.6895	0.1345	0.4312	0.0000	0.0000	48.0650	0.0168	0.0157	106.3547
17.9104	39.4720	0.6506	0.1366	0.5042	0.0000	0.0000	47.9336	0.0160	0.0451	106.6684



**Sample: M042**

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
17.3939	39.1049	0.7285	0.2177	0.3874	0.0000	0.0000	48.1745	0.0084	0.0414	106.0566
17.5297	38.7635	0.6607	0.2477	0.3702	0.0000	0.0000	47.8456	0.0219	0.0368	105.4760
17.4064	38.4961	0.7023	0.3034	0.3637	0.0000	0.0000	47.6162	0.0143	0.0414	104.9437
17.1155	39.1212	0.6392	0.2606	0.3486	0.0000	0.0000	48.8260	0.0160	0.0203	106.3473
17.2585	38.8647	0.6548	0.2687	0.3118	0.0000	0.0000	48.6871	0.0118	0.0212	106.0785
16.8830	38.5418	0.6544	0.2267	0.3085	0.0000	0.0000	46.9347	0.0403	0.0359	103.6252

**Sample: M027**

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
17.4194	38.7963	0.7556	0.1390	0.2197	0.0000	0.0000	47.9897	0.0235	0.0295	105.3727
17.4865	38.9408	0.7206	0.1322	0.2735	0.0000	0.0000	47.7947	0.0319	0.0359	105.4160
17.7179	38.9774	0.6864	0.1859	0.3034	0.0000	0.0000	47.2601	0.0177	0.0341	105.1829
17.6059	39.3754	0.6428	0.2192	0.3477	0.0000	0.0000	47.5507	0.0294	0.0487	105.8197
17.4217	39.5122	0.6450	0.1635	0.4340	0.0000	0.0000	47.0013	0.0277	0.0542	105.2596
17.5663	39.7118	0.4901	0.1271	0.5560	0.0000	0.0000	47.1836	0.0302	0.0423	105.7073

**Sample: M041**

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
17.6538	39.0005	0.6364	0.1716	0.4130	0.0000	0.0000	46.8756	0.0084	0.0230	104.7822
17.4204	38.6509	0.6576	0.1781	0.4099	0.0000	0.0000	46.6383	0.0059	0.0148	103.9759
18.2251	39.2061	0.6275	0.2001	0.3907	0.0000	0.0000	48.5331	0.0008	0.0257	107.2091
18.6780	38.6133	0.7226	0.1881	0.3630	0.0000	0.0000	47.0886	0.0152	0.0156	105.6843
18.5156	39.0053	0.7059	0.1933	0.3705	0.0000	0.0000	47.9442	0.0000	0.0018	106.7365
18.1299	38.2186	0.6840	0.2083	0.3466	0.0000	0.0000	46.7339	0.0008	0.0138	104.3359

**Sample: M016**

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.4141	38.7299	0.7544	0.1787	0.3108	0.0000	0.0000	46.9951	0.0303	0.0395	105.4527
18.0656	38.3061	0.7265	0.1931	0.3026	0.0000	0.0000	47.7074	0.0269	0.0257	105.3539
18.5588	38.8143	0.7075	0.1739	0.3116	0.0000	0.0000	47.6145	0.0211	0.0377	106.2393
18.0868	38.2046	0.7171	0.1831	0.3033	0.0000	0.0000	47.0495	0.0210	0.0460	104.6114
18.2201	38.4662	0.7476	0.1883	0.3144	0.0000	0.0000	47.8515	0.0177	0.0193	105.8251
18.3106	38.4270	0.7517	0.1829	0.3116	0.0000	0.0000	47.6462	0.0084	0.0340	105.6723

Sample: M012

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.0747	38.0391	0.6676	0.2125	0.3512	0.0000	0.0000	47.4662	0.0093	0.0487	104.8692
17.8800	38.0776	0.6455	0.2067	0.3801	0.0000	0.0000	46.3972	0.0000	0.0202	103.6072
18.5623	38.8417	0.6670	0.2227	0.4090	0.0000	0.0000	47.3173	0.0000	0.0460	106.0659
18.0782	38.7859	0.6217	0.2123	0.4855	0.0000	0.0000	46.1206	0.0000	0.0505	104.3546
18.3084	37.9846	0.5980	0.2106	0.4761	0.0000	0.0000	46.7203	0.0135	0.0368	104.3482
18.5416	38.8380	0.5081	0.1373	0.6238	0.0000	0.0000	45.9613	0.0000	0.0211	104.6311

Sample: M022

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.9018	38.1494	0.7089	0.4057	0.2956	0.0000	0.0000	47.2216	0.0000	0.0276	105.7105
18.6642	38.3168	0.7016	0.2872	0.3282	0.0000	0.0000	47.2569	0.0000	0.0230	105.5779
18.3521	37.8213	0.6816	0.2593	0.3141	0.0000	0.0000	47.3135	0.0008	0.0101	104.7527
18.3786	38.3922	0.6559	0.2692	0.3220	0.0000	0.0000	48.5694	0.0000	0.0230	106.6102
18.7003	34.4399	0.6770	0.2496	0.3408	0.0000	0.0000	48.3430	0.0076	0.0105	102.7686
18.2386	38.7169	0.7063	0.3435	0.3491	0.0000	0.0000	46.6486	0.0126	0.0370	105.0525

Sample: M030

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
17.9329	38.7237	0.7587	0.2413	0.2328	0.0000	0.0000	47.4801	0.0000	0.0241	105.3935
17.4484	37.6343	0.7456	0.2218	0.2737	0.0000	0.0000	44.6506	0.0034	0.0553	101.0331
17.3972	36.3000	0.7600	0.2182	0.2509	0.0000	0.0000	46.2872	0.0135	0.0260	101.2529
14.7926	28.6072	0.6839	0.1994	0.2081	0.0000	0.0000	45.1165	0.0000	0.0000	89.6077
16.1659	31.2796	0.6696	0.2185	0.2428	0.0000	0.0000	45.4179	0.0000	0.0253	94.0197
18.3265	38.4317	0.7039	0.2944	0.3269	0.0000	0.0000	46.4043	0.0000	0.0242	104.5118

Sample: M003

Ca	Na	Mg	Cl	N	F	O	S	K	Total	
18.7619	39.1338	0.6957	0.1752	0.4362	0.0000	0.0000	46.6895	0.0101	0.0747	105.9771
18.5867	39.0519	0.6984	0.1690	0.4062	0.0000	0.0000	46.8238	0.0126	0.0196	105.7681
18.7917	39.1456	0.6707	0.1777	0.4269	0.0000	0.0000	46.5465	0.0042	0.0526	105.8158
17.1347	35.2549	0.7041	0.1512	0.3550	0.0000	0.0000	46.2802	0.0000	0.0463	99.9265
17.5874	35.6373	0.6975	0.1649	0.3747	0.0000	0.0000	45.4832	0.0000	0.0583	100.0032
18.4031	39.0671	0.7034	0.1847	0.4098	0.0000	0.0000	46.1472	0.0008	0.0370	104.9530

Appendix I: Plotted Graphs for Microprobe Data



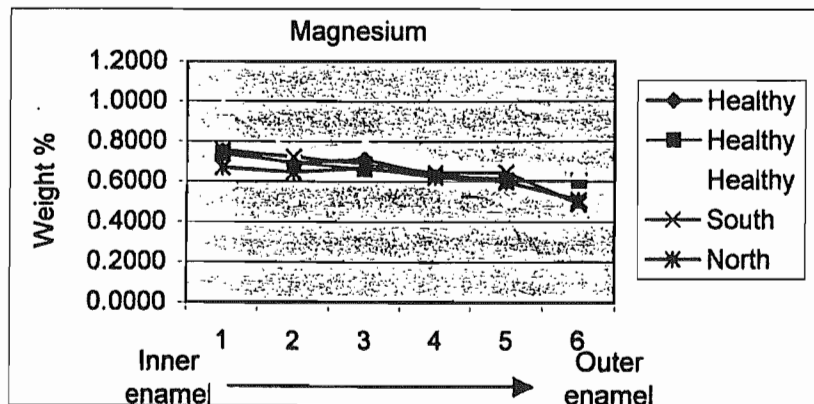
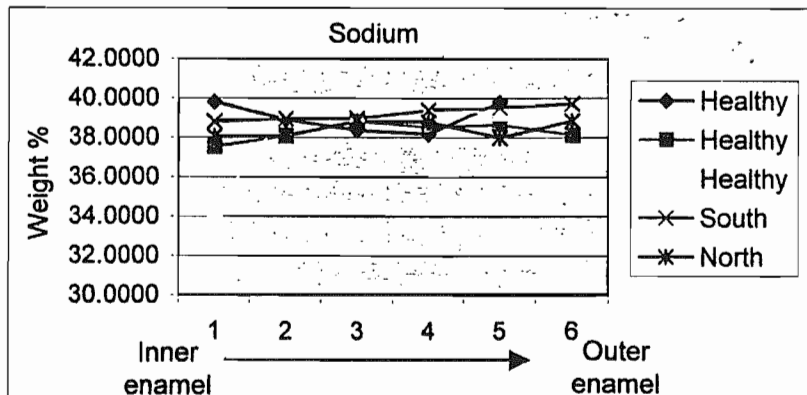
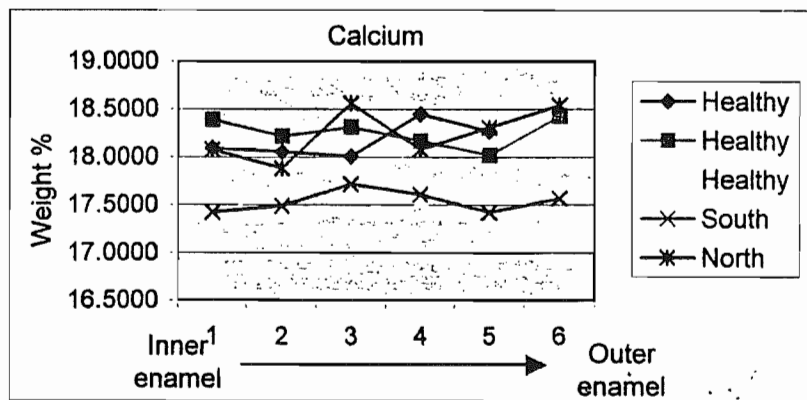
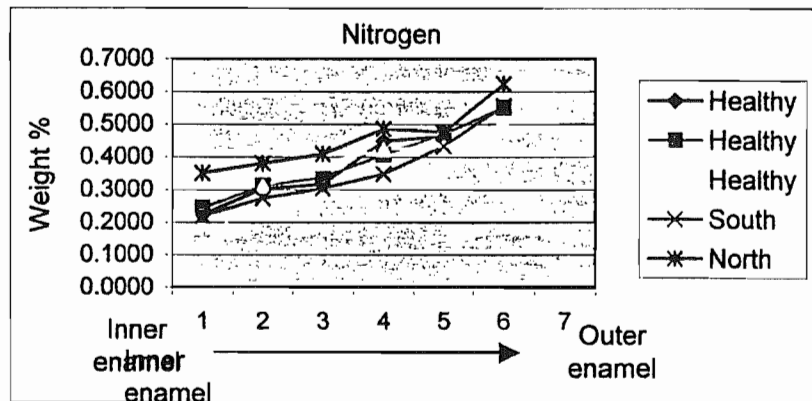
February 26, 2004

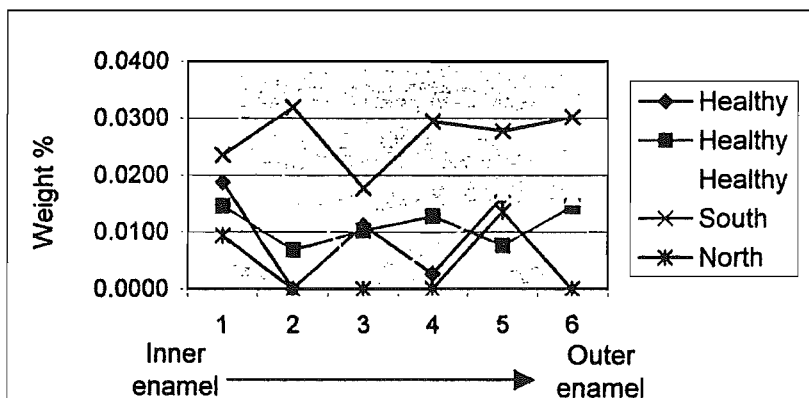
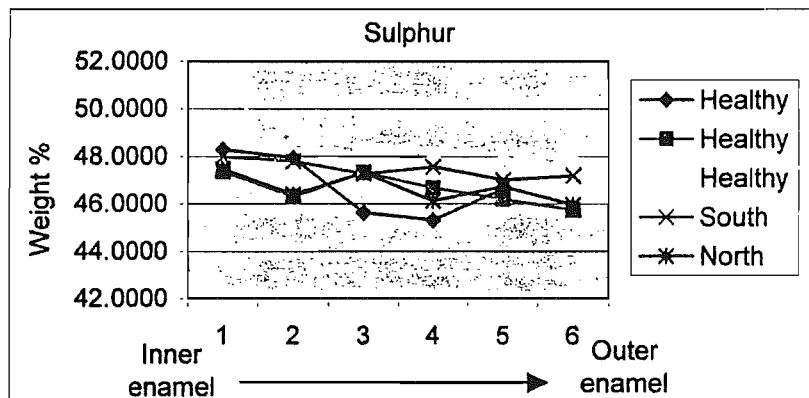
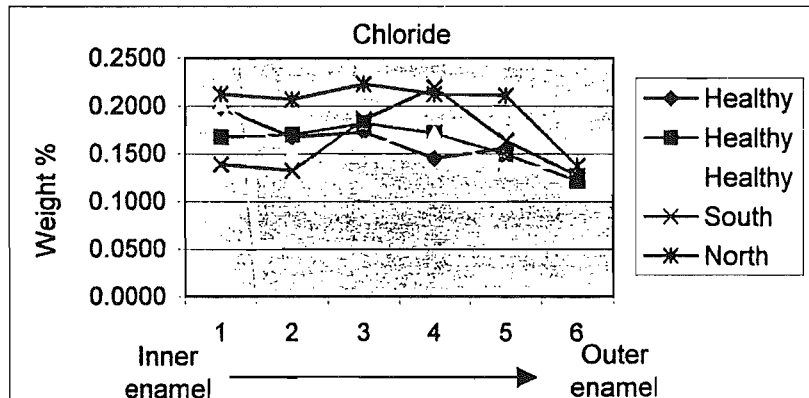
Department of Earth Sciences  
Dalhousie University

Attention: Dr. M. Zentilli


Re: Results of analysis on submitted samples.

	Sample					
	T-1	T-2	T-3	T-4	T-5	T-6
Si (%)	0.15	0.84	0.24	0.22	0.21	0.11
Ti (%)	<0.009	<0.009	<0.009	<0.009	<0.009	<0.009
Al (%)	0.025	0.046	0.021	0.042	0.017	0.023
Mg (%)	0.34	0.27	0.34	0.44	0.46	0.53
Ca (%)	26.00	24.94	26.95	27.11	27.46	29.10
Na (%)	0.24	0.22	0.25	0.26	0.27	0.36
P (%)	9.82	9.43	9.78	9.73	9.95	9.99
C(Total)(%)	18.37	18.56	17.93	17.88	17.61	16.74
As (ppm)	4	4	2	5	4	4
Ba (ppm)	260	220	230	240	220	240
Be (ppm)	<1	<1	<1	<1	<1	<1
Cd (ppm)	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2
Co (ppm)	<1	<1	<1	<1	<1	<1
Cr (ppm)	14	30	19	15	18	22
Cu (ppm)	4	5	5	3	5	5
Fe (ppm)	844	1950	1200	838	1060	1170
K (ppm)	95	87	84	67	65	134
Li (ppm)	4	3	5	5	5	5
Mn (ppm)	37	62	59	88	89	42
Mo (ppm)	<1	<1	<1	<1	<1	<1
Ni (ppm)	2	4	3	2	1	1
Pb (ppm)	6	5	<2	<2	6	<2
Rb (ppm)	6	8	5	6	3	5
Sr (ppm)	285	264	315	318	331	313
V (ppm)	8	9	9	8	9	7
Zn (ppm)	113	104	101	100	100	100





	Sample					
	W-1	W-2	W-3	W-4	W-5	W-6
Si (%)	0.12	0.34	0.29	0.28	0.21	0.18
Ti (%)	<0.009	<0.009	<0.009	<0.009	<0.009	<0.009
Al (%)	0.035	0.030	0.013	0.011	0.011	0.014
Mg (%)	0.30	0.26	0.26	0.28	0.29	0.31
Ca (%)	16.70	15.92	15.31	17.45	10.57	17.80
Na (%)	0.19	0.16	0.18	0.28	0.30	0.36
P (%)	6.81	6.37	5.78	6.33	5.94	6.76
C(Total)(%)	27.12	29.74	30.09	28.67	29.80	28.11
As (ppm)	2	3	2	3	8	2
Ba (ppm)	380	380	340	340	350	350
Be (ppm)	<1	<1	<1	<1	<1	<1
Cd (ppm)	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2
Co (ppm)	<1	<1	<1	<1	<1	<1
Cr (ppm)	12	24	24	29	19	19
Cu (ppm)	4	5	6	5	4	3
Fe (ppm)	568	1470	1610	1830	1180	1010
K (ppm)	104	75	158	288	293	309
Li (ppm)	3	3	3	3	3	3
Mn (ppm)	14	17	18	17	17	15
Mo (ppm)	<1	<1	<1	<1	<1	<1
Ni (ppm)	<1	<1	2	3	5	2
Pb (ppm)	4	4	2	3	2	3
Rb (ppm)	2	2	2	4	4	4
Sr (ppm)	366	358	361	403	381	408
V (ppm)	8	5	6	9	8	7
Zn (ppm)	113	109	96	99	100	103



Cyril Cole