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ELPHIDIUM EXCAVATUM (TERQUEM): PALEOBIOLOGICAL AND

STATISTICAL INVESTIGATIONS OF INFRASPECIFIC

VARIATION

by

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B. Sc. (Hons. Geol. Sci.) (Queen's)

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submitted in partial fulfillment of the requirements for the
degree of Master of Science at Dalhousie University, Halifax,
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DALHOUSIE UNIVERSITY

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ABSTRACT

Detailed study of large sympatric populations and fossil assemblages of the highly variable species Elphidium excavatum (Terquem) collected from 20 widely spaced locations indicates that a variety of morphotypes of Elphidium can be linked to one another in a number of interlocking intergradational series. Ten morphotypes are recognized and grouped as formae (ecophenotypes) of Elphidium excavatum (Terquem); these morphotypes were previously considered as 22 independent taxa by various authors.

To test the hypothesis that these ecophenotypes are distinct morphologically, the ten ecophenotypes were separated into groups based on differences in external morphology; 15 of the characters by which the groups are distinguished were measured and or scored on 721 individuals (11-163 per forma). Discriminant and classification functions were calculated from these character measurements using the SPSS computer program DISCRIMINANT. To illustrate the derivation of these functions, two examples (2 groups and 2 variables; 3 groups and variables), were calculated and explained step by step using the MINITAB interactive statistical package.

Fifteen analyses, using either one sample or split sample approaches, and simultaneous or stepwise analytic methods, classify 84-90% of the specimens into the subjectively defined formae to which they were assigned. Either morphotype (forma) or location was treated as the dependent variable. The analyses showed that there is no strong relationship between formae and geographic location, thus strengthening the subjective conclusion that these are ecophenotypes and not subspecies.

Although all of these formae belong to the same species, it is suggested that the distinction among them should be retained because of their potential as a valuable interpretive tool in paleo-ecological and biostratigraphic studies of Holocene and Pleistocene sediments.

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INTRODUCTION

AN IDENTIFICATION PROBLEM

Applied micropaleontology often views purely taxonomic studies as being of academic interest only. Without a uniform and clearly defined taxonomy, however, it becomes very difficult to compare data from different sources. At the same time, the indicator value of the species is diminished if several different names are applied to the same taxonomic unit (Medioli and Scott 1978). This problem becomes most acute in areas where infraspecific variability is highest (i.e., nearshore and shelf environments).

The species Elphidium excavatum (Terquem), which is extremely important for the interpretation of post-glacial events, has been plagued by a myriad of synonyms. The first to recognize the real nature of the problem was Feyling-Hanssen (1972), who grouped several common species as ecophenotypes of E. excavatum. The variations within this species have been interpreted either as ecophenotypic (Bartlett 1965b, Feyling-Hanssen 1972, Cronin 1979, Poag 1978) or as a subspecific (Wilkinson 1979). It has been noted, but not illustrated, that E. excavatum (= E. olavatum of many workers) appears to grade into a number of different species (e.g. Parker 1952b, Weiss 1954, Brodniewicz 1965, Bartlett 1965b, Buzas 1965b, Cronin 1979).

During a geochemical study of E. excavatum tests from a Labrador Shelf sediment core (Miller 1979), it became apparent that specimens of two different Elphidium species could be arranged into an intergradational series covering their spectrum of morphological

variability. Miller et al. (1982) continued this investigation and arranged five distinct "species" in three similar intergradational series. Thus, the biologic principle of conspecificity of specimens arranged in an intergradational series, as enunciated by Mayr et al. (1953) and applied to foraminifera by Medioli and Scott (1978), was used in an attempt to solve the taxonomic problems of the E. excavatum group.

Demonstration of an intergradational series between taxonomic units previously defined as species is a useful tool for illustrating the range of variation that may be encompassed by different samples of a taxon which is probably a single biological species in the sense of Mayr et al. (1953). When morphological variants of the species appear to convey important information about the environment, however, it becomes necessary to delimit the range of morphology encompassed by the different environmental indicators, and to assign infraspecific names to these morphotypes.

When two or more morphotypes are linked through an intergradational series, there are no set limits to each morphotype and the "morphological boundaries" are arbitrary and artificial. As pointed out by Scott (1974), subjective assessment of variation is alone inadequate, in principle and practice, for scientific communication. The problem becomes that of where to draw boundaries along the morphological gradient in order to produce workable, identifiable, taxonomic units, and how to classify these units within the species.

AIMS OF THIS WORK

1) The overall aims of this work are to investigate the polytypic nature of the foraminiferal species Elphidium excavatum (Terquem) using both conventional biological criteria and multivariate statistical methods; and to compare the results of the two methods of analysing variation within a species. 2) Within the Linnean hierarchy (as best as possible) the species is first "split" into morphotypes and the intergradation between morphotypes is investigated and documented. The taxonomic (and apparent biological) relationships among the morphotypes are also investigated and their corresponding ecological and geographical ranges noted.

3) Once the patterns of infraspecific variation have been noted and the presence or absence of intermediate forms documented, the kind of biological variation present can be determined and an appropriate classification adopted.

4) These same morphotypes are then analysed statistically using the multivariate technique of discriminant analysis. This is to determine if the morphotypes are statistically distinct and to determine, using the classification phase of the analyses, how well these morphotypes are delineated based on the information (morphological characteristics) included in the analysis. How well these morphotypes are delineated is also a test on the placement of the arbitrary boundaries between morphotypes, which split the morphologic range of the species into more rigid discrete units.

5) Finally, three other species are included in the statistical analysis to determine how well this E. excavatum group is delineated

within a larger framework.

In general, then, the purpose of this work is to develop a classification scheme for the species E. excavatum (Terquem) which meets the following criteria: (1) fits into both the biologic and paleontologic frameworks (the Linnean hierarchy) and consequently is acceptable to workers in both fields; (2) is statistically valid; (3) is repeatable by other workers; and consequently objectively solves the taxonomic problems of the group.

Only when there are standard taxonomic units can an accurate description of patterns of infraspecific variation be formulated. Clear definition of taxonomic variants in turn is a prerequisite for the explanation of the species variation in terms of geographical and ecologic isolation.

ELPHIDIUM EXCAVATUM (TERQUEM): ECOPHENOTYPIC VERSUS SUBSPECIFIC
VARIATION

METHODS OF STUDY

Different standard methods of preparation and preservation were employed, due to the various types of samples under study. None of them is sufficiently unusual to warrant description.

Samples from various localities (Table 1) were made available by numerous colleagues. Most of these samples were dry residues, except those collected by Scott and Medioli, which had been stained with Rose Bengal and stored in a mixture of denatured ethanol and water.

All samples studied contained large assemblages or populations of E. excavatum. In this study, a distinction is made between populations of stained individuals (i.e., individuals stained by Rose Bengal, hence alive at the moment of collection) and assemblages of empty tests. Stained individuals of the same species in the same sample clearly represent a "population", i.e., potentially interbreeding individuals. An assemblage of empty tests could contain reworked individuals and does not contain anything that could be potentially interbreeding; as such it does not constitute a "population".

The principle of intergradation is normally affixed to "populations", but that is mainly for lack of consideration of fossil assemblages. Thus, the individuals of populations are potentially interbreeding but, in the vast majority of cases, no one has bothered to check if, in fact, they do interbreed. Usually, a visual appraisal

Location	Latitude	Longitude	Collected By	Age	Formae Identified	Sample No. Reference	Type of Environment
Beaufort Sea Canada	60°56'50''N	134° 33'00''W	Vilks, 1970	recent assemblage	<i>clavata</i> , <i>excavata</i> <i>magna selseyensis</i>	853 Vilks et al. 1979	arctic nearshore with estuarine influence. salinity: 28-31‰ water depth: 10m
Beaufort Sea Canada	?	?	?	Holocene assemblage	<i>clavata</i> , <i>excavata</i>	F2255, made available by Bartlett; no reference	arctic nearshore
Hirtshals Denmark	57°36'N	9°58'W	Vilks, 1979	Late Pleistocene assemblage 14,000-35,000 Y.B.P.	<i>clavata</i> , <i>excavata</i>	Zone C of the older Yoldia clay, Andersen 1971	arctic shallow water
Labrador Sea " "	54°36'30''N	56°15'00''W	Vilks, 1977	Late Pleistocene (Wisconsinian) assemblage > 22,000 Y.B.P.	<i>clavata</i> , <i>excavata</i>	Core 12 826-831cm Vilks and Mudie 1978; Miller 1979; Vilks 1980; Miller et al. 1982	melting ice margin (?) basin environment; no modern analogue
Champlain Sea, Quebec (St. Alme Guillaume Quebec)	45°53'N	72°55'W	Guilbault, 1980 (Service de Geotechniques Resources, Quebec)	Late Pleistocene assemblage 11,500 Y.B.P.	<i>magna</i> , <i>clavata</i> <i>excavata</i> , <i>lidoensis</i>	YAM-F13-80A > 106A 24.5m in boring. Guilbault 1980	Facies B cold bottom water Salinity 25-35‰ Guilbault suggests the deeper part of the Baltic Sea as a modern analogue (i.e. Lutze, 1974.)
Bay of Chaleur Gulf of St. Lawrence Canada	1) 47°54'29''N 2) 48°00'12''N 3) 48°04' N	65°50'20''W 65°21'19''W 66°19' W	Shafer and Cole, May 1971	recent population	<i>excavata</i> , <i>clavata</i> <i>williamsoni</i> , <i>magna lidoensis</i> , <i>cuvillieri</i>	1) SRA-52 2) SQA-52 3) SRQ-53 Schafer and Cole 1978	Shallow water, nearshore, estuarine.
Miramichi Estuary, N.B., Canada	47°07'05''N	65°06'05''W	Scott, 1976	recent population	<i>clavata</i> , <i>lidoensis</i> <i>excavata</i> , <i>magna</i>	station 6; Scott, Schafer and Medioli 1977, 1980	open bay zone of estuary Salinity: 20-25‰; water depth 5-10m.
Baie Verte, Northumberland Strait, Canada	46°02'20''N	63°42'80''W	Medioli, 1979	Holocene assemblage 4,000-5,000 Y.B.P.	<i>selseyensis</i> , <i>clavata lidoensis</i> , <i>gunteri magna</i> , <i>excavata</i> <i>cuvillieri</i> , <i>galvestonesis</i>	Navicula core 2, 60-62cm Prime 1980	mid-Holocene hypsithermal (?) warm, temperate to sub-tropical, water depth < 5m, this fauna does not exist there today.
Annapolis Basin, N.S. Canada	1) 45°40'19''N 2) 44°38'09''N	65°38'53''W 65°45'08''W	Bartlett, 1968	recent population	<i>selseyensis</i> , <i>magna lidoensis</i> , <i>clavata excavata</i>	1) BQ-68-AB-8 2) BQ-68-AB-16 no reference	shallow, partially restricted estuary. Tidal range > 12m salinity probably 25-32‰
Chezetocook Inlet, N.S.	44°41'N	63°14'W	Scott and Medioli, 1977	recent population	<i>magna</i> , <i>selseyensis</i> <i>clavata</i> , <i>lidoensis</i> <i>excavata</i> , <i>williamsoni</i> <i>cuvillieri</i>	stations 49-55; Scott 1977, Scott Medioli 1980, Scott, Schafer and Medioli, 1980	nearshore turbulent zone subtidal; salinity: 25-32‰ water depth: 3-5m

Table 1: Listing of samples studied. The listing under "Formae Identified" is given in order of abundance, at the time of collection. **Illustrated in Miller et al. (1982).

Maine-New Brunswick Estuary (near Eastport, Maine)	44°45'N (approx.)	67° W (approx.)	no reference	recent population	<u>williamsoni</u> , <u>clavata</u> <u>lidoensis</u>	B-I-71 Schafer 1971	shallow water, nearshore estuarine
Scotian Shelf (off Liverpool, N.S.) Canada	43°59'N	64°39'W	Medioli and Scott, 1978	recent population	<u>magna</u> , <u>clavata</u>	Station 134 Miller, Scott and Medioli, unpub. data	nearshore turbulent zone 40m deep normal marine conditions
Long Island Sound, N.Y. U.S.A.	40°57'N	73°30'W	Schafer, 1965	recent population	<u>selseyensis</u> , <u>lidoensis</u> <u>excavata</u> , <u>clavata</u>	Field No. 722 Schafer 1968, 1970.	outer estuary, temperate salinity 17-20‰ water depth: 10-20m.
San Francisco Bay, CA U.S.A.	37°36'N	122°21'W	1969 (borehole transect across San Francisco Bay)	pre-Disconsinian, 7 Sangamonian assemblage 80,000-100,000 Y.B.P. oxygen isotope stage 5	<u>lidoensis</u> , <u>excavata</u> <u>gunteri</u> , <u>selseyensis</u> , <u>clavata</u>	230-11 43m below present sea level	Unit B. subtidal mudflat and channel environments. salinity: 20-32‰ temperature: 8-20° annually
San Diego Bay, CA, U.S.A.	33°34'N	117°11'W	Bradshaw, 1972	recent population	<u>lidoensis</u> , <u>lucidum</u> <u>clavata</u> , <u>selseyensis</u> <u>gunteri</u>	IX-1, IX-2 Scott et al. 1976	inner bay assemblage. salinity: 31-35‰ water depth: 3-5m
San Antonio Bay, Gulf of Mexico, TX U.S.A.	28°26'N	96°53'W	Poag, 1972	recent population	<u>gunteri</u> , <u>lidoensis</u> , <u>gylveslonensis</u> , <u>cuyillieri</u> , <u>clavata</u>	sample 13 Poag 1976, 1978	mudflat; salinity 5-15‰ (over sampling period) water temperature: 12-23°C
Wadden Sea, The Netherlands	?	?	Hofker, 1975, 1976	recent or Holocene assemblage	<u>williamsoni</u> , <u>gunteri</u>	no sample number; made available by A. Fortuin. Hofker 1977	probably tidal flat.
Venice Lagoon, Italy	45°30'N	12°26'E	Petrucchi, 1981	recent population	<u>gunteri</u> , <u>cuyillieri</u> , <u>lidoensis</u> .	Sample 1	Albani and Serandrei Barbero 1982, water temperature: 6-25°C salinity: 32-35‰ in lagoon itself
Bay of Izmir Turkey	38°44'06''N	26°33'00''E	Piper and Aksu, 1979	late Pleistocene (Wurm) assemblage oxygen isotope upper stage 2	<u>cuyillieri</u> , <u>lidoensis</u> , <u>excavata</u> , <u>williamsoni</u>	core 79-Iz-2 sample 4 30-32cm sample 10 92-94cm Piper and Aksu 1981	current water depth: 110m Pleistocene pro-delta slope sea level lowered 110m.

Table 1: continued.

of morphological similarities is considered more than adequate to make the assumption that two individuals do or do not interbreed. Similarly, a visual appraisal of a fossil assemblage should be more than adequate and equally valid to decide whether or not two individuals would have been capable of interbreeding had they existed at the same time. However, to be prudent, the assemblages studied in this thesis were carefully chosen from those lacking evidence of reworking.

Representative specimens covering all aspects of morphological variation of the species were selected from each sample and prepared for standard scanning electron microscope (SEM) observation. A total of 1057 SEM photos of 810 E. excavatum specimens were taken either at Dalhousie University (Bausch and Lomb Nanolab 2000) or the Bedford Institute (Cambridge 180) using both Polaroid NP/52 and NP/55 film.

Samples were collected from a wide variety of environments as well as geographical range (Table 1): Beaufort Sea, Labrador Shelf, Champlain Sea, Bay of Chaleur, Miramichi Estuary, Baie Verte, Annapolis Basin, Chezzetcook Inlet, and the Scotian Shelf, Canada; a Maine-New Brunswick Estuary, Long Island Sound, San Francisco Bay, San Diego Bay, and San Antonio Bay, U.S.A.; Hirtshals, Denmark; Wadden Sea, Netherlands; Venice, Italy; and Bay of Izmir, Turkey. The age range of the samples is late Pleistocene to recent.

HISTORICAL REVIEW

Before proceeding with the observations of this study, it is necessary to examine the complex history of Elphidium excavatum

(Terquem) in some detail, in order to place this study in proper perspective. A schematic outline of the history is illustrated in Figure 1 (back pocket). As far as possible, the following review is presented in chronological order. The letters in parenthesis indicate the position of the information on the schematic diagram (Figure 1).

D'Orbigny described three species relevant to this study:

Polystomella oceanensis (1826, p. 285) (A on Figure 1) a recent species from the French coast; (C) P. poeyana (1839a, p. 55, Pl. 6:25-26) from Cuba; and (B) P. articulata (1839b, p. 30, Pl. 3:9,10) from recent material from the Falkland Islands. It appears that d'Orbigny did not figure or choose a holotype of P. oceanensis, so it remained a nomen nudum until figured by Fornasini (1904, p. 13, Pl. 3:10). The remaining two species were figured; but the type of P. articulata is lost (if it was ever designated) and a lectotype of P. poeyana was designated by Loeblich and Tappan in 1964 (CC).

In England in 1858, Williamson described a species (D) Polystomella umbilicatula (Walker and Jacob), (1858, p. 42-44, Pl. 3:81-82), a species later shown to be quite distinct from Walker and Jacob's. In the same publication, Williamson also described a variety of this species. Williamson called this species P. umbilicatula var. incerta, and it has become known as Elphidium incertum (Williamson).

Terquem (1876) described Polystomella excavata (E) from the shore of Dunkerque, French south coast.

Heron-Allen and Earland (1909) identified a species as Polystomella striatopunctata (Fichtel and Moll) variety (p. 695, Pl.

21:2); in 1911 the same authors designated this species as P. striatopunctata var. selsevensis (p. 488), from the shore sands of southeast England (F).

Cushman (1930, p. 18, Pl. 7:8-9) described and figured a large white opaque Elphidium, which he referred to as E. incertum (Williamson) (G). In the same paper, he described and figured a variety, E. incertum var. clavatum (p. 20, Pl. 7:10) which was smaller, translucent orange-brown and often, but not always, with a knobby boss or bosses occupying the umbilicus (H). Unfortunately, in the writer's opinion, Cushman (1930, p. 21, Pl. 8:4-7) also appears to have mistaken Williamson's species (P. umbilicatula - D) for E. excavatum, even while displaying Terquem's original figures (I) of E. excavatum (Cushman, 1930, p. 21, Pl. 8:1-3). This apparent error has persisted and may have confused workers in Europe, the eastern U.S., and Canada up until the 1970's (e.g. Todd and Low 1961, Richter 1961, 1964a, 1967, Haake 1962, 1967, Feyling-Hanssen 1964, Murray 1965a, 1968, Scott et al. 1977).

Compounding this, Heron-Allen and Earland (1932) described E. (Polystomella) excavatum (Terquem) (p. 439, Pl. 16:22-23) from the Falkland Islands (J) and placed it in synonymy (K) with the E. excavatum (Terquem) of Cushman, 1930 (Pl. 8:4-7) i.e. Williamson's species. Heron-Allen and Earland (1932) clearly did not place these specimens in synonymy with their P. striatopunctata var. selsevensis.

In the next 10 years (1930-1940) five species were described by North American workers which have become widely reported in the literature. These were: (L) Elphidium gunteri Cole, 1931 (p. 34, Pl.

4:9-10) from Pliocene (later shown to be Pleistocene by Poag 1978) deposits in Florida; (M) a variety of this, E. gunteri var. galvestonense (= E. galvestonensis) Kornfeld 1931 (p. 87, Pl. 15:1a-b) from Texas and Louisiana; (N) E. lidoense Cushman, 1936 (p. 86, Pl. 15:6a-b) from beach sands in Venice, Italy, and two species described by Natland (1938) from the California coast: (O) E. translucens (p. 144, Pl. 5:3,4) and (P) E. tumidum (p. 144, Pl. 5:5,6).

In addition, nine other species were described from 1930 to 1951. Two were described by Shupack (1934): (Q) E. brooklynense (p. 10, opp. p. 9, Figs. 7a-b) and (R) E. florentinae (p. 9, opp. p. 9, Figs. 5a-b) from Long Island Sound and New York Harbour. Five were described by Cushman and Brönnimann (1948) from Trinidad: (S) Criboelphidium trinitatense (p. 20, Pl. 4:8), (T) C. limnosum (p. 19, Pl. 4:7), (U) C. vadescens (p. 18, Pl. 4:5), (V) C. salsum (p. 19, Pl. 4:6) and (W) C. kugleri (p. 18-19, Pl. 4:4). The remaining two were (X) E. littorale Le Calvez and Le Calvez, 1951 (p. 251, Fig. 5:a-b) and (Y) E. guntheri (sic) var. waddense van Voorthuysen 1951 (p. 25, Pl. 2:16a-b).

In 1966, Levy described a common shallow water Mediterranean form he called (Z) E. cuvillieri (p. 5-6, Pl. 1:6a-c, Pl. 2:2).

Thus, from 1930-1970, 11 different species which the writer assigns to the E. excavatum group were described and figured in North American literature. 1) Elphidium excavatum of Cushman's concept (i.e. the species of Williamson)(I), was reported by Cushman (1930, 1939, 1949), Todd and Low, (1961), and Adams and Frampton (1965); 2) E. incertum of Cushman (not Williamson's taxon) (G) was described by

Cushman (1930, 1939, 1944, 1949), Parker (1948, 1952a, 1952b), Todd and Low (1961), and Bartlett (1965b); 3) E. clavatum Cushman (AA), and E. incertum var. clavatum (H) were described by Cushman (1930, 1939, 1948), Loeblich and Tappan (1953), Todd and Low (1961), Cooper (1964), and Buzas (1965b, 1966). 4) E. poeyana (d'Orbigny) (CC) was described by Cushman (1930), Parker et al. (1953), Todd and Brönninann (1957), Todd and Low (1961), and Loeblich and Tappan (1964); 5) E. gunteri Cole (DD) was described by Cushman (1939), Parker et al. (1953), Phleger (1954, 1960a, 1960b), Bandy (1956), Lehmann (1957), and Lankford (1959); 6) E. galvestonense Kornfeld (EE) was described by Parker et al. (1953), Phleger (1954, 1960a), Lehmann, (1957), Parker and Athearn (1959), and Todd and Low, (1961); 7) E. oceanense (d'Orbigny) (FF) was reported by Cushman (1939); 8) E. selsevense (Heron-Allen and Earland) (GG) was described by Cushman (1939) and Parker (1952b); 9) E. lidoense was described by Cushman (1936, 1939) (N); 10) E. translucens Natland (HH) was reported by Bandy (1953), Parker et al. (1953), Phleger (1954, 1964), Todd and Bronnimann (1957), Todd and Low (1961); and 11) E. tumidum Natland (II) was reported by Parker et al. (1953), Todd and Bronnimann (1957), Lehmann (1957), and Phleger (1960b).

In addition, seven of the nine other species mentioned previously were also reported in the literature. Shupack's two species were referred to by Weiss (1954) and Cushman and Bronnimann's five species were referred to by Todd and Brönnimann (1957).

Loeblich and Tappan (1953) elevated E. incertum var. clavatum Cushman to the specific rank E. clavatum Cushman (AA); and showed that

the E. incertum of Cushman was quite distinct from that of Williamson's taxon, Polystomella umabilicatula variety incerta Williamson (1858). Buzas (1966) proved, through statistical analysis and wall structure investigation, that E. clavatum Cushman, and E. incertum of Cushman could not be separated into distinct species (JJ); a view shared by Parker (1952b) and Bartlett (1965b) based solely on external morphology. Buzas (1966) initiated the investigation because he had observed morphological gradation from one species to the other (Buzas 1965b, 1966). In addition to other differences, Buzas (1966) showed that Williamson's E. incertum had an optically granular wall structure, instead of the optically radial wall structure of Cushman's specimens. Thus, Buzas (1966) concluded that both of Cushman's forms did belong to E. clavatum Cushman (AA).

At the same time, European workers were describing and figuring seven species under six names: 1) E. excavatum (Terquem) of Cushman's concept (i.e. Williamson's species) (I) was recorded by Rottgardt (1952), van Voorthuysen (1957, 1960), Jarke (1961) Richter (1961, 1964a, 1967), Haake (1962, 1967), Feyling-Hanssen (1964), Brodniewicz (1965) and Murray, (1965a, 1968, 1970); 2) E. excavatum (Terquem) (sensu stricto) (KK) was discussed by Lutze (1965, 1968), Lévy (1966), Haake (1967), Lévy et al. (1969, 1975), Murray (1971) and Knudsen (1973b); 3) E. articulatum (d'Orbigny) (BB) was recorded by Lutze (1968), Murray (1971), Rosset-Moulinier (1972) and Knudsen (1973a, 1973b); 4) E. gunteri Cole (DD) was discussed by van Voorthuysen (1957, 1960), Haake (1962), Richter (1964a), Lévy (1966) and Lévy et al. (1969); 5) E. selseyense (Heron-Allen and Earland) (LL) was

discussed by Brand (1941), van Voorthuysen (1957, 1960), Richter (1961, 1964a) and Haake (1962); 6) E. clavatum Cushman (NN) was discussed by Hansen (1965) and Knudsen (1971a, 1971b); and 7) E. lidoense Cushman (MM) by Accordi and Socin (1951), Lévy (1966), Cita and Premoli-Silva (1967) and Lévy et al. (1969).

During the 1960's, a few workers started grouping some of these species. Haake (1962) was one of the first to place E. selseyense (Heron-Allen and Earland) (LL) and E. clavatum (AA) Cushman in synonymy (OO); Haake used the name E. selseyense because he considered E. clavatum to be a junior synonym. Both Hansen (1965) and Knudsen (1971b) could not separate E. clavatum and E. selseyense (PP). In 1965 Lutze studied topotype material of E. excavatum and concluded that E. excavatum (Terquem) and E. clavatum Cushman were the same species (QQ). However, Lutze (1965) did differentiate the two at a subspecific level (E. excavatum excavatum, E. excavatum clavatum).

By 1970 then, various workers had combined E. selseyense and E. clavatum (PP) (Hansen 1965, Knudsen 1971); or E. clavatum and E. excavatum (QQ) (Lutze 1965), or E. excavatum and E. selseyense (RR) (Haake 1967, Lévy et al. 1969, 1975, and [later] Feyling-Hanssen 1972 and Banner and Culver 1978). Lévy et al. (1975) reported that Terquem's holotype had been lost; they redescribed E. excavatum (Terquem) and erected a neotype. Haake (1967), and von Daniels (1970) had also placed E. (Cribrononion) lidoense in synonymy with E. excavatum (SS).

In addition, van Voorthuysen (1957) placed his E. gunteri var. waddense back into E. gunteri (TT); Lévy (1966), and later Hansen and

Lykke-Andersen (1976) and Poag (1978) put E. oceanense and E. littorale into E. gunteri (UU) because they considered the former to be a nomen nudum, and Murray placed E. gunteri into E. oceanense (VV).

In 1972 Feyling-Hanssen completed a comprehensive study of these species and concluded that there was only one highly variable species: E. excavatum (Terquem) (WW). Feyling-Hanssen's species is comprised of E. excavatum (Terquem), E. clavatum Cushman, E. incertum var. clavatum Cushman (not Williamson), E. selsevense (Heron-Allen and Earland) and E. lidoense Cushman. Feyling-Hanssen (1972) noted a pattern to the variability and concluded that there are four ecophenotypes of this one species. He differentiated these ecophenotypes on a "forma" level, so that earlier specific names were retained as formae names of E. excavatum. He suggested that the distribution of these ecophenotypes was environmentally controlled but also implied some geographical restrictions. The four formae designated by Feyling-Hanssen (1972) are: (1) E. excavatum forma clavata Cushman, for the small translucent orange-brown form with (or without) the knobby bosses, which he observed dominating assemblages in arctic environments; (2) E. excavatum (Terquem) and E. selsevense (Heron-Allen and Earland) which he placed in synonymy as E. excavatum forma selseyensis (Heron-Allen and Earland) and defined as a larger, orange-brown to white form found in boreal environments (North Sea region and western Baltic); (3) E. excavatum forma lidoensis Cushman which was described as a strongly ornamented, knobby form found in the Lusitanian regions (the west coasts of France and Portugal); and (4) for the white form, which he believed Cushman identified as E.

incertum (Williamson), he suggested the name E. excavatum forma alba. The same author found E. excavatum forma alba in foraminiferal zone E of early Holocene sediments in the Oslo Fjord area.

In 1969, Lévy et al. reported on Cushman's error (in mistaking Williamson's species for E. excavatum) and they returned to Terquem's original definition of the species. For the form that Cushman and subsequent workers had identified as E. excavatum, Lévy et al. returned to Williamson's specific epithet umbilicatula (XX). However, Haynes (1973) pointed out that this name is invalid because it is occupied by Walker and Jacob's species, so Haynes suggested the name E. williamsoni (YY) for Williamson's species.

Haynes (1973) also commented on the differences between specimens of E. selseyense (Heron-Allen and Earland) in the British Museum and E. excavatum (Terquem's type figure). He stated that E. selseyense (ZZ) can be distinguished, and should remain distinct from E. excavatum (Terquem) (KK) as redefined by Lévy et al. (1969, 1975). Both Haynes (1973) and Lévy et al. (1969, 1975) state that the sutures are non-granular, but the toptype illustrated (Lévy et al. 1975, p. 176, Pl. 3:5-6) shows granular material in the sutures. This discrepancy was pointed out by Wilkinson (1979) who placed the neotype of Lévy et al. (1975, p. 176, Pl. 3:1-2) in E. excavatum (Terquem), and referred their toptype to E. clavatum selseyense (Wilkinson, 1979). Wilkinson (1979) studied E. excavatum (Terquem) and E. clavatum Cushman; and his results differ markedly from those of Feyling-Hanssen (1972). Wilkinson (1979) concluded that he was dealing with Cushman's E. clavatum, a species distinct from E.

excavatum. He also defined eight subspecies of E. clavatum (AAA) which, Wilkinson states, form a gradational sequence based on umbilical characteristics. One of these subspecies is E. clavatum selseyense, which is an invalid name (as pointed out by Haynes, 1982 pers. comm., as selseyense takes priority).

Other workers were beginning to recognize ecophenotypy within the genus Elphidium. Poag (1978) recognized four ecophenotypes of two species, E. gunteri (BBB and CCC) and E. galvestonense (DDD). In 1981 Haynes recognized the polymorphic nature of E. excavatum (EEE). He pointed out that the species could be regarded as a "superspecies", including E. clavatum and E. selseyense and related forms which he referred to as "siblings" of Elphidium. ex gr. excavatum. Haynes (1981) also states that E. excavatum (Terquem) can be regarded as one polytypic species (sensu Beckner 1959) which he called the E. excavatum subsp. gr. Haynes includes E. incertum (Williamson), E. clavatum Cushman, E. selseyense (Heron-Allen and Earland), E. williamsoni Haynes and E. cf. advenum sensu Todd and Low in the group, but he did not include Cushman's E. lidoense.

In 1982 Rodrigues and Hooper voiced the opinion that Terquem's original concept of E. excavatum included E. williamsoni Haynes. They also followed Wilkinson's (1979) lead in distinguishing among the species E. excavatum, E. selseyense, E. lidoense and E. clavatum. Rodrigues and Hooper based this decision on the fact that since (1982, p. 415) "...no morphological series relating modern specimens of E. clavatum to either E. selseyense or E. lidoense has been adequately documented in the literature, we choose to regard E. clavatum as

distinct from E. selsevense and E. lidoense." A month later, intergradation of these four species in material from eight locations was reported and illustrated by Miller et al. (1982) (FFF). Miller et al. (1982) designated five ecophenotypes of E. excavatum. Two of these formae are the same as Feyling-Hanssen's (E. excavatum f. clavata and f. lidoensis), two formae (E. excavatum f. excavata and f. selsevensis) result from the splitting of Feyling-Hanssen's E. excavatum forma selsevensis; and one is a new ecophenotype previously undescribed (E. excavatum forma magna).

OBSERVATIONS

MORPHOTYPIC VARIATION

The study of this material has allowed delineation of ten morphotypes, nine of which are interpreted as being conspecific with E. excavatum as documented by the intergradational series illustrated in the plates.

These morphotypes, which are considered here as "formae", are easily recognised under low power microscopy. Photoplates 20-28 illustrate these formae. For the sake of clarity the formae are designated such that they correspond to conventional species previously described in the literature as much as possible and take their names from the "species" they represent. The salient characteristics of the formae are outlined below.

E. excavatum forma excavata (pl. 20) has lobate chambers, and straight sutures extending unconstricted into the umbilicus. The pore

density is greater in this forma than in forma clavata, giving the test a hazy appearance. Ponticuli are typically strongly developed.

E. excavatum forma williamsoni (pl. 21) is an inflated, rotund form, with smooth peripheral outline and rounded periphery. It has a flat umbilicus, with the chambers extending completely into the umbilicus. The ponticuli are very regular and well developed, covering up to half the chamber width. The test walls are finely and densely perforate.

E. excavatum forma selseyensis (pl. 22) is recognised by its large size; smooth to lobate peripheral outline; sub-acute periphery; and greatly convex walls, giving the umbilicus a raised appearance. The sutures are slightly backwards curved to straight, with irregular, indistinct to strongly developed ponticuli, and often with papillae filling the sutures. The umbilicus contains granular material, or bosses, or both.

E. excavatum forma clavata (pl. 23) is small, disc-shaped, orange-brown, translucent, often with an umbilical boss; and always with an imperforate (complete or incomplete) collar surrounding the umbilicus. The sutures are generally backwards curved, with a few narrow, often incomplete ponticuli.

E. excavatum forma gunteri (pl. 24) is a small to medium sized form, rather rotund, with a coarsely perforate wall. The sutures are straight, not depressed, and marked by many regular, raised rectangular shaped ponticuli, often longer than the chambers are wide. The umbilicus contains papillae/irregular bosses (irregular lateral extensions of the ponticuli and chambers).

E. excavatum forma galvestonensis (pl. 25) is a large, many chambered (13-18) form with a large, very raised umbo (boss or bosses) in the umbilicus and many regular distinct ponticuli. There may be a ring of papillae surrounding the boss or in the sutures. The wall is heavily calcified and very finely perforate giving the test a porcelaneous appearance. The periphery is sub-acute.

E. excavatum forma lidoensis (pl. 26) is a small form, with a large open umbilicus filled with papillae/bosses. The sutures are backwards curved, distinctly broadening towards the umbilicus, and also filled with papillae; ponticuli are not generally well developed. Within this forma, two "subforma" are observed, a phenomenon that will be detailed in both the Discussion (p. 30) and Systematic Paleontology (p. 155).

E. excavatum forma tumidum (pl. 27), is a large, ornamented form resembling forma selseyensis. However, the ornamentation and ponticuli are much more regular on forma tumidum. The umbilicus is large, circular, depressed and filled with papillae/bosses. The chamber extensions into the umbilicus are truncated sharply. The periphery is usually rounded and the chambers inflated.

E. excavatum forma cuvillieri (pl. 28), is a smooth, round disc shaped Elphidium about the same size as forma clavata. The peripheral outline can range from smooth to very lobate. The sutures are straight or gently backwards curved, and characterized by very regular rows of sutural pores.

E. excavatum forma magna (pl. 27) is recognised by its larger size, smooth peripheral outline, sub-acute periphery; and strongly

convex walls, which give the umbilicus a raised appearance. The umbilicus is usually large and filled with one knobby boss. The sutures are backwards curved, and some (or all) may be constricted before reaching the umbilicus.

ECOPHENOTYPIC VARIATION

Having delineated the ten distinct morphotypes, analysis of their distribution in the study areas is necessary to ascertain if they are ecophenotypes or subspecies. This information is also important for the study of paleo-ecology and biostratigraphy of Pleistocene and Holocene deposits.

Beaufort Sea: Two series of morphotypes are illustrated, one from a Holocene surface sample (pl. 2:1-8) and one from a Holocene core sample (pl. 3:1-12). Both samples are assemblages of empty non-living tests. The first assemblage is almost entirely E. excavatum forma clavata. However, the second contains four formae: clavata, excavata, magna and selseyensis.

Hirtshals Denmark: Two formae were recognized in this late Pleistocene assemblage: E. excavatum forma clavata (pl. 2:9-17, pl. 23:1) and forma excavata (pl. 2:18-20). E. excavatum forma clavata comprised greater than 95% of the E. excavatum population.

Champlain Sea: Four formae are recognized from this late Pleistocene assemblage: f. clavata (pl. 4:1,6-9; pl. 23:2-3), f. excavata (pl. 4:10-11), f. magna (pl. 4:2-5, pl. 27:9-10) and f. lidoensis (pl. 4:12). Specimens of three of the formae (magna, clavata, and excavata) can be assembled into an intergradational

series. Forma lidoensis appears to be rare and no link was observed joining this forma to the series. This sample exhibits more variability and contains more formae than any other post-glacial/arctic sample studied.

Scotian Shelf: Two formae, E. excavatum forma clavata (pl. 4:13-16) and magna (pl. 4:17-20; pl. 27:5-8,11) were observed and assembled into an intergradational series from this recent sample of live specimens (a population). Forma magna exhibits more variability here than observed elsewhere. Intermediate specimens, linking the two formae (i.e. pl. 4:15-17) were also observed.

Miramichi Estuary: Three formae have been recognised and assembled into an intergradational series from a live population from Miramichi Estuary: E. excavatum forma clavata (pl. 5:1-7; pl. 23:4-7), forma lidoensis (pl. 5:8-12; pl. 26:7-9) and forma excavata (pl. 5:13-16). The specimens from Miramichi exhibited a wider ranged of variability than seen in the previous post-glacial/arctic samples and a larger number of the specimens could be considered to be morphologically intermediate between formae. Specimens of E. excavatum forma excavata from Miramichi were more irregular than those seen previously. These specimens greatly resemble the neotype illustrated by Lévy et al. (1975, Pl. 3:5-6).

Bay of Chaleur: An intergradational series was assembled from three live populations. Six formae are present, and this sample exhibits, on the whole, more ornamentation than any other sample studied, especially specimens of E. excavatum forma excavata (pl. 6:8-16,19). These specimens have many regular ponticuli and granular

material in the sutures and umbilici. E. excavatum forma excavata is linked through intermediate specimens to forma lidoensis (pl. 6:7), forma williamsoni (pl. 6:17-18; pl. 21:10-11) and forma cuwillieri (pl. 6:20). E. excavatum forma clavata (pl. 6:3-6; pl. 23:13) and forma magna (pl. 6:1-2) are also present, grading into one another, and forma clavata is linked to forma lidoensis (pl. 6:6-7).

Annapolis Basin: An intergradational series was assembled with specimens from two recent assemblages collected in the Annapolis Basin. The E. excavatum population exhibits a wide range of morphological variation in these samples, with many intermediate specimens.

Elphidium excavatum forma excavata (pl. 7:13-16; pl. 20:3) in these samples closely resembles the neotype described by Lévy et al. (1975, Pl. 3:5-6). E. excavatum forma selsevensis (pl. 7:1), forma magna (pl. 7:2-3), forma clavata (pl. 7:4-9; pl. 23:12) and forma lidoensis (pl. 7:10-12; pl. 26:1-2) were also observed.

Chezzetcook Inlet: An intergradational series was assembled from seven live populations collected from the mouth of Chezzetcook Inlet. Seven formae were recognized, five displaying a wide range of variability. Elphidium excavatum forma clavata (pl. 8:4-8), was present throughout the estuary in very low numbers. E. excavatum forma selsevensis (pl. 8:10-13) was also present throughout in low numbers, but became more prominent in the outer estuary. These specimens were the most irregular of the group; each with a large umbilicus, filled with bosses and papillae. The representative specimen bears a marked resemblance to specimens of E. selsevense

collected by Heron-Allen and Earland and illustrated by Banner and Culver (1978, Pl. 9:12-14).

E. excavatum forma magna (pl. 8:1-3; pl. 27:1-4) was first observed at this location (=Cribrononion excavatum incertum (Cushman, [not Williamson]) of Scott 1977, Scott et al. 1980) and is dominant in the nearshore turbulent zone. This forma best exhibits its characteristics in this area which is hereby designated the type area for E. excavatum f. magna.

Elphidium excavatum forma excavata (pl. 8:14-16; pl. 20:12) became more prominent in the intertidal zone; and E. excavatum forma lidoensis (pl. 8:9; pl. 26:3) was also present in low numbers throughout the area.

Isolated specimens of E. excavatum forma cuvillieri (pl. 28:23) were also observed. E. excavatum forma williamsoni (pl. 21:5-6) is rare in these particular samples but it is the dominant form in intertidal areas of the marsh (=Cribrononion umbilicatula (Williamson) of Scott 1977, and C. williamsoni (Haynes) of Scott and Medioli 1980).

A Maine-New Brunswick Estuary: Three formae were observed from a live population at this location and no intermediate specimens were observed. E. excavatum forma williamsoni was the dominant form (pl. 9:1-16; pl. 21:1-3) and some specimens with fewer chambers may be juveniles of this forma. Isolated specimens of E. excavatum forma lidoensis (pl. 9:17) and E. excavatum forma clavata (pl. 9:18) were also observed.

Long Island Sound: This live population contained an abundant (greater than 75%) E. excavatum with four intergradational formae

present, the dominant one being E. excavatum forma selsevensis (pl. 6,10:9-13; pl. 22:1-13). These specimens were large and irregular, and closely resemble specimens of E. selsevense from the Dovey Marshes illustrated by Haynes (1973, Pl. 22:3-4; Pl. 24:11; Pl. 26:4,7,9).

Elphidium excavatum forma excavata (pl. 10:6-8,14; pl. 20:4-7), forma lidoensis (pl. 10:15-16; pl. 26:3-5,10-11) and forma clavata (pl. 6:1-2) are present as minor constituents of the population.

San Diego Bay: Six formae were observed from a live population at this location. One of the dominant forms was E. excavatum forma tumidum (pl. 11:1-4, 15-16; pl. 27:13-19). Forma tumidum was linked by intermediate specimens to E. excavatum forma selsevensis (pl. 11:5), forma lidoensis (pl. 11:9-13, pl. 26:20-22,27) and forma clavata (pl. 11:6-8; pl. 23:20-21). The specimens of forma clavata are more irregular and ornamented than those found in colder environments.

Two other formae were also observed. E. excavatum forma gunteri (pl. 11:17-18; pl. 24:13) was present but no intermediate specimens linking them to the remainder of the group were found. Three other specimens, tentatively identified as E. excavatum forma galvestonensis (pl. 11:19-21; pl. 25:15-16) (identifications based on enlargements, pl. 25:15-16) were also observed. No intermediate specimens linking these to the remainder of the group were found.

San Francisco Bay: Seven formae were recognized from a Pleistocene core assemblage from San Francisco Bay. The dominant forms were E. excavatum forma excavata (pl. 12:1-3,12; pl. 20:8-10) and forma lidoensis (pl. 12:4-5,7-10; pl. 26: 21-24) which were linked

to forma clavata (pl. 12:11) and forma selsevensis (pl. 12:14-15).

One specimen was tentatively identified as E. excavatum forma tumidum (pl. 12:6), and another as E. excavatum forma williamsoni (pl. 12:19).

E. excavatum forma gunteri was also observed in this sample (pl. 12:16-18; pl. 24:21). As with the San Diego Bay sample, no intermediate specimens were observed here linking this forma to other members of the E. excavatum group.

Baie Verte, Northumberland Strait: This assemblage was the most unusual and interesting studied in terms of the E. excavatum fauna. Eight formae were observed and all could be assembled into an intergradational series (two series were assembled and illustrated here). Many intermediate specimens were observed; this coupled with the fact that some specimens were badly etched made identifying many of them to the forma level difficult and very subjective.

Elphidium excavatum forma gunteri (pl. 13:1-2; pl. 14:1-3; pl. 24:14-20) and E. excavatum forma galvestonensis (pl. 13:17-20; pl. 14:19-20; pl. 25: 11-14) were observed, with intermediate specimens (pl. 14:3 and pl. 13:17) linking these two formae to the remainder of the group.

E. excavatum forma clavata (pl. 13:3-5,8-9,16; pl. 14:4-5,13-14) was the most dominant form present, and exhibited considerable variability. Specimens of forma selsevensis (pl. 13:12-15; pl. 14:8-9,18) were similar to those seen in Long Island Sound. E. excavatum forma excavata (pl. 13:10; pl. 14:11-12), forma lidoensis (pl. 13:11; pl. 14:10; pl. 26:12-13) forma magna (pl. 14:15-17) and forma cuvillieri (pl. 13:6-7; pl. 14:6-7) were also observed.

San Antonio Bay: Four formae were identified from this live population and the two dominant formae were assembled into an intergradational series. E. excavatum forma gunteri (pl. 15:8-15; pl. 24:1-12) and forma lidoensis (pl. 15:1-7,16-20; pl. 26:14-19) were linked through numerous intermediate specimens. Both formae exhibit more variability than observed in specimens (of the same formae) from other locations. Some specimens of forma gunteri exhibit extreme variability in the development of the ponticuli (i.e. pl. 15:9; pl. 24:1-3); some ponticuli are not really as well developed on the ultimate and penultimate sutures. The specimens of forma lidoensis exhibit the key characteristics of the forma, but they resemble more those specimens found along the west coast of North America and the Mediterranean. The wall perforations are coarser, the peripheral outline more lobate, the sutures more depressed, and the papillae/bosses in the sutures and umbilicus are larger and more variable in these "Lusitanian" specimens versus the "boreal" environment specimens from maritime Canada and New England.

Two other formae were observed from this location, but they are morphologically isolated from the remainder of the group. E. excavatum forma galvestonensis (pl. 16:1-8; pl. 25:1-10) was observed to be more common from this location than from any other, and these specimens best exhibited the characteristics of the forma. E. excavatum forma cuvillieri was also observed, (pl. 16:9-14; pl. 28:22) more common at this location than at any other North American location. These specimens resemble those observed in the European samples. An unidentified species of Elphidium (pl. 16:15) was also

observed.

Wadden Sea: Two formae of E. excavatum were observed from this Wadden Sea Holocene assemblage. E. excavatum forma williamsoni (pl. 17:1-12; pl. 21:7-9,12-24) comprised over 95% of the E. excavatum population and was very variable. E. excavatum forma gunteri (pl. 17:13-15) was also observed. No intermediate specimens were observed.

Venice Lagoon: Four formae were identified (and one other tentatively identified) from a Venice Lagoon live population; two formae were assembled into an intergradational series. This sample contained E. excavatum forma gunteri (pl. 18:1-4, pl. 24:22-24) and forma lidoensis (pl. 18:5-7, pl. 26:23-25) which are similar morphologically to those specimens observed from San Antonio Bay, particularly the specimens of forma lidoensis, and the intermediate forms present (i.e. pl. 18:4). E. excavatum forma cuvillieri (pl. 18:8-15; pl. 28:11,13-14,16-21) and forma williamsoni (pl. 18:17) were also identified. The specimens of forma cuvillieri are extremely variable, particularly the sutures, ponticuli, and umbilical regions. Two specimens were observed that may be E. excavatum forma galvestonensis (pl. 18:16,18). These were identified by comparison with specimens from San Diego Bay.

Bay of Izmir, Turkey: Three formae were observed from a late Pleistocene assemblage from this location. All three are morphologically isolated and no intermediate specimens were observed. E. excavatum forma lidoensis (pl. 19:1-7; pl. 26:28-32), forma cuvillieri (pl. 19:8-15; pl. 28:1-10,12,15) and forma williamsoni (pl. 19:16-17) were found. The specimens of forma lidoensis are the

"Lusitanian" form; and specimens of forma cuvillieri are similar to those found living in Venice Lagoon.

DISCUSSION

The observations presented above suggest patterns in the degree of variability within the range of the E. excavatum group. Samples from colder waters display less variability than their counterparts from more temperate environments. Samples from nearshore estuarine locations display a wider range of variability and contain a larger number of intermediate forms than samples from more stable environments.

The range of variability observed in the three samples from Europe is much narrower. The formae are quite isolated and distinct at two of the locations (Wadden Sea and Bay of Izmir), no intermediate specimens linking any of the formae have been observed. At the third European location, Venice Lagoon, only E. excavatum forma gunteri and forma lidoensis can be linked, all other formae are isolated, as is the case with the sample from San Antonio Bay. These four samples come from locations where the annual climatic ranges are not extreme. The specimens displaying the widest range of variation are those from eastern North American temperate estuaries which are subject to extremes in climate and environmental conditions. It is only in this particular region that E. excavatum forma gunteri, galvestonensis, williamsoni and cuvillieri can be linked to the core formae (i.e. formae clavata and excavata) of the group. This apparent lack of intermediate forms may be one reason why some European workers (i.e.

Haynes 1973, 1982 pers. comm.; Murray 1979, 1982 pers. comm.) have not grouped some of these morphotypes (i.e. forma gunteri, williamsoni, or cuvillieri) with the remainder of the group. Another reason for not grouping may be the variation within the forma. For example E. excavatum forma lidoensis can be split into two "subforma": a "boreal" environment form from areas with extremes in climatic variation, i.e. Miramichi Estuary, Annapolis Basin and Long Island Sound; and a "Lusitanian" environment form, from areas with a narrower climatic range, i.e. San Diego Bay, San Antonio Bay, Venice Lagoon and Bay of Izmir. Both forms exhibit the key characteristic of the forma: sutures filled with papillae, broadening towards the umbilicus, giving the umbilicus a star shaped appearance. The "boreal" form found along the North Atlantic seaboard resembles, and can be linked to E. excavatum forma excavata; the wall perforations are fine and the papillae small. The ponticuli are more strongly developed on this form. The "Lusitanian" form resembles, and can be linked to E. excavatum forma gunteri; the periphery is rounded, wall perforations coarse, papillae more variable in size and a larger number of bosses present in the umbilicus. This "Lusitanian" form is the one most often seen by European workers, and in the European samples examined, the link to the core of the E. excavatum group is not apparent.

It is difficult to draw definite conclusions about the environmental preferences of some of the morphotypes. As Myers (1943) pointed out, there are many ecological parameters acting simultaneously upon several phases of the life cycle of foraminifera, making it difficult to estimate the possible effect of a single

variable while comparable changes in magnitude are taking place in other conditions. As noted by Raup and Stanley (1971), the same morphological variables may be under genetic control, or under control of the environment, making separation of genetic and non-genetic factors especially difficult. Separation of such factors is impossible in a situation suggested by Jardine and Sibson (1971), when the extent to which variables are environmentally modifiable is in itself under genetic control.

In very general terms, however, the following useful observations can be emphasized: 1) Elphidium excavatum forma clavata, the dominant member of the group and cosmopolitan form, is found in cold, normal marine waters or slightly reduced salinities; 2) forma excavata (a cosmopolitan form) is found as a minor constituent of the population in the intertidal zone; 3) forma williamsoni a very cosmopolitan form, is the dominant intertidal/marsh form where there is little wave action; 4) forma selseyensis is a temperate to polar water (1-16°C) estuarine form on both sides of the Atlantic; 5) forma lidoensis, also present on both Atlantic seaboard and the Pacific coast, is a warm to temperate water estuarine and lagoon form; 6) forma gunteri a cosmopolitan form, appears to replace forma clavata in temperate to tropical waters; 7) forma galvestonensis, a tropical, nearshore, lagoon form preferring normal to hyper-salinities, appears to be geographically isolated, being present mainly along eastern North America; a tropical, nearshore, lagoon form preferring normal to hyper-salinities; 8) forma cuvillieri appears to be a subtidal temperate to tropical normal marine form, common in the Mediterranean

and along the European (Atlantic) coast but occurring in the Gulf of Mexico too; 9) forma tumidum is observed only along the western North American coastline and 10) forma magna appears to be an arctic to temperate water nearshore turbulent zone form.

Feyling-Hanssen (1972) reported the following occurrences: 1) Elphidium excavatum forma clavata in arctic and subarctic waters from moderate depths; 2) forma selsevensis (= forma excavata) in the boreal environment; and 3) forma lidoensis in the Lusitanian regions. He noted that (1972 p. 339): "In all environments, variation in shape and sculpture of this species occur, but a certain pattern in the distribution of different forms is recognizable..... This pattern must also be of paleo-ecological significance, and for these reasons it must be considered of some importance to maintain a taxonomic separation between the major variations within the species".

Similar observations were made earlier by Bartlett (1963, 1964, 1965a, 1965b). Bartlett studied the occurrence of Elphidium incertum "complex" (= E. excavatum) of Tracadie Bay, Prince Edward Island (1965a) and the Scotian Shelf (1963, 1964); observing that differences in external morphology were apparently related to environmental parameters (Bartlett, 1965b). Bartlett found large opaque forms (forma magna) associated with turbulent nearshore environments (as did Scott 1977, Scott et al. 1980) or the outer shelf. In normal marine environments, such as inner shelves and open bays, Bartlett (1965b) observed translucent biumbonate forms with one or more umbilical bosses and translucent biumbilicate specimens (=forma clavata?). Back-bay and lagoonal specimens appeared to be smaller, extremely

variable in external morphology and often the umbilicus had a depressed, slit-like appearance (=forma excavata).

Wilkinson (1979) was of the opinion that he was dealing with two distinct species when he pointed out that the type descriptions of Elphidium excavatum and E. clavatum differ markedly. He compared the type figure of Polystomella excavata Terquem (1876), and the neotype described by Lévy et al. (1975) with Cushman's E. clavatum, and concluded that the morphological differences between the two were taxonomic and sufficient to justify specific separation.

Wilkinson (1979) reported the geographical distribution of the different morphological variants as having very little or no overlap and consequently designated them as subspecies. According to Mayr et al. (1953, p. 30): "Subspecies are geographically defined aggregates of local populations which differ taxonomically from other such subdivisions of a species. Not more than one subspecies of any one polytypic species can exist in breeding condition in any one area". In the populations studied, seven morphological variants of E. excavatum, including those that Wilkinson would regard as subspecies, are found concurrently living in Bay of Chaleur; six are present in San Diego Bay; five were observed in the Annapolis Basin, Chezzetcook Inlet, and San Antonio Bay; four were present in Long Island Sound and Miramichi Estuary; three were observed in the Maine-New Brunswick Estuary and Venice Lagoon; two were observed in the Beaufort Sea and off Liverpool, Nova Scotia. In addition, of the assemblages studied eight morphotypes were noted in the Baie Verte core sample; seven were present in San Francisco Bay core; four were observed in the Champlain

Sea sample; three were present in the Bay of Izmir core sample; and two formae were present in the Labrador Shelf core, and in the Hirtshals, Denmark and Wadden Sea samples.

Hence these distributions provide strong evidence that the morphotypes examined in this study do not fulfill Mayr's definition of subspecies. These morphotypes appear to be ecophenotypes, which are the result of non-genetic modification of the phenotype to specific ecologic conditions (Mayr et al. 1953). A similar interpretation was proposed by Feyling-Hanssen in 1972.

Wilkinson (1979) noted that E. excavatum (Terquem) and E. clavatum Cushman were morphologically distinct. Rodrigues and Hooper (1982, p. 415) also stated: "because no morphological series relating modern specimens of E. clavatum to either E. selsevensis or E. lidoense have been adequately documented in the literature, we choose to regard E. clavatum as distinct from E. selsevense and E. lidoense". As shown in this thesis however, these four "species" and six other often reported taxonomic units (E. williamsoni, E. gunteri, E. galvestonense, E. cuvillieri, E. tumidum and E. excavatum forma magna) as well as numerous other "species", can be accommodated into various intergradational series, which indicates only one highly variable species is present, E. excavatum (Terquem), which includes at least 10 ecophenotypes. Since the ecophenotypes of E. excavatum have paleo-ecological rather than taxonomic significance, it is suggested that Feyling-Hanssen's (1972) trinomial terminology, inclusive of an epithet for the forma, be retained.

Poag (1978) has studied ecophenotypy in the genera Ammonia,

Elphidium, Palmerinella, and Ammotium from San Antonio Bay, Texas.

(The San Antonio Bay samples studied here are some of the same samples, kindly made available by Dr. Poag). Poag's conclusions about ecophenotypy in Elphidium are different than those drawn here, probably for two reasons. One is the subjectivity of the methods used to delimit the ecophenotypes. When two or more morphotypes can be linked through an intergradational series, there is no set limits to each morphotype and the "morphological boundary" between two morphotypes is up to the author's discretion. The other reason may be the limited range of variability present in Poag's sample relative to the scope of the variability observed in the twenty samples described in this work. No one sample contains the whole spectrum of the species (indeed, neither does twenty samples) but the range of variability in twenty is usually greater than that observed in one. Consequently, the boundaries between morphotypes fall in different places. Arnold (1968) has emphasized that large natural populations must be studied throughout as much of the geographic range as possible to determine the incidence of each suspect variant.

Haynes (1981, p. 61-62) states that E. excavatum can be regarded as a "superspecies" comprising E. clavatum and E. selseyense and their allies as siblings of E. ex gr. excavatum. Alternatively, this group may be viewed as one polytypic species, E. excavatum subsp. gr.

Haynes states that intermediates do exist and that the distribution patterns he has observed suggest (p. 62): "a morphological continuum with distinctive 'end members' in different geographical areas that are conveniently regarded as separate species.". Haynes (1981)

continues: "... it is difficult, if not impossible to distinguish between 'subspecies' and 'ecophenotypes' which in any case are potential if not actual subspecies."

The observations made in this study contradict those of Haynes; they indicate that eight of the formae studied are not geographically isolated (a hypothesis that will be further investigated in the statistical study). Only two formae, tumidum and galvestonensis do not have a widespread occurrence throughout both Europe and North America. As stated previously, the presence of intergradational series makes it highly unlikely that these morphotypes are subspecies.

It has been suggested (see Haynes 1981) that live cultures are the only definite method of solving genetic vs. non-genetic variation problems. For some groups, this method has been successful (e.g. Ammonia beccarii, Schnitker 1974). However, the intergradational series technique applied to live populations is also effective because, in a sense, an "in situ" culture is being investigated. This technique has the advantage of examining a population from a natural environment rather than the artificial environment provided by laboratory cultures. It is difficult to duplicate all the conditions of the natural environment, and with so many variables, Myers (1943) considers it is virtually impossible to determine which one or combination of influences determines morphological variation. Hence, it is felt that the intergradational series techniques for this kind of taxonomic study is a useful and valid tool.

CONCLUSIONS

- 1) Information from the literature and observations here indicate that Elphidium excavatum (Terquem) is a highly variable species comprising at least nine (possibly ten) distinct ecophenotypes (formae). No definite conclusions are drawn at this time about the tumidum form; though it is treated as a forma of E. excavatum, no definite conclusions can be based on 11 specimens from two locations.
- 2) Distribution patterns of the formae suggest association with environmental variables rather than simple geographic locality (many formae can live in one area at times). These observations lead to the rejection of the subspecies ranks proposed by Wilkinson (1979) and Haynes (1981).
- 3) The designation of "forma" has been retained from Feyling-Hanssen (1972) and ten formae can be recognised with the binocular microscope:
 - 1) E. excavatum forma excavata (= forma selseyensis of Feyling-Hanssen) found as a constituent of populations in intertidal zones;
 - 2) forma williamsoni is the dominant intertidal/marsh form where there is little wave action;
 - 3) forma clavata, the dominant member of the group found in cold normal marine waters or slightly reduced salinities;
 - 4) forma selseyensis (not sensu Feyling-Hanssen), a temperate to polar water estuarine morphotype;
 - 5) forma gunteri, which appears to replace forma clavata in temperate to tropical waters;
 - 6) forma galvestonensis, possibly geographically isolated along the eastern North American coast, a tropical, nearshore lagoon form, preferring normal to hyper-salinities;
 - 7) forma lidoensis, a warm to temperate water estuarine and lagoon form;
 - 8) forma cuvillieri appears to be a subtidal temperate to tropical normal marine form;
 - 9)

forma tumidum which has been observed only along the western North American coast and 10) forma magna a nearshore, turbulent zone morphotype.

Variability in the E. excavatum group (number of formae and the percentage of intermediate forms present) appears to increase when each, or a combination of, the following variables is increased: water temperature, proximity to shore, estuarine influence and the range of annual climatic variation.

STATISTICAL INVESTIGATION OF INFRASPECIFIC VARIATION

PREVIOUS WORK

The earliest attempts to apply numerical methods to taxonomy date from the rise of biometrics in the last century (Sokal and Sneath 1963). As early as 1898 Heinke used a measure of phenetic distance to distinguish between races of herring. It was realized early on that biometrics could be applied to systematics (Sokal and Sneath 1963).

Foraminiferal taxonomy is based almost exclusively on characters of the shell, and foraminiferal shell structures have been most commonly analysed by univariate methods. These include linear measurements, enumerator data, ratios, and relative variability, all of which are explained by Scott (1974). Bivariate analysis, usually pairs of shell measurements analysed by linear regression, have also been used. Scott (1974) cites many examples of both univariate and bivariate methods (as applied to foraminifera); a good example is described by Gradstein (1974).

However, a form as complex as the foraminiferal shell requires many variates for its quantitative representation. The shell is an integrated structure and variates need to be considered simultaneously rather than in pairs. Multivariate methods are capable of doing this, and the development of electronic computational methods and equipment, makes multivariate analysis possible.

The many variates quantitatively representing the foraminiferal test can often be envisaged as a cluster of points in a multidimensional space. New variables "canonical variates" (plotted on canonical axes), are computed which are linear combinations of the

original variables, and are so oriented that the sample means are now at maximum distances apart (Scott 1974). Buzas (1966) computed three canonical variates based on invariant characters of four species of Elphidium; these canonical variates graphically represent a large proportion of intersample variance. Buzas (1966) proved that these four species were only three species statistically; two species, Elphidium clavatum Cushman and E. incertum Cushman (not Williamson) were not statistically separable.

To differentiate infraspecific populations exhibiting ecologic or geographic patterns to their distribution, various techniques of discriminant analysis (Fisher 1936) may be used to test the hypothesis that they can be differentiated (Jardine and Sibson 1971). Discriminant analysis constructs canonical variates from multivariate normal populations with common variances and covariances and allocates individuals among known populations. These new variates are linear and the populations are at maximum distances apart. Discriminant analysis is particularly applicable if the specimens to be allocated belong to one of the populations represented in the computation of the function; there is no provision for a specimen not belonging to one of the populations in question (Scott 1974).

Another advantage of discriminant analysis is that it has been successfully used previously in similar cases, where there is no convenient breaks in the sequence to allow the delineation of distinct morphotypes. Ashton et al. (1957) and Pritchard (1960) have applied discriminant analysis to infraspecific variation in anthropology and botany respectively; and Reyment (1973) in paleontology.

DISCRIMINANT ANALYSIS

WHAT IS IT?

Discriminant analysis is a statistical method for deriving one or more discriminant functions, each of which is a linear combination of two or more independent variables that will discriminate best between the a priori defined groups. This derivation is achieved by using a statistical decision rule to determine the maximum between-group variance relative to the within-group variance, i.e. to obtain the largest ratio of the between-group to within-group variance (Hair et al. 1979).

The general equation of a discriminant function is:

$$Z = w_1x_1 + w_2x_2 + w_3x_3 + \dots w_px_p \quad (\text{equation 1})$$

or

$$Z = \underline{x}'\underline{w} \quad (\text{equation 2})$$

where:

Z is the value of the discriminant function or discriminant score, for a defined group

w_p is the coefficient for the p^{th} variable, x_p

$\underline{x} = (x_1, x_2, x_3, \dots, x_p)'$ is the independent variables vector, and

$\underline{w} = (w_1, w_2, w_3, \dots, w_p)'$ is the coefficients vector.

To obtain the discriminant function, each independent variable is multiplied by its corresponding weight and these products are added together. If there are more than two groups, more than one discriminant function is needed to separate the groups. In general, there are a maximum of $g-1$ functions (where g is the number of groups). The result is a single composite discriminant score for each function for each individual in the analysis. The discriminant

function tests the hypothesis that the means of the two (or more) groups are equal. By averaging the discriminant scores for all the individuals within a particular group, the group mean, called the group centroid, is obtained. The test for statistical significance of the discriminant function is a generalized measure of the distance between the group centroids, and is computed by comparing the statistical distribution of the discriminant scores in each group. If the overlap in the distributions is small, the discriminant function separates the groups well. If the overlap is large, the function is a poor discriminator between the groups (Hair et al. 1979).

A good mathematical description of discriminant analysis is given in Sneath and Sokal (1973).

There are a few assumptions made in the application of discriminant analysis. One is that there is multivariate normality of the distributions and equal dispersion and covariance structures for the groups. Scott (1974) says that the assumption of equal covariance matrices seldom holds for foraminiferal data; causes and reasons for non-homogeneity are discussed by Reyment (1962, 1969). Another assumption is that there are equal a priori group probabilities. However, discriminant analysis is not overly sensitive to violations of these assumptions unless the violations are extreme (Hair et al. 1979). The assumption that there is equal probability of an unknown sample belonging to any group is the most difficult to justify when taxonomic data are used (Davis 1973), but tests for this probability are beyond the scope of this work.

GEOMETRIC REPRESENTATION

A graphical illustration of a two-group analysis may help elucidate the procedure just described. Figure 2 represents a scatter diagram and the projection resulting from computation of a discriminant function.

There are two groups, a and b, and two variables, x_1 and x_2 , measured for each specimen of the two groups. The x_1 , x_2 values are plotted for each specimen. By finding a linear combination of the original variables x_1 and x_2 , the result can be projected onto a new axis, representing the discriminant function, Z, drawing a straight line through the two points where the ellipses encircling the data for each group intersect. When the data points for groups a and b are projected onto the new Z axis, they condense the information about group differences into a set of points on a single axis (Hair et al. 1979). The overlap between the univariate distribution a' and b' is smaller than that obtained by any other line drawn through the scatter plots (Green and Tull 1975).

PROCEDURE

The application of discriminant analysis can be divided into three major stages: (1) derivation, (2) validation and (3) interpretation (Hair et al. 1979). The derivation stage involves determining whether or not a discriminant function which is statistically significant can be derived to separate the groups. The validation stage involves developing a classification matrix to further evaluate the predictive accuracy of the discriminant function. The interpretation stage involves determining which of the independent variables contribute the most toward discriminating between the groups

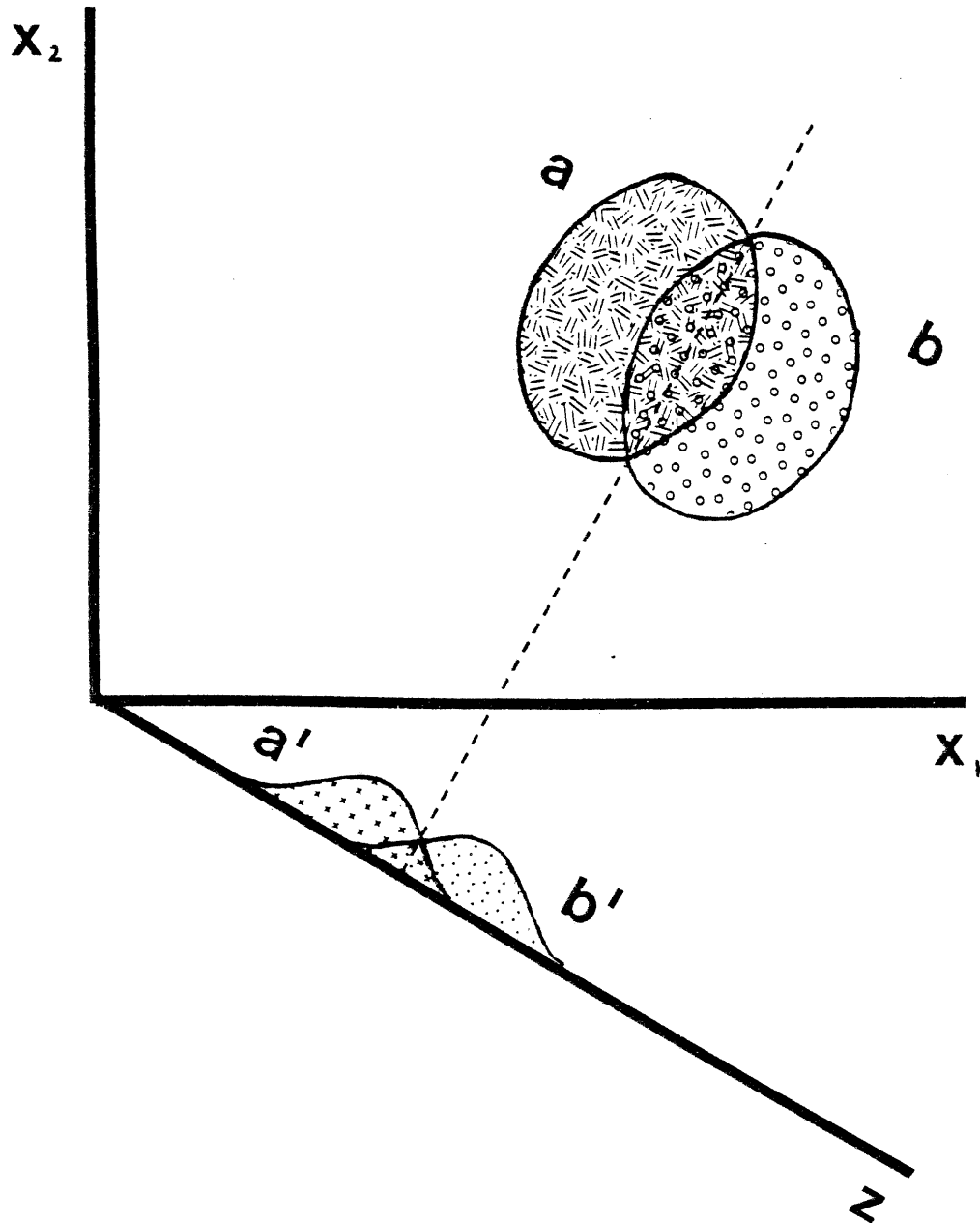


Figure 2: A two group analysis. A scatter diagram for characters x_1 and x_2 , obtained for all the individuals in groups a and b ; and the projection resulting from the computation of a discriminant function (from Hair et al. 1979).

(Hair et al. 1979).

DERIVATION

This stage consists of several steps: a) variable selection, b) sample division, c) computational methods, and d) statistical significance (Hair et al. 1979).

Variable selection: To apply discriminant analysis, the analyst must first specify which variables are to be independent variables and which is to be the dependent variable. The dependent variable should be chosen first. There can be two or more categories or groups of the dependent variable but these groups must be mutually exclusive and exhaustive. The independent variables must then be chosen. These variables can be selected in two ways. Variables identified from previous research or from a theoretical model (which is the underlying basis of the research question) can be employed. The second method of choosing variables is intuitive, based on trying to extend the researcher's knowledge. In both methods, those variables are selected which logically might be related to predicting the groups for the dependent variable (Hair et al. 1979).

Sample division: The discriminant function (or functions) must be tested for statistical validity. One procedure (the split sample method) involves developing the discriminant function(s) on one data set and then testing it (them) on another (Frank et al. 1965). The first data set, the analysis sample, is used to develop the discriminant function. The second set, the holdout sample, is used to test the discriminant function.

Frank et al. (1965) point out that an upward bias will occur in

the prediction accuracy of the discriminant function if the same individuals are used in developing the classification matrix as were employed in computing the discriminant function. If the split sample method is not used, then the classification accuracy will be higher than is valid for the discriminant function.

There are no definite rules for dividing the data into analysis and holdout samples. One can employ a 50 - 50, 60 - 40, or 75 - 25 split between the two groups, respectively. However, when selecting individuals for the two samples, a proportionally stratified sampling procedure is usually followed. If the categorical groups of the dependent variable are not equally represented in the total sample, then the size of the groups within the holdout sample should be proportional to that group's representation within the total sample (Hair et al. 1979).

The most frequent procedure utilized in validating the discriminant function is to divide the groups randomly and run the analysis only once. Frank et al. (1965) suggest that greater confidence could be placed on the validity of the function(s) if the above procedure were followed several times.

There are other more sophisticated methods for validating discriminant functions, a summary of these methods can be found in Crask and Perreault (1977).

Computational method: There are two computational methods that can be utilized in deriving the discriminant functions: the simultaneous (direct) method, and the stepwise method (Hair et al. 1979).

The simultaneous method involves computing the discriminant function so that all of the independent variables are considered

concurrently. The discriminant function(s) is computed using the entire set of independent variables, regardless of the discriminating power of each. The simultaneous method is appropriate for the initial analysis, when the function(s) is derived. Once the discriminating power of the function(s) is ascertained, the stepwise method might be employed to see the intermediate results based on only the most discriminating variables (Nie et al. 1975).

The stepwise method involves entering the independent variables into the discriminant function(s) one at a time, based on their discriminant power. The first step is for the computer to determine the single best discriminating variable. This initial variable is then paired with each remaining variable until a second variable is chosen that best improves the discriminating power of the function in combination with the first variable. The third and subsequent variables are selected in a similar manner. As additional variables are included, some previously selected variables may be removed if the information they contain about group differences is available in some combination of other included variables. Eventually, all variables will have been included in the function(s), or excluded if they do not contribute significantly to the discriminating power of the function(s) (Nie et al. 1975).

The mathematical derivation of the discriminant function(s) (as derived from first principles) is given in Appendix B.

Statistical significance: After the discriminant function has been derived, its level of significance must be assessed. If the function is not significant at or beyond the 0.05 level, there is little justification for continuing to the validation and interpretation

stages because there is little likelihood that the function will classify accurately (Hair et al. 1979).

VALIDATION

Once the discriminant function(s) have been developed, the statistical significance of the function(s) must be determined. In the SPSS program, the statistic used is a chi-square, χ^2 (Nie et al. 1975). However, the level of significance of this statistic is a poor indication of the function's ability to discriminate between the groups (Hair et al. 1979). With large sample sizes, the group means (centroids) could be almost identical and there would still be a statistically significant difference with the χ^2 test. With sufficiently large sample size, there could be a significant (χ^2) difference between two (or more) groups and yet, for example, only 53 percent would be correctly classified (when chance is 50 percent for two equal sized groups) (Morrison 1969). For reliable classification the classification matrices should be developed to provide a more accurate assessment of the discriminating power of the function (Hair et al. 1979).

The validation stage involves several steps. These include: a) construction of classification matrices, b) using chance models to determine the expected percent of correctly classified specimens and c) assessing the classification accuracy relative to the chance of random group assignment (Hair et al. 1979).

Construction of classification matrices: To validate the discriminant functions, classification functions are developed and evaluated. As mentioned earlier the analysis sample was used to compute the

discriminant function(s). The holdout or validation sample was retained for use in developing the classification matrix. The general equation for a classification function is:

$$\ln \frac{1}{g} + (\mathbf{x} - \bar{\mathbf{x}}_i)' \mathbf{W}^{-1} \bar{\mathbf{x}}_i \quad (\text{equation 3})$$

where:

g is the number of groups

\mathbf{x} is the independent variable vector (for the specimen being tested)

$\bar{\mathbf{x}}_i$ is the mean vector for the independent variables of the i th group, and

\mathbf{W} is the within groups variance-covariance matrix.

Derivation of the general equation is given in Appendix B. The classification rule is to evaluate these g functions for each specimen, and classify the specimen with independent variables \mathbf{x} into the group which gives the largest function value. The $g \times g$ classification matrix tabulates for each group the number of specimens classified into each group (correctly classified when assigned to its own group and incorrectly classified when assigned to another group). The hit ratio, which is the proportion correctly classified, is the sum of the diagonal entries on this matrix divided by the sample size. For a two group case, it can be shown that the classification functions and discriminant functions are essentially the same (Appendix B).

Chance models: Another factor that must be considered is the percentage of specimens that would be correctly classified by chance. When the sample sizes of the groups are equal, the determination of the chance classification statistic (c) is simply obtained by dividing

1.0 by the number of groups (ie: $c=1/g$). For the case where the group sizes are unequal, c can be based on the sample size of the largest group, referred to as the maximum chance criterion (Hair et al. 1979). When the sample sizes are unequal and all specimens are to be classified, the discriminant function defies the odds when classifying a specimen into a smaller group; a factor which should be taken into account (Morrison 1969).

If the percentage of correct classifications is significantly larger than would be expected by chance, an attempt can be made to interpret the discriminant functions. Hair et al. (1979) suggest that as a rough estimate, the classification accuracy should be at least 25 percent greater than by chance; this criterion is easy to apply to equal sized groups (Hair et al. 1979).

INTERPRETATION

If the discriminant function is statistically significant and the classification accuracy acceptable, then the results can be interpreted. The discriminant functions should be examined to determine the relative importance of each of the independent variables in discriminating between groups. Three methods have been suggested for determining the relative importance of these variables: a) standardized discriminant weights, b) discriminant structure correlations, and c) partial F-values (Hair et al. 1979).

Discriminant weights: The traditional approach to interpreting discriminant functions involves examining the sign and magnitude of the standardized discriminant weights (or function coefficients), calculated for each variable in computing the discriminant functions.

Independent variables with relatively larger weights contribute more to the discriminating power of the function than do variables with smaller weights regardless of the sign (+ or -) Hair et al. 1979). There are two drawbacks associated with emphasizing discriminant weights. A small weight may either mean that its corresponding variable is irrelevant in determining a relationship; or that the variable has been partialled out of the relationship because of a high degree of multicollinearity (Hair et al. 1979).

Discriminant loadings: Discriminant loadings (or structure correlations) measure the simple linear correlation between each independent variable and the discriminant function(s). The discriminant loadings reflect the variance the independent variables share with the discriminant function, and can be interpreted like factor loadings in assessing the relative contribution of each independent variable to the discriminant function (Hair et al. 1979).

Partial F-values: As discussed earlier, there are two computational approaches that can be utilized in deriving the discriminant function(s) - simultaneous and stepwise. When the stepwise method is selected, the relative discriminating power of the independent variables is measured through the use of partial F - values. Large F-values would indicate greater discriminating power.

ANALYSIS OF FORAMINIFERA DATA

COMPUTER PROGRAMS

The statistical analysis was completed using a FORTRAN language

computer program from SPSS - "Statistical Package for the Social Sciences". It is a prepared program that performs a specified set of operations, under the control of a simplified set of instructions (Klecka et al. 1975). The major features of SPSS: data collection, control cards, data and system files, etc., are explained in the SPSS Primer (Klecka et al. 1975). A complete description of the SPSS system including the subprogram DISCRIMINANT is given in "the SPSS manual", "SPSS: Statistical Package for the Social Sciences" (Nie et al. 1975).

To aid in understanding the SPSS output, an illustrative example was devised (see Illustrative Example - TEST) and most of the SPSS computations for this TEST example were duplicated using another statistical package, MINITAB. MINITAB is a step-by-step interactive package (Ryan et al. 1981). The computations reproduced from the SPSS output using MINITAB will be outlined in detail later (see Appendix C: TEST Calculations).

The computing was done on the CDC Cyber 170 computer at Dalhousie University.

DATA COLLECTION

There are many comprehensive textbooks on numerical taxonomy (among them Sokal and Sneath 1963, Jardine and Sibson 1971, and Sneath and Sokal 1973). These all discuss the philosophy behind numerical taxonomy and the theoretical considerations of data collection, and character selection, measurements, coding and ranking, etc. which will not be discussed here. Scott (1974) discusses character selection and measurement in detail for foraminiferal biometric studies and the

reader wishing to pursue the matter is referred to these works or those of his choice.

The first step was to choose the dependent variable and it was decided, based on the observations and subjective taxonomy to make "morphotype" the dependent variable (group) for most of the analyses. However, morphotype was treated as an independent variable in two of the analyses, and for these two analyses, location was the dependent variable (otherwise location was treated as an independent variable). Selection and measurement of the independent variables will be described in detail in the following section.

There were two alternatives available for the actual measurement of the independent variables. One was to take the measurements directly from the specimens observed under a microscope, a method employed by Buzas and Culver (Buzas 1966, Buzas and Culver 1981 pers. comm.). The other method was to obtain scanning electron microscope (SEM) photographs of each specimen and make the measurements from those photographs. Scott (1974) discusses the advantages of this method and lists authors who have had success completing biometric studies with the aid of the SEM. This second method was chosen because it is easier to return to a photograph to verify measurements than to relocate a specimen under a microscope. However, the disadvantage of photographs is that photos are often distorted, because the specimen was not centered, or because the specimen was damaged during the preparation for or during photographing. (In this study, photos of 89 specimens were rejected for these reasons.) If the first method had been employed, the measurements may have been more accurate, but also more difficult to make and reproduce.

VARIABLE SELECTION and MEASUREMENT

In this study, the dependent variable is usually the "morphotype" or group, to which specimen belongs. As described in the first part of the thesis, ten morphotypes (ecophenotypes) of Elphidium excavatum were recognized using a subjective method. Even without exact knowledge of the degree of overlap between the typological units and those to be generated in the analysis, the practice of Gradstein (1974) was followed, that of retaining the ten epithets, of the "subjectively" defined morphotypes and using these names for the "statistically" defined morphotypes. This has the advantage that a set of labels is already available (Gradstein 1974).

Each specimen was assigned to one of these ten morphotypes (1-10) as outlined on Table 2. In addition, for three of the analyses, three other species were added (11-13, Table 2). These were Elphidium bartletti, E. subarcticum and Haynesina orbiculare. The group codes for these species are also given on Table 2.

Throughout the remainder of the statistical investigation, the morphotypes will be referred to as groups, and the groups will usually be referred to by code number only. The Fortran code for group is FORM (from 'forma'). All other variables were also assigned a code name. Sixteen independent variables were chosen, in consultation with Dr. S.J. Culver, Dr. M.A. Buzas (Culver and Buzas 1981 pers. comm.), and Dr. C.T. Schafer (Schafer 1982 pers. comm.). In addition, variables were chosen with reference to previous work by Buzas (Buzas 1966). The data were measured (ranked/scored) so they could be utilized directly in the SPSS system .

<u>Number (FORM)</u>	<u>Morphotype or Group</u>
1	<u>clavata</u>
2	<u>excavata</u>
3	<u>selseyensis</u>
4	<u>lidoensis</u>
5	<u>magna</u>
6	<u>gunteri</u>
7	<u>galvestonensis</u>
8	<u>cuvillieri</u>
9	<u>williamsoni</u>
10	<u>tumidum</u>
11	<u>H. orbiculare</u>
12	<u>E. subarcticum</u>
13	<u>E. bartletti</u>

Table 2: Code numbers used for each of the ten morphotypes (or groups) of Elphidium excavatum and three other Elphidiidae species in the analysis.

Ten of the independent variables are qualitative, six are quantitative. Kendall and Stuart (1966) state that a set of mixed variables (some qualitative, some quantitative) can't be processed satisfactorily by discriminant functions, but Nie et al. (1975) make no mention of these restrictions; in fact, their examples contain both.

The first qualitative variable is the location (LOC) the specimen was collected from. Specimens were from 20 samples from 19 different locations (see Table 1) and were assigned code numbers for the purpose of the analyses, as outlined on Table 3.

The remainder of the qualitative variables were observations made from the SEM photographs, and were given arbitrary scores or ranks, as explained below. Table 4 lists all the variables, their codes, and possible ranks or scores. Table 5 refers to illustrations of the variables.

PAP denotes the presence (scored as 1) or absence (scored as 0) of papillae anywhere on the test surface other than directly within the suture. UMC0 denotes the presence (1) or absence (0) of an imperforate collar of test material surrounding the umbilicus.

Seven variables were given arbitrary rankings. The depression of the umbilicus (DEUM), was measured as depressed (1), flush with the test wall (2), or raised (3). The density of the wall pores (POR) was ranked as very fine (4-barely seen in photo), fine (3), medium (2), and coarse (1). The angle of the margin (AOMA) was ranked as acute (1) or subacute (2). The peripheral outline (PERO) was ranked as completely smooth (1); slightly lobate or the last few chambers lobate (2); or markedly lobate or more than one half the chambers in the

<u>Number (LOC)</u>	<u>Location</u>
1	Beaufort Sea - Vilks
2	Beaufort Sea - Bartlett
3	Hirtshals, Denmark
4	Labrador Shelf
5	Labrador Shelf
6	Miramichi Estuary
7	Northumberland Strait
8	Annapolis Basin
9	Chezzetcook Inlet
10	Long Island Sound
11	Bay of Chaleur
12	San Diego Bay
13	Bay of Izmir (Turkey)
14	Champlain Sea
15	Wadden Sea
16	Venice Lagoon
17	Maine-New Brunswick Estuary
18	San Antonio Bay (Texas)
19	Liverpool, Nova Scotia
20	San Francisco Bay

Table 3: Code numbers for the variable locations. Location 4 refers to a gradational sequence from a Labrador Shelf core, Location 5 is an intergradational series from the same core. Both of these are discussed and illustrated elsewhere (Miller 1979, Miller et al. 1982).

dependent or independent	discrete or continuous	qualitative or quantitative	code	possible scores or rankings
dependent	discrete	qualitative	FORM	1 - 10
independent	discrete	qualitative	LOC PAP UMCO AOMA SUT DEUM PERO DEPO REPO POR	1 - 20 0, 1 0, 1 1, 2 1, 2 1, 2, 3 1, 2, 3 0, 1, 2, 3 0, 1, 2, 3, 4 1, 2, 3, 4
		quantitative	CHAM PONT NOBO	count count count, then classed 0, 1, 2, 3, 4
			POSU	ratio, then classed
	continuous	quantitative	GSD GS90	0, 1, 2, 3, 4 measurement measurement

Table 4: Summary of variables used in the analysis.

Table 5: Listing of specimens illustrating possible independent variable ranks or scores. Plate and figure listings refer to this work.

Variable and code	Rank/Score	Illustration
Forma (FORM)	1-10	See Table 2
Location (LOC)	1-20	See Table 3
Papillae (PAP)	0 1	pl. 24: 11,13; pl. 29: 1,8,22; pl. 24: 12,15; pl. 27: 3,13,32
Imperforate (UMCO) Collar	0 1	pl. 21: 2,4,6; pl. 27: 5,17 pl. 24: 2,6,13,16,19
Depression of (DEUM) Umbilicus	1 2 3	pl. 21: 1,4; pl. 24: 1,3 pl. 24: 6,14; pl. 29: 6,9,20 pl. 25: 9,11; pl. 28: 3,7,11
Wall Pore (POR) density	1 2 3 4	pl. 25: 5,13; pl. 27: 14,17 pl. 24: 2,21; pl. 29: 6,9,22 pl. 24: 11,14; pl. 28: 14,15 pl. 22: 3,6,14,20
Angle of (AOMA) Margin	1 2	pl. 21: 5,9; pl. 24: 2,13,15 pl. 25: 9,11; pl. 28: 3,4,7
Peripheral (PERO) Outline	1 2 3	pl. 24: 2,11; pl. 28: 1,10 pl. 27: 5,23; pl. 29: 8,15 pl. 23: 1,3; pl. 29: 16
Curvature of (SUT) Sutures	1 2	pl. 24: 4,8,19; pl. 28: 2,5 pl. 27: 12,13; pl. 29: 16,22
Development of (DEPO) ponticuli	0 1 2 3	pl. 27: 21,23,26 pl. 23: 14; pl. 27: 5,7,13 pl. 21: 1,2; pl. 24: 12,13 pl. 24: 8,15; pl. 29: 8,10,20
Regularity of (REPO) ponticuli	0 1 2 3 4	pl. 24: 2; pl. 27: 21,23 pl. 21: 12; pl. 24: 3; pl. 27: 19 pl. 25: 13,22; pl. 28: 5,7,14 pl. 25: 23; pl. 28: 12,17 pl. 22: 5,8,16; pl. 29: 12
Number of (NOBO) bosses (classed)	0 1 2 3 4	pl. 27: 3,8; pl. 29: 8,10,22 pl. 24: 2,4,8; pl. 26: 4,13 pl. 24: 11; pl. 27: 19, 32 pl. 25: 11; pl. 27: 14; pl. 28: 12,16 pl. 27: 24,28; pl. 28: 14
Number of ponticuli/ suture (POSU)	0 1 2 3 4	pl. 24: 2; pl. 27: 26 pl. 21: 12; pl. 24: 3; pl. 27: 21 pl. 23: 2,13; pl. 27: 19 pl. 25: 11,14; pl. 28: 7,17 pl. 22: 6,11,14; pl. 25: 9

final whorl lobate (3). The sutures (SUT) were ranked as straight or with more than one half the sutures straight (1) or more than one half the sutures curved (2). The ponticuli (DEPO) were ranked as absent (0), poorly developed or indistinct (1), distinct but not completely spanning the suture (2) or more completely developed and extending all the way across the suture (3). The regularity of the ponticuli (REPO) were ranked as absent (0), very irregular (with some sutures having ponticuli absent, others having many ponticuli) (1), medium regular (2), very regular (3), and extremely regular (4).

Four of the quantitative variables were actual counts or measurements taken from the photographs. Two were discrete counts: the number of chambers (CHAM) and the number of ponticuli observed on the final whorl (PONT). Two variables were size measurement readings, the greatest spiral diameter (GSD - the largest diameter measured through the umbilicus and usually through the final chamber), and the diameter measured 90° to the greatest spiral diameter (GS90).

Finally, there were two measurements used that were highly variable, and in some cases difficult to measure. For these two variables, classes were erected. One variable was the number of bosses (NOBO). It was often difficult to distinguish between an umbilical boss and papillae in the umbilicus. Consequently the number of bosses per specimen was counted and classed accordingly: 0 bosses = 0; 0 - 2 bosses = 1; 2 - 4 bosses = 2; 4 - 8 bosses = 3; and 8 - 16 bosses = 4. This same class system was applied to the ratio POSU (= number ponticuli / suture). In effect, this is a measurement of REPO, and might make REPO redundant. If the ratio was less than 1, the class is 0, 1 to 2 = 1; 2 - 4 = 2; 4 - 8 = 3; and 8 - 16 = 4.

This system of classing variables, and of classing these variables in particular has been employed by Buzas (1966).

For five of the analyses indicator or dummy variables were created; dummies were created for location in four analyses and for forma in one analysis. In this instance each location (or forma) was represented by a new independent variable with value one if the specimen was from that particular location (or of that forma), and with value zero otherwise. When location was coded as dummy variables (DUM1 to DUM20) each with score one, LOC was not included in the analysis. Similarly when forma was treated as dummy variables, there were 10 additional independent variables (DUM21 to DUM30) in the analysis and FORM was not included.

VARIABLE TRANSFORMATIONS

The quantitative independent variables were tested for constant variance by plotting each group mean (for each variable) against its standard deviation. PONT, did not have a constant variance. However, the square root transformation of PONT, POSQ did have a constant variance and was used in place of PONT in all analyses.

There is a general tendency in biological populations for variance to be a function of the mean (Scott 1974).

Some of the quantitative variables are related to ontogenetic development, or the growth stage of the organism. These include CHAM, GSD and GS90. The measurements of the continuous variables depend on the ultimate growth stage which the individual attained (Gradstein 1974). CHAM, GSD, and GS90 become greater with increasing test size, preventing general conclusions from being drawn from the comparison of

means based on the raw data. If possible, the effects of ontogenetic development should be removed (Gould 1970). Because only adult specimens were chosen for this study (based on Buzas's [1966] criteria for designating a specimen "adult"), the effects of age were minimized in this work. The two variables measuring size, GSD and GS90 were highly correlated so a new variable GSR, the ratio $GS90/GSD$ was computed and used in all analyses, in place of GSD and GS90. None of the other variables (qualitative or quantitative) correlated highly with one another.

ANALYTICAL FEATURES OF SUBPROGRAM 'DISCRIMINANT'

For each analysis a set of basic or core operations were performed. Some of these operations are referred to as "statistics", others are referred to as "options". The statistics and options calculated are given on Table 6.

Subprogram DISCRIMINANT always prints the standardized discriminant function coefficients. They are used to compute the discriminant score for a case in which the original discriminating variables are in standard form (Z scores). The coefficients have been derived in such a way that the discriminant scores produced are in standard form (Nie et al. 1975). For each function the overall mean is zero and the standard deviation is one. Discriminating variables are not coded in standard form, and standardized function coefficients may not be very useful for computational purposes. Option 11 prints unstandardized function coefficients, which when multiplied by the raw values of the independent variables give the unstandardized

Table 6: Statistics and options calculated/performed in the analyses (from Nie et al. 1975).

Statistics/options calculated in all analyses	
Statistic/Option	Operation/calculation performed.
Statistic 1	Group means - means of all values of the dependent variable and for the total set of cases.
Statistic 2	Standard deviations for each group and the total set of cases.
Statistic 3	Pooled within - groups covariance matrix.
Statistic 6	Univariate F ratios. One-way analysis of variance test for equality of group means on a single discriminating variable. An F is printed for each variable.
Option 1	All missing value declarations are ignored. All cases included during the stepwise and analysis phases, provided they satisfy the GROUPS specification. All cases are classified regardless of their group assignment.
Option 5	Print classification results table indicating for each group the number of cases classified into each of the groups and the percent correct classifications for the known groups.
Option 6	Print discriminant scores and classification information for each case. This includes case identification (subfile name and sequence number); group number of the group the case actually belongs to; group number (G) of the closest group; the probability of a case in group G being that far from the group centroid denoted by $P(X/G)$; the probability of the case being in group $P(G/X)$; if the probability of membership in the second closest group is greater than .0005, that probability and the number of the second closest group is provided; and the discriminant scores.
Option 7	Print a single plot of cases. For one function, this plot is a histogram of the distribution of cases along the function. For two or more functions, a scatter plot of the first two discriminant functions is printed.
Option 8	Print a separate plot for each group
Option 10	Print territorial map. Again for more than two functions, a plot for the first two discriminant functions is printed.
Option 11	Print unstandardized discriminant function coefficients. These are the coefficients to be used in computing the discriminant scores from raw data. The constant to be added as an adjustment for the variable means is also printed.
Option 12	Print classification functions. These produce classification scores when used with raw data from the discriminating variables. The constant to be added as an adjustment for the variable means is also printed.
Option 17	Output discriminant scores.
Option 18	Output membership probabilities for all groups.
Statistics and Options used in some of the analyses	
Statistic 7	Test for equality of group covariance matrices. This is Box's M and its associated F test. This statistic is computed for the covariance matrices based on the discriminating variables. If Option 14 is in effect this statistic is also computed for the covariance matrices based on the discriminant functions.
Option 14	Use individual group covariance matrices for classification. Instead of the pooled within groups covariance matrix in computing the probabilities of group membership.

discriminant score. When the overall mean is adjusted to zero (by addition of a constant) the standardized score is obtained.

It is not necessary that all variables be included in all analyses. Those variables included are given on the the VARIABLES LIST. All the independent variables were included in the analyses except where specified.

Using the DIRECT method all the variables specified are entered concurrently into the analysis.

In the STEPWISE procedure, the independent variables are selected for entry into the analysis on the basis of their discriminating power, either by the subprogram itself, or by specifying the order (by inclusion levels) with the ANALYSIS card.

For most of the STEPWISE analyses the ANALYSIS card was not used. The analyses where the entry order was specified will be discussed later.

There are five stepwise selection criteria. Only one method, the Wilks' lambda was used for stepwise analysis, because this method was the most similar to the DIRECT method. When METHOD = WILKS, the criterion is the overall multivariate F ratio for the test of differences among the group centroids. The variable which maximized the F ratio also minimized Wilks' lambda, a measure of group discrimination (Nie et al. 1975).

When a STEPWISE analysis is performed it is necessary to specify six other parameters or allow them to default. For all of the STEPWISE analyses four of the parameters were allowed to default. These were: TOLERANCE (tolerance level for stepwise selection, default value .001); MAXSTEPS (maximum steps for the stepwise procedure

(default is twice the number of variables in the analysis); FIN (minimum F to enter, default value 1.0); and FOUT (minimum F to avoid removal, default is 1.0). Two other parameters, PIN (maximum significance level of F-to-enter, default value 1.0) and POUT (maximum significance level of F-to-remove to avoid removal, default is 1.0) were allowed to default on some analyses, and specified on others (which will be discussed later). All of these parameters are explained fully in Nie et al. 1975.

There are additional controls which can be imposed on the analysis. Two parameters which must be specified are the number of discriminant functions to be derived and the percentage of variation that these functions must account for. These are specified under FUNCTIONS.

The maximum number of discriminant functions to be derived is either one less than the number of groups (i.e. $g-1$) or equal to the number of variables, whichever is smaller. The dependence on the number of original variables is due to the mathematical impossibility of creating more new variables. The importance of the number of groups stems from geometric principles, that the maximum number of dimensions needed to completely describe a set of points is one less than the number of points (Nie et al. 1975). In all the analyses, the maximum number of functions were allowed.

The subprogram DISCRIMINANT provides two measures for judging the importance of the discriminant functions. One is through the eigenvalue computed in the process of deriving the discriminant functions. The sum of the eigenvalues is a measure of the total variance existing in the discriminating variables. A single

eigenvalue expressed as a percentage of the total sum of the eigenvalues is a measure of the relative importance of the associated function. Discriminant functions are derived in order of importance, and the process can be stopped when the relative importance is judged too small (Nie et al. 1975). In these analyses the program was instructed to account for all the variance, i.e. no function was judged too small.

A second criterion for eliminating discriminant functions is to test for the significance of discriminating information not yet accounted for in the derived functions. As each function is derived, starting with zero functions, Wilks' lambda is an inverse measure of the discriminating power in the original variables which has not yet been removed by the discriminant functions - the larger lambda is, the less information remaining. Lambda can be transformed into a chi-square statistic for an easy test of statistical significance. Functions that are not statistically significant can be removed. However, if the number of functions are specified, that number of functions is computed regardless of the values for the relative percentage and the significance level (Nie et al. 1975).

In the classification phase of the analysis, there is one parameter to be specified, and this is the PRIORS specification. This refers to the a priori estimate of group membership. There are three ways of inputting the a priori probabilities. When PRIORS = EQUAL (or PRIORS not included) the a priori probabilities are considered equal and no adjustments made. When PRIORS = SIZE, adjustments are made on the basis of prior probabilities being proportional to the number of cases in each group, i.e. more cases will be assigned to a larger

group. Alternatively, a set of prior probabilities can be provided. PRIORS was allowed to default (to EQUAL) for all of the analyses.

ILLUSTRATIVE EXAMPLE - TEST

It was realized fairly early on that a massive data set, consisting of 15 independent variables (plus the two dependent/independent variables, location and forma) for each specimen, and specimens representing 10 different morphotypes (groups) would produce very complicated calculations that may be difficult to validate and interpret. Validation and interpretation of the results would not be complete without understanding the derivation and calculation of the discriminant and classification function(s). However, understanding and duplicating the calculation of up to nine functions each containing up to 15 independent variables is a major undertaking in itself. Instead, an illustrative example was designed, one that was representative of the population data set, but was simple enough so that the discriminant and classification functions could be derived, understood, and the calculations duplicated. This illustrative example is called TEST. The objectives of TEST were to concentrate on the first stage, the derivation stage of the discriminant analysis, and to explain and duplicate the calculation of the various discriminant and classification functions on the SPSS output, using MINITAB.

TEST CALCULATIONS

A sample population (TEST) was set up consisting of 190 specimens (114 of group one [Elphidium excavatum forma clavata], 29 of group

three [forma selseyensis], and 47 of group four [forma lidoensis]). The selection of these three groups was arbitrary. Three independent variables were chosen, two qualitative (NOBO and POSQ) and one quantitative (GSD). There were four TEST analyses, which are outlined in Table 7. All TEST data are given in Appendix A, Table A1.

Test Case	Groups	Variables
TEST 1	1, 3	NOBO, GSD
TEST 2	1, 3	NOBO, GSD, POSQ
TEST 3	1, 3, 4	NOBO, GSD
TEST 4	1, 3, 4	NOBO, GSD, POSQ

Table 7: Summary of TEST analyses.

One of the objectives was to understand and duplicate, using MINITAB, the calculation of the various functions and matrices given on the SPSS output. These included: the pooled within groups variance-covariance matrix (W); the two sets of discriminant functions referred to as unstandardized (Z standardized only) and standardized (Z and independent variable standardized); the Z values for the group centroids (\bar{Z} for a group), the critical Z values; the classification functions; the discriminant scores (for the two group analyses); and the classification scores (for the three group analyses).

The analyses were carried out using the simultaneous rather than

the stepwise method because at this stage the relative importance of each variable in the analysis was not relevant. The split sample approach was not taken because it was not necessary to test the validity of the functions at this stage but merely to determine how the functions were calculated. The statistical significance of the functions was calculated and observed to be significant in all TEST analyses; therefore, this aspect was pursued no further at this time. The classification due to chance and classification accuracy were not examined at this time because most of these aspects fall under stage two, validation of the analysis. The emphasis of the TEST section is on stage one, derivation. In the analysis of the Elphidium data stages two and three are also discussed.

All four TEST analyses were completed, but for the sake of brevity only TEST One and TEST Four are described in Appendix C. TEST Two and TEST Three contained no information not illustrated in TEST One and TEST Four.

ANALYSIS OF ELPHIDIUM EXCAVATUM POPULATION

METHODS

Of the 810 Elphidium excavatum specimens photographed, 721 were used in the analysis. The number of specimens of each forma from each location used in the analyses is given in Table 8. The complete data set is given in Appendix A, in Table A2. There are only a few specimens of forma galvestonensis (7) and forma tumidum (11) for the analyses but no more specimens of these two formae were available. The data for the three additional species is given in Appendix A, in

FORMA	1	2	3	4	5	6	7	8	9	10	TOTAL
LOCATION											
1	9	-	-	-	-	-	-	-	-	-	9
2	10	2	-	-	-	-	-	-	-	-	12
3	10	3	-	-	-	-	-	-	-	-	13
4	10	2	-	-	-	-	-	-	-	-	12
5	14	1	-	-	-	-	-	-	-	-	15
6	12	4	-	12	1	-	-	-	-	-	29
7	39	5	15	9	10	11	9	6	-	-	104
8	9	3	2	8	2	-	-	-	-	-	24
9	8	3	5	6	8	-	-	1	3	-	34
10	8	20	20	10	-	-	-	-	-	-	58
11	8	10	1	2	2	-	-	1	4	-	28
12	7	1	6	15	-	2	-	-	-	10	41
13	-	1	-	25	-	-	-	39	-	-	65
14	14	1	-	1	16	-	-	-	-	-	32
15	-	-	-	-	-	3	-	-	44	-	47
16	-	-	-	8	-	16	-	12	-	-	36
17	1	-	-	1	-	-	-	-	48	-	50
18	1	-	-	24	-	34	11	8	-	-	79
19	2	-	-	-	7	-	-	-	-	-	9
20	1	5	2	12	-	3	-	-	1	1	25
	163	61	51	134	46	69	20	67	100	11	721

Table 8: Number of specimens of each forma from each location used in the analyses.

Table A3.

Thirty-two analyses were completed. Twelve were preliminary analyses on a data set of approximately 300 specimens and these cases were classified to see how effective the discriminating variables are and to decide the value of continuing the work. Based on these preliminary results 20 other analyses were completed. Some of these analyses proved redundant or did not produce information relevant to the study. Consequently, only the results of 15 analyses will be discussed. One 'core' or 'basic' analysis will be discussed in detail. Then the options/statistics/features of the other analysis will be listed and explained as needed.

ANALYSES COMPLETED

The 15 analyses can be divided into five groups as outlined on Table 9.

Group A analyses: Four analyses were completed in this group. FORM was the dependent variable and the data contained in one data set; this one data set was both analysed and classified. The DIRECT (or simultaneous) method was used. The main purposes of this group of analyses were to determine the importance or effect of location, and to compare the subjective classification results to those generated by the computer. In analysis A-1 LOC was not included and in A-3 it was included as dummy variables.

Statistic 7 was added to analysis A-1, and indicated that four of the group covariance matrices were not equal.

Consequently, in analysis A-2 option 14 was exercised, and the classifications were based on the separate group covariance matrices,

Table 9: Summary of analyses, and overall percent correctly classified in each analysis.

Group of Analyses	Analyses Name	Description	Percent correctly classified
A		- FORM as dependent variable - one (complete) data set - 721 specimens -DIRECT (simultaneous) methods	
	A-1	LOC not included addition of statistic 7	84.88%
	A-2	addition of option 14	89.99%
	A-3	LOC coded as dummy variables and included as independent variables	85.66%
B		-LOC as dependent variable -one (complete) data set -DIRECT (simultaneous) method	
	B-1	FORM not included	55.05%
	B-2	FORM coded as dummy variables and included as independent variables	54.79%
C		-FORM as dependent variable -one (complete) data set -STEPWISE method-WILKS LAMBDA	
	C-1	LOC not included	85.16%
	C-2	LOC included	85.30%
	C-3	LOC coded as dummy variables and included as independent variables. ANALYSIS feature exercised DUM1 to DUM20 out at step 7	87.66%
D		-FORM as dependent variable -SPLIT-SAMPLE approach (two data sets, ANALYSIS and HOLDOUT)	
	D-1	LOC not included DIRECT method	A-83.76 H-83.80
	D-2	LOC not included DIRECT method addition of Option 14	A-89.43 H-86.19
	D-3	LOC coded as dummy variables, and included as independent variables. DIRECT method	A-87.28% H-88.09%
	D-4	LOC coded as dummy variables and included as independent variables. Addition of Option 14. STEPWISE method WILKS LAMBDA. FIN,POUT specified at .05.	A-87.28% H-89.04%
E		-FORM as dependent variable -one data set with addition of three Elphidiidae species (90 specimens, total 811)	
	E-1	DIRECT method	85.70%
	E-2	DIRECT method addition of Option 14	90.60%
	E-3	STEPWISE method-WILKS LAMBDA addition of Option 14.	90.60%

rather than the pooled matrix. The importance of the assumption that these matrices are equal can be determined by comparing the classification results with and without exercising option 14, as well as exercising statistic 7.

Group B analyses: To further test the influence or effect of location, LOC was treated as the dependent variable. The data was contained in one set which was both analysed and classified. In analysis B-1, FORM was not included and in B-2 it was included as dummy variables.

Group C analyses: FORM was the dependent variable and the one data set analysed and classified. The main feature of these three analyses was that the STEPWISE method of Wilks' lambda was used to determine the order of importance of the independent variables. Analysis C-1 did not include LOC. Another analysis, C-2, was completed with LOC, to determine the relative importance of location (i.e. at what step LOC was removed). Then analysis C-3 was completed with location coded as dummy variables; these dummies were removed simultaneously (using the ANALYSIS feature) at step 7 (the step where LOC was removed in analysis C-2).

Group D analyses: In this group FORM was again the dependent variable but the split-sample approach was taken. The one data set was randomly split into the analysis sample (511 specimens) and holdout sample (210 specimens) to test the classification accuracy of the functions. (No specimens of groups 7 and 10 were placed in the hold out sample because of their low numbers in the analysis sample).

The DIRECT method was used for three of the analyses. Analyses D-1 and D-2 did not include LOC. Option 14 was added to D-2. In

analyses D-3 and D-4 location was included as dummy variables. In D-4 a STEPWISE analysis was completed, option 14 added, and PIN and POUT specified at .05.

Group E analyses: Three analyses were completed on a larger data set, which contained 90 specimens of three other species in addition to the other 721 E. excavatum specimens. The main objectives here were to see the changes in the classification results when three other species were added. The DIRECT method was used without (E-1) and with (E-2) option 14. A stepwise analysis was computed (E-3) to determine if the relative importance of the variables changed when three other species were introduced.

ANALYSIS RESULTS

Group A analyses: Analysis A-1 is treated as a core or basic analysis and the complete results in the form of the computer printout (output) are given in Appendix D. Statistics 1, 2, and 3 are straight forward and do not warrant discussion. Calculation of statistic 3 and option 11, are demonstrated in Appendix C.

Five of the nine discriminant functions account for 95% of the variance (see p. 326). All of the functions are highly significant, though the last four functions don't each account for more than 2% of the variance. The magnitude of the standardized discriminant function coefficients indicate the importance of each independent variable within each function, and those with greatest magnitude are circled on the printout (p. 326). Of the coefficients for the first five functions, AOMA is the most important variable in function 1; UMCO and

POR in function 2; PAP, UMCO and NOBO in function 3; UMCO, POR, CHAM and POS in function 4; and PAP, UNCO, AOMA and DEPO in function 5. Because function 1 accounts for almost 37% of the variance AOMA is the single most discriminating variable. The univariate F-ratios (p. 325) show that AOMA, POR, POSQ, REPO, POSU, and UMCO have large F-values, and are the major group discriminators.

Scatter plots (p. 332-341) of the discriminant scores of the first two functions are given for each group (option 8) and for all cases (option 7) (p. 331); from these plots a territorial map (p. 330) is constructed (option 10). The territorial map is a visual representation of group membership probability based on the two discriminant function scores. The distance (or difference in function scores) between any two group centroids on the map is a measure of the overall difference in morphology between these two groups. The map shows that groups 1, 4, and 8 are closely related (or poorly discriminated by these variables). Group 2 is close to these three groups. Groups 10 and 7 are the next two most closely related groups, followed by groups 10 and 6, and 2 and 3. Group 9 is the most morphologically distinct. All nine discriminant scores, group membership probabilities, and actual group memberships for each case are given (options 6, 17, and 18) on p. 342-367.

The discriminant scores for the group centroids are given on a separate table (p. 327).

The remainder of the output pertains to the classification phase of the analysis. All cases are classified regardless of their group assignment (option 1). As there are 10 groups, there are 10 classification functions (see Appendix B) and these functions (p. 328)

are given on the output (option 12). Statistic 7 tests for the equality of the group covariance matrices. Usually the pooled within-groups covariance matrix is used in calculating the classification scores from the classification functions. When option 14 is exercised, the individual group covariance matrices are used for each group, respectively. In this analysis groups 7, 8 and 9 did not have equal matrices and group 10 did not have enough cases to be accurately tested (p. 329).

The analysis does not discriminate group 4 well, it classifies 16.4% of the group 4 cases in group 1, and 15.7% of the cases in group 2. It also places a few specimens of group 5 in groups 3 and 7 (and vice versa). The overall percent of group cases correctly classified is 84.88%. Of the 15.12% incorrectly classified, at least half are group 4 specimens.

In analysis A-2 option 14 was exercised; this uses the individual group covariance matrices for classification, instead of the pooled within group covariance matrix and is employed when the individual matrices are inhomogenous. The overall classification results are given in Table 10. The overall percent correctly classified has improved to 89.99%. Most of the improvement is from the group 4 cases, 76% of the group 4 cases have been correctly classified, instead of 62% as in analysis A-1.

The A-2 territorial map (Figure 3) is markedly different from the A-1 map; the centroids for groups 1, 2, 4, and 8 are very close together. Group 9 now falls within group 2, group 7 within group 3, group 1 within group 4, and group 8 within groups 2 and 1.

In analysis A-3 location, coded as 20 dummy variables, was

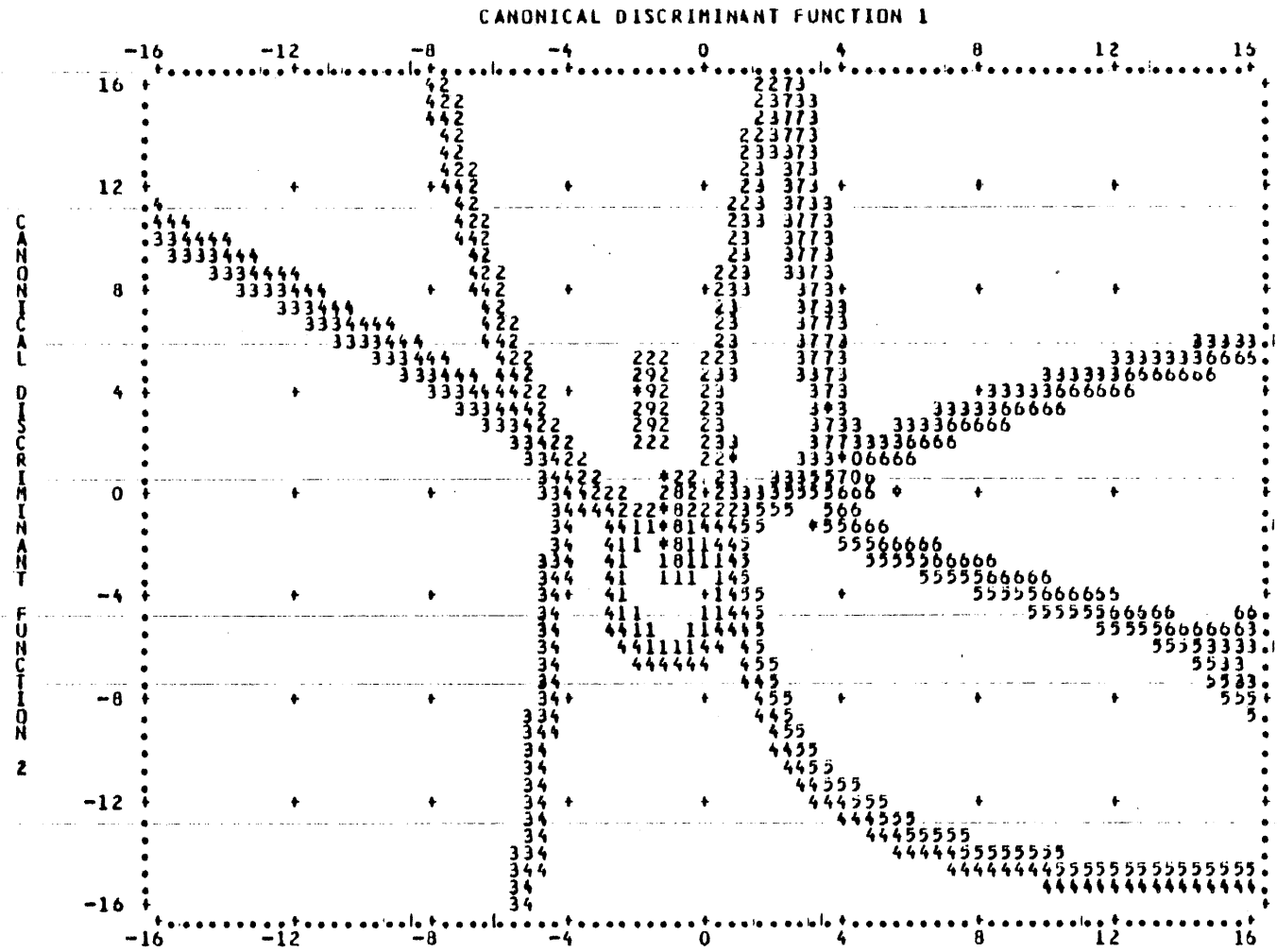


Figure 3: Territorial map, analysis A-2. It is assumed that all functions but the first two are zero. * Indicates a group centroid.

group	no. of specimens	1	2	3	4	5	6	7	8	9	10
1	163	153 93.9	3 1.8	0	5 3.1	1 .6	0	0	1 .6	0	0
2	61	3 4.9	51 83.6	0	5 8.2	0	1 1.6	0	1 1.6	0	0
3	51	1 2.0	3 5.9	42 82.4	2 3.9	1 2.0	0	1 2.0	0	0	1 2.0
4	134	8 6.0	17 12.7	6 4.5	103 76.9	0	0	0	0	0	0
5	46	1 2.2	0	1 2.2	0	43 93.5	0	1 2.2	0	0	0
6	69	0	0	1 1.4	0	0	68 98.6	0	0	0	0
7	20	0	0	0	0	0	0	20 100	0	0	0
8	67	3 4.5	0	0	0	0	0	0	64 95.6	0	0
9	100	1 1.0	2 2.0	1 1.0	0	0	0	0	2 2.0	94 94.0	0
10	10	0	0	0	0	0	0	0	0	0	10 100.0

Table 10: Classification results, analysis A-2. The top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified - 89.88%

included as an independent variable. Looking at the univariate F ratios (Table 11) some of these dummy variables (i.e., DUM10, DUM12 to DUM15, DUM17 and 18) have significant F-values. However, at some of these locations one group comprises over 80% of the cases and the analysis is in fact discriminating on form. The overall classification results have improved only slightly, to 85.66%.

Group B analyses: To further test the influence of location two analyses were completed with LOC as the dependent variable. FORM was treated as an independent variable. The classification results are quite poor only 55.05% (Table 12). Comparing Tables 8 and 12, it can be seen that the number correctly classified at each location corresponds closely to the dominant group present at each location. Coding FORM as dummy variables (analysis B-2) changes the results only slightly to 54.79%. (In an earlier analysis on LOC, FORM was not included and the results were essentially the same - 54.79%).

Group C analyses: To further determine the relative importance of each independent variable, three analyses were completed on one data set with FORM as the dependent variable but using the STEPWISE method. LOC was not included in analysis C-1. Nine functions were calculated (see Table 13) but the last function was close to not being significant. The univariate F-ratios and Wilks' lambda for each independent variable are given on Table 14. Note that these values are the same as on the bottom of Tables 11 and 16. GSR is the only variable that is not significant at the .01 level. The summary table (Table 15) lists the variables in order in which they were removed. The variables are not removed in order of descending F-values. This is because some of the differences among groups in a variable are

WILKS LAMBDA (U-STATISTIC) AND UNIVARIATE F-RATIO WITH 9 AND 711 DEGREES OF FREEDOM			
VARIABLE	WILKS LAMBDA	F	SIGNIFICANCE
DUM1	.955673	3.372	.0002
DUM2	.959388	3.345	.0005
DUM3	.958755	3.349	.0004
DUM4	.959335	3.345	.0005
DUM5	.938226	3.148	.0009
DUM6	.961355	3.179	.0009
DUM7	.907077	3.829	.0001
DUM8	.978944	1.694	.0854
DUM9	.953221	3.018	.0013
DUM10	.793122	2.223	.0001
DUM11	.953333	3.857	.0001
DUM12	.730331	2.917	.0003
DUM13	.835088	4.520	.0001
DUM14	.824333	4.525	.0001
DUM15	.826133	4.717	.0001
DUM16	.839720	4.694	.0001
DUM17	.579099	11.422	.0001
DUM18	.717577	3.033	.0003
DUM19	.590033	9.761	.0001
DUM20	.957333	3.519	.0003
PAP	.448222	97.255	.0001
UACD	.851133	14.660	.0001
DEUM	.470233	69.600	.0001
PEUM	.257455	227.880	.0001
ADY	.144344	92.622	.0001
ZBDA	.510221	75.844	.0001
PEBDA	.911339	7.860	.0001
PEBDA	.290855	18.540	.0001
PEBDA	.290855	18.540	.0001
PEBDA	.677755	37.560	.0001
PEBDA	.310998	73.601	.0001
PEBDA	.321977	55.844	.0001
PEBDA	.321977	55.844	.0001
PEBDA	.296977	37.000	.0001
PEBDA	.973339	2.160	.0230

Table 11: Wilks' lambda and univariate F-ratios for the independent variables, analysis A-3.

Table 12: Classification results, analysis B-1. The number to the left of the slash in each square is the actual number of specimens classified into that group. The number to the right of the slash is the percent the actual number represents. Overall percent correctly classified -55.05%.

Group		1	2	3	4	5	6	7
	no. of specimens							
1	9	8/88.9	0	0	0	0	1/11.1	0
2	12	1/8.3	8/66.7	1/8.3	0	0	1/8.3	0
3	13	2/15.4	0	9/69.2	0	0	0	1/7.7
4	12	3/25.0	0	1/8.3	5/41.7	2/16.7	0	0
5	15	1/6.7	0	1/6.7	2/13.3	10/66.7	1/6.7	0
6	29	2/6.9	1/3.4	1/3.4	1/3.4	1/3.4	11/37.9	1/3.4
7	104	0	5/4.8	1/1.0	3/2.9	6/5.8	6/5.8	33/31.7
8	24	0	1/4.2	0	1/4.2	0	0	1/4.2
9	34	0	2/5.9	1/2.9	1/2.9	0	2/5.9	1/2.9
10	58	0	1/1.7	5/8.6	2/3.4	2/3.4	2/3.4	1/1.7
11	28	0	2/7.1	2/7.1	1/3.6	2/7.1	1/3.6	1/3.6
12	41	2/4.9	0	0	1/2.4	3/7.3	2/4.9	0
13	65	1/1.5	0	0	0	0	0	0
14	32	5/15.6	0	0	2/6.3	1/3.1	3/9.4	1/3.1
15	47	0	0	0	0	0	0	0
16	36	0	0	0	0	0	0	0
17	50	0	0	0	0	0	1/2.0	0
18	78	1/1.3	0	0	0	0	0	3/3.8
19	9	0	0	0	0	0	0	1/11.1
20	25	0	0	0	0	0	2/8.0	0

Group		8	9	10	11	12	13	14
	no. of specimens							
1	9	0	0	0	0	0	0	0
2	12	0	1/8.3	0	0	0	0	0
3	13	0	0	1/7.7	0	0	0	0
4	12	0	0	0	1/8.3	0	0	0
5	15	0	0	0	0	0	0	0
6	29	0	0	5/17.2	2/6.9	0	0	1/3.4
7	104	6/58	10/9.6	6/5.8	2/1.9	1/1.0	4/3.8	0
8	24	8/33.3	0	1/4.2	5/20.8	0	0	1/4.2
9	34	4/11.8	7/20.6	4/11.8	1/29	0	2/5.9	0
10	58	6/10.8	0	34/58.6	1/1.7	0	0	0
11	28	2/7.1	0	5/17.9	7/25.0	0	2/7.1	0
12	41	4/9.8	0	1/2.4	0	22/53.7	0	0
13	65	0	0	1/1.5	0	4/6.2	53/81.5	0
14	32	0	0	0	0	1/3.1	0	18/56.3
15	47	0	0	0	0	0	0	0
16	36	0	0	0	0	6/16.7	7/19.4	0
17	50	0	0	0	0	0	0	1/2.0
18	78	0	5/6.4	0	1/1.3	1/1.3	7/9.0	1/1.3
19	9	1/11.1	0	0	0	0	0	0
20	25	1/4.0	0	4/16.0	0	1/4.0	0	0

Group		15	16	17	18	19	20	
	no. of specimens							
1	9	0	0	0	0	0	0	
2	12	0	0	0	0	0	0	
3	13	0	0	0	0	0	0	
4	12	0	0	0	0	0	0	
5	15	0	0	0	0	0	0	
6	29	0	0	0	0	1/3.4	2/6.9	
7	104	0	1/1.0	0	10/9.6	8/7.7	2/1.9	
8	24	0	0	0	0	2/8.3	4/16.7	
9	34	1/2.9	1/2.9	1/2.9	1/2.9	4/11.8	1/2.9	
10	58	0	0	0	0	0	4/6.9	
11	28	1/3.6	0	1/3.6	0	1/3.6	0	
12	41	0	3/7.3	0	1/2.4	0	2/4.9	
13	65	0	3/4.6	0	3/4.6	0	0	
14	32	0	0	0	0	0	1/3.1	
15	47	39/83.0	0	5/10.6	3/6.4	0	0	
16	36	0	16/44.4	0	5/13.9	0	2/5.6	
17	50	9/18.0	0	39/78.0	0	0	0	
18	78	1/1.3	5/6.4	1/1.3	51/65.4	0	1/1.3	
19	9	0	0	0	0	7/77.3	0	
20	25	1/4.0	4/16.0	0	0	0	12/48.0	

CANONICAL DISCRIMINANT FUNCTIONS										
FUNCTION	EIGENVALUE	PERCENT OF VARIANCE	CUMULATIVE PERCENT	CANDONICAL CORRELATION	AFTER FUNCTION	WILKS	LAMBDA	CHI-SQJARED	D.F.	SIGNICANCE
1*	5.45389	36.93	36.93	.9192086	-	0	.0012479	4736.1	117	0
2*	3.85739	26.13	63.06	.8911387	-	2	.0080665	3415.0	95	0
3*	2.69360	18.22	81.22	.8539678	-	3	.0371919	2293.2	77	0
4*	1.68264	11.38	92.60	.7917804	-	4	.1447222	1307.5	60	0
5*	.49423	3.34	95.95	.5751154	-	5	.3382377	673.34	45	0
6*	.29001	1.95	97.91	.4741438	-	6	.2801153	361.40	32	0
7*	.16270	1.10	99.01	.3747799	-	7	.7483547	205.38	21	0
8*	.12315	.83	99.84	.3311314	-	8	.8701149	98.574	12	.0000
9*	.02326	.15	100.00	.1507623	-		.9772737	16.290	5	.0061

* MARKS THE 9 FUNCTION(S) TO BE USED IN THE REMAINING ANALYSIS.

Table 13: Statistics for the nine discriminant functions calculated for analysis C-1.

WILKS LAMBDA (U-STATISTIC) AND UNIVARIATE F-RATIO WITH 9 AND 711 DEGREES OF FREEDOM				
VARIABLE	WILKS LAMBDA	F	SIGNIFICANCE	
PAP	.44822	97.25	0	
UMCO	.35113	146.0	0	
DEUM	.47023	89.00	0	
POR	.25749	227.8	0	
ADMA	.19494	326.2	0	
NOBO	.51021	75.84	0	
PERO	.91139	7.680	0	
REPO	.29881	185.4	0	
SUT	.67775	37.56	0	
CHAM	.51698	73.81	0	
POSU	.32197	156.4	0	
DEPO	.52885	70.38	0	
POSQ	.29697	187.0	0	
GSR	.97339	2.160	.0230	

Table 14: Univariate F-ratios and Wilks' lambda for the independent variables, analysis C-1.

STEP	ENTERED	ACTION REMOVED	VAR IN	WILKS LAMBDA	SIG.
1	ADMA		1	.194942	0
2	POR		2	.052769	0
3	REPO		3	.018151	.0000
4	UMCO		4	.006893	0
5	PAP		5	.003955	0
6	NOBO		6	.002802	0
7	DEUM		7	.002237	0
8	CHAM		8	.001823	0
9	SUT		9	.001560	0
10	POSU		10	.001420	0
11	DEPO		11	.001322	.0000
12	POSQ		12	.001283	0
13	PERO		13	.001250	.0000

----- VARIABLES NOT IN THE ANALYSIS AFTER STEP 13

VARIABLE	TOLERANCE	MINIMUM TOLERANCE	F TO ENTER	WILKS LAMBDA
GSR	.9550541	.1342956	.9461	.00123

F LEVEL OR TOLERANCE OR VIN INSUFFICIENT FOR FURTHER COMPUTATION.

Table 15: Summary table for the stepwise analysis C-1.

explained by variables already included in the analysis. GSR was not removed after step 13 (and hence not included in the analysis) because F-to-enter fell below the default value. The classification results have improved only slightly (compared to analysis A-1) and have increased to 85.16% due to the omission of GSR.

In analysis C-2, the relative importance of LOC was examined. Of the 14 variables LOC's F-ratio is twelfth in magnitude (52.16 - see Table 16) but as the variables are removed, the relative values of the F-ratio and Wilks' lambda change; at step 7 LOC had the smallest Wilks' lambda and was removed at this step. At the end of the analysis, after step 14, GSR was the only variable not included (because F-to-enter fell below the default value). Note again that the last variables are exactly the same as on the bottom of Tables 11 and 14. The classification results - 85.30% - improved by only one specimen, (compared to the previous [C-2] analysis).

LOC was coded as dummy variables in analysis C-3 and using the ANALYSIS feature, the dummies were removed simultaneously at step 7. The variables removed in steps 1-6 remained the same as in the previous two analyses. The dummy variables were then removed in steps 7 through 26 (DUM1 to DUM19 respectively). DUM20 was not removed, and due to PIN and POUT specifications, DUM20, POSU and GSR were not included in the analyses. A summary is given on Table 17. Coding LOC as dummy variables did improve the classification results by 17 specimens, to 87.66% (see Table 18). Group 4 classification improved the most (16% within that group, followed by group 2 - 9.9% and group 3 - 9.8% within those groups, respectfully).

Group D analysis: The three previous groups of analyses have

WILKS LAMBDA (U-STATISTIC) AND UNIVARIATE F-RATIO WITH 9 AND 711 DEGREES OF FREEDOM			
<u>VARIABLE</u>	<u>WILKS LAMBDA</u>	<u>F</u>	<u>SIGNIFICANCE</u>
LOC	.60233	52.16	0
PAP	.44822	97.25	0
UMCO	.35113	146.0	0
DEUM	.47023	89.00	0
POR	.25749	227.8	0
AOMA	.19494	326.2	0
NUBO	.51023	75.84	0
PERO	.91139	7.680	0
REPO	.29881	185.4	0
SUT	.67775	37.56	0
CHAM	.51698	73.81	0
POSU	.32197	166.4	0
DEPO	.52885	70.38	0
POSQ	.29697	187.0	0
GSR	.97339	2.160	.0230

Table 16: Univariate F-ratio and Wilks' lambda for the independent variables, analysis C-2.

STEP	ACTION	VAR	WILKS	SIG.
ENTERED	REMOVED	IN	LAMBDA	
1	ADMA	1	.194942	0
2	POB	2	.052789	0
3	REPO	3	.018151	.0000
4	UMCO	4	.006898	.0000
5	PAP	5	.003955	.0000
6	NOBO	6	.002802	.0000
7	DUM1	7	.002691	.0000
8	DUM2	8	.002587	.0000
9	DUM3	9	.002496	.0000
10	DUM4	10	.002425	.0000
11	DUM5	11	.002325	.0000
12	DUM6	12	.002247	.0000
13	DUM7	13	.002051	.0000
14	DUM8	14	.001923	.0000
15	DUM9	15	.001833	.0000
16	DUM10	16	.001243	.0000
17	DUM11	17	.001051	.0000
18	DUM12	18	.000740	.0000
19	DUM13	19	.000579	.0000
20	DUM14	20	.000499	.0000
21	DUM15	21	.000428	.0000
22	DUM16	22	.000373	.0000
23	DUM17	23	.000155	.0000
24	DUM18	24	.000131	.0000
25	DUM19	25	.000120	.0000
26	DUM	26	.000095	.0000
27	SUT	27	.000088	.0000
28	CHAM	28	.000079	.0000
29	PCSO	29	.000072	.0000
30	DEPC	30	.000067	.0000
31	PERD	31	.000065	.0000

VARIABLES NOT IN THE ANALYSIS AFTER STEP 31

VARIABLE	TOLERANCE	MIN. VARIATION TOLERANCE	SIGNIF. OF F TO ENTER	WILKS	LAMBDA
DUM20	.0000000	.0000000			
POBU	.3280172	.1552729	.0976	.00000	
GSR	.8899841	.1931397	.2152	.00000	

F LEVEL OR TOLERANCE OR VAR INSUFFICIENT FOR FURTHER COMPUTATION.

Table 17: Summary table for the stepwise analysis C-3.

group		1	2	3	4	5	6	7	8	9	10
	no. of specimens										
1	163	153 93.9	1 .6	1 .6	5 3.1	1 .6	0 0	0 0	1 .6	1 .6	0 0
2	61	4 6.6	50 82.0	3 4.9	3 4.9	0 0	1 1.6	0 0	0 0	0 0	0 0
3	51	1 2.0	2 3.9	41 80.4	1 2.0	3 5.9	0 0	0 0	0 0	0 0	3 5.9
4	134	16 11.9	15 11.2	1 .7	99 73.9	1 .7	1 .7	0 0	0 0	1 .7	0 0
5	46	1 2.2	0 0	0 0	0 0	43 93.5	1 2.2	1 2.2	0 0	0 0	0 0
6	69	0 0	0 0	1 1.4	0 0	0 0	64 92.8	0 0	1 1.4	1 1.4	2 2.9
7	20	0 0	0 0	0 0	0 0	1 5.0	1 5.0	18 90.0	0 0	0 0	0 0
8	67	5 7.5	0 0	0 0	0 0	0 0	0 0	0 0	62 92.5	0 0	0 0
9	100	1 1.0	3 3.0	2 2.0	0 0	0 0	0 0	0 0	2 2.0	92 92.0	0 0
10	10	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	10 100.0

Table 18: Classification results, analysis C-3. Top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified, 87.66%.

been performed on only one data set. To test the classification accuracy of the functions the split sample approach was taken. Analysis D-1 was performed to compare the classification results of the two samples. The results of the analysis sample were 83.76% correctly classified. The results of the holdout sample are given on Table 19 - 83.80%. The consistency of the classification results for the two samples indicates that the functions are good discriminators. When option 14 is added (analysis D-2) the percent correctly classified jumps to 89.43% for the analysis sample (comparable to analysis A-2), and to 86.19% for the holdout sample (Table 20). When LOC (as dummy variables) is included as independent variables the results for the analysis sample are 87.28% and for the holdout sample 88.09% (Table 21). Taking this analysis further, using the STEPWISE method, and specifying PIN and POUT at .05 (analysis D-4), the results are 87.28% correctly classified (analysis sample) and 89.04% correctly classified (Table 22-holdout sample). On Table 23, the first 6 variables removed are the same as for analysis C-3. In this instance, DUM20 was not removed from the analysis (unlike analysis C-3) but DUM1 to DUM5 were removed in addition to PERO, POSQ, and GSR (Table 23). Comparing Tables 19 and 22, the improvements in the correctly classified are largest for groups 3, 4 and 6.

Group E analyses: The last three analyses were completed on one data set, containing the 721 Elphidium excavatum specimens plus 90 other specimens, comprising of 30 specimens each of Elphidium subarcticum, E. bartletti, and Haynesina orbiculare. Analysis E-1 was completed using the DIRECT method. There are now 12 (i.e. 13-1) discriminant functions, of which the first five account for 95% of the

group		1	2	3	4	5	6	7	8	9	10
	no. of specimens										
1	48	47 97.9	0 0	0 0	0 0	0 0	0 0	0 0	1 2.1	0 0	0 0
2	19	2 10.5	16 84.2	5.3 5.3	0 0	0 0	0 0	0 0	0 0	0 0	0 0
3	16	0 0	3 18.8	8 50	2 12.5	2 12.5	0 0	1 6.2	0 0	0 0	0 0
4	41	7 17.1	7 17.1	2 4.8	24 58.5	0 0	0 0	0 0	0 0	0 0	1 2.4
5	13	0 0	0 0	0 0	0 0	13 100	0 0	0 0	0 0	0 0	0 0
6	22	0 0	0 0	0 0	0 0	0 0	19 86.4	0 0	0 0	0 0	3 13.6
8	21	1 5	0 0	0 0	0 0	0 0	0 0	0 90	19 5	1 5	0 0
9	70	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0	70 100	0 0

Table 19: Classification results of the holdout sample, analysis D-1. Top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified: 83.80%.

group		1	2	3	4	5	6	7	8	9	10
	no. of specimens										
1	48	45 93.8	2 4.2	0 0	1 2.0	0 0	0 0	0 0	0 0	0 0	0 0
2	19	1 5.3	18 94.7	0 0	0 0	0 0	0 0	0 0	0 0	0 0	0 0
3	16	0 0	4 25.0	8 60.0	3 18.8	1 6.2	0 0	0 0	0 0	0 0	0 0
4	41	1 2.4	5 12.2	3 7.4	32 78.0	0 0	0 0	0 0	0 0	0 0	0 0
5	13	0 0	0 0	0 0	0 0	11 84.6	2 15.4	0 0	0 0	0 0	0 0
6	22	0 0	0 0	2 9.1	0 0	0 0	20 90.9	0 0	0 0	0 0	0 0
8	21	3 14.3	0 0	0 0	0 0	0 0	0 0	0 0	18 85.7	0 0	0 0
9	70	0 0	0 0	1 1.4	0 0	0 0	0 0	0 0	0 0	69 98.6	0 0

Table 20: Classification results of the holdout sample, analysis D-2. The top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified: $181/210=86.19\%$.

group	no. of specimens	1	2	3	4	5	6	7	8	9	10
1	48	46 95.8	0	0	1 2.1	1 2.1	0	0	0	0	0
2	19	1 5.3	16 84.1	1 5.3	1 5.3	0	0	0	0	0	0
3	16	0	2	12	1	1	0	0	0	0	0
4	41	0 5 12.3	12.5 6 14.6	75 1 2.4	6.2 29 70.7	6.2 0	0	0	0	0	0
5	13	0	0	0	0	13 100	0	0	0	0	0
6	22	0	0	0	0	0	21 95.5	0	0	1 4.5	0
8	21	0 2 10	0	0	0	0	0	0	19 90	0	0
9	70	0	1 1.4	1 1.4	0	0	0	0	0	68 97.2	0

Table 21: Classification results of the holdout sample, analysis D-3. The top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified: 185/210 = 88.09%.

group		1	2	3	4	5	6	7	8	9	10
	no. of specimens										
1	48	46 95.8	0	0	1	1	0	0	0	0	0
2	19	1 5.2	16 84.2	1	1	0	0	0	0	0	0
3	16	0	2 12.5	12 75.0	1	1	0	0	0	0	0
4	41	6 14.6	3 7.3	1 2.5	31 75.6	0	0	0	0	0	0
5	13	0	0	0	0	13 100	0	0	0	0	0
6	22	0	0	0	0	0	22 100	0	0	0	0
8	21	2 9.5	0	0	0	0	0	0	19 90.5	0	0
9	70	0	1 1.4	1 1.4	0	0	0	0	0	68 97.1	0

Table 22: Classification results of the holdout sample, analysis D-4. Top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified, 193/210 = 89.04%.

STEP	ENTERED	ACTION	VAR'S	WILKS	SIG.
		REMOVED	IN	LAMBDA	
1	ACMA		1	.1206463	.0
2	POPO		2	.033952	.0
3	REPO		3	.017954	.0
4	UMCO		4	.006810	.0
5	PAP		5	.004252	.0000
6	NOBO		6	.002913	.0
7	DUM12		7	.002005	.0000
8	DUM13		8	.001432	.0
9	DUM17		9	.001081	.0000
10	DUM15		10	.000415	.0
11	DUM12		11	.000325	.0000
12	DUM16		12	.000255	.0
13	DEFUM		13	.000205	.0000
14	CHAM		14	.000181	.0
15	SUT		15	.000162	.0000
16	DUM14		16	.000145	.0
17	DUM19		17	.000132	.0
18	DUM10		18	.000121	.0
19	POSU		19	.000112	.0000
20	DUM7		20	.000105	.0
21	DUM9		21	.000100	.0
22	DUM6		22	.000094	.0
23	DUM20		23	.000089	.0
24	DUM11		24	.000083	.0
25	DUM8		25	.000073	.0000
26	DEPO		26	.000071	.0

VARIABLES NOT IN THE ANALYSIS AFTER STEP 25

VARIABLE	TOLERANCE	MINIMUM TOLERANCE	SIGNIF. OF F TO ENTER	WILKS	LAMBDA
DUM1	.8378283	.2519759	.2050	.00007	.00007
DUM2	.8395924	.2503417	.4375	.00007	.00007
DUM3	.8089443	.2591995	.9878	.00007	.00007
DUM4	.8480981	.2580673	.9999	.00007	.00007
DUM5	.7654429	.2517952	.9008	.00007	.00007
PERCO	.8883032	.2569464	.6391	.00007	.00007
POSU	.1752483	.1752483	.0557	.00007	.00007
GSR	.9019759	.2551720	.4789	.00007	.00007

F LEVEL OF TOLERANCE OR VIN INSUFFICIENT FOR FURTHER COMPUTATION.

Table 23: Summary table for the stepwise analysis D-4.

variance (Table 24). The last function is not significant. The univariate F-ratios and Wilks' lambda are given for each independent variable (Table 25); comparing Table 25 with the same table for analysis A-1 (in Appendix D p. 329) the relative importance of each variable has not changed.

The territorial map for this analysis (Figure 4) is quite different than that for analysis A-1 (see p. 330). The group centroids for B (E. subarcticum) and C (E. bartletti) appear to be superimposed. The centroids for the formae of E. excavatum are spatially, closer together. The maximum distance between any two morphotype centroids is less than from any morphotype centroid to that for the two other Elphidiidae species. The farthest distance is from the Haynesina orbiculare (A) centroid to any morphotype centroid.

The overall classification results (percent correctly classified) are 85.70% (Table 26). The functions have placed one group 2 (E. excavatum f. excavata) and two group 4 (E. excavatum f. lidoensis) specimens into E. subarcticum and one group 4 specimen into E. bartletti. One E. subarcticum specimen has been classed as E. bartletti and three specimens vice versa. All Haynesina specimens have been correctly classified and no specimens incorrectly classed as Haynesina.

Exercising option 14 improved the classification results (analysis E-2, 90.60%).

A STEPWISE analysis was completed to further determine the importance of each independent variable. The variables were removed in the same order (Table 27) as in analysis C-1 (Table 15) except that the importance of REPO decreased, (it came out at step 12 instead of

CANONICAL DISCRIMINANT FUNCTIONS										
FUNCTION	EIGENVALUE	PERCENT JF VARIANCE	CUMULATIVE PERCENT	CANONICAL CORRELATION	AFTER FUNCTION	MILKS	LAMBDA	CHI-SQUARED	D.F.	SIGNIFICANCE
1*	7.24046	35.89	35.89	.9371620	-	0	.0032377	6407.4	168	0
2*	5.21434	25.85	61.74	.9163141	-	1	.0024532	4797.3	143	0
3*	3.51735	17.44	79.17	.8824010	-	2	.0152449	3332.2	120	0
4*	2.35553	11.73	90.90	.8369734	-	3	.0688565	2131.1	99	0
5*	.97720	4.84	95.74	.7033189	-	4	.2317723	1154.9	80	0
6*	.31454	1.55	97.30	.4881629	-	5	.4582514	621.52	63	0
7*	.23638	1.17	98.47	.4372463	-	6	.6021729	403.99	48	0
8*	.12511	.63	99.10	.3345456	-	7	.7445119	234.99	33	0
9*	.09759	.43	99.58	.2981880	-	8	.8384327	140.39	24	.0000
10*	.04270	.21	99.79	.2023737	-	9	.9202255	66.218	15	.0000
11*	.04200	.21	100.00	.2007659	-	10	.9595229	32.911	8	.0001
12*	.00010	.03	100.03	.0133161	-	11	.9998227	.14123	3	.9863

* MARKS THE 12 FUNCTION(S) TO BE USED IN THE REMAINING ANALYSIS.

Table 24: Statistics for the 12 discriminant functions calculated for analysis E-1.

WILKS LAMBDA (U-STATISTIC) AND UNIVARIATE F-RATIO WITH 12 AND 798 DEGREES OF FREEDOM				
VARIABLE	WILKS	LAMBDA	F	SIGNIFICANCE
PAP	.45027		81.19	.0000
UMCO	.32271		139.6	.0000
DEUM	.41106		95.28	.0000
POR	.19771		259.4	.0000
AOMA	.15540		343.1	.0000
NOBO	.45795		79.71	.0000
PERO	.85120		11.63	.0000
REPO	.22609		227.6	.0000
SUT	.57538		31.96	.0000
CHAM	.50200		65.97	.0000
POSU	.24661		203.2	.0000
DEPD	.35321		121.8	.0000
POSD	.22599		227.8	.0000
GSR	.94089		4.178	.0000

Table 25: Wilks' lambda and univariate F-ratios for the independent variables, analysis E-1.

STEP	ENTERED	ACTION	REMOVED	VAR IN	WILKS LAMBDA	SIG.
1	AOMA			1	.156397	.0000
2	POR			2	.033531	.0000
3	POSU			3	.007013	.0000
4	UMCO			4	.002559	.0000
5	PAP			5	.001595	.0000
6	NOBO			6	.000925	.0000
7	DEUM			7	.000654	.0000
8	CHAM			8	.000522	.0000
9	SUT			9	.000445	.0000
10	REPO			10	.000395	.0000
11	PERO			11	.000357	.0000
12	DEPD			12	.000323	.0000
13	POSD			13	.000311	.0000
14	GSR			14	.000295	.0000

Table 27: Summary table for the stepwise analysis E-3.

group		1	2	3	4	5	6	7	8	9	10	11	12	13
	no. of specimens													
1	163	156 95.7	1 .6	0	1 .6	1 .6	0	0	4 2.5	0	0	0	0	0
2	61	7 11.5	43 70.5	3 4.9	3 4.9	0	1 1.6	0	1 1.6	1 1.6	0	0	1 1.6	0
3	51	1 2.0	3 5.9	36 70.6	1 2.0	4 7.8	0	4 7.8	0	0	2 3.9	0	0	0
4	134	22 16.4	20 14.9	5 3.7	80 59.7	1 .7	0	0	0	0	2 1.5	0	3 2.2	1 .7
5	46	1 2.2	0	0	0	43 93.5	0	2 4.3	0	0	0	0	0	0
6	69	0 0	2 2.9	1 1.4	0	0	63 91.3	0	1 1.4	0	2 2.9	0	0	0
7	20	0 0	0	0	0	2 10.0	0	18 90.0	0	0	0	0	0	0
8	67	2 3.0	0	0	0	0	0	0	64 95.5	0	1 1.5	0	0	0
9	100	0 0	0	1 1.0	0	0	0	0	3 3.0	96 96.0	0	0	0	0
10	10	0 0	0	0	0	0	0	0	0	0	10 100	0	0	0
11	30	0 0	0	0	0	0	0	0	0	0	0	27 90.0	3 10.0	0
12	30	0 0	0	0	0	0	0	0	0	0	0	1 3.3	29 96.7	0
13	30	0 0	0	0	0	0	0	0	0	0	0	0	0	30 100

Table 26: Classification results, analysis E-1. The top number in each square is the actual number of specimens classified in that group, bottom number the percent the above number represents. Overall percent correctly classified: 85.70%.

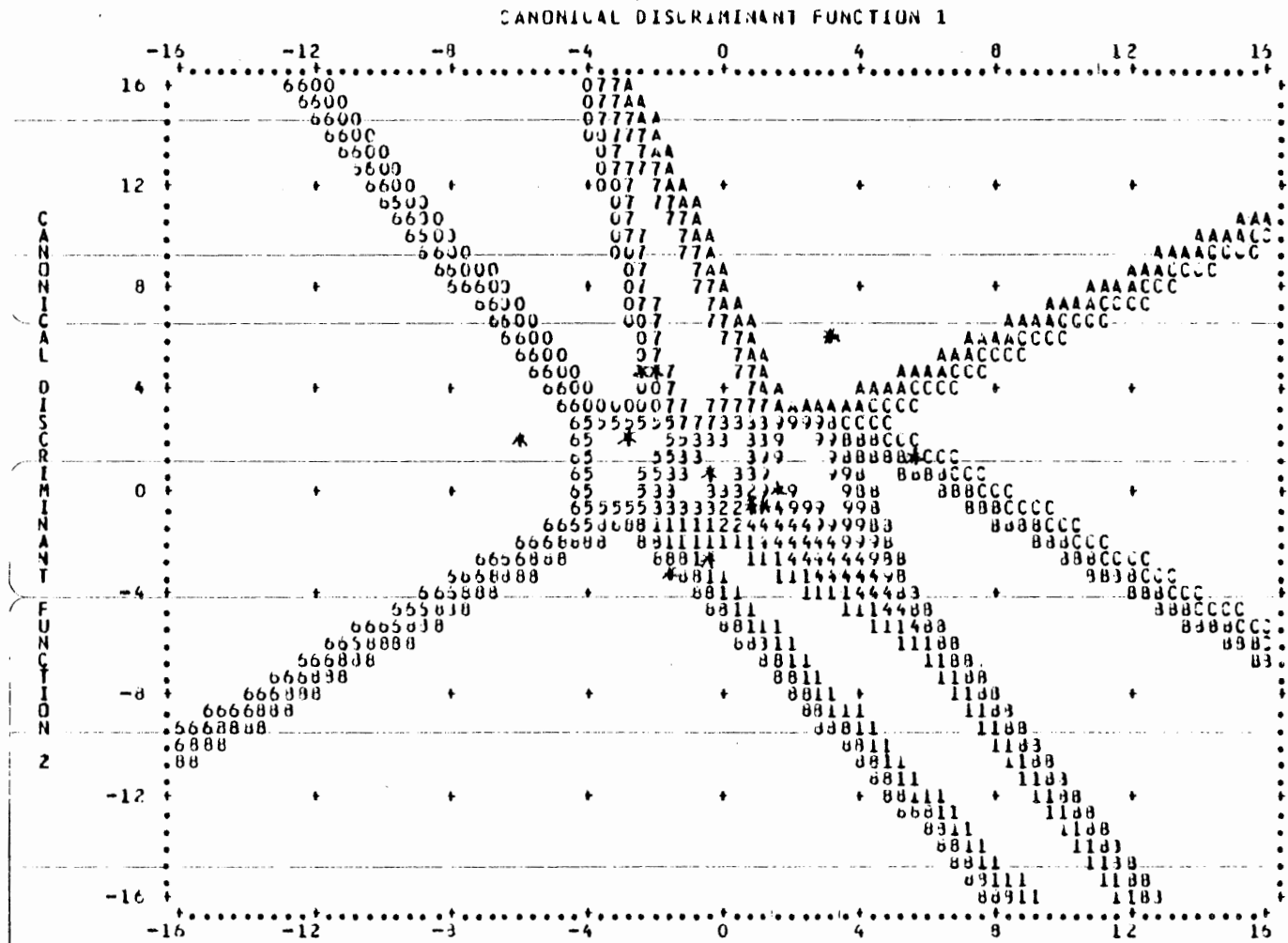


Figure 4: Territorial map, analysis E-1. It is assumed that all functions but the first two are zero. *Indicates a group centroid.

step 3). Reciprocally, POSQ came out at step 3 instead of step 12. GSR was included in the analysis, at the final step, step 14. The classification results (overall percent correctly) remained at 90.60%.

DISCUSSION OF ANALYSIS RESULTS

The overall percent of correctly classified specimens, that is, specimens classified into the group to which they were subjectively assigned, is a measure of how well the functions discriminate the groups based on the information (morphological characteristics, independent variables) supplied. The consistency of these results plus the high percentage of correctly classified specimens for all analysis (84-91%) indicates that the functions are good discriminators, and the independent variables included are good group discriminators.

Group 4, forma lidoensis, is not as well defined as the other formae. Many group 4 specimens are incorrectly classified as belonging to groups 1 or 2 in particular. Only 62% of the group 4 specimens are correctly classified in analysis A-1. Of the variables measured, there is no unique combination that exclusively defines group 4. Subsequent observation shows that the feature of the sutures broadening towards the umbilicus is exclusively present in this form and not included in the analysis. Including this feature would probably increase the discriminating power of the functions in relation to group 4.

There are also other patterns to group misclassification. The functions do not discriminate well between groups 1 and 2, groups 3 and 5, and groups 5 and 7. Groups 1 and 2, forma clavata and forma

excavata are the two most similar formae. The only variable generally present in clavata and absent in excavata is UMC0. The formae are also similar in size and shape. Groups 3 and 5, forma selsevensis and forma magna both have a subacute periphery and a raised umbilicus, as well as similar shape and sizes. They are discriminated on development and regularity of the ponticuli and the umbilical ornamentation. There are similar problems with groups 5 and 7, forma magna and forma galvestonensis. In terms of size and shape they are very similar, only POR and PAP are consistently different, the pores being much denser and finer on galvestonensis and papillae being present more often on this form.

Option 14 was exercised due to non-homogeneity of the individual variance-covariance matrices. The SPSS program test for homogeneity is calculating statistic 7; a test is outlined by Reyment (1962) and some examples given there. The main reason for non-homogeneous variance-covariance matrices in this case is that many of the independent variables are binary or categorical. For a binary variable the mean and standard deviation are directly related. The mean is the proportion of specimens in one of the two categories and the standard deviation is a direct function of this proportion. Unfortunately it is not possible to correct this problem using a simple transformation, but using individual variance-covariance matrices in the classification functions effectively compensates for this. Non-homogeneity is not uncommon for biologic data (Reyment 1962, Scott 1974). Reyment (1962) does point out that non-homogeneity of these matrices, calculated from a sample of microfossils from a single layer of sediment may represent a homogeneous fraction in the

development of the species, or it may be a mixture of thousands of generations, depending on the rate of sedimentation incidence of reworking or whether or not the species burrows into the sediment. These causes are unlikely in this case as only samples carefully examined and found lacking evidence of reworking were chosen for inclusion; and all the populations studied were from surface samples and would not represent a large time span paleontologically. The assemblages studied possibly representing a longer time span contained very few specimens from those groups with non-homogeneous matrices.

Another cause of non-homogeneity may be growth invariance (Burnaby 1966, Reyment 1969). Growth invariance is a major factor to be dealt with in multivariate analysis of paleontologic data, particularly for analysis of organisms which do not have a terminal growth size (Burnaby 1966). Though an effort was made to minimize the influence of growth stages by not including variables dependent on growth stage and by choosing only adult specimens. As best as could be ascertained only two variables related to growth, CHAM and GSR, were included in the analysis and these two variables are not solely dependent on growth. Neither CHAM or GSR had significant F values GSR was not included in the STEPWISE analysis.

The importance of LOC (location) as an independent variable was investigated in the group A and B analyses; and the results indicate that LOC is not a factor in correctly classifying specimens. The results improve only very slightly from 84.88% (A-1) to 85.66% (A-3) when LOC, as dummy variables, is included. When LOC is treated as the dependent variable (B-1, B-3) the results are very poor, only 55% correctly classified. There are no patterns between formae present

and location, which further strengthens the conclusion drawn from the subjective investigation, that these are not subspecies.

The variables with large partial *F*-values are the variables which are almost exclusively present or absent within each forma (as determined using the STEPWISE method, group C analyses). These included AOMA (angle of margin), UMC0 (presence of an imperforate umbilical collar), POR (density of the porosity) and NOBO (number of bosses). Generally, at least five functions are required to account for 95% of the variance. This indicates that no one variable (or variables) alone discriminate the groups, but various unique combinations of these variables are the best discriminators.

When the split sample approach was used, (group D analyses) the classification results were very consistent, varying no more than 3.5% for the two samples (analysis and holdout) for the four split-sample analyses. This further indicates that the functions are good discriminators.

The Group E analyses investigated the changes in the functions when the taxonomic framework was enlarged by including three other species of Elphidiidae. The results are represented graphically by the territorial map (Figure 4). The group centroids of the E. excavatum morphotypes are spatially closer together than the distance from any morphotype centroid to that for the two other Elphidium species. The farthest distance is from the Haynesina orbiculare centroid to any morphotype centroid. The group centroids for E. subarcticum and E. bartletti appear to be superimposed. These results graphically represent the classification hierarchy derived from subjective taxonomy. Intraspecific distances are smaller than

specific ones, which in turn are smaller than generic ones. However the spatially close relationship between E. bartletti and E. subarcticum is probably due to the variables the discrimination is based on; the variables were chosen within the context of E. excavatum and are not good discriminators for these two species. These results do indicate that distances in terms of discriminant function scores may represent distances within the conventional classification hierarchy.

CONCLUSIONS

- 1) The consistent high value of the overall percent correctly classified indicates the functions calculated are good discriminators; which in turn indicates that the morphological characters observed and included in the analysis are those characteristic of each morphotype and those necessary for defining each form. This is strengthened by the consistency of the split-sample analyses results.
- 2) Location is not a factor in determining morphotype and there is no pattern between morphotype occurrences and geographical location. This strengthens the conclusion from the subjective investigation that these morphotypes are not subspecies but are ecophenotypes (formae).
- 3) Adding three species to the analysis indicates that the spatial distance (represented graphically) between the group mean scores (centroids) of the first two discriminant functions for any two groups is related to their taxonomic relationship. The infraspecific distance is less than an ecophenotype - species (to E. bartletti or E. subarcticum) distance which is less than an ecophenotype - genera distance (to H. orbiculare). Refinement of the system may make it

possible to determine taxonomic relationships from mean function score differences and graphical distances. This would be a help particularly in determining if variants are species or infraspecific variants; or in determining if variants are local (isolated and unique) versus cosmopolitan.

SUMMARY OF RESULTS

The biological and statistical investigations indicate that Elphidium excavatum (Terquem) is a polytypic species comprising at least nine (possibly ten) distinct ecophenotypes (formae). These formae are first subjectively defined and then statistically tested using discriminant analysis. The results show that 84-91% of the specimens are classified into the groups to which they were subjectively assigned. The functions and the morphological characteristics on which the functions are based are good discriminators.

The presence of intergradation, and the lack of a pattern between morphotype occurrences and geographical location indicates that these are ecophenotypes rather than subspecies. The distribution patterns of the formae, however, suggest association with environmental variables rather than simple geographic locality (many formae can live in one area at times). These observations lead to the rejection of the subspecies ranks proposed by Wilkinson (1979) and Haynes (1981). The designation of "forma" has been retained from Feyling-Hanssen (1972) and ten formae can be recognized with the binocular microscope: 1) E. excavatum forma excavata (= forma selsevensis of Feyling-Hanssen) found as a constituent of populations in intertidal zones; 2) forma williamsoni is the dominant intertidal/marsh form where there is little wave action; 3) forma selsevensis (not sensu Feyling-Hanssen) a temperate to polar water estuarine morphotype; 4) forma clavata, the dominant member of the group found in cold normal marine waters or slightly reduced salinities; 5) forma gunteri, which appears to replace forma clavata

in temperate to tropical waters; 6) forma galvestonensis, appearing to be geographically isolated along eastern North America, a tropical, nearshore lagoon form, preferring normal to hyper-salinities; 7) forma lidoensis, a warm to temperate water estuarine and lagoon form; 8) forma cuvillieri appears to be a subtidal temperate to tropical normal marine form; 9) forma tumidum which has been observed only along the western North American coast and 10) forma magna a nearshore, turbulent zone morphotype.

Variability in the E. excavatum group (number of formae and the percentage of intermediate forms present) appears to increase when each, or a combination of, the following variables is increased: water temperature, proximity to shore, estuarine influence, and the range of annual climatic variation.

The classification results indicate that it is possible to recognize, objectively and consistently, morphotypes within species, and to draw artificial but practical morphological boundaries between morphotypes, in order to identify and classify them, and create workable taxonomic units. With a clearly objectively defined taxonomy, it is possible to compare data from different sources and to draw more concise conclusions about the paleo-ecological implications and significance of the ecophenotypes.

The statistical analysis completed here is a simple procedure with the aid of computational facilities, but the data collection techniques time consuming, elementary, crude, and they introduce operator error. These methods will become obsolete as more sophisticated equipment is developed capable of more complex analyses. Biometry on foraminifera is heavily constrained by inadequate data on

shape, particularly chamber shape (Scott 1974). The solution to these problems may be micro-analysers - microcomputer based image analysis systems such as the one described by Granlund and Hermelin (1983). These systems eliminate operator error, minimize the time required for data collection and provide a vastly superior system for collection, storage, and retrieval of morphological data (Granlund and Hermelin 1983).

SYSTEMATIC PALAEOLOGY

Identified material was supplied to the author by Prof. G. Lutze, Prof. R. Feyling-Hanssen, Prof. J. R. Haynes, Prof. J. W. Murray, Prof. D. Sloan, Dr. M. A. Buzas, Dr. S. W. Snyder, and Miss R. Todd. Dr. G. Vilks, Dr. C. Schafer, Dr. G. Bartlett and Dr. C. W. Poag assisted the authors with some of the identifications. The primary and secondary specimens of a new forma have been deposited at the Smithsonian Institution together with representative specimens of other formae. In addition, representative collections have been deposited with various persons/departments; a listing is given in Appendix E.

The following collections were examined: primary and secondary types of Elphidium and related genera housed at the Smithsonian Institution (including identified material from Dr. A. Lévy and Dr. F. W. Haake); specimens of Natland's at Scripps Institute of Oceanography; and Heron-Allen and Earland's specimens from the shores of Selsey Bill, Sussex, at the British Museum of Natural History.

Listed below, for each forma, is a synonymic list, selected from available references with clear illustrations. Original references are cited, regardless of the clarity of the illustrations.

It must be emphasized that the formae are here described using the conventional format for species, although they all belong to the same species.

In two instances the term "ab" is used (i.e. Elphidium excavatum forma williamsoni Haynes ab Williamson and E. excavatum forma oceanensis Fornasini ab d'Orbigny). The term was first used in this

sense by Medioli and Scott (in press) and designates a binomen that was first published in a way that did not make it available (e.g. a nomen nudum) and only a later work made it available. The author preceeding "ab" is the legal author of the species, but the author following "ab" published the binomen first.

In the synonymies below, figured specimens of one species, recorded in the reference cited may be placed in more than one forma. This has been designated as such: E. excavatum forma clavata: E. clavatum Cushman. BUZAS, 1966, (part), p. 591, pl. 71:1-2; E. excavatum forma selseyensis: E. clavatum Cushman. BUZAS, 1966, (part), p. 591, pl. 71:3-4.

Order FORAMINIFERA	Eichwald, 1830
Suborder ROTALINA	Delage and Herouard, 1896
Superfamily ROTALIACEA	Ehrenburg, 1839
Family ELPHIDIIDAE	Galloway, 1933
Genus ELPHIDIUM	de Montfort, 1808
Type species <u>Elphidium</u>	
<u>macellum</u>	Fichtel and Moll, 1798

Genus Elphidium
de Montfort, 1808.

The genus description as published and cited by Loeblich and

Tappan (1964) does not warrant repeating (see also Hansen and Lykke-Andersen [1976], and Rosset-Moulinier [1976]).

However, some debate has arisen as to which genus (Elphidium, Criboelphidium, Cribrononion) the species Elphidium excavatum (Terquem) belongs. The majority of workers have retained it in the genus Elphidium (Feyling-Hanssen 1972, Lévy et al. 1975, Hansen and Lykke-Andersen 1976, Rosset-Moulinier 1976). Elphidium and Criboelphidium have an areal aperture (which is absent in Cribrononion); all three genera have a row of pores at the base of the apertural face. Loeblich and Tappan (1964) differentiate the genera on the presence (in Elphidium) or absence (in Criboelphidium and Cribrononion) of retral processes; Lévy et al. (1975), Hansen and Lykke-Andersen (1976) and Rosset-Moulinier (1976) report finding retral processes and consequently retain the species in the genus Elphidium. There is also the genus Elphidiononion described by Hofker (1951) based on Polystomella poevana d'Orbigny, in which this author has also placed Heron-Allen and Earlands' species P. selsevense (Hofker 1977). Hansen and Lykke-Andersen (1976) consider the three genera Cellanthus, Criboelphidium and Cribrononion synonymous with Elphidium.

Scott and Medioli (1980) have discussed the validity of using the apertures for generic differentiation; they have placed Criboelphidium in synonymy with Cribrononion, and have retained the species E. excavatum as part of the genus Cribrononion, based on differences with the type species of Elphidium (also Haman 1973).

As Hansen and Lykke-Andersen (1976) point out, in the past the

term "retral process" has had varied definitions. Hansen and Lykke-Andersen (1976) follow Wade's (1957) terminology: returning to Carpenter's (1862, p. 278-279) definition of a retral process as an extension of the chamber lumen found on the inside of the chambers. Often, the term retral process has been applied to include the surrounding wall that spans the suture. Hansen and Lykke-Andersen (1976, p. 4) suggest the term "ponticulus" (Latin: small bridge) to characterize the "sutural bridge" or prolongation of the wall that spans the suture. Thus, a ponticulus may be hollow (if it delineates a retral process), or it may be solid.

Carpenter's (1862) definition of a retral process, and Hansen and Lykke-Andersen's (1976) definition of a ponticulus apply in this work.

Elphidium excavatum (Terquem) forma excavata
Terquem, 1876

pl. 1:1:1; pl. 2:18-20; pl. 3:7-8; pl. 4:10-11; pl. 5:
13-16; pl. 6:8-16, 19; pl. 7:13-16; pl. 8:15-16; pl.
10:6-8,14; pl.12:12; pl.13 :10; pl.14:11-12; ?pl.16:
15; pl. 20:1-12

Polystomella excavata. TERQUEM, 1875, p. 20, pl.2:20f (Non vide, not available). TERQUEM, 1876, p. 429, pl. 2:2a-f. (fide Cushman 1930, 1939; Lévy et al. 1969, 1975; Feyling-Hanssen 1972).

Elphidium excavatum (Terquem). Cushman, 1930, (part), p. 21, pl. 8:1-3. (figures after Terquem). CUSHMAN, 1939, p. 58, pl. 16:7-9 (figures after Terquem). CUSHMAN, 1944, p. 26, pl. 3:40. PARKER, 1952a, p. 412, pl. 5:8. PARKER, 1952b, p. 448, pl. 3:13. LÉVY, 1966, p. 4, pl. 1:8a-b. MURRAY, 1971, p. 159, pl. 66:1, 2, 4, 6, 7. KNUDSEN, 1973b, p. 279, pl. 2:3. LÉVY ET AL., 1975, p. 176-178,

text-fig. 9 (figures after Terquem), pl. 3:1, 2, 5, 6.

ROSSET-MOULINIER, 1976, (part), p.38, pl. 2:14-15. KNUDSEN, 1979, (part), p. 208-209, pl. 1:15-18; pl. 5:1-2.

Elphidium translucens. NATLAND, 1938, p. 144, pl. 5:3-4. CUSHMAN, 1939, p. 65, pl. 20:7a-b. BANDY, 1953, p. 176, pl. 22:9a-b. PARKER ET AL., 1953, p. 9, pl. 3:27. PHLEGER, 1954, p. 639, pl. 2:10. TODD and BRONNIMAN, 1957, p. 39, pl. 7:6a-b. TODD and LOW, 1961, p. 20, pl. 2:4. PHLEGER and EWING, 1962, p. 178, pl. 4:7. PHLEGER, 1964, p. 388, pl. 2:1.

Elphidium incertum (Williamson). PHLEGER and WALTON, 1950, (part), p. 277, pl. 2:17, 18.

Elphidium incertum (Williamson) var. clavatum Cushman. PHLEGER, 1952b, p. 83, pl. 14:10.

Elphidium hispidulum Cushman. TODD and BRÖNNIMANN, 1957, p. 39, pl. 2:1.

Elphidium poeyana (d'Orbigny). LANKFORD, 1959, p. 2083, pl. II:5a-b.

Elphidium spinatum Cushman and Valentine var. translucens Natland. UCHIO, 1960, p. 62, pl. 4:23, 24.

Elphidium selseyense (Heron-Allen and Earland). HAAKE, 1962 (part), p. 49, pl. 6:1. JONES and ROSS, 1979, text-fig. 7:A-B.

Elphidium clavatum Cushman. LOEBLICH and TAPPAN, 1953, (part), p. 98, pl. 19:9-10. HANSEN, 1965, p. 325, text-figs. 5:6. BRODNIEWICZ, 1965, (part), p. 210-213, pl. 10:3. KNUDSEN, 1971b, p. 273, pl. 20:5-6

Elphidium cf. clavatum Cushman. MICHELSEN, 1967, p. 237-238, pl. 4:7a-b.

Cribrononion excavatum (Terquem). HAAKE, 1967, (part), p. 13-27, pl. 1:4-5. LÉVY ET AL., 1969, p. 93, pl. 1:1-3 (figures after Terquem); pl. 1:4.

Elphidium incertum (Williamson) variant. MURRAY, 1968, p. 83-96, pl. 1:7a-b.

Cribrononion incertum (Williamson). VON DANIELS, 1970, p.88, pl. 7:12.

Elphidium excavatum (Terquem) forma selsevensis (Heron-Allen and Earland). FEYLING-HANSSSEN, 1972, p. 341, pl. 5:1-4. KNUDSEN, 1976, p. 431-449. FEYLING-HANSSSEN, 1976b, p. 177. FEYLING-HANSSSEN, 1976c, p. 355. MILLER, 1979, p. 33, pl. 1:1-2, pl. 2:1-2, pl. 3:1, pl. 4:4-5, pl. 5:4-5. SNYDER and KATROSH, 1979, p. 254, pl. 1:1-2. SLOAN, 1981, p. 275, pl. 1:5. KNUDSEN, 1982, p. 170, fig. 14:14:5-6.

Cribroelphidium spinatum var. transluscens (Natland). SCOTT, 1976, p. 170. SCOTT ET AL., 1976, p.74.

Cribroelphidium excavatum selsevensis (Heron-Allen and Earland). SCOTT, 1977, p. 170, pl. 6:3. SCOTT ET AL., 1977, p. 1579, pl. 5:3.

Elphidium excavatum forma clavata Cushman. KNUDSEN, 1978, (part) p. 34, pl. 5:4.

Elphidium clavatum nudum. WILKINSON, 1979, p. 638, pl. 1:6, pl. 2:8.

Cribrononion excavatum forma selsevensis (Heron-Allen and Earland). SCOTT and MEDIOLI, 1980, p. 35, pl. 5:6. SCOTT ET AL., 1980, p. 228.

Elphidium ex gr. excavatum (Terquem). Elphidium excavatum (Terquem) sensu stricto. HAYNES, 1981, p. 61-62, pl. 8:9.

Elphidium sp. group B. CANN and DE DEKKER, 1981, (part), p. 663-664, pl. 1:3-5, 17.

Elphidium excavatum forma excavata (Terquem). MILLER ET AL., 1982, p. 128-130, pl. 1:9-12, pl. 2:1-2, pl. 3:1-2, pl. 4:13-16, pl. 5:15-16, pl. 6:6-8, 14.

Elphidium excavatum (Terquem) forma alba Feyling-Hanssen. KNUDSEN, 1982, p. 170, fig. 14:12:5, fig. 14:14:8-9.

Types:

Holotype: Polystomella excavata (specimen lost) Terquem, 1875, p. 20, pl. 2:2a-f, (not available, private publication of the author; fide Ellis and Messina, 1940). Terquem, 1876, p. 429, pl. 2:2a-f (repeat of the 1875 MS).

Deposited in: Musee de Dunkerque, France.

Neotype: Elphidium excavatum (Terquem). Lévy et al., 1975, p. 176-178, pl. 3:1-2 No. FG 447.

Neoparatypes: No. FG 448.

Deposited in: l'Institute de Paleotologie du Museum National d'Historie Naturelle de Paris.

Topotype: Elphidium excavatum (Terquem). Levy et al., 1975, p. 176-178, pl. 3:5-6, also topotypes deposited at the Smithsonian Institution, USNM No. 343319.

Types of junior synonyms:

Elphidium translucens:

Holotype: Natland, 1938, p. 144, pl. 5:3-4, USNM No. 22549.

Paratype: USNM No. 23332.

Deposited in: Smithsonian Institution.

Representative plesiotype: Miller, 1979, pl. 4:4, pl. 5:4; Miller et al. 1982, pl. 1:10; USNM No. 312511.

Diagnostic characteristics: Generally small, though larger than forma clavata, and with chambers more lobate and sutures straighter, extending unstricted into the umbilicus. Elphidium excavatum forma excavata generally has a greater pore density than E. excavatum forma clavata; giving the test a hazy appearance. The pore perforations extend to the umbilicus. It is interesting to note that Terquem used for this species the epithet excavata (excavated umbilicus?).

Description: As described by Feyling-Hanssen in 1972 (E. excavatum forma selseyensis), and Lévy et al. in 1975 (E. excavatum).

Maximum Diameters: 0.35 mm - 0.55 mm (Terquem 1876, p. 429); 0.25 mm - 0.65 mm (Lutze, 1965, text-fig. 8); 0.38 mm - 0.66 mm (Bartlett, 1965b, p. 17).

Remarks: Elphidium excavatum forma excavata (Terquem) is the name retained for the original specimens described by Terquem (Polystomella excavata) and the neotype of Lévy et al., 1975.

Feyling-Hanssen (1972) indicates that many European workers (among them van Voorthuysen 1960, Brodniewicz 1965, Hansen 1965, Lutze 1968) grouped this form E. excavatum forma excavata with E. clavatum (= E. excavatum forma clavata). Some workers included it in synonymy

with E. selseyense (= E. excavatum forma selseyensis), among them Brand 1941, Haake 1962, Lévy et al. 1969, Feyling-Hanssen 1972.

Many European workers were following the lead of Cushman's (1930, 1939, 1949) and identifying a form that appears to be the "species" of Williamson, Polystomella umbilicatula 1858 = (E. williamsoni Haynes, 1973) as E. excavatum (i.e. van Voorthuysen 1957, 1960, Richter 1961, 1964a, 1964b, 1967, Jarke 1961, Woszydlo 1962, Haake 1962, 1967, Feyling-Hanssen 1964, Brodniewicz 1965, and Murray 1965a, 1968, 1970). Lutze (1968) Murray (1971) and Haynes (1973) have all commented on the confusion arising from Cushman's work, and they too placed Cushman's E. excavatum in synonymy with Williamson's taxon.

As pointed out by Wilkinson (1979), Terquem's original material was lost, and Lévy et al. (1975) , designated a neotype (E. excavatum of Lévy et al. = E. excavatum forma excavata).

Rodrigues and Hooper (1982) suggest that of the three specimens of Polystomella excavata illustrated by Terquem (1875, 1876), one (Pl. 2:2c-d) was of Polystomella umbilicatula. Of Lévy et al.'s (1975) specimens of E. excavatum Rodrigues and Hooper (1982) refer the neotype (1975, Pl. 3:1-2) and topotype (1975, Pl. 3:5-6) to P. striatopunctata var. selseyense, thus they do not believe E. excavatum sensu Terquem and E. excavatum sensu Lévy et al. to be conspecific. After examining the Heron-Allen and Earland collection at the British Museum, and topotypic material of Lévy's at the Smithsonian Institution, the conclusion is reached that Lévy's material is not P. striatopunctata var. selseyense (sensu Heron-Allen and Earland). Rodrigues and Hooper's (1982) suggestion that Terquem included E.

williamsoni in his species may be correct; but they base this on the observations of an umbilical boss (in one) and ponticuli on the periphery (on both) of Terquem's two illustrations. Some specimens observed here have an umbilical boss and/or ponticuli on the periphery and yet have been attributed to E. excavatum forma excavata, not E. excavatum forma williamsoni (i.e. pl.6:12,15, pl. 20: 3,5,6,8; also Miller et al., 1982, Pl. 1:10, Pl.2:2, Pl. 4:14). The other suggestion of Rodrigues and Hooper's (1982), that E. excavatum sensu Terquem and E. williamsoni Haynes are conspecific, is not a new observation (it was made by van Voorthuysen in 1957). It must be remembered, however, that what van Voorthuysen (1957) identified as E. excavatum has been listed in synonymy as E. williamsoni by Lutze 1968, Murray 1971 and Haynes 1973. Because Terquem's material has been lost, it is suggested here that 1) the concept of E. excavatum s.s. (or forma excavata) be based on Lévy et al.'s neotype; 2) the concept of forma williamsoni be based on Williamson's and Haynes' "species", and 3) the concept of forma selseyensis be based on Heron-Allen and Earland's "species".

Here, Elphidium translucens Natland is considered a junior synonym.

Distribution: The form as described here has often been placed in synonymy with either E. excavatum forma clavata or forma selseyensis, or confused with E. excavatum forma williamsoni (= E. williamsoni Haynes ab Williamson), and consequently the distribution of this form is difficult to determine. It appears to be a nearshore form becoming

slightly more common in intertidal and shallow subtidal environments.

Elphidium excavatum forma williamsoni
Haynes, 1973, ab Williamson, 1858.

pl. 6:17-18; pl. 9:1-16; pl. 12:19; pl. 17:1-12;
?pl. 18:17; pl. 19:16-17; pl. 22:1-24

?Polystomella articulata. d'Orbigny, 1839b, p. 30, pl. 3:9, 10 (fide Murray, 1971).

Polystomella umbilicatula. WILLIAMSON, 1858, p. 42-44, pl. 3:81-82.
(non Nautilus umbilicatula Walker and Jacob). TERQUEM, 1875, p. 25,
pl. 2 : 3a-b. (fide Lévy et al., 1969; Haynes, 1973). TERQUEM, 1876,
p. 429 (fide Lévy et al., 1969; Haynes, 1973).

Polystomella striatopunctata (Fichtel and Moll) variety. HERON-ALLEN
and EARLAND, 1909, (part), p. 695, pl. 21:2a-c.

Nonionina depressula Walker and Jacob sp. HERON-ALLEN and EARLAND,
1909, (part), p. 692.

Polystomella striatopunctata (Fichtel and Moll) var. selsevensis.
HERON-ALLEN and EARLAND, 1911, (part), p. 448.

Elphidium excavatum (Terquem). CUSHMAN, 1930, (part), p. 21, pl.
8:4-7. HERON-ALLEN and EARLAND, 1932, p. 439, pl.16:21-23. CUSHMAN,
1939, p. 58, pl. 16:10 (figure after Williamson), pl.16:11-12.
CUSHMAN, 1949, p. 28, pl. 6:2a-b (fide Haynes, 1973). VAN
VOORTHUYSEN, 1957, p. 31, pl. 23:8a-b. VAN VOORTHUYSEN, 1960, p. 255.
JARKE, 1961, p. 21-36, pl. 2:2. RICHTER, 1961, p. 163-170, pl. 1.
TODD and LOW, 1961, p. 19, pl. 2:5. HAAKE, 1962, (part), p. 47-48,
pl. 5:5. WOSZIDLO, 1962, p. 74-75, pl. 3:8. FEYLING-HANSSSEN, 1964,

p. 344, pl. 20:7-8. RICHTER, 1964a, p. 343-353, text-fig. 3-4. ADAMS and FRAMPTON, 1965, p. 58, pl. 5:7. BRODNIEWICZ, 1965, p. 214, pl. 7:5, pl. 11:4. MURRAY, 1965a, p. 513, pl. 1:6, 6 (fide Haynes, 1973). MICHELSEN, 1967, p. 238, pl. 5:2. RICHTER, 1967, p. 291-335. MURRAY, 1968, p. 83-96, pl. 1:12a-b. HAMAN, 1969, p. 139-142 (Hamen, 1981, pers. comm.). MURRAY, 1970, p. 484. BOLTOVSKOY and VIDARTE, 1977, (part), p. 38, pl. 2:12.

Elphidium articulatum (d'Orbigny). CUSHMAN, 1930, p. 22, pl. 10:6-8. BANDY, 1953, p. 176, pl. 22:5a-b. MURRAY, 1971, pl. 153, pl. 63:1-7. KNUDSEN, 1973a, p. 188, pl. 3:13. KNUDSEN, 1973b, p. 278, pl. 2:2. KNUDSEN, 1976, p. 431-449. ROSSET-MOULINIER, 1976, p. 89, pl. 1:1-4. APTHORPE, 1980, (part), p. 225, p. 2:6. BOLTOVSKOY ET AL., 1980, p. 29, pl. 13:1-4.

Elphidium alvarezianum (d'Orbigny). HERON-ALLEN and EARLAND, 1932, p. 440, pl. XVI:24-25.

Criboelphidium cf. koeboense (LeRoy). LEHMANN, 1957, p. 348, pl. 2:21-24.

Elphidium clavatum Cushman. JARKE, 1961, p. 21-36, pl. 2:2. BRODNIEWICZ, 1965, (part), p. 210-213, pl. 10:2, 4, 7.

Ephidium sp. 1. HAAKE, 1962, pl. 5:9.

Cribrononion cf. alvarezianum (d'Orbigny). LUTZE, 1965, p. 101-102, pl. 15:4-6.

Elphidium oceanicum (Cushman). LÉVY, 1966, p. 5, pl. 1:2a-b.

Elphidium kusiroense Asano. MATOBA, 1967, p. 254, pl. 27:9a-b.

Elphidium aff. semistriatum (d'Orbigny). CITA and PREMOLI-SILVA, 1967, (part), p. 35, pl. 2:10.

- Elphidium sp. CITA and PREMOLI-SILVA, 1967, (part), pl. 2:14-15.
- Cribrononion excavatum (Terquem). HAAKE, 1967, (part), p. 13-27, pl. 1:7-8.
- Cribrononion articulatum (d'Orbigny). LUTZE, 1968, p. 27, pl. 1:1-2.
- Elphidium umbilicatum (Williamson). LÉVY ET AL., 1969, p. 96, pl. 1:6, pl. 2:1-2. KNUDSEN, 1971b, p. 281-282, pl. 13:8-11, pl. 23:1-4.
- Cribroelphidium articulatum (d'Orbigny). ROSSET-MOULINIER, 1972, p. 176, pl. 14:1-5.
- Elphidium incertum (Williamson). SHEENAN and BANNER, 1972, p. 31-40, pl. 1 1-5.
- Elphidium williamsoni. HAYNES, 1973, p. 207, pl. 24:7, pl. 25:6, 9, pl. 27:1-3. HANSEN and LYKKE-ANDERSEN, 1976, p. 9, pl. 5:1 - 6. CULVER and BANNER, 1978, p. 53-72. KNUDSEN, 1979, p. 210, pl. 1:19, pl. 5:6-7. MURRAY, 1979, p. 52, fig. 16:C-D. SCOTT and MEDIOLI, 1980, p. 40, pl. 5:4. SCOTT ET AL., 1980, p. 228. KNUDSEN, 1982, p. 170, fig. 14:12:19, fig. 14:14:10-11.
- Elphidium hughesi Cushman and Grant. BERGEN and O'NEILL, 1976, p. 1290, pl. 1:1-2.
- Cribroelphidium excavatum (Terquem). SCOTT, 1977, p. 169, pl. 6:1. SCOTT ET AL., 1977, p. 1578, pl. 5:4.
- Elphidium sp. group A. CANN and DE DEKKER, 1981, p. 663, pl. 1:7-16, 18-23.
- Elphidium ex gr. excavatum (Terquem). Elphidium williamsoni Haynes. HAYNES, 1981, p. 61-62, pl. 8:11.

Types:

Holotype: Polystomella umbilicatula, (non Nautilus umbilicatus, Walker and Jacob); Williamson, 1858, p. 42-44, pl. 3.:81-82.

Hypotypes: Elphidium williamsoni Haynes, 1973, p. 207-209, pl. 24:7, pl. 25:6, 9.

Deposited in: British Museum - Natural History, Stub 1970:II:26:597; slide 197 0:II:26:431:432; sections 1970:II:26:507, 1970:II:26:508.

Types of junior synovynms:

?Polystomella articulata.

Holotype: lost-d'Orbigny, 1839b, p. 30, pl. 3:9, 10.

Deposited in: ? Laboratoire de Paleontologie du Museum de l'Histoire Naturelle, Paris.

Representative plesiotype: pl.21:1-2.

Diagnostic characteristics: An inflated, rotund form, with smooth peripheral outline and rounded periphery. A flat umbilicus on each side, chambers extending completely to the umbilicus; a boss/papillae may be present. Ponticuli very regular and well developed, covering up to half the chamber width. Wall very finely and densely perforate.

Description: As described by: Williamson (Polystomella umbilcatula) in 1858, Haynes (E. williamsoni) in 1973, and Hansen and Lykke-Andersen (E. williamsoni) in 1976.

Maximum diameters: Diameter 0.48 mm, width approx. 0.20 mm (Haynes, 1973, p. 208).

Remarks: The specific name williamsoni was suggested by Haynes, (1973) for the inflated, many chambered marsh and estuarine species first referred to by Williamson (1858, p. 42-44, Pl.3:81-82) as Polystomella umbilicatula (Walker and Jacob) and widely referred to by many European workers as Elphidium excavatum (Terquem) (i.e.: van Voorthuysen 1957, 1960, Jarke 1961, Richter 1961, 1964a, 1964b, 1967, Woszidlo 1962, Haake 1962, 1967, Feyling-Hanssen 1964, Brodriewicz 1965, and Murray 1965a, 1968, and 1970). Lévy et al. (1969) refer to this marsh species as E. umbilicatula (Williamson) but, as pointed out by Haynes (1973) this latter designation contravenes Article 49 of the ICZN (Stoll et al. 1961) because Williamson placed his species in synonymy with Nautilus umbilicatus Walker and Jacob, a species clearly distinct from his. Consequently, Haynes suggested the designation E. williamsoni, a nomen novum (ICZN, Appendix E, Paragraph 21; Stoll et al. 1961) for this form because it was clearly Williamson's species. The epithet williamsoni has been followed here. However, Haynes designated his own material as holotype of the new nominal species; thus contravening Article 72D of the ICZN (Stoll et al. 1961) which specifically states that in this case Williamson's specimen should have been designated as holotype. Consequently, Williamson's specimen is designated here as the only valid holotype; and Haynes' specimens (holotype and paratypes) should be demoted to hypotypes. Haynes (1981) included this form in the E. excavatum

"polytypic species" group.

Rodrigues and Hooper (1982) are of the opinion that Terquem's original concept of E. excavatum included E. williamsoni.

This species has also been referred to as Elphidium articulatum (d'Orbigny). The relationship between E. articulatum and E. williamsoni is very confusing in the literature and there are many contradictory opinions.

E. articulatum was described by d'Orbigny (1839b) from the coast of Patagonia and the Falkland Islands. Williamson (1858, p. 43) remarked on the similarity between his species and d'Orbigny's figure of P. articulata, stating that the only difference between the two was that d'Orbigny described his specimens as having numerous septal apertures dispersed irregularly over the entire plane, instead of being confined to a single row as in his specimens.

Cushman (1930) distinguished between E. articulatum and E. excavatum (= E. williamsoni?) and maintained the distinction later.

Heron-Allen and Earland (1932) also reported finding E. articulatum from the Falkland Islands - and distinguished it from E. excavatum. Heron-Allen and Earland reported some confusion about the type when they examined d'Orbigny's collection in Paris. The specimen identified there as P. articulata was sharp edged with 12 chambers and in their opinion (p. 439-440) was possibly P. flexosa.

Parker (1952a) considers E. articulatum closely related to E. bartletti Cushman, and that young specimens of the two are identical. Parker says (1952a, p. 411): "A comparison with specimens from the

Falkland Islands shows the Portsmouth species to be almost identical although slightly less compressed." She further states: "It is possible that E. bartletti represents the Arctic development of E. articulatum which is not reported from that area." Loeblich and Tappan (1953, p. 98) report that d'Orbigny's type figure shows a sharp, acutely angled periphery, rather than the broadly rounded periphery of all other specimens referred to his species, and present in E. bartletti.

Haynes (1973), regards the type specimen of E. articulatum as lost but says that the type figure of E. articulatum shows a lobate species with inflated chambers, small septal pits, and areal apertures, which is different from the flat sided E. excavatum with sutural bridges. These observations of the type figure are in agreement with the concept of E. bartletti.

Rosset-Moulinier (1976, p. 89) has examined topotypic material (i.e. from the Falklands and Patagonia) of E. articulatum supplied by Boltovskoy and states E. articulatum is conspecific with E. williamsoni.

Murray (1982 pers. comm.) examined the d'Orbigny collection in Paris in the early 1960's and said that the state of preservation of the specimen was so poor as to render it useless.

Haynes (1982 pers. comm.) has suggested that the name E. articulatum be regarded as a nomen dubium; and due to the conflicting opinions in the literature on the concept of E. articulatum, Haynes suggestion has been taken here. The only unambiguous epithet for this form is williamsoni Haynes ab Williamson.

Distribution: A widely distributed form, extensively documented in the literature. Reported from shallow subtidal and intertidal zones in outer estuaries, lagoons, tidal flats and in particular marshes, where the salinity is near normal and there is no wave action, only tidal flux. It is the dominant Elphidium in intertidal areas of marshes along the European and North American (particularly north of New York) Atlantic seaboard. It is also found in glacio-fluvial and post-glacial late Pleistocene-Holocene sediments (i.e. Knudsen 1979), and has been reported as far back as the Eemian in Europe (van Voorthuysen 1957).

Elphidium excavatum (Terquem) forma selseyensis
Heron-Allen and Earland, 1911 (emended Brand, 1941)

pl.1:2:2; pl.7:1; pl. 8: 10-13; pl.10: 9-13; pl.11:15;
pl.12:14-15; pl.13: 12-15; pl.14: 8-9, 18; pl. 22: 1-9

Polystomella striatopunctata (Fichtel and Moll) variety. HERON-ALLEN and EARLAND, 1909, (part), p. 695, pl. 21:2a-c.

Nonionina depressula Walker and Jacob sp. HERON-ALLEN and EARLAND, 1909 (part) p. 92.

Polystomella striatopunctata (Fichtel and Moll) var. selseyensis. HERON-ALLEN and EARLAND, 1911 (part), p. 448.

Elphidium incertum (Williamson). CUSHMAN and COLE, 1930, p. 96, pl. 13:6,7. CUSHMAN, 1930, (part), p. 18, pl. 7:5, 8, 9. CUSHMAN, 1944, p. 25, pl. 3:28-31. PHLEGER and WALTON, 1950, (part), p. 277, pl. 2:17, 18. SCHNITKER, 1971, p. 198, pl.7:4a-b.

Elphidium incertum (Williamson) var. clavatum Cushman. CUSHMAN and COLE, 1930, p. 96, pl. 13:8-9. CUSHMAN, 1944, p. 25, pl.3:32-33. PARKER, 1952a, (part), p. 412, pl. 5:10-11. TODD and BRÖNNIMANN, 1957, (part), p. 39, pl. 6:10.

Elphidium brooklynense. SHUPACK, 1934, p. 10, opp. p. 9, figs. 7a-b.

Elphidium discoidale (d'Orbigny). SHUPACK, 1934, p. 11, opp. p. 9, figs. 9a-b.

Elphidium gunteri Cole. SHUPACK, 1934, (part), p. 11, opp. p. 9, figs. 6-b. CUSHMAN, 1944, p. 27, pl. 3:42. TODD and LOW, 1961, p. 19, pl. 2:10.

Elphidium clavatum Cushman. SHUPACK, 1934, p. 11, opp. p. 9, figs. 8a-b. WEISS, 1954, p. 159, pl. 32:14. BUZAS, 1965b, (part), p. 58-59, pl. 2:6-7, pl. 3:1-2. SCHNITKER, 1971, p. 198, pl.7:5a-b.

Elphidium selseyense (Heron-Allen and Earland). CUSHMAN, 1939, p. 59-60, pl. 16:26-27 (figures after Heron-Allen and Earland). RICHTER, 1961, p. 163-170, pl. 1. HAAKE, 1962, (part), p. 49, pl. 5:12-15; pl. 6:2-4. RICHTER, 1964a, p. 343-353, text-fig. 5. ATKINSON, 1969, p. 538. MURRAY, 1970, p. 484. HAMAN, 1973, p. 134. HAYNES, 1973, (part), p. 204-205, pl. 22:3-4, pl. 26:4, 7, 9, pl. 29:1-3. CULVER and BANNER, 1978, p. 53-78.

Elphidium incertum selseyensis (Heron-Allen and Earland). BRAND, 1941, p. 65-66. FEYLING-HANSEN, 1954, p. 142, pl. 2:12a-b.

Elphidium tumidum Natland. CUSHMAN and TODD, 1947, (part), p. 14, pl. 2:21.

Criboelphidium limnosum. CUSHMAN and BRÖNNIMANN, 1948, p. 19, pl. 4:7. TODD and BRÖNNIMANN, 1957, p. 39, pl. 16:13.

- Elphidium incertum (Williamson) variants. PARKER, 1952b, (part), p. 448, pl. 3:14, 17, pl. 4:1-2.
- Elphidium gunteri Cole var. galvestonense Kornfeld. BANDY, 1954, (part), p. 136, pl. 30:2.
- Elphidium florentinae Shupack. WEISS, 1954, p. 159, pl. 32:8-11.
- Elphidium strattoni (Applin) var. joaquinensis. BANDY and ARNAL, 1957, p. 55, pl 7:6.
- Elphidium yadescens (Cushman and Bronnimann). TODD and BRÖNNIMANN, 1957, p. 39, pl. 7:10-11.
- Elphidium galvestonense Kornfeld. TODD and LOW, 1961, p. 19, pl. 2:9.
- Elphidium longipontis Stschedrina. BRODNIEWICZ, 1965, (part), p. 213, text-fig. 33, pl. 7:5, 7, 8.
- Cribrononion excavatum (Terquem). HAAKE, 1967, (part), p. 13-27, pl. 1:6, 10-11.
- Elphidium excavatum (Terquem). BOLTOVSKOY and BOLTOVSKOY, 1968, (part), p. 148, pl. 1:16a-b. BANNER and CULVER, 1978, p. 177-207, pl. 9:12-14.
- Criboelphidium selseyense (Heron-Allen and Earland). BACHHUBER and MCCLELLAND, 1977, p. 259, text-fig. 3:B.
- Elphidium selseyense (Heron-Allen and Earland). HOFKER, 1977, p. 257, pl. 8:8-9, pl. 9:1.
- Elphidium clavatum selseyense (Heron-Allen and Earland). WILKINSON, 1979, p. 638, pl. 1:5.
- Elphidium excavatum forma album (Feyling-Hanssen). POAG ET AL., 1980, pl. 1:9.
- Elphidium articulatum (d'Orbigny). APHORPE, 1980, (part), p. 225,

pl.28:7.

Elphidium ex gr. excavatum (Terquem). Elphidium selsevense

(Heron-Allen and Earland). HAYNES, 1981, p. 61-62, pl. 8:13.

Elphidium excavatum forma selseyensis (Heron-Allen and Earland).

MILLER ET AL., 1982, p. 132-133, pl. 1:13-16, pl. 5:10-13, pl. 6:9-13.

KNUDSEN, 1982, (part), p. 170, fig.:14:12:11-12, fig.:14:14:7.

Types:

(?)Syntypic series: Polystomella striatopunctata (Fichtel and Moll)

var. Heron-Allen and Earland, 1909, p. 695, pl.21:2a-c.

Deposited in: Heron-Allen and Earland collection, (from Selsey Bill, Sussex), British Museum (Natural History), slide nos: 51, 54, 56, 57, 61, 65, 66, 67.

Lectotype: designated by Brand, 1941, p. 66, as: Polystomella striatopunctata variety selseyensis Heron-Allen and Earland, 1909, p. 695, pl. 21:2a, 2c; not 2b.

Metatype, (possible syntype): designated by Banner and Culver, 1978, as: Elphidium excavatum pl. 9:12-14.

Deposited in: Heron-Allen and Earland collection, British Museum (Natural History), BM(NH) No. ZF3833.

Types of junior synonyms:

Elphidium brooklynense.

Holotype: Shupack, 1934, p. 10, opp. p.9:7a-b

Deposited in: American Museum of Natural History (New York) No. 695

Criboelphidium limnosum.

Holotype: Cushman and Brönnimann, 1948, p. 19, pl. 4:7

Deposited in: Cushman Collection, Smithsonian Institution, USNM No. 56645.

Paratype: USNM No. 56747 (as above).

Elphidium strattoni (Applin) var. joaquinensis.

Holotype: Bandy and Arnal, 1957, p. 55-56; pl. 7:6.

Deposited in: Smithsonian Institution, USNM No. 237448.

Representative plesiotype: pl. 8:13, USNM No. 312512.

Diagnostic characteristics: Test generally larger than E. excavatum forma clavata and forma excavata; the peripheral outline is smooth to lobate, with a subacute periphery. The test thicker is through the umbilicus, consequently the umbilicus appears raised. Sutures slightly backwards curved or straight; with irregular indistinct to strongly developed ponticuli and often papillae filling the sutures. The umbilicus contains papillae, or bosses (irregular lateral extensions of the chambers), or both.

Description: As described by: Heron-Allen and Earland (P. striatopunctata variety) in 1932 and Haynes in 1973 (E. selseyense).

Maximum Diameters: 0.43 mm (Haynes, 1973, p. 205); 0.25 mm - 0.64 mm (Lutze, 1965, text-fig. 8).

Remarks: The name E. excavatum forma selseyensis is retained for the specimens described by Heron-Allen and Earland (1909, 1911), collected from the shores of Selsey Bill. There are two species of particular

interest in their collection: Polystomella striatopunctata (Fichtel and Moll) variety selseyensis, and Nonionina depressula Walker and Jacob variety selseyensis. Specimens of the former, in the opinion of the author, actually belonged to two formae, most specimens are Elphidium excavatum forma williamsoni, the remainder are forma selseyensis (Heron-Allen and Earland) (this work). Most specimens of the latter species appeared to the author to be formae of Elphidium excavatum, mainly forma selseyensis. The remaining specimens appeared not to belong to the genus Elphidium. It should be noted that Heron-Allen and Earland did not designate any "type" specimens of their new variety of Polystomella striatopunctata; and although there are specimens in the collection labelled Nonionina depressula var. selseyensis, in the 1909 publication Heron-Allen and Earland refer only to "Nonionina depressula Walker and Jacob sp." (p. 692).

Brand (1941) published an emended description of this species and he designated Heron-Allen and Earland's 1909 figure (Pl. 21:2a, 2c; not 2b) as lectotype.

The E. excavatum forma selseyensis described here does not include the form described by Feyling-Hanssen (1972) as E. excavatum forma selseyensis.

Haynes (1973) presents SEM photographs of specimens which "closely resemble" (Haynes, 1973, p. 205) those collected by Heron-Allen and Earland and housed in the British Museum (E. selseyense of Haynes, 1973).

Banner and Culver (1978, Pl. 9:12-14) illustrate a specimen of E. excavatum which is a metatype (possible syntype) identified by

Heron-Allen and Earland as Polystomella striatopunctata variety selseyensis. This specimen (# ZF3833) illustrated by Banner and Culver (1978) is considered here to belong to Elphidium excavatum forma selseyensis.

Haynes, (1981) included this form in his E. excavatum "polytypic species" group.

Junior synonyms of this form are: Elphidium brooklynense Shupack (1934), Elphidium strattoni var. joaquinensis Bandy and Arnal, (1957), and Criboelphidium limnosum Cushman and Bronnimann (1948).

Distribution: Preliminary observations indicate that E. excavatum forma selseyensis occurs nearshore, especially in shallow, temperate waters (reaching 10-15°C) under estuarine influence along European and North American coastlines.

Elphidium excavatum (Terquem) forma clavata
Cushman, 1930 (emended, Loeblich and Tappan, 1953).

pl.1:1:3; pl.2:1-8, 9-17; pl. 3:1-6, 9-12; pl. 4:1,
6-9, 13-14; pl. 5:1-7; pl. 6:3-6; pl. 7:4-9; pl. 8:
4-8, 14; pl. 9:18; pl. 10:1-5; pl. 11:5-8; pl. 12:
11; pl. 13:3-5, 8-9, 16; pl. 14:4-5, 13-14; pl. 23:
1-21.

Elphidium incertum (Williamson) var. clavatum. CUSHMAN 1930, p. 20,
pl. 7:10a-b. CUSHMAN, 1939, p. 57, pl. 16:1a-b. CUSHMAN, 1948, p.
57, pl. 6:8. VAN VOORTHUYSEN, 1949, p. 65, pl. 1:4b-c. PHLEGER,
1952a, p. 83, pl. 14:10.

Elphidium incertum (Williamson). CUSHMAN, 1939, p. 57, pl. 15:23-24.
TEN DAM and REINHOLD, 1941, p. 52, pl. 3:8a-b. PARKER, 1948, p. 248,

pl. 5:7. PHLEGER, 1952a, p. 83, pl. 14:7. WEISS, 1954, p. 159, pl. 32:7. ADAMS and FRAMPTON, 1965, p. 58, pl. 5:6. SLOAN, 1981, p. 281, pl. 2:1a-b.

Elphidium incertum (Williamson) and variants. PARKER, 1952b, (part), p. 448, pl. 3:16.

Elphidium clavatum Cushman. LOEBLICH and TAPPAN, 1953, (part), p. 98, 101-102, pl. 19:8. WEISS, 1954, (part), p. 159, pl. 32:13. TODD and LOW, 1961, (part), p. 18, pl. 2:1. ANDERSON, 1963, p. 315. COOPER, 1964, p. 95-96, pl. 6:5-7. BRODNIEWICZ, 1965, (part), p. 210-213, pl. 10:1, 6, 7, 8. BUZAS, 1965a, p. 23, pl. 3:3a-b. NAGY, 1965, p. 124, pl. 2:21 (fide Feyling-Hanssen, 1972). BUZAS, 1966, (part), p. 591, pl. 7:1-2, 5-8. MATOBA, 1967, p. 254, pl. 27:8a-b. MICHELSEN, 1967, p. 236-237, pl. 4:6a-b. TODD and LOW, 1967, p. A33. MURRAY, 1969, (part), p. 416. MATOBA, 1970, p. 51, pl. 6:11a-b. KNUDSEN, 1971b, p. 273, pl. 11:10-13, pl. 20:7-8. SCHNITKER, 1971, (part), p. 198. SEN GUPTA, 1971, p. 89, pl. 2:28-29. BARTLETT and MOLINSKY, 1972, p. 1204-1215, pl. 1:1a-b. CULVER and BANNER, 1978, p. 56. BERGEN and O'NEIL, 1979, p. 1290, pl. 1:5-6. LAGOE, 1979a, p. 260, pl. 1:3, 5, 6. RODRIGUES and HOOPER, 1982, (part), p. 411-416, text-fig. 2:G-P, text-fig. 3:A-P.

Elphidium incertum clavatum Cushman. FEYLING-HANSSSEN, 1954, p. 141, pl. 2:11. BOLTOVSKOY, 1954, p. 275, pl. 24:7. PHLEGER, 1960b, pl. 5:11. FEYLING-HANSSSEN, 1964, p. 345, pl. 20:11-15. COLE and FERGUSON, 1975, p. 21, pl. 7:9-10.

Elphidium incertum incertum (Williamson). FEYLING-HANSSSEN, 1954, p. 141, pl. 2:10. FEYLING-HANSSSEN, 1964, p. 344, pl. 19:16-17.

Elphidium selsevense (Heron-Allen and Earland). HAAKE, 1962, (part), p. 49-50, pl. 6:1. WOSZIDLO, 1962, p. 74-75, pl. 3:10.

Elphidium excavatum (Terquem). TODD and BRÖNNIMANN, 1957, (part), p. 39, pl. 6:12. MURRAY, 1971, p. 159, pl. 66:3, 5. HANSEN and LYKKE-ANDERSON, 1976, p. 10, pl. 6:1-6. BUZAS ET AL., 1977, p. 95. BOLTOVSKOY and VIDARTE, 1977, p. 38, pl. 2:11-12. SCHAFER and COLE, 1978, p. 27, pl. 9:7. KNUDSEN, 1979, (part), p. 208-209, pl. 1:13-14, pl. 4:6-7, pl. 5:3-6. COLE, 1981, p. 100, pl. 15:1-12.

Elphidium incertum (Williamson) "COMPLEX". BARTLETT, 1965b, p. 14, pl. 1:4-12.

Cribrononion excavatum excavatum (Terquem, 1875). LUTZE, 1965, p. 96, pl. 15:39.

Cribrononion excavatum clavatum (Cushman, 1930). LUTZE, 1965, p. 96, pl. 15:40-41.

Elphidium clavatum Cushman "Complex". GREGORY, 1970, p. 226, pl. 14:1.

Elphidium incertum (Williamson) forma clavatum Cushman. WAGNER, 1970, p. 24, pl. 2:3-5.

Elphidium excavatum (Terquem) forma clavata Cushman. FEYLING-HANSEN, 1972, p. 339, pl. 1:1-9, pl. 2:1-9. KNUDSEN, 1973a, p. 188, pl. 5:4. FEYLING-HANSEN, 1976a, p. 92, fig. 8:13-14. FEYLING-HANSEN, 1976b, p. 177. FEYLING-HANSEN, 1976c, p. 355. KNUDSEN, 1976, p. 431-449, pl. 2:14-17. KNUDSEN, 1977, text-fig. 7:15-18, text-fig. 8:5-6. KNUDSEN, 1978, (part), p. 34, pl. 3:3-5, pl. 5:1-3. MILLER, 1979, p. 27, pl. 1:3-8, pl. 2:3-8, pl. 3:2-8, pl. 4:1-3, 6-16, pl. 5:1-3, 6-8, pl. 6:9-16. MILLER ET AL., 1982, p. 124-127, pl. 1:5-8, pl. 2:3-8,

pl. 3: 3-8, pl. 4:1-6, pl. 5:4-8, 14, pl. 6:1-5. VILKS ET AL., 1982, pl. 1:18. KNUDSEN, 1982, p. 170, fig. 14:12:6-8, fig. 14:12:1-4.

Elphidium excavatum (Terquem) forma alba. FEYLING-HANSSSEN, 1972, p. 340, pl. 3:1-9. FEYLING-HANSSSEN, 1976c, pl. 4:4. VILKS ET AL., 1982, pl. 1:16-17.

Cribrorhynchium sp. cf. C. clavatum (Cushman). LANKFORD and PHELEGER, 1973, p. 117, pl. 3:24.

Cribrorhynchium excavatum clavatum Cushman. SCOTT, 1977, p. 169, pl. 6:2.

Elphidium clavatum clavatum Cushman. WILKINSON, 1979, p. 634, pl. 1:1.

Elphidium clavatum subclavatum Gudina. WILKINSON, 1979, p. 639, pl. 1:2.

Elphidium clavatum terminatum. WILKINSON, 1979, p. 639, pl. 1:7, pl. 2:2-4.

Elphidium clavatum lobatum. WILKINSON, 1979, p. 637, pl. 1:3, pl. 2:7.

Elphidium excavatum (Terquem) forma clavatum Cushman. POAG ET AL., 1980, pl. 1:12.

Cribrononion excavatum (Terquem) forma clavatum Cushman. SCOTT and MEDIOLI, 1980, p. 35, pl. 5:5. SCOTT ET AL., 1980, p. 228.

Elphidium excavatum forma lidoensis Cushman. SLOAN, 1981, p. 277, pl. 1:6.

Elphidium ex gr. excavatum (Terquem). E. clavatum Cushman. HAYNES, 1981, p. 61-62, pl. 8:8.

Types:

Holotype: Elphidium incertum (Williamson) variety clavatum. Cushman, 1930, p. 20, pl. 7:10a-b.

Deposited in: Cushman Collection, Smithsonian Institution, USNM No. 10403.

Hypotypes: Elphidium clavatum Cushman. Loeblich and Tappan, 1953, p. 98, 101-102, pl. 19:8-10.

Deposited in: Smithsonian Institution, USNM Nos. P2024a-b, P2025.

Representative plesiotype: Miller, 1979, pl. 1:4, pl. 2:4; Miller et al., 1982, pl. 1:5, pl. 2:4, pl. 3:4; USNM No. 36-312510

Diagnostic characteristics: Generally: small, disc-shaped, orange-brown translucent form; often with an umbilical boss, and always with an imperforate (complete or incomplete) collar surrounding the umbilicus. The sutures are generally backwards curved, with few narrow often incomplete, ponticuli. It is interesting to note that Cushman named this form clavatum (clavical, from collar? from imperforate collar?).

Description: As described by: Loeblich and Tappan (E. clavatum) in 1953 and Feyling-Hanssen (E. excavatum forma clavata) in 1972.

Maximum diameters: 0.23 -0.77 mm (Loeblich and Tappan, 1953, p. 98); 0.17 - 0.48 mm (Feyling-Hanssen, 1964, p. 187; 1972, p. 345); 0.34 - 0.84 mm (Bartlett, 1965b, p. 16)

Remarks: The name Elphidium excavatum forma clavata is retained for the small disc-shaped, orange-brown translucent form figured by Cushman (1930, p.20, Pl. 7:10). However, Cushman's description may cover not only forma clavata, but forma selseyensis and forma magna as well.

Haake (1962) was the first to consider E. clavatum a junior synonym of E. selseyense.

Brodniewicz (1965) pointed out first that Cushman may have been describing two forms. Cushman's collection at the Smithsonian includes specimens belonging to four formae (clavata, excavata, magna and selseyensis) under the name E. incertum var. clavatum. Forma clavata was elevated to specific rank by Loeblich and Tappan (1953), and first designated an ecophenotype of Elphidium excavatum by Feyling-Hanssen (1972). Four of Wilkinson's (1979) subspecies are considered junior synonyms. Haynes (1981) included it in his E. excavatum "polytypic species" group.

Distribution: Elphidium excavatum forma clavata is the central member of this E. excavatum group; it is reported in recent and Holocene sediments from estuaries, nearshore zones, bays, and continental margins from the high arctic to tropical environments. It is perhaps best known for its occurrence (where it is often dominant) in Weichselian (Wurm-Wisconsin) pro-glacial, interstadial and late glacial deposits (Feyling-Hanssen 1972). However, it did not first appear in the Weichselian; it has been recorded as far back as the

earliest marine Pleistocene on both sides of the Atlantic (e.g. Snyder and Katrosh 1979, van Voorthuysen 1949, 1951)

Elphidium excavatum (Terquem) forma gunteri
Cole, 1931

pl. 11:17-18; pl. 12:16-18; pl. 13:1-2; pl. 14:1-3;
pl. 15:8-15; pl. 17:13-15; pl. 18:1-3; pl. 24:1-24.

?Polystomella oceanensis. FORNASINI, 1904, p. 13, pl. 3:10 (ab
d'Orbigny, 1826, p. 285; described but not illustrated or figured).
Elphidium gunteri. COLE, 1931, p. 34, pl. 4:9, 10 (fide Parker et al.,
1953). CUSHMAN, 1939, p. 49-50, pl. 13:10 (figure after Cole).
PARKER ET AL., 1953, p. 8, pl. 3:18-19. PARKEF, 1954, (part), p.
508, pl. 6:16. PHLEGER, 1954, p. 639, pl. 2:3-4. BANDY, 1956,
(part), p. 194, pl. 30:19a-b. LEHMANN, 1957, p. 348, pl. 3:1-3. VAN
VOORTHUYSEN, 1957, p. 32, pl. 23:11a-b. LANKFORD, 1959, p. 2083, pl.
II:7 a-b. PARKER and ATHEARN, 1959, p. 342, pl. 50 :36. PHLEGER,
1960a, p. 277, pl. 3:6,22, pl. 4:12. PHLEGER, 1960b, pl. 7:18; pl.
9:1,17. VAN VOORTHUYSEN, 1960, p. 255. HAAKE, 1962, p. 48, pl.
5:3-4. RICHTER, 1964a, p. 343-353, text-fig. 7. LÉVY, 1966, p. 4,
pl. 1:1a-b. CITA and PREMOLI-SILVA, 1967, p. 35-46, pl. 2:4-5.
BOLTOVSKOY and BOLTOVSKOY, 1968, (part), p. 148, pl. 1:15a-b. LÉVY
ET AL., 1969, p. 94. KNUDSEN, 1971b, p. 277, pl. 12:9, 10; pl.
21:4-7. 1973b, p. 279, pl. 2: 5. ROSSET-MOULINIER, 1976, p. 92,
pl. 1:10, 11; pl. 2:1-4. BUZAS ET AL., 1977, p. 95. KNUDSEN, 1979,
p. 209, pl. 6:4-7. SNYDER and KATROSH, 1979, pl. 2:1-2. BOLTOVSKOY
ET AL., 1980, p. 30, pl. 13:1 5-18. SLOAN, 1981, p. 285, pl. 2:4a-b.

BUZAS and SEVERIN, 1982, p. 37, pl. 8:4.

Elphidium gunteri var. galvestonense. KORNFIELD, 1931, (part), p. 87, pl. 15:2a-3b. CUSHMAN, 1939, p. 60, pl. 16:25 (figure after Kornfeld). PHLEGER, 1951, p. 46. PHLEGER and PARKER, 1951, p. 10, pl. 5:13-14. BANDY, 1954, (part), p. 136, pl. 30:2a-b.

Elphidium oceanense (d'Orbigny). CUSHMAN, 1939, p. 56, pl. 15:8a-b (figures after Fornasini ab d'Orbigny).

Criboelphidium trinitatense. CUSHMAN and BRÖNNIMANN, 1948, (part), p. 20, pl. 4:8.

Elphidium littorale. LE CALVEZ and LE CALVEZ, 1951, p. 251, fig. 5:a-b.

Elphidium guntheri (sic) var. waddensis. VAN VOORTHUYSEN, 1951, p. 25, pl. 2:16a-b.

Elphidium rugulosum Cushman and Wickenden. BANDY, 1956, p. 194, p. 30:21a, b.

Elphidium trinitatense (Cushman and Brönnimann). TODD and BRÖNNIMANN, 1957, p. 39, pl. 7:12.

Elphidium tumidum Natland. PHLEGER and EWING, 1962, p. 178, pl. 4:18.

Elphidium oceanensis (d'Orbigny). MURRAY, 1968, p. 83-96, pl. 1:10a-b. MURRAY, 1971, (part), p. 165, pl. 69:3-7. BOLTOVSKOY and VIDARTE, 1977, p. 38, pl. 3:1. MURRAY, 1979, p. 52, fig. 16:A-B. APHORPE, 1980, pl. 28:1-2.

Elphidium cf. reticulosum Cushman. MATOBA, 1970, p. 52, pl. 6:12a-b.

Elphidium sagra (d'Orbigny). TODD and LOW, 1971, p. C16, pl. 3:7a-b. SYNDER and KATROSH, 1979, pl. 2:3.

Criboelphidium gunteri (Cole). ROSSET-MOULINIER, 1972, p. 178, pl.

18:1-5.

Elphidium waddense van Voorthuysen. HAYNES, 1973, p. 206-207, pl.

24:4,10, pl. 26:1, pl. 28:10-11. CULVER and BANNER, 1978, p. 53-72.

Elphidium guntheri (sic) Cole. HANSEN and LYKKE-ANDERSEN, 1976, p.

12, pl. 8:10-12, pl. 9:1-3.

Elphidium vadescens (Cushman and Brönnimann). HANSEN and

LYKKE-ANDERSEN, 1976, p. 12, pl. 7:12, pl. 8:1-9.

Elphidium gunteri Cole forma salsum. POAG, 1978, (part), p. 402, pl.

2:11-12.

Elphidium gunteri Cole forma typicum. POAG, 1978, p. 402, pl.

2:13-16. POAG, 1981, p. 61, pl. 37:1, pl. 38:1a-1b.

Cribrononion gunteri (Cole). PRIME, 1980, p. 30, pl. 1:13.

Cribrononion granulorum (d'Orbigny). ALBANI and SERANDREI BARBERO,

1982, p. 238, pl. 1:3.

Types:

Holotype: Elphidium gunteri. Cole, 1931, p. 34, pl. 4:9-10.

Deposited in: Florida State Geological Museum, S-2103.

Hypotypes: designated by Parker et al., 1953, p. 8, as: Elphidium

gunteri variety galvestonense Kornfeld, 1931, (part), p. 87, pl.

15:2a-3b.

Deposited in: Stanford University Paleontological Type Collection,
types no. 689 and 692 (microspheric form).

Types of junior synonyms:

?Polystomella oceanensis:

Original designation: Fornasini, 1904, p. 13, pl. 3:10 (ab d'Orbigny, 1826, p. 285; described but not illustrated or figured).

Deposited in: ? Laboratoire de Paleontologie du Museum de l'Histoire Naturelle, Paris. (no designated type specimen, Y. Le Calvez, pers. comm. to Hansen and Lykke-Andersen, 1976).

Criboelphidium trinitatense:

Holotype: Cushman and Brönnimann, 1948, p. 20, pl. 4:8, USNM No. 56646.

Paratype: USNM No. 56748.

Deposited in: Cushman Collection, Smithsonian Institution.

Elphidium guntheri (sic) var. waddensis:

Holotype: van Voorthuysen, 1951, p. 25, pl. 2:16a-b.

Deposited in: Netherlands Geological Survey, Haarlem, no. F. 1810.

Elphidium littorale.

Holotype: Le Calvez and Le Calvez, 1951, p. 251, fig. 5a-b.

Deposited in: not given.

Representative plesiotype: pl. 15:9, pl. 24:1-3.

Diagnostic characteristics: A small to medium sized Elphidium, rather rotund, with a coarsely perforate wall. The sutures are straight, not depressed, and marked by many regular, raised, rectangular shaped ponticuli, often longer than the chambers are wide. The umbilicus contains papillae/irregular bosses (irregular lateral extensions of

the ponticuli and chambers).

Description: As described by Cole in 1931 (E. gunteri).

Maximum diameters: diameter 0.44 mm, thickness 0.26 mm (Cole, 1931, p. 34); 0.35 - 0.43 mm (Kornfeld, 1931, p. 87, microspheric form); diameter 0.39 mm, thickness 0.18 mm (van Voorthuysen, 1951, p. 25).

Remarks: E. excavatum forma gunteri is the name retained for the round, thick, coarsely perforate form described from the Pliocene of Florida by Cole (1931). The type locality was later determined to be Pleistocene in age (Poag 1978). Included in this morphotype is the microspheric form of E. gunteri var. galvestonense described by Kornfeld (1931). The holotype of E. gunteri cannot be found at the moment and Buzas and Culver (1981 pers. comm.) suggest that one of Kornfeld's specimens (#692, designated as a hypotype of E. gunteri by Parker et al. 1953) become the neotype of E. gunteri.

Feyling-Hanssen (1972, p. 344) remarked on the similarity between E. gunteri and E. excavatum forma lidoensis; but he pointed out that they could be separated on the basis of the distinct rectangular and numerous sutural bridges (gunteri) and the broadening of the sutures towards the umbilicus (lidoensis).

Poag (1978) splits E. gunteri into two ecophenotypes, forma salsum and forma typicum. Those he assigns to forma typicum are specimens with ponticuli having length equal to or greater than the chamber width (perforate wall width between sutures). There is some

confusion here because, of Poag's photos (1978, Pl. 2): he places figures 1 - 12 in forma salsum (= forma lidoensis of this author) and figures 13 - 16 in forma typicum, (=forma gunteri of this author) and yet only figures 15 and 16 show ponticuli greater in length than the adjacent chamber wall. Instead, here, the two morphotypes are split on the basis of the sutures broadening towards the umbilicus (= forma lidoensis) and ponticuli spanning the suture completely (= forma gunteri). Consequently, Poag's E. gunteri forma salsum (Pl. 2:1-10) is placed in forma lidoensis; his forma salsum (Pl. 2:11-12) and his forma typicum are placed in forma gunteri.

It is probable that P. oceanensis, described by d'Orbigny (1826) is the same morphotype as E. excavatum forma gunteri. However, the validity of both the name P. oceanensis and the species itself is in doubt. (When transferred to the masculine genus Elphidium the epithet should have become oceanense, but many workers retained the feminine epithet oceanensis).

D'Orbigny (1826) described, he never figured or illustrated, the species. Hansen and Lykke-Andersen (1976) on a personal communication from Y. Le Calvez, state that there is no (and probably never was a) holotype of the species. Both Hansen and Lykke-Andersen (1976, p. 12), and Poag (1978, p. 402) consider E. oceanensis to be a nomen nudum. However, E. oceanensis (from d'Orbigny's collection) was figured by Fornasini (1904, Pl. 3:10), so that the authorship of the species is "Fornasini 1904 ab d'Orbigny 1826". Fornasini's figure does not show the rounded periphery or coarse perforations of the

gunteri morphotype and could be a figure of either forma selseyensis or forma tumidum.

Murray (1971, 1979) has illustrated specimens he has identified as E. oceanensis (some that would be included here in forma lidoensis); Murray (1982 pers. comm.) does not think that E. oceanensis is conspecific with E. excavatum because besides morphological differences, it has a different environmental preferences.

Haynes (1982 pers. comm.) has also remarked that the type of E. oceanensis is unrecognizable based on Fornasini's figure - and that a new figure is required. Haynes suggests (1982 pers. comm.) that the name gunteri could be retained on the basis of usage.

Junior synonyms of this morphotype also include E. trinitatense (Cushman and Brönnimann), and a variety of E. gunteri, E. gunteri var. waddensis described by van Voorthuysen (1951) and later put in synonymy with E. gunteri by the same author (van Voorthuysen 1957). Haynes (1973) has elevated van Voorthuysen's variety to specific rank and illustrates specimens of his E. waddense (1973, p. 206-207, Pl. 24:4, 10, Pl. 26:1, Pl. 28:10-11) that resemble Murray's specimens of E. oceanensis (1971, Pl. 69:1-7); Murray's specimens are considered here to be intermediate between forma lidoensis and forma gunteri but based on the broadening sutures and indistinct ponticuli they are placed in forma lidoensis.

Also considered a junior synonym is Elphidium littorale Le Calvez and Le Calvez (1951).

Distribution: A widely distributed form, extensively documented in the literature. A warm to tropical water nearshore form tolerating hypo- to hyper-salinities. Found on both the Atlantic and Mediterranean coasts of Europe along beaches and in lagoons, as well as open bays. Also found throughout the Gulf of Mexico, and along the California coast. Reported from late Pleistocene shallow, brackish, glacio-fluvial deposits on both sides of the Atlantic (Knudsen 1973a, 1973b, 1979, Snyder and Katrosh 1979).

Elphidium excavatum (Terquem) forma galvestonensis
Kornfeld, 1931

?pl. 11:19-21; pl. 13:17-20; pl. 14:19-20; pl. 16:1-8;
?pl. 18:18; pl. 25:1-16

Elphidium discoidale (d'Orbigny). CUSHMAN, 1930, p. 22, pl. 8:9a-b. CUSHMAN and COLE, (1930), p. 97, pl. 13:10a-b. PHLEGER, 1951, p. 46. PHLEGER and PARKER, 1951, (part), p. 10, pl. 5:11. TODD and BRÖNNIMANN, 1957, (part), p. 39, pl. 6:8.

Elphidium gunteri Cole var. galvestonense. KORNFELD, 1931, (part), p. 87, pl. 15:1a-b (not 2a-3b).

Elphidium galvestonense Kornfeld. PARKER ET AL., 1953, p. 8, pl. 3:15-16. PHLEGER, 1954, p. 639, pl. 2:1-2. LEHMANN, 1957, p. 348, pl. 2:37-40. PARKER and ATHEARN, 1959, p. 342, pl. 50:33-35. PHLEGER, 1960a, p. 277, pl. 3:19

Elphidium gunteri Cole. PARKER, 1954, (part), p. 508, pl. 6:16. BANDY, 1956, (part), p. 194, pl. 30:19a-b.

Elphidium excavatum (Terquem). TODD and BRÖNNIMANN, 1957, (part), p.

39, pl. 6:11.

Elphidium morenoi Bermudez. TODD and LOW, 1971, p. 16, pl. 3:6.

Cellanthus galvestonense (Kornfeld). SCOTT, 1976, p. 170

Elphidium galvestonense Kornfeld forma typicum. POAG, 1978, p. 404,
pl. 3: 12-16, 19-21. POAG, 1981, p. 60, pl. 35:3, pl. 36:3a-3b.

Elphidium ex gr. excavatum (Terquem). Elphidium cf. advenum sensu
Todd and Low. HAYNES, 1981, p. 61-62, pl. 8:12.

Types :

Holotype: Elphidium gunteri var. galvestonense. Kornfeld, 1931,
(part), p. 87, pl. 15:1a-b (not 2a-3b).

Lectotype: designated by Parker et al., 1953, p. 8, as: Elphidium
gunteri var. galvestonense, Kornfeld, 1931, p. 87, pl. 15:1a-b.

Deposited in: Stanford University Paleontological Type Collection,
Type No. 691 (megalospheric form).

Representative plesiotype: pl. 16:3, pl.25:1-3.

Diagnostic characteristics: A large, many chambered (13-18) form with
a large very raised boss (or bosses) at the umbilicus and many
regular, distinct ponticuli. There may be a ring of papillae
surrounding the boss or in the sutures. The wall is heavily calcified
and very finely perforate, giving the test a porcelaneous appearance.
The periphery is subacute.

Description: As described by: Kornfeld in 1931 (E. gunteri var.

galvestonense megalospheric form); Parker et al. in 1953 (E. galvestonense) and Poag in 1978 (E. galvestonense forma typicum).

Maximum diameters: diameter 0.48 to 0.57 mm, thickness 0.28 mm (Kornfeld, 1931, p. 87, megalospheric form).

Remarks: E. excavatum forma galvestonensis is the morphotype originally described by Kornfeld (1931) as the megalospheric form of E. gunteri var. galvestonense. One of Kornfeld's specimens (1931, p. 87, Pl. 15:1a-b, No. 691) has been designated as lectotype by Parker et al. (1953). Poag (1978) has split E. galvestonense into two formae, forma typicum and forma mexicanum. Here, only forma typicum is included in this morphotype. Poag's other forma, forma mexicanum appears to be Kornfeld's E. incertum var. mexicanum a species whose relationship to E. excavatum is uncertain.

Distribution: E. excavatum forma galvestonensis occurs in low frequencies and is most abundant in the Northumberland Strait core sample and the Gulf of Mexico samples. It was not positively identified in any of the European samples, and no occurrences from Europe are documented in the literature. It has not been reported as older than Holocene. It is a nearshore (bay, beach and marsh) form and prefers waters that are warm and saline and that exhibit little fluctuation in these parameters.

Elphidium excavatum (Terquem) forma lidoensis
Cushman, 1936

pl. 1:2:1; pl. 4:12; pl. 5:8-12; pl. 6:7; pl. 7:10-12;
pl. 8:9; pl. 9:17; pl. 10:15-16; pl. 11:9-13; pl.
12:4-5, 7-10; 13; pl. 13:11; pl. 14:10; pl. 15:1-7,
9-14; pl. 18:4-7; pl. 19:1-7; pl. 26:1-32

?Polystomella arctica (Parker and Jones). TERQUEM, 1876, p. 428, pl.
2:1 (fide Feyling-Hanssen 1972).

Elphidium florentinae. SHUPACK, p. 1934, p. 9, opp. p.9:5a-b.

Elphidium lidoensis. CUSHMAN, 1936, p. 86, pl. 15:6. CUSHMAN, 1939,
p. 62-63, pl. 17:17. ACCORDI and SOCIN, 1950, p. 12, 15, pl. 1: 8.
CITA and PREMOLI-SILVA, 1967, p. 35-36, pl. 2:1-2. LÉVY, 1966, p. 5,
pl. 1:5a-c.

Criboelphidium salsum. Cushman and Brönnimann, 1948, p. 19, pl. 4:6.
TODD and BRÖNNIMANN, 1957, p. 39.

Criboelphidium vadescens. Cushman and Brönnimann, 1948, p. 18, pl.
4:5. LEHMANN, 1957, p. 348, pl. 2:28-29.

Elphidium clavatum Cushman. WEISS, 1954, (part), p. 159, pl. 32:12.
MURRAY, 1969, (part), p. 416, pl. 17:1-4. SCHNITKER, 1971, (part), p.
198, pl. 7:5a-b .

Elphidium gunteri Cole var. galvestonense Kornfeld. BANDY, 1954,
(part), p. 136, pl. 30:2.

Elphidium tumidum Natland. TODD and BRÖNNIMANN, 1957, (part), p. 39,
pl. 7:9.

Elphidium oceanicum Cushman. LEHMANN, 1957, p. 348, pl. 3:9-12.

Elphidium sp. A. VAN VOORTHUYSEN, 1957, p. 31, pl. 23:10.

Elphidium cf. E. minimum (Segwenza). PARKER, 1958, p. 271, pl. 4:8-9.

Elphidium granulorum (d'Orbigny). PARKER, 1958, p. 270, pl. 4:10-11.

Elphidium incertum (Williamson) var. PHLEGER, 1964, p. 383, pl. 2:2.

Elphidium kozlowskii. BRODNIEWICZ, 1965, p. 205, text-fig. 29, pl. 7:4, pl. 9:1-6.

Nonion depressulus (Walker and Jacob). MURRAY, 1965b, p. 148-149, pl. 25:6-7; pl. 26:7-8.

Elphidium excavatum (Terquem). CITA and PREMOLI-SILVA, 1967, p. 35-36, pl. 2:3. ROSSET-MOULINIER, 1976, (part), p. 89-92, pl. 1:5-9, 12. MURRAY, 1979, p. 50, figs. 15:C-D. BUZAS and SEVERIN, 1982, p. 37, pl. 8:2.

Cribrononion excavatum (Terquem). HAAKE, 1967, (part), p. 13-27, pl. 1:1-3, 9, 12-14. VON DANIELS, 1970, p. 87, pl. 7:11.

Cribrononion lidoense (Cushman). LÉVY ET AL., 1969, p. 94, pl. 1:9a-b.

Elphidium oceanensis (d'Orbigny). MURRAY, 1971, (part), p. 165, pl. 69:1-2.

Criboelphidium excavatum (Terquem). ROSSET-MOULINIER, 1972, p. 177, pl. 16:1-4, pl. 17:1-4.

Elphidium excavatum (Terquem) forma lidoensis Cushman.

FEYLING-HANSEN, 1972, p. 344, pl. 6:1-7. SNYDER and KATROSH, 1979, p. 254, pl. 2:4. MILLER ET AL., 1982, p. 134-136, pl. 1:17-20, pl. 4:7-12, pl. 5:9, pl. 6:15-16.

Elphidium selseyense (Heron-Allen and Earland). HAYNES, 1973, (part), p. 204-205, pl. 22:1-2, pl. 24:11, pl. 26:5, 10.

Criboelphidium excavatum clavatum (Cushman). SCOTT ET AL., 1977, p. 1579, pl. 5:2.

Elphidium gunteri Cole forma salsum. POAG, 1978, (part), p. 402, pl.

2:1-10. POAG, 1981, p. 61, pl. 37:2, pl. 38:2a-2b.

Elphidium clavatum lidoense Cushman. WILKINSON, 1979, p. 637, pl. 1:4.

Elphidium excavatum var. A. SLOAN, 1981, p. 278, pl. 2:2a-b.

Elphidium incertum var. A. SLOAN, 1981, p. 282, pl. 2:3a-b.

Types:

Holotype: Elphidium lidoense Cushman, 1936, p. 86, pl. 15:6a-b, USNM No. 23201.

Paratypes: USNM Nos. 19176, 23020, 39935.

Deposited in: Cushman Collection, Smithsonian Institution.

Types of junior synonyms:

Elphidium florentinae:

Holotype: Shupack, 1934, p. 9, opp. p. 9: 5a-b.

Deposited in: American Museum of Natural History (New York) No. 696

Cribroelphidium salsum:

Holotype: Cushman and Brönnimann, 1948, p. 19, pl. 4:6, USNM No. 56644.

Paratype: USNM No. 56750.

Deposited in: Cushman Collection, Smithsonian Institution.

Cribroelphidium vadeszens:

Holotype: Cushman and Brönnimann, 1948, p. 18, pl. 4:5, USNM No. 56643.

Paratype: USNM No. 56649.

Deposited in: Cushman Collection, Smithsonian Institution.

Elphidium kozlowskii:

Holotype: Brodniewicz, 1965, p. 205, text-fig. 29, pl. 7:4, pl. 9:1-6. (pl.9:5).

Deposited in: Paleozoological Institute of the Polish Academy of Sciences (Warsaw) No. F. VIII / 89-94.

Representative plesiotypes: "boreal" water form, pl. 7:11, USNM No. 312513; "Lusitanian" water form, pl. 25:14-16.

Diagnostic characteristics: Test compressed, sutures backwards curved, distinctly broadening towards the umbilicus, filled with papillae; ponticuli often absent (or not visible); umbilicus open and large, filled with papillae, or irregular bosses, or both.

Description: As described by: Cushman in 1936 (E. lidoense) and by Feyling-Hanssen in 1972 (E. excavatum forma lidoensis).

Maximum Diameters: 0.50 - 0.60 mm (Cushman, 1936, p. 86); 0.20 - 0.40 mm (Brodniewicz, 1965, p. 205).

Remarks: The name E. excavatum forma lidoensis is the name retained for the form described by Cushman (E. lidoense, 1936) and Feyling-Hanssen, (E. excavatum forma lidoensis, 1972). Within this

forma, two "subforma" are observed: a "boreal" environment form from areas of extremes in climatic variation, i.e. Miramichi Estuary, Annapolis Basin, and Long Island Sound; and a "Lusitanian" environment form, from areas with a narrower climatic range, i.e. San Diego Bay, San Antonio Bay, Venice, and Bay of Izmir. Both forms exhibit the key characteristics of the forma. However, the "boreal" form can be linked to E. excavatum forma excavata; the wall perforations are fine and the papillae small. The "Lusitanian" form resembles, and can be linked to, E. excavatum forma gunteri; the periphery is rounded, wall perforations coarse, papillae more variable in size, and a larger number of bosses present in the umbilicus.

Feyling-Hanssen (1972, p. 344) remarked on the similarity of E. gunteri and E. lidoense; but he pointed out that they could be separated on the basis of the distinct rectangular and numerous sutural bridges (gunteri) and the broadening of the sutures towards the umbilicus (lidoensis).

As mentioned earlier, part of Poag's (1978) E. gunteri forma salsum is included in this form.

The placement of E. oceanense (d'Orbigny) is in doubt, but specimens assigned to this form by some workers (i.e. Murray 1971, [part]) either belong to this form or are intermediate between the two formae gunteri and lidoensis.

Junior synonyms include Elphidium florentinae Shupack (1934), Criboelphidium salsum and C. vadescens (both) Cushman and Brönnimann (1948), and E. kozlowskii Brodniewicz, (1965).

Distribution: E. excavatum forma lidoensis was first collected by Cushman, (E. lidoense, Cushman 1936, 1939) from Venice, Italy; where it was later (again) recorded by Cita and Premoli-Silva (1967) as E. lidoense.

The occurrence of E. excavatum forma lidoensis in European and North American waters is difficult to document; it appears that this form has been identified by other species names.

The occurrences of E. excavatum forma lidoensis recorded in the literature suggest a shallow, subtidal estuarine environment with waters attaining a summer temperature of at least 20°C. It appears to be a marginal marine form; almost all the occurrences are areas under estuarine influence (among them Miramichi Estuary, Chezzetcook Inlet, Annapolis Basin, Long Island Sound, San Antonio Bay, Venice, and Dovey Estuary, [U. K.]) or lagoons and mangrove swamps (southern California and Trinidad).

Elphidium excavatum (Terquem) forma tumidum
Natland, 1938.

pl. 11:1-4, 14-15; pl. 12:6; pl. 27:12-19.

Elphidium tumidum. NATLAND, 1938, p. 144, pl. 5:5-6. CUSHMAN, 1939, p. 65, pl. 20:8a-b. TODD and BRÖNNIMANN, 1957, (part), p. 39, pl. 6:7a-9b. LEHMANN, 1957, p. 348, pl. 3:15-16.

Elphidium sp. cf. E. tumidum Natland. PARKER ET AL., 1953, p. 9, pl. 3:28-29.

Elphidium incertum var. clayatum Cushman. TODD and BRÖNNIMANN, 1957,

(part), p. 39, pl. 6:10a-b.

Elphidium cf. E. tumidum Natland. PHLEGER, 1960a, p. 277, pl. 3:24.

Cellanthus tumidum (Natland). SCOTT, 1976, p. 170. SCOTT ET AL.,
1976, p. 74.

Types:

Holotype: Elphidium tumidum, Natland, 1938, p. 144, pl. 5:5-6.

Deposited in: Smithsonian Institution, USNM No. 22550.

Representative plesiotype: pl.11:16, pl. 27:15.

Diagnostic characteristics: A large, ornamented form resembling forma selsevensis. However, the ornamentation and ponticuli are much more regular on forma tumidum. The umbilicus is large, circular, depressed and filled with papillae/bosses. The chamber extensions into the umbilicus are truncated sharply. The periphery is broadly rounded, and the chambers inflated.

Description: As described by Natland (E. tumidum, 1938).

Maximum diameters: diameter 0.50 mm, thickness 0.22 mm (Natland, 1938, p. 144).

Remarks: The name E. excavatum forma tumidum is given to the morphotype described by Natland (E. tumidum, 1938). Natland (1938) reported it to be distinct from, but related to E. articulatum. Scott (1982 pers. comm.) has observed intergradation between this form and

other morphotypes of E. excavatum in California lagoons and marshes, though intergradation is not fully documented here, and this form was not found at any other locations.

Distribution: Natland (1938) reported this form to be common off southern California; but there have been relatively few occurrences documented in the literature under this name. No evidence was found that this form has been widely documented under another name. From the literature, it can be concluded that this form is restricted to the west coast of the United States and the Gulf of Mexico.

Elphidium excavatum (Terquem) forma cuvillieri
Levy, 1966.

pl. 6:20; pl. 13:6-7; pl. 14:6-7; pl. 16:9-14;
pl. 18:8-15; pl. 19:8-15; pl. 28:1-23.

?Polystomella poeyana. D'ORBIGNY, 1839a, p. 55, pl. 6:25-26.

Elphidium poeyanum (d'Orbigny). CUSHMAN, 1930, p. 25, pl. 10:4-5.

PARKER ET AL., 1953, p. 9, pl. 3:26. PARKER, 1954, p. 509, pl. 6:17.

BANDY, 1954, (part), p. 136, pl. 30:6a-b. PHLEGER, 1954, p. 639, pl.

2:8-9. TODD and BRÖNNIMANN, 1957, p. 39, pl. 7:2-4. PHLEGER, 1960a,

p. 277, pl. 3:17, pl. 5:10. TODD and LOW, 1961, p. 20, pl. 2:7. TODD

and LOW, 1971, p.C16, pl.3:8. HANSEN and LYKKE-ANDERSEN, 1976, p. 13,

pl. 9:9-12; pl. 10:1-5. POAG, 1981, p. 63, pl. 39:3, pl.40:3a-3b.

Elphidium cf. articulatum (d'Orbigny). CUSHMAN, 1944, (part), p. 26,

pl. 3:41.

Criboelphidium kugleri. CUSHMAN and BRÖNNIMANN, 1948, p. 18-19, pl.

4:4a-b. TODD and BRÖNNIMANN, 1957, p. 39. LEHMANN, 1957, p. 348, pl.

2:25-27.

Elphidiononion poevanum (d'Orbigny). HOFKER, 1951, p. 356 (fide
Loeblich and Tappan, 1964, p. C635).

Cribroelphidium poevana (d'Orbigny). LOEBLICH and TAPPAN, 1964, p.
C635, fig. 508:3a-4b.

Elphidium cuvillieri. LEVY, 1966, p. 5-6, pl. 1:6a-c, pl. 2.
ROSSET-MOULINIER, 1972, p. 177, pl.15:1-4. HAYNES, 1973, p. 197, pl.
24:17-18, pl. 26:12. ROSSET-MOULINIER, 1976, p. 93, pl. 3:4-8.
MURRAY, 1979, p. 50, fig. 14:E-F.

Cribrononion cuvillieri (Levy). LEVY ET AL., 1969, p. 93, pl.
1:10a-11.

Cribrononion translucens (Natland). VON DANIELS, 1970, p. 88, pl.
7:13. ALBANI and SERANDREI BARBERO, 1982, (part), p. 240, pl.1:7-9
(not 10).

Elphidium translucens (Natland). HANSEN and LYKKE-ANDERSEN, 1976, p.
11, pl. 7:1-11.

Elphidium kugleri (Cushman and Brönnimann). BUZAS ET AL., 1977, p.
95. BUZAS and SEVERIN, 1982, p. 37, pl. 8:5.

Elphidium discoidale (d'Orbigny) forma translucens. POAG, 1981, p.
59, pl. 35:2, pl. 36:2a-2b.

Elphidium discoidale (d'Orbigny) forma typicum. POAG, 1981, p. 59,
pl. 35:1, pl. 36:1a-1b.

Types:

Holotype: Elphidium cuvillieri, Lévy, 1966, p. 5-6, pl. 1:6a-c, pl. 2.

Deposited in: not given

Types of junior synonyms:?Polystomella poeyana:

?Holotype: d'Orbigny, 1839a, p. 55, pl. 6:25-26.

Lectotype: designated by Loeblich and Tappan, 1964, p. C635, Fig. 508:3a-b

Deposited in: Laboratoire de Paleontologie du Museum de l'Histoire Naturelle; Paris.

Cribrorhynchium kugleri:

Holotype: Cushman and Brönnimann, 1948, p. 18-19, p. 4:4a-b.

Deposited in: Cushman Collection, Smithsonian Institution, USNM No. 56642.

Paratype: USNM No. 56746 (as above).

Representative plesiotype: pl. 26:8.

Diagnostic characteristics: A smooth, round, disc shaped Elphidium about the same size as forma clavata. The peripheral outline can range from smooth to very lobate. The sutures are straight or gently backwards curved, and characterized by very regular rows of sutural pores. Papillae are completely absent in this form; the umbilicus is slightly depressed (perforate or imperforate) and closed by a glassy plate of fused chamber ends.

Description: As described by: Lévy in 1966 (E. cuvillieri), Lévy et al. in 1969 (Cribrononion cuvillieri) and Haynes in 1973 (E. cuvillieri).

Maximum diameters: 0.5 mm (Lévy, 1966, p. 6); 0.36 - 0.47 mm (Rosset-Moulinier, 1976, p. 93).

Remarks: The name E. excavatum forma cuvillieri is the name retained for the species described by Lévy (1966) as E. cuvillieri. He reported this form to be a common shallow water form in the Mediterranean. This leads to the question as to why it had not been previously recognized or described. One possible explanation is that has been included by other authors with E. poeyana (d'Orbigny). E. poeyana was described by d'Orbigny from Cuba (1839a).

Williamson (1858) remarked on the similarity between his P. umbilicatula (= E. excavatum forma williamsoni) and d'Orbigny's P. poeyana, stating the only difference was in the alleged arrangement of septal apertures.

Loeblich and Tappan 1964, designated one of d'Orbigny's specimens as lectotype (Loeblich and Tappan, 1964, p. C365, Figure 508:3a-4b). The material at the Smithsonian Institution identified by Loeblich and Tappan is considered very similar and possibly conspecific with the morphotype cuvillieri.

This form was probably described by Cushman and his co-workers as Criboelphidium kugleri (Cushman and Brönnimann 1948, Todd and Brönnimann 1957, and Lehmann 1957).

Levy (1966, p. 6) remarks on the similiarity between his species (E. cuvillieri) and d'Orbigny's. Lévy (1966) did not compare his specimens directly with d'Orbigny's, but with specimens from the

"Stampien" supplied by Poignant which are considered equivalent with topotypes from Cuba (Cuvillieri and Szakall 1949). Lévy (1966) also remarked on the similarity of his species to E. lidoense Cushman.

The name cuvillieri is retained until d'Orbigny's material can be examined. The name cuvillieri is an unambiguous designation for the form described by Lévy; the uncertainty lies in its relationship to d'Orbigny's species (E. poeyana).

Distribution: The distribution of this form is difficult to document because of the relatively few (and those quite recent) references to E. cuvillieri. It is a warm shallow water form preferring near normal salinities. It has probably been reported from both sides of the Atlantic as E. poeyana and from North America as E. (or C.) kugleri; from warm, shallow, near normal marine environments.

Elphidium excavatum (Terquem) forma magna
Miller, Scott and Mediolini, 1982.

pl. 1:1-2; pl. 4:2-5; 15-20; pl. 6:1-2; pl. 7:2-3;
pl. 14:15-17; pl. 27:1-11.

Elphidium discoidale (d'Orbigny). CUSHMAN, 1939, p. 54, pl. 15:7.
PHLEGER and PARKER, 1951, (part), p. 10, pl. 5:10. PARKER ET AL.,
1953, p. 7, pl. 13:13-14. LEHMANN, 1957, p. 348, pl. 2:18-20.

Elphidium incertum (Williamson) var. clavatum Cushman. CUSHMAN, 1948
(part), p. 57, pl. 6:8. PARKER, 1952a, (part), p. 412, pl. 5:11.

Elphidium clavatum Cushman. LOEBLICH and TAPPAN, 1953, (part), p. 98.
TODD and LOW, 1961, (part), p. 18-19, pl. 2:1. TODD and LOW, 1967,
(part), p. A33, pl. 4:16-17. "Complex". GREGORY, 1970, (part), p.
226, pl. 14. RODRIGUES and HOOPER, 1982, (part), p. 411-416,
text-fig. 2:A-F.

Cribrorhynchium incertum (Cushman, [not Williamson]). SCOTT, 1977, p.
170, pl. 6:4-5.

Elphidium excavatum (Terquem) forma clavata Cushman. KNUDSEN, 1979,
(part), p. 208-209, pl. 4:4-5.

Cribrononion excavatum incertum (Cushman, [not Williamson]). SCOTT
ET AL., 1980, p. 228, pl. 4:4-5.

Elphidium galvestonense (Kornfeld). BOLTOVSKOY ET AL., 1980, p. 29,
pl. 13:12.

Elphidium excavatum (Terquem) forma magna. Miller, Scott and Mediolini,
1982, p. 138-139, pl. 1:4-5, pl. 5:1-3.

Primary representative plesiotype: pl. 8:1; USNM No. 312508

Secondary representative plesiotypes: pl. 7:2, pl. 8:2, pl. 8:3;
USNM No. 312509.

Stratigraphic Age: recent

Type Locality: Chezzetcook Inlet, N. S., Canada; Station 53a (Scott 1977, Scott and Mediolli 1980, Scott et al. 1980).

Derivation of Specific Name: magnus; Latin, meaning large.

Diagnostic characteristics: Test often large, peripheral outline smooth to slightly lobate, periphery subacute, and walls greatly convex giving the umbilicus a raised appearance. Umbilicus large, usually filled with one large knobby boss; sutures smooth, backwards curved, with ponticuli and some papillae in the sutures. Some (or all) of the sutures constricted before reaching the umbilicus, forming the imperforate collar around the umbilicus. Differs from forma clavata only in size, shape and environment where found.

Description: Test free, planispiral, involute, biumbonate central boss of clear shell material, usually large, sometimes absent, in a few cases subdivided; walls convex, periphery subacute, peripheral outline smooth to slightly lobate in the latest part of the test, chambers 10-13 (usually 11 or 12) in the final whorl, gradually increasing in size as added. The sutures are depressed, backwards curved, usually closed (but from one to all may remain open) before

reaching the umbilicus, as a result of fusing of the chamber ends; forming a complete (or imcomplete) imperforate collar or ring around the umbilical area. Sutures with a single row of apertural pores; and from few to many short, narrow, distinct ponticuli, often not extending entirely across the sutures. Wall usually thick, calcareous, orange-brown to colourless to white; transparent to opaque with progressive chamber overlap; with radiate structure. Wall distinctly perforate; pores round, septa and apertural face with few pores; tendency towards developing fewer or no pores in the central extensions of the chamber walls; aperture a single row of pores at the base of the apertural face.

The subacute periphery is illustrated on Plate 27:4.

Maximum Diameters: 0.30 mm - 0.60 mm (perhaps larger).

Remarks: This name is given to the forma found in turbulent nearshore zones.

It may be the form identified by Cushman as the large opaque nearshore form he called Elphidium incertum (Williamson). There are specimens of this forma in the Cushman collection, identified by Cushman as E. incertum, E. incertum var. clavatum, and E. discoidale. Feyling-Hanssen (1972) has found opaque specimens of early Holocene age which he has named E. excavatum forma alba; and he placed Cushman's white forms in synonymy with his. However, after examining material supplied by Prof. Feyling-Hanssen; the conclusion was reached that specimens identified as E. excavatum forma alba were etched

specimens of E. excavatum forma clavata and forma excavata. This same conclusion had previously been reached by Scott et al. (1977). The processes and possible cause of etching have been discussed by Murray (1967) and other possible cause have been covered by Walker (1971) and Mageau and Walker (1976).

Distribution: E. excavatum forma magna has been found in recent sediments from nearshore turbulent zones, mainly in the Maritime Provinces of Canada. Gregory (1970, E. clavatum "complex") found it in Bedford Basin, Scott (Criboelphidium incertum, 1977) and Scott et al. (Criboelphidium excavatum incertum, 1980) found it the dominant form in the turbulent zone of Chezzetcook Inlet, Nova Scotia.

It has also been identified in samples from the Annapolis Basin, Nova Scotia, and the Beaufort Sea.

E. excavatum forma magna has been reported as E. clavatum (Loeblich and Tappan 1953, Todd and Low 1961, 1967) or E. incertum (Parker 1952a) by these and probably other authors as well. Bartlett (E. incertum "COMPLEX", 1965b) reported large opaque forms commonly associated with turbulent, nearshore environments or the outer shelf.

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PLATES 1 - 29

PLATE 1

Photographs of five formae of Elphidium excavatum (Terquem).

Photos taken through a low power dissecting microscope (magnifications unknown, approximately 30-50 x).

1:1. E. excavatum forma excavata (Terquem). Note the depressed umbilicus; straight, depressed sutures and slight to markedly lobate peripheral outline. Specimens from (left to right): Bay of Chaleur SRA-52 (2), and Chezzetcook Inlet station 54a₁.

1:2. E. excavatum forma magna Miller, Scott and Medioli. This form is larger with a subacute periphery and raised umbo. Note the large boss on the specimen on the right. Specimens from: Chezzetcook Inlet station 54a₁.

1:3. E. excavatum forma clavata Cushman; the small, flat, disc-shaped form. Specimens from: Labrador core 12, 825-830 cm.

2:1. E. excavatum forma lidoensis Cushman. Note the star shaped pattern of papillae in the umbilicus. Specimens from (left to right): Long Island Sound No. 722, Miramichi Estuary 6a, Bay of Chaleur SRQ-52, and Beaufort Sea F2257.

2:2. E. excavatum forma selsevensis (Heron-Allen and Earland). Pink colour denotes living material stained with Rose Bengal. Note the ornamentation in the large umbilicus. All three specimens from Long Island Sound, No. 722.



PLATES 2-19

These plates illustrate the ten morphotypes of Elphidium excavatum (and the intergradational series) present in the samples studied from the locations listed on Table 1. Magnifications are given in diameters, not in terms of area. Side views of whole specimens. Each specimen is identified by its specimen number (SN).

PLATE 2

1-8. Specimens of E. excavatum forma clavata Cushman, from a Holocene assemblage from the Beaufort Sea. All specimens have an incomplete or complete imperforate collar or umbilical area; though the boss is incomplete or absent in some cases. 1. SN67 x 125. 2. SN68 x 145. 3. SN69 x 149. 4. SN70 x 118. 5. SN71 x 104. 6. SN72 x 183. 7. SN73 x 131. 8. SN74 x 104.

9-20. An intergradational series of the two formae clavata and excavata from a late Pleistocene assemblage from Hirtshals Denmark.

9-17. E. excavatum forma clavata Cushman; the umbilical boss may be complete, incomplete, or absent but the imperforate collar or imperforate umbilical area is always present. 9. SN77 x 82. 10. SN78 x 99. 11. SN79 x 76. 12. SN80 x 127. 13. SN81 x 100. 14. SN82 x 97. 15. SN83 x 132. 16. SN84 x 97. 17. SN85 x 129.

18-20. E. excavatum forma excavata (Terquem). 18. An intermediate form, approaching forma excavata SN86 x 89. 19. SN87 x 123. 20. SN88 x 84.

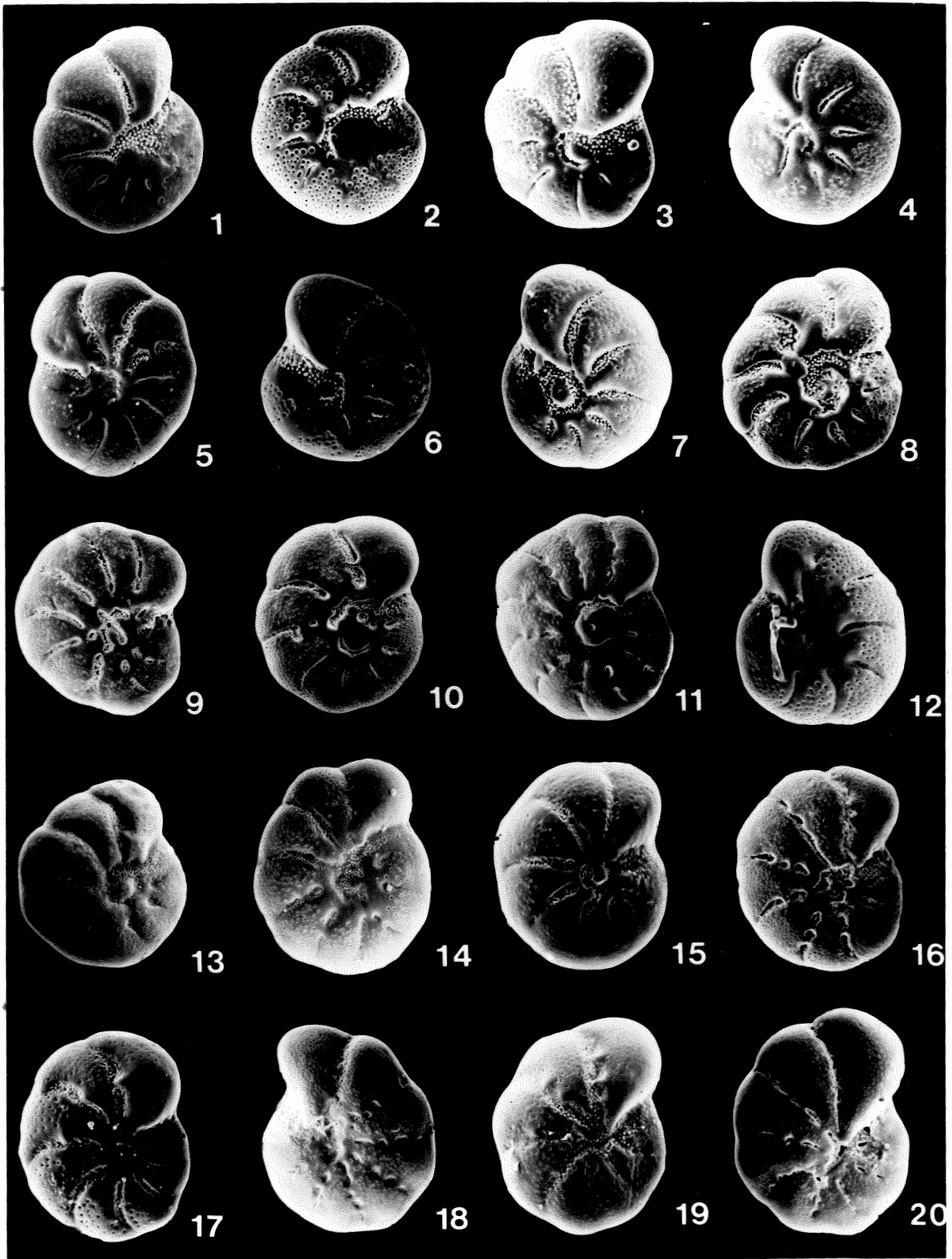


PLATE 3

Elphidium excavatum (Terquem), an intergradational series of two formae clavata and excavata from a Holocene assemblage from the Beaufort Sea.

1-6. Typical specimens of E. excavatum forma clavata Cushman.

1. SN186 x 174. 2. SN187 x 133. 3. SN188 x 166. 4. SN189 x 143. 5. SN190 x 184. 6. SN191 x 230.

7-8. E. excavatum forma excavata (Terquem). 7. SN192 x 104. 8. SN193 x 92.

9-12. E. excavatum forma clavata, ornamented specimens. 9. SN194 x 102. 10. SN195 x 84. 11. SN196 x 94. 12. SN197 x 103.

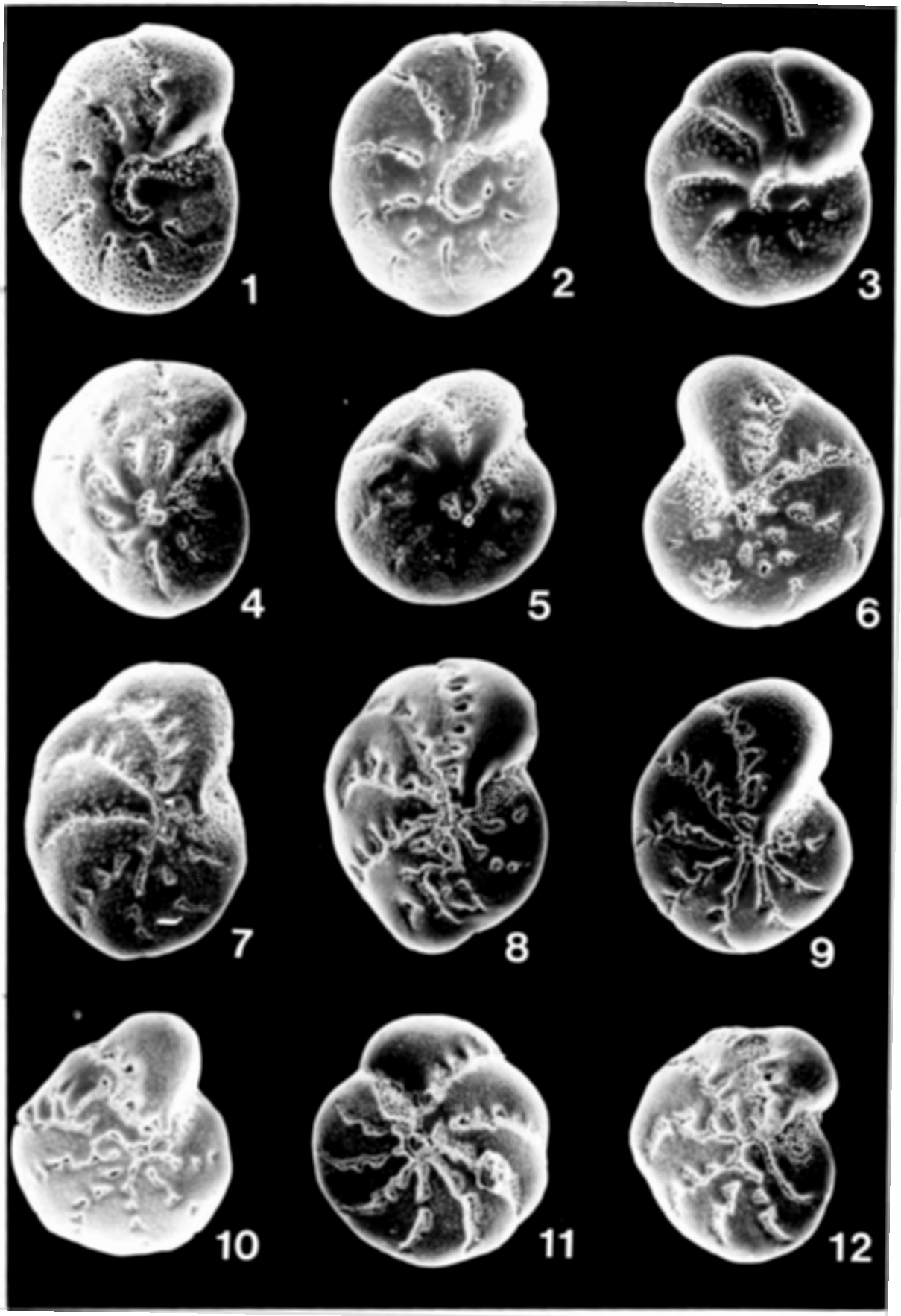


PLATE 4

1-12. Elphidium excavatum (Terquem), an intergradational series from a Pleistocene assemblage from the Champlain Sea. Four formae are present.

1. Elphidium excavatum forma clavata Cushman SN596 x 115.

2-5. Elphidium excavatum forma magna Miller, Scott and Medioli. Recognized by its subacute periphery and the raised umbilical area. If a boss is present it is usually large. 2. SN490 x 147. 3. SN501 x 170. 4. SN483 x 176. 5. SN491 x 129.

6-9. Elphidium excavatum forma clavata Cushman. 6. SN488 x 123. 7. SN497 x 159. 8. SN502 x 145. 9. SN493 x 125.

10-11. Elphidium excavatum forma excavata (Terquem). Two specimens intermediate between formae clavata and excavata. The umbilicus is quite depressed and (for the most part) the sutures open in the umbilicus (as in forma excavata), but the sutures are curved and there is an incomplete imperforate collar present (as in forma clavata). 10. SN495 x 136. 11. SN500 x 144.

12. Elphidium excavatum forma lidoensis Cushman. The sutures are curved, opening towards the umbilicus, and the ponticuli absent or poorly developed, SN489 x 164.

13-20. Elphidium excavatum (Terquem) an intergradational series from a population from the Scotian Shelf (off Liverpool, N.S.) Canada.

13-14. Elphidium excavatum forma clavata Cushman. Two typical (though broken) specimens. 13. SN901 x 95. 14. SN771 x 108.

15-20. Elphidium excavatum forma magna Miller, Scott and Medioli. 17-20. These specimens have the large, raised boss typical

of this forma. 15. SN908 x 61. 16. SN906 x 72. 17. SN905 x 69. 18.
SN903 x 71. 19. SN909 x 66. 20. SN907 x 66.

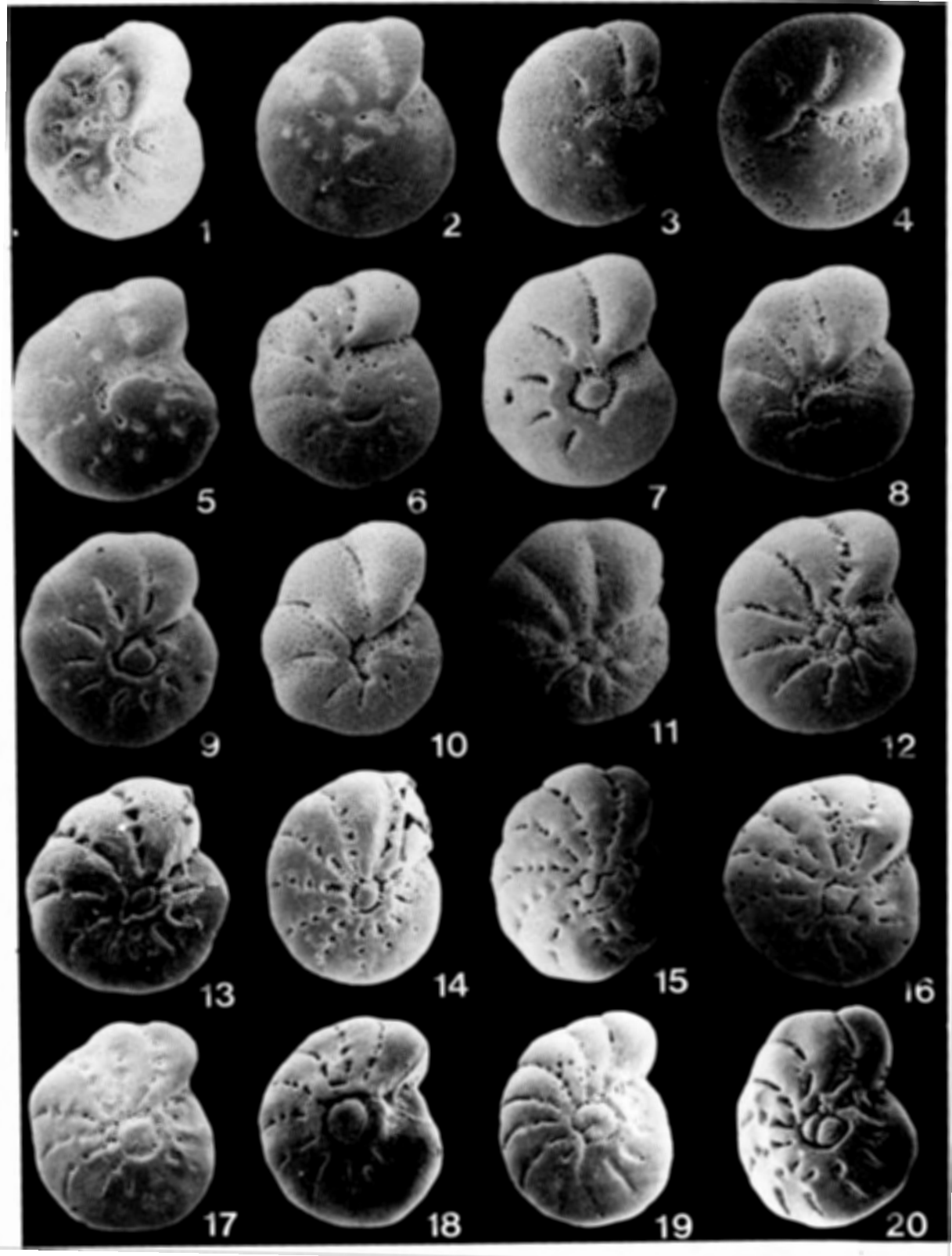


PLATE 5

Elphidium excavatum (Terquem), an intergradational series assembled from a population from Miramichi Estuary, New Brunswick, Canada. Collected from the open bay zone (Scott et al. 1980). Note the wider range of variability of the group (three formae present), and the large degree of ornamentation.

1-7. E. excavatum forma clavata Cushman. 1. SN108 x 75. 2. SN109 x 143. 3. SN110 x 117. 4. SN111 x 97. 5. SN112 x 126. 6. SN113 x 127. 7. An intermediate form, approaching E. excavatum forma lidoensis Cushman, SN114 x 91.

8-12. E. excavatum forma lidoensis Cushman. 8. SN115 x 96. 9. SN116 x 81. 10. SN117 x 98. 11. SN118 x 99. 12. SN119 x 102.

13-16. E. excavatum forma excavata (Terquem). In warmer, less saline waters, this forma has a larger umbilicus and the umbilicus and sutures contain granular material and papillae. These specimens greatly resemble the neotype illustrated by Lévy et al. (1975). 13. SN120 x 88. 14. SN121 x 93. 15. SN122 x 83. 16. SN123 x 97.

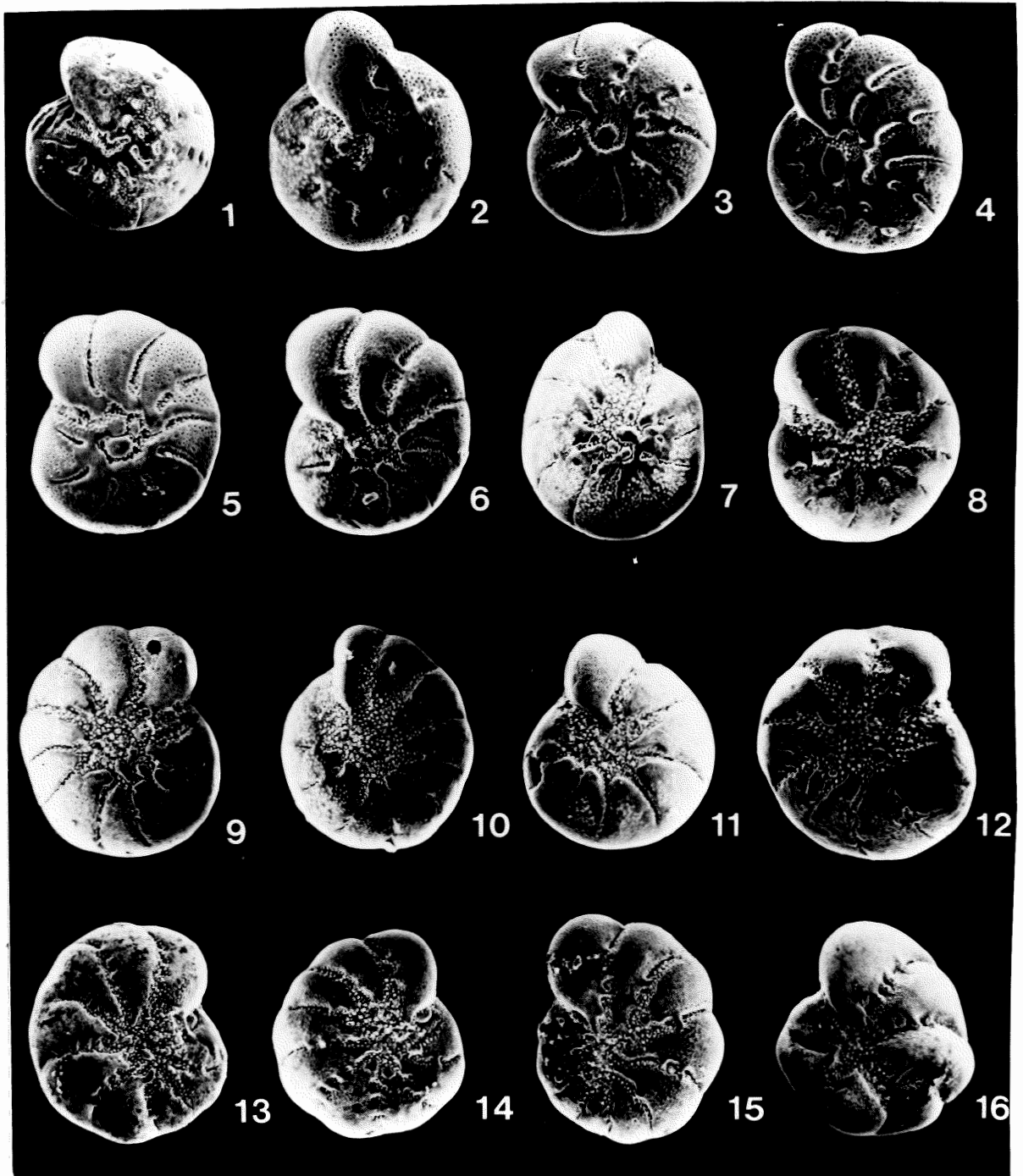


PLATE 6

Elphidium excavatum (Terquem), an intergradational series assembled from a population from the Bay of Chaleur, Gulf of St. Lawrence. There is wide variability to the group (six formae present) and a large degree of ornamentation and irregularity to many of the specimens.

1-2. Elphidium excavatum forma magna Miller, Scott and Mediolli.

1. SN269 x 70. 2. SN287 x 87.

3-6. Elphidium excavatum forma clavata Cushman. 3. SN294 x 141.

4. SN291 x 95. 5. SN282 x 88. 6. SN283 x 117.

7. Elphidium excavatum forma lidoensis Cushman. An intermediate specimen between forma clavata and forma lidoensis, SN293 x 100.

8-16. Elphidium excavatum forma excavata (Terquem). Notice the regularity, frequency, and development of the ponticuli, the lobate peripheral outline, and the papillae. The wall pores are so fine they are in some cases indistinct. 8. SN286 x 90. 9. SN278 x 69. 10. SN266 x 57. 11. SN280 x 77. 12. SN277 x 72. 13. SN267 x 72. 14. SN262 x 77. 15. An intermediate specimen, approaching forma williamsoni, SN274 x 64. 16. SN272 x 73.

17-18. Elphidium excavatum forma williamsoni Haynes ab Williamson. The ponticuli are not quite as well developed as in a "typical" forma williamsoni. 17. SN263 x 69. 18. SN275 x 67.

19. Elphidium excavatum forma excavata (Terquem). An intermediate specimen, approaching forma cuvillieri, SN288 x 80.

20. Elphidium excavatum forma cuvillieri Lévy. Note the imperforate but continuous umbilicus, SN285 x 105.

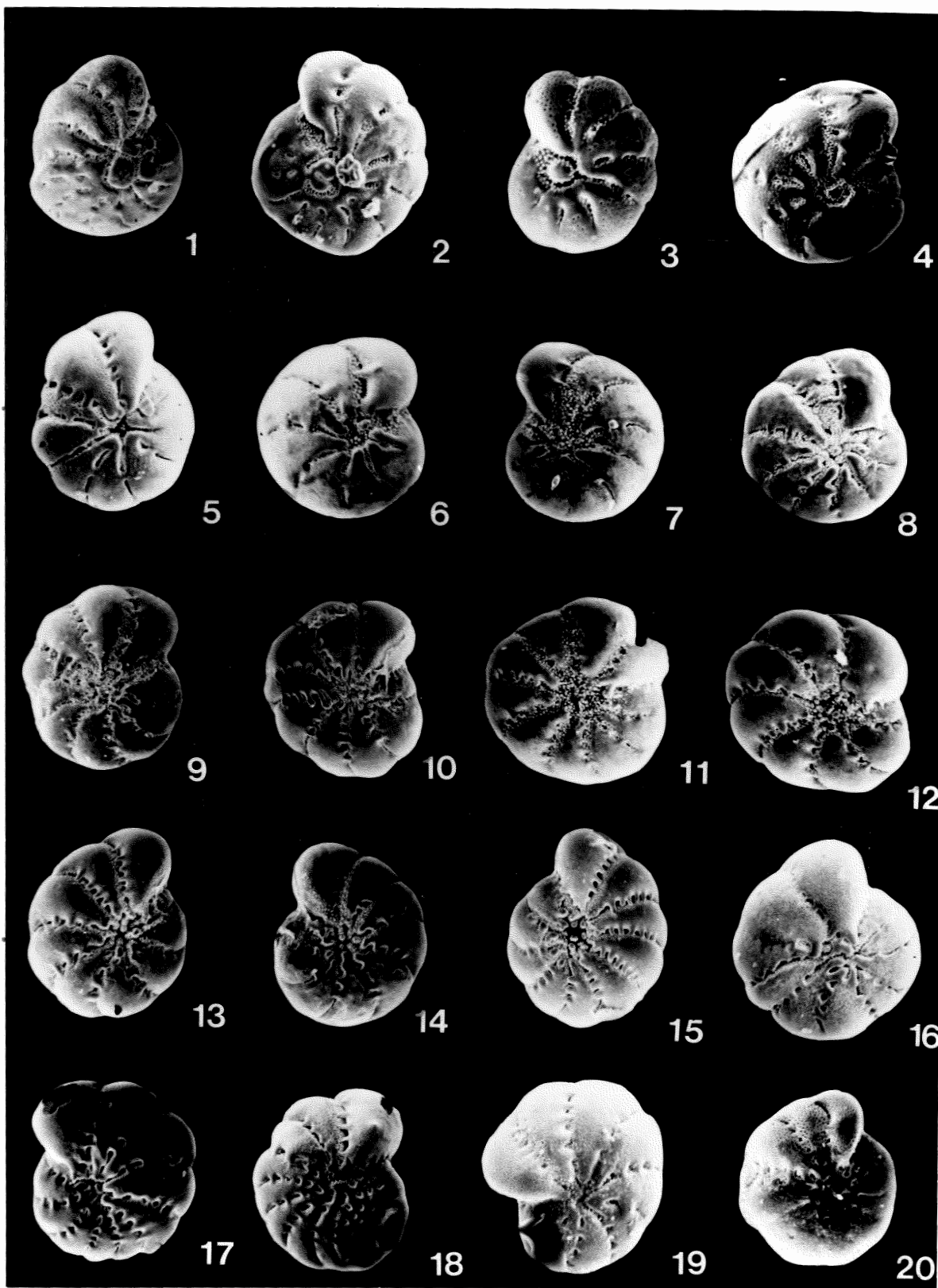


PLATE 7

Elphidium excavatum (Terquem), an intergradational series assembled from an assemblage from the Annapolis Basin, Nova Scotia, Canada. Note the wide degree of variability (five formae present).

1. E. excavatum forma selseyensis (Heron-Allen and Earland), from sample 8 SN50 x 90.

2-3. E. excavatum forma magna Miller, Scott and Medioli, from sample 8. 2. Secondary plesiotype, SN51 x 96. 3. SN52 x 103.

4-9. E. excavatum forma clavata Cushman. 4,8. From sample 8. 5-7,9. From sample 16. 4. SN53 x 94. 5. SN54 x 142. 6. SN55 x 92. 7. SN56 x 117. 8. SN57 x 105. 9. SN58 x 141.

10-12. E. excavatum forma lidoensis Cushman, from sample 16. 10. An intermediate specimen between forma clavata and forma lidoensis, SN59 x 112. 11. Representative plesiotype, SN60 x 104. 12. SN61 x 119.

13-16. E. excavatum forma excavata (Terquem). 13-14. These specimens from sample 16 resemble the neotype of Lévy et al. (1975). 13. SN62 x 104. 14. SN63 x 104. 15-16. These specimens resemble the specimens from arctic environments, from sample 8. 15. SN64 x 99. 16. SN65 x 118.

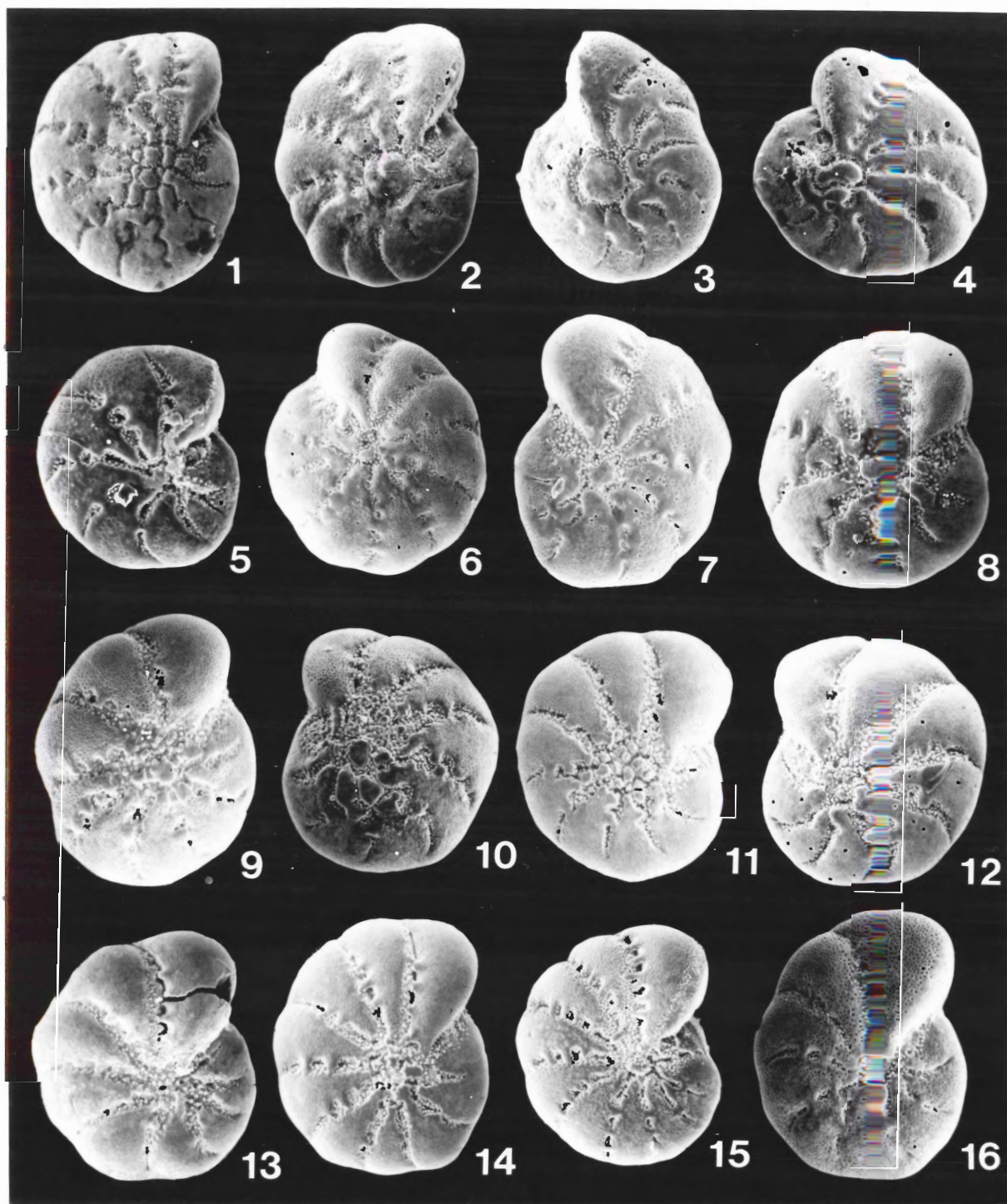


PLATE 8

Elphidium excavatum (Terquem), an intergradational series assembled from a population from the nearshore zone (Scott et al. 1980) of Chezzetcook Inlet, Nova Scotia. Notice the wide variability of the group (five formae present), and the large degree of ornamentation and irregularity of many of the specimens.

1-3. E. excavatum forma magna Miller, Scott and Medioli. 1. "Primary" representative plesiotype from station 51a₁, SN28 x 52. 2. "Secondary" representative plesiotype from 51a₁, SN29 x 53. 3. "Secondary" representative plesiotype from 53a₁, SN30 x 53.

4-8. E. excavatum forma clavata Cushman. 4. From 51a₁, SN31 x 84. 5. From 54a₂, SN32 x 90. 6. From 54a₂, SN33 x 80. 7. From 51a₁, SN34 x 68. 8. From 50a₁, SN35 x 86.

9. E. excavatum forma lidoensis Cushman. 9. From 50a₂, SN36 x 66.

10-13. E. excavatum forma selseyensis (Heron-Allen and Earland). 10. From 49a₁, SN37 x 74. 11. From 52a₂, SN38 x 142. 12. From 54a₁, SN39 x 70. 13. Representative plesiotype, from 55a₂, SN40 x 51.

14. E. excavatum forma clavata Cushman, from 50a₂, SN41 x 86.

15-16. E. excavatum forma excavata (Terquem), these specimens resemble the neotype of Lévy et al. (1975). 15. From 50a₁, SN42 x 74. From 55a₁, SN43 x 78.

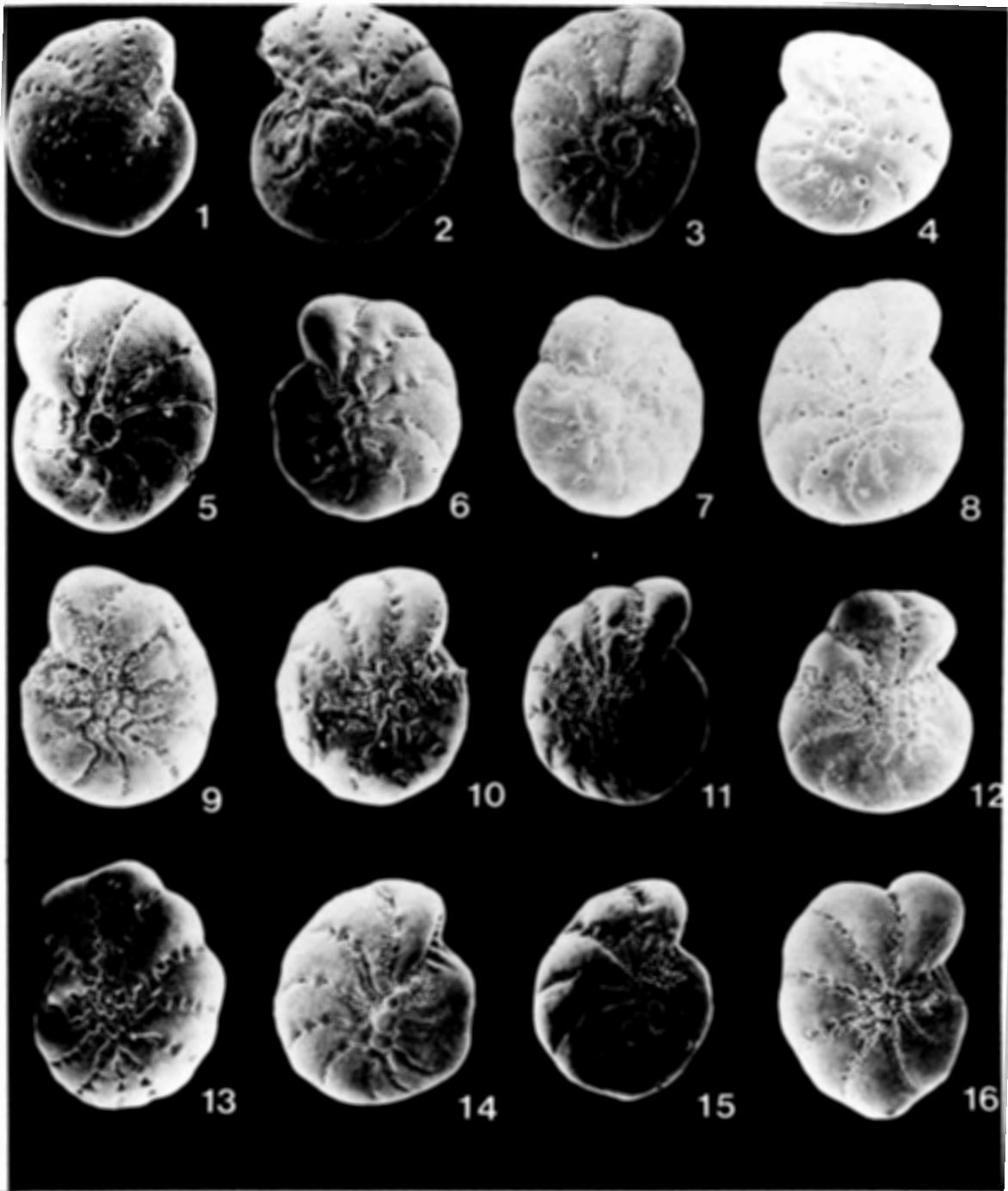


PLATE 9

1-18. Specimens of E. excavatum from a population from a Maine-New Brunswick estuary. Three formae are present, one is very dominant.

1-16. Typical specimens of E. excavatum forma williamsoni Haynes ab Williamson. The ponticuli are very regular and extend across the periphery. The walls are densely perforate; and an umbilical boss may or may not be present. 1. SN438 x 130. 2. SN461 x 124. 3. SN439 x 110. 4. SN453 x 114. 5. SN443 x 112. 6. SN456 x 95. 7. SN441 x 100. 8. SN452 x 94. 9. SN451 x 83. 10. SN449 x 85. 11. SN427 x 77. 12. SN428 x 76. 13. SN436 x 80. 14. SN442 x 72. 15. SN458 x 80. 16. SN462 x 75.

17. E. excavatum forma lidoensis Cushman, SN477 x 134.

18. E. excavatum forma clavata Cushman, SN445 x 81.

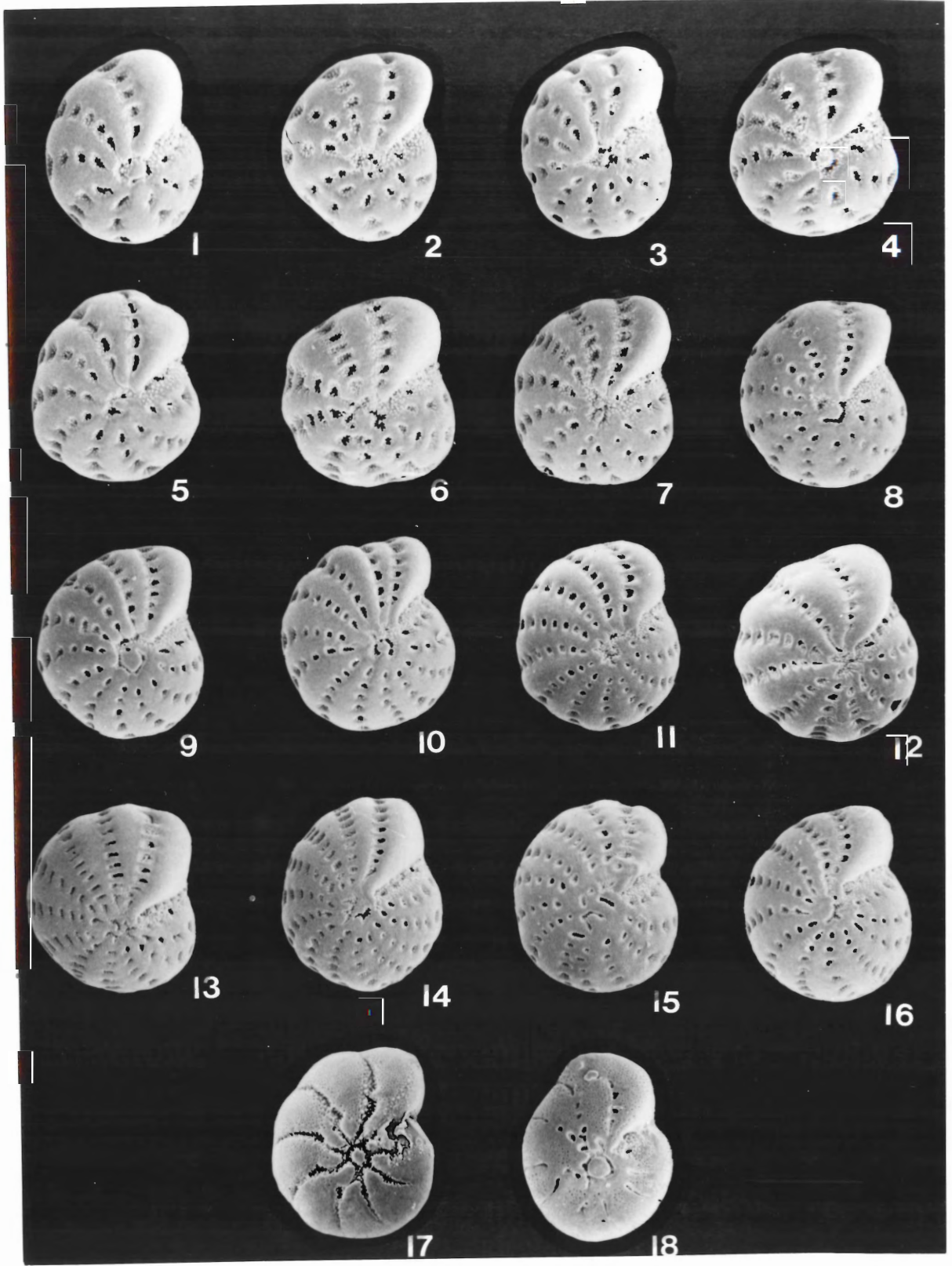


PLATE 10

1-16. Elphidium excavatum (Terquem) an intergradational series assembled from a population from Long Island Sound. Notice the wide range of variability (four formae present) and the large degree of ornamentation and irregularity of many of the specimens.

1-3. E. excavatum forma clavata Cushman (juveniles?) 1. SN90 x 249. 2. SN91 x 175. 3. An intermediate specimen, SN92 x 148.

4-5. E. excavatum forma clavata Cushman. 4. SN93 x 134. 5. SN94 x 90.

6-8. E. excavatum forma excavata (Terquem). 6. SN95 x 99. 7. SN96 x 103. 8. SN97 x 96.

9-13. E. excavatum forma selsevensis (Heron-Allen and Earland). 9. SN98 x 62. 10. SN99 x 65. 11. SN100 x 57. 12. SN101 x 83. 13. SN102 x 64.

14. E. excavatum forma excavata (Terquem). An intermediate specimen between forma excavata and forma selsevensis, SN103 x 85.

15-16. E. excavatum forma lidoensis Cushman. 15. SN104 x 93. 16. SN105 x 94.

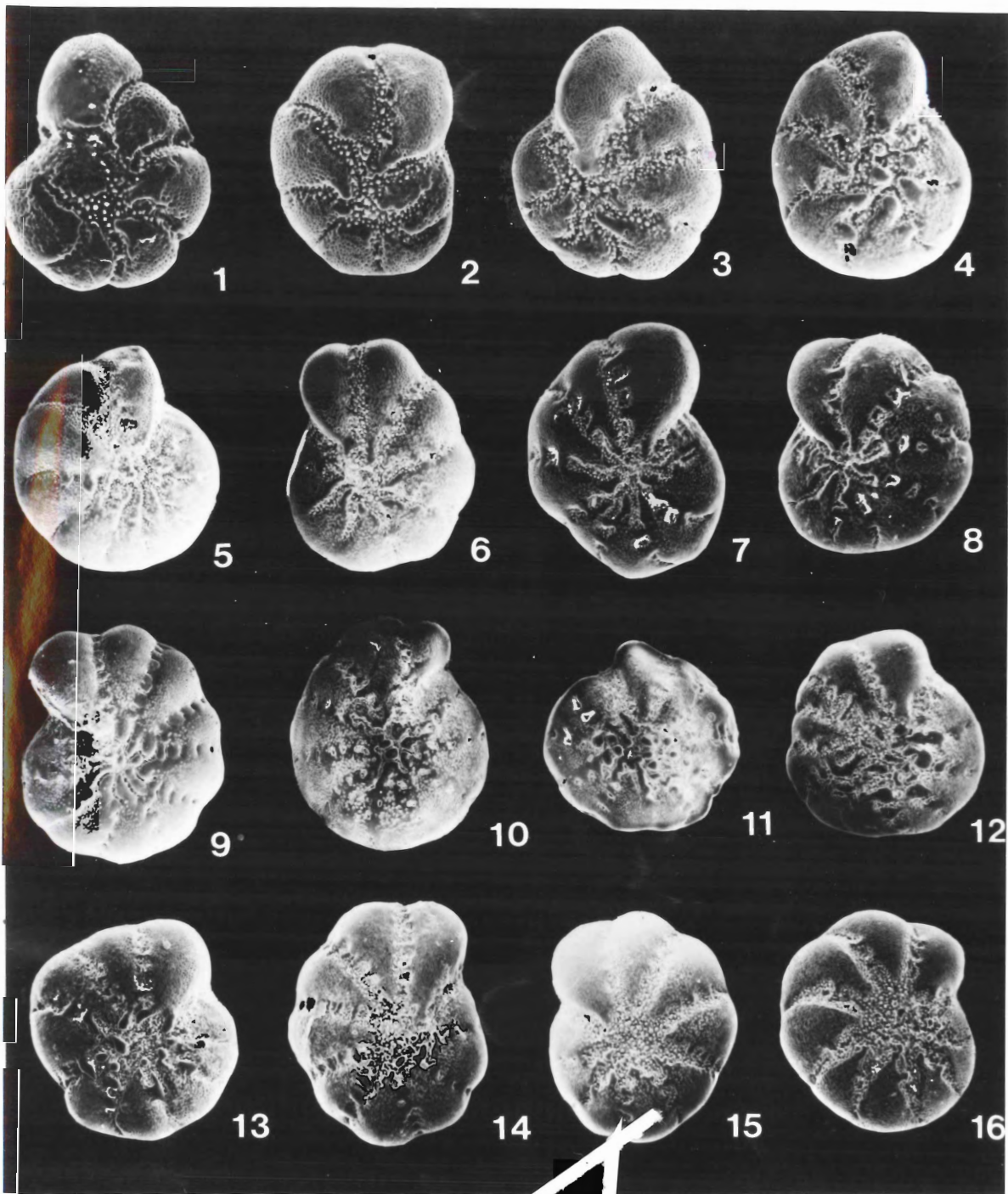


PLATE 11

Elphidium excavatum (Terquem), an intergradational series of four formae (six present in total) from a population from San Diego Bay, California. This is the only location where the morphotype tumidum was observed as a dominant form.

1-4. E. excavatum forma tumidum Natland. The ponticuli are wide and regularly spaced, the umbilicus is large, circular and filled with papillae/bosses. The chambers end abruptly against the umbilicus. 1. SN786 x 160. 2. SN784 x 141. 3. SN321 x 70. 4. SN316 x 89.

5. E. excavatum forma selsevensis (Heron-Allen and Earland), SN352 x 102.

6-8. E. excavatum forma clavata Cushman. These specimens are irregular and ornamented. 6. SN313 x 84. 7. SN356 x 90. 8. SN323 x 63.

9-13. E. excavatum forma lidoensis Cushman. 9. SN309 x 108. 10. SN310 x 141. 11. SN789 x 167. 12. SN791 x 224. 13. SN306 x 166.

15-16. E. excavatum forma tumidum Natland. 15. SN305 x 130. 16. Representative plesiotype, SN359 x 140.

17-18. E. excavatum forma gunteri Cole. No intermediate specimens linking this forma to the other formae were observed. 17. SN304 x 83. 18. SN360 x 81.

19-21. Specimen believed to be E. excavatum forma galvestonensis Kornfeld. These are not typical specimens; and were tentatively identified with the aid of enlargements (pl. 26:15-16). No intermediate specimens were observed linking this forma to the other formae from this location. These specimens were not included in the

statistical analysis. 19. SN325 x 84. 20. SN326 x 147. 21. SN324 x
126.

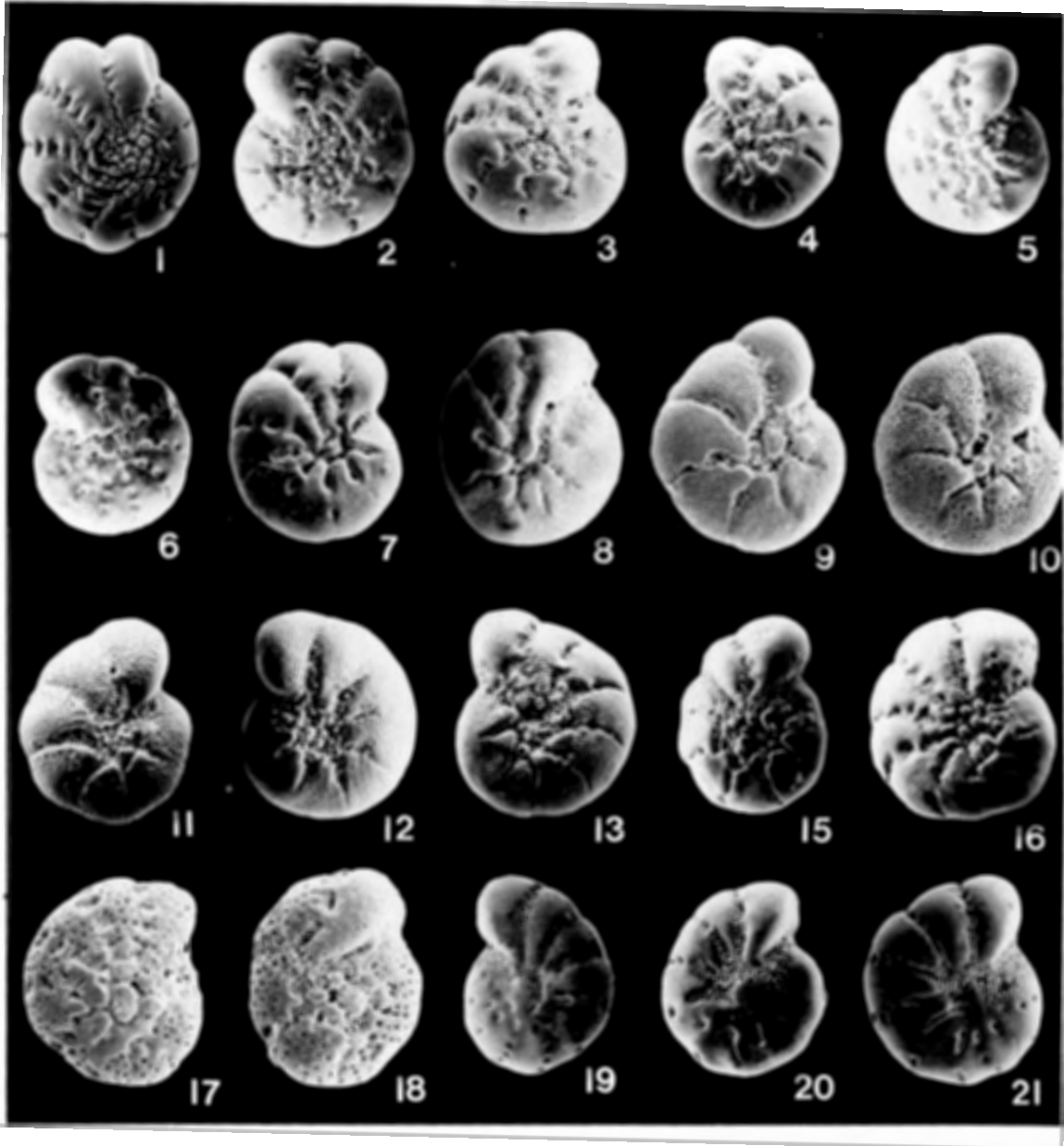


PLATE 12

Elphidium excavatum (Terquem) an intergradational series of five formae from a Pleistocene assemblage from San Francisco Bay. There is wide variability to the group (seven formae present) and a large degree of ornamentation and irregularity to many of the specimens.

1-3. E. excavatum forma excavata (Terquem). 1. SN751 x 87. 2. SN765 x 85. 3. SN767 x 88.

4-5. E. excavatum forma lidoensis Cushman. 4. SN756 x 98. 5. SN781 x 170.

6. E. excavatum forma tumidum Natland. An intermediate specimen between forma lidoensis and forma tumidum, with the circular umbilicus and the ponticuli beginning to develop as for the latter form, SN754 x 90.

7-10. E. excavatum forma lidoensis Cushman. 7-8. Two typical specimens. 7. SN760 x 143. 8. SN764 x 189. 9-10. Two irregular ornamented specimens. 9. SN779 x 96. 10. SN753 x 106.

11. E. excavatum forma clavata Cushman, a specimen with an imperforate umbilical collar and sutures not extending externally to the periphery, SN765 x 108.

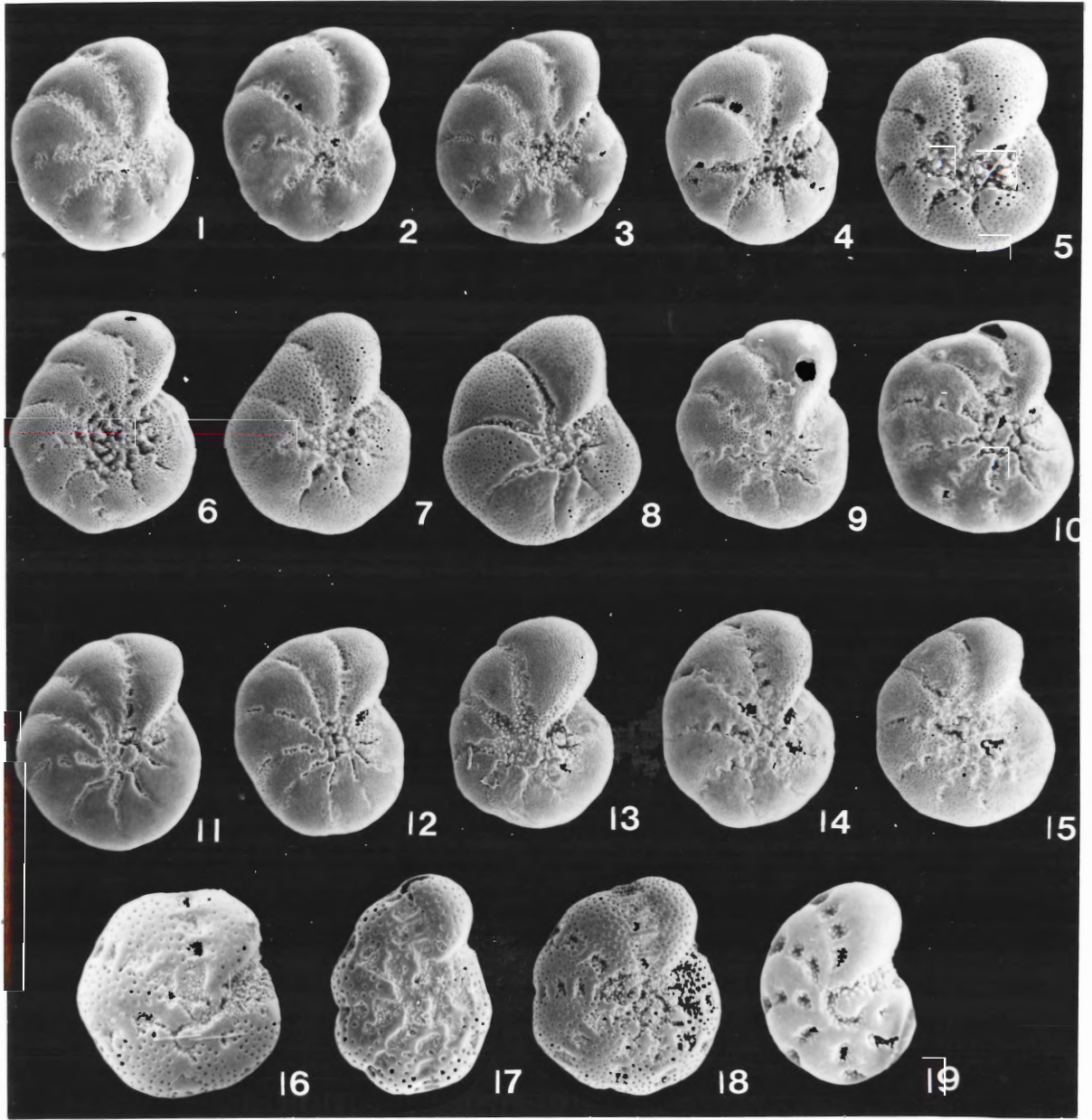
12. E. excavatum forma excavata (Terquem), SN752 x 89.

13. E. excavatum forma lidoensis Cushman. An irregular specimen, SN778 x 103.

14-15. E. excavatum forma selseyensis (Heron-Allen and Earland). Two irregular specimens. 14. SN761 x 95. 15. SN776 x 95.

16-18. E. excavatum forma gunteri Cole. Irregular specimens. 16. SN773 x 144. 17. SN763 x 130. 18. SN759 x 108.

19. Specimen tentatively identified as E. excavatum forma williamsoni Haynes ab Williamson, when compared to specimens from a Maine-New Brunswick estuary (pl. 10:1) and from Bay of Izmir, Turkey (pl. 19:16-17). Possibly a juvenile specimen, SN774 x 187.



PLATES 13 - 14

Elphidium excavatum (Terquem) two intergradational series (eight formae present) from a mid-Holocene assemblage from Baie Verte, Northumberland Strait, Canada. At no other location has such variability or ornamentation been observed in the E. excavatum group. This location also had the largest percentage of intermediate forms.

PLATE 13

1-2. E. excavatum forma gunteri Cole. 1. SN154 x 107. 2. SN153 x 105.

3-5. E. excavatum forma clavata Cushman, ornamented specimens. 3. SN128 x 62. 4. SN129 x 93. 5. SN130 x 93.

6-7. E. excavatum forma ouvillieri Lévy, specimens lacking the smooth umbilicus typical of the morphotype. 6. SN131 x 68. 7. SN132 x 63.

8-9. E. excavatum forma clavata Cushman. 8. SN133 x 76. 9. SN134 x 72.

10. E. excavatum forma excavata (Terquem), SN135 x 57.

11. E. excavatum forma lidoensis Cushman, SN136 x 93.

12-15. E. excavatum forma selseyensis (Heron-Allen and Earland).

12. A specimen approaching forma tumidum, SN137 x 81. 13. SN138 x 61.

14. SN139 x 82. 15. An intermediate specimen approaching forma excavata, SN140 x 96.

16. E. excavatum forma clavata Cushman, SN143 x 82.

17-20. E. excavatum forma galvestonensis Kornfeld. 17. An intermediate specimen between forma clavata and forma galvestonensis.

The pores are of intermediate density and the ponticuli not strongly developed. The umbo is not fully raised as typical for forma galvestonensis, SN142 x 94. 18-20. Typical specimens. 18. SN160 x 82. 19-20. Etched and broken specimens. 19. SN163 x 59. 20. SN164 x 69.

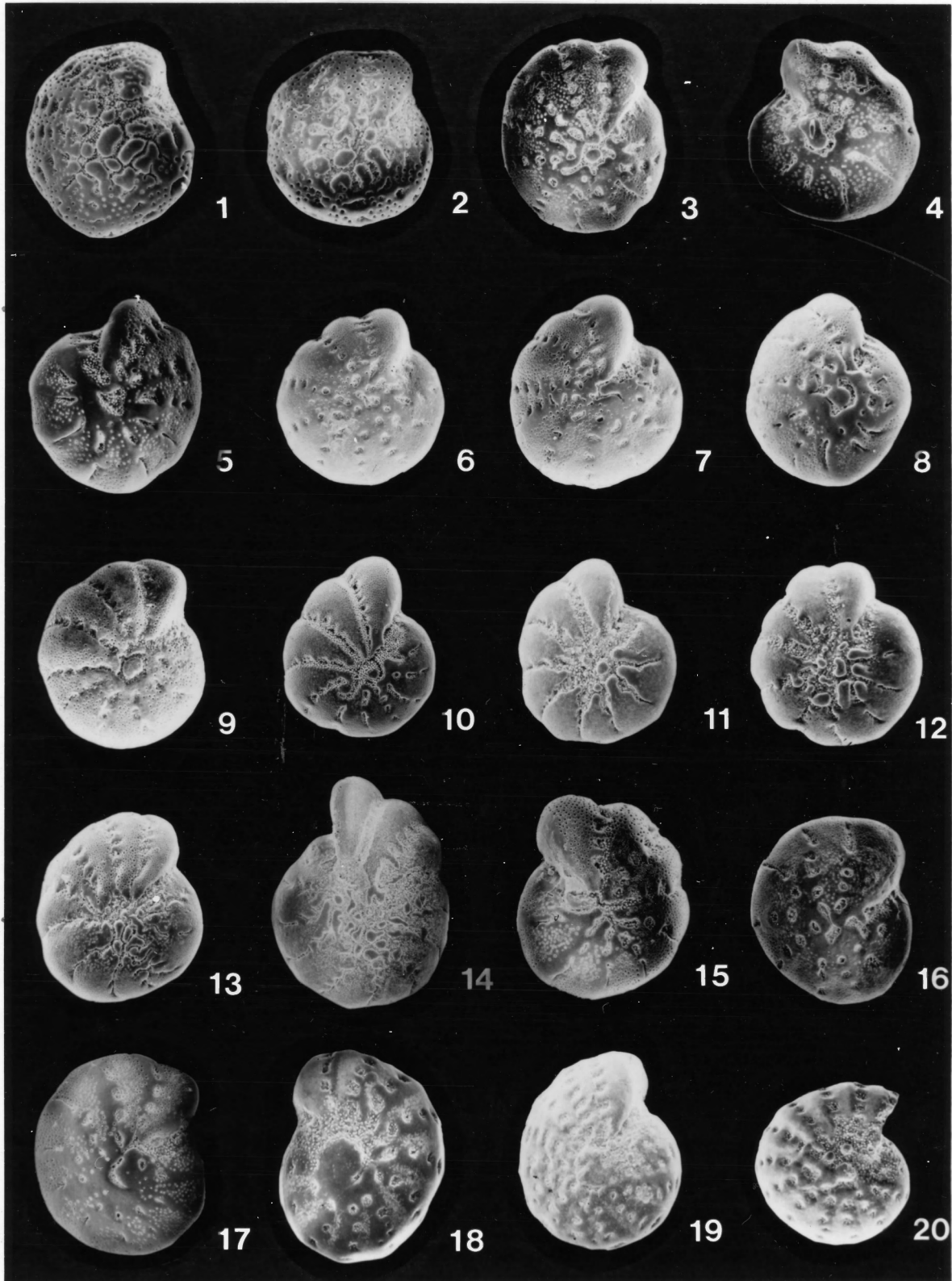


PLATE 14

1-3. E. excavatum forma gunteri. 1-2. Typical specimens. 1. SN176 x 94. 2. SN184 x 121. 3. Intermediate specimen, between forma gunteri and forma clavata, SN231 x 96.

4-5. E. excavatum forma clavata Cushman. 4. SN252 x 119. 5. SN234 x 88.

6-7. E. excavatum forma cuvillieri Lévy. 6. An intermediate specimen between forma clavata and forma cuvillieri, SN167 x 86. 7. SN245 x 80.

8-9. E. excavatum forma selseyensis (Heron-Allen and Earland). 8. SN170 x 70. 9. SN239 x 77.

10. E. excavatum forma lidoensis Cushman, an ornamented specimen, SN255 x 94.

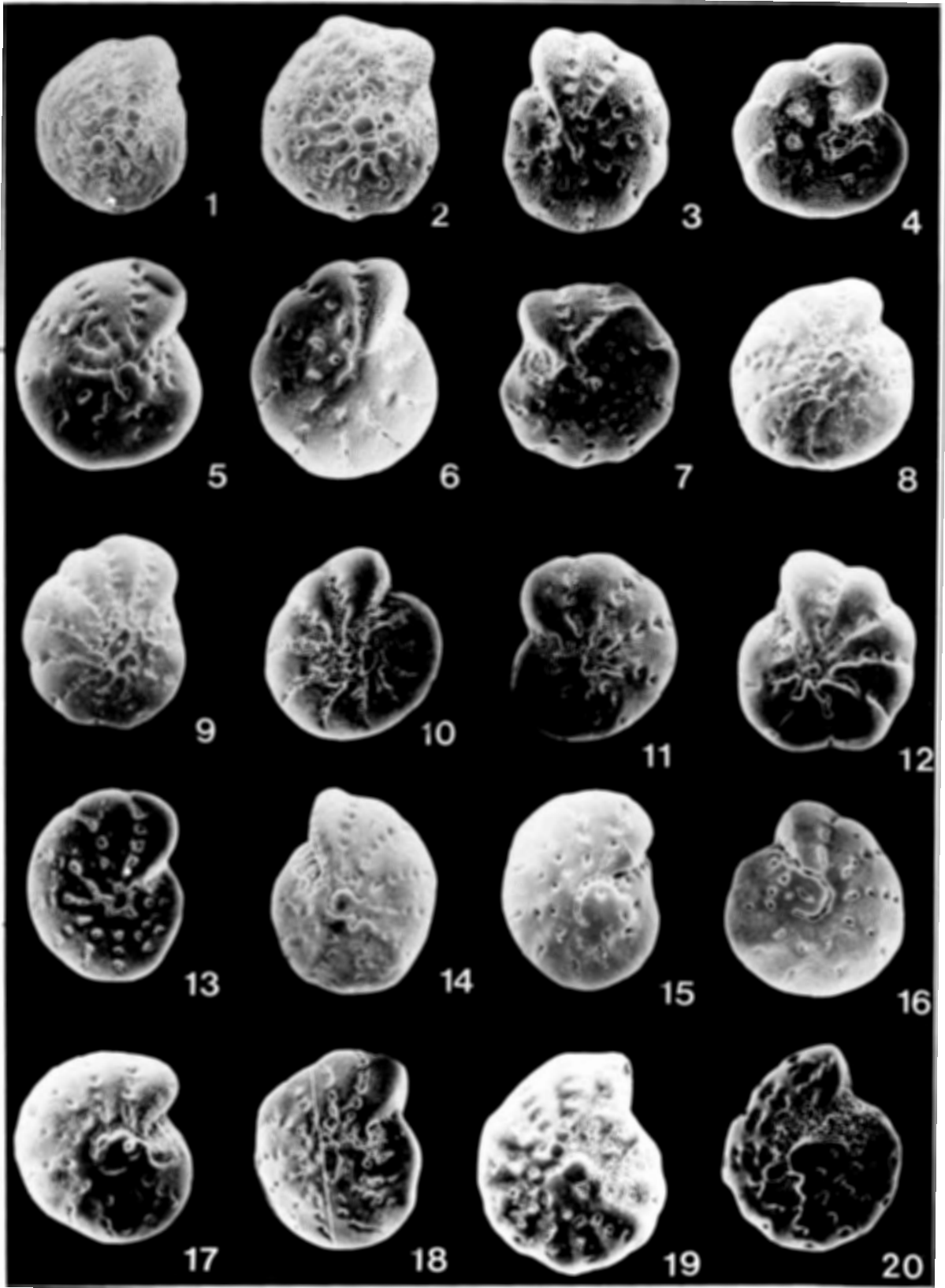
11-12. E. excavatum forma excavata (Terquem), irregular specimens. 11. SN243 x 87. 12. SN232 x 99.

13-14. E. excavatum forma clavata Cushman. 13. SN246 x 99. 14. SN240 x 63.

15-17. E. excavatum forma magna Miller, Scott and Medioli. 15. SN237 x 81. 16. SN242 x 74. 17. SN230 x 89.

18. E. excavatum forma selseyensis (Heron-Allen and Earland), SN249 x 56.

19-20. E. excavatum forma galvestonensis Kornfeld. 19. SN179 x 83. 20. SN260 x 132.



PLATES 15 - 16

Elphidium excavatum (Terquem), from a population from San Antonio Bay, Texas.

PLATE 15

An intergradational series of the two dominant formae, E. excavatum forma gunteri and forma lidoensis. Note the gradual morphological changes from one forma to the other, through the specimens illustrated. This location has the largest number of intermediate forms linking these two formae.

1-7. E. excavatum forma lidoensis Cushman. These warmer water specimens are more coarsely perforate than those specimens seen in maritime Canada and the northeastern United States. 1. SN705 x 98. 2. SN718 x 126. 3. SN680 x 94. 4. SN728 x 118. 5-7. Intermediate specimens exhibiting characteristics of both formae. If not for the coarse perforations, these three specimens could be identified as forma excavata. 5. SN750 x 149. 6. SN727 x 133. 7. SN724 x 107.

8-15. E. excavatum forma gunteri Cole. Typical specimens of the forma. 8. SN741 x 101. 9. Note the variation in the size and shape of the ponticuli from one suture to another, representative plesiotype SN744 x 101. 10. SN748 x 62. 11. SN729 x 66. 12. SN703 x 91. 13. SN723 x 108. 14. SN733 x 126. 15. SN719 x 115.

16-20. E. excavatum forma lidoensis Cushman. 16. SN697 x 189. 17. SN691 x 157. 18. SN676 x 107. 19. SN735 x 118. 20. SN786 x 117.

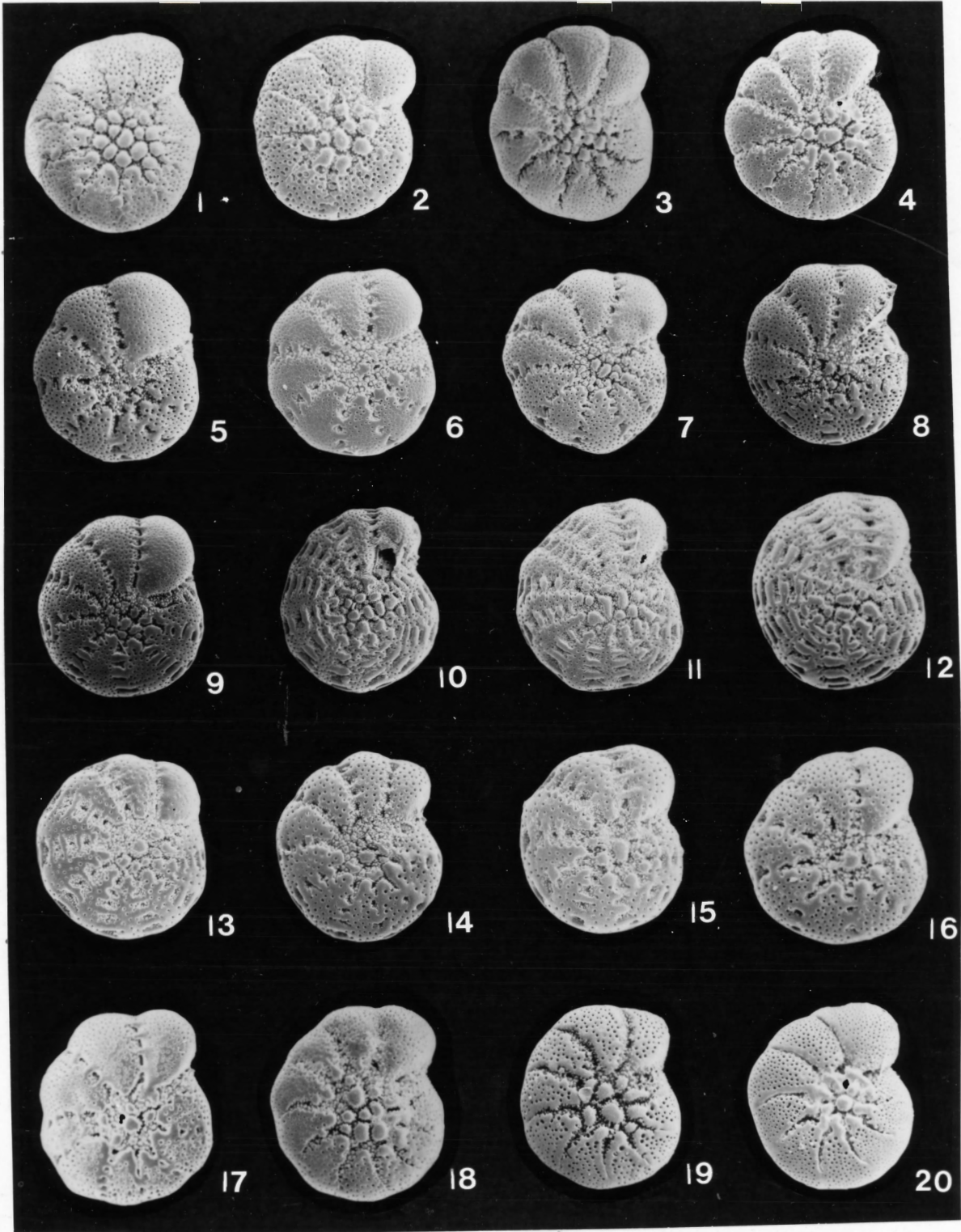


PLATE 16

The two other formae of E. excavatum identified in San Antonio Bay samples. No intermediate specimens were found linking these two formae to the other forma present or the remainder of the E. excavatum group.

1-8. E. excavatum forma galvestonensis Kornfeld. Typical specimens of this forma. 1. SN701 x 77. 2. SN797 x 83. 3. Representative plesiotype, SN795 x 49. 4. SN796 x 54. 5. SN900 x 62. 6. SN700 x 66. 7. SN720 x 65. 8. SN748 x 63.

9-14. E. excavatum forma cuvillieri Lévy. 9. SN690 x 142. 10. SN737 x 146. 11. SN725 x 123. 12. SN726 x 115. 13. SN696 x 144. 14. SN742 x 127.

15. Elphidium sp., not included in the analysis, SN710 x 134.

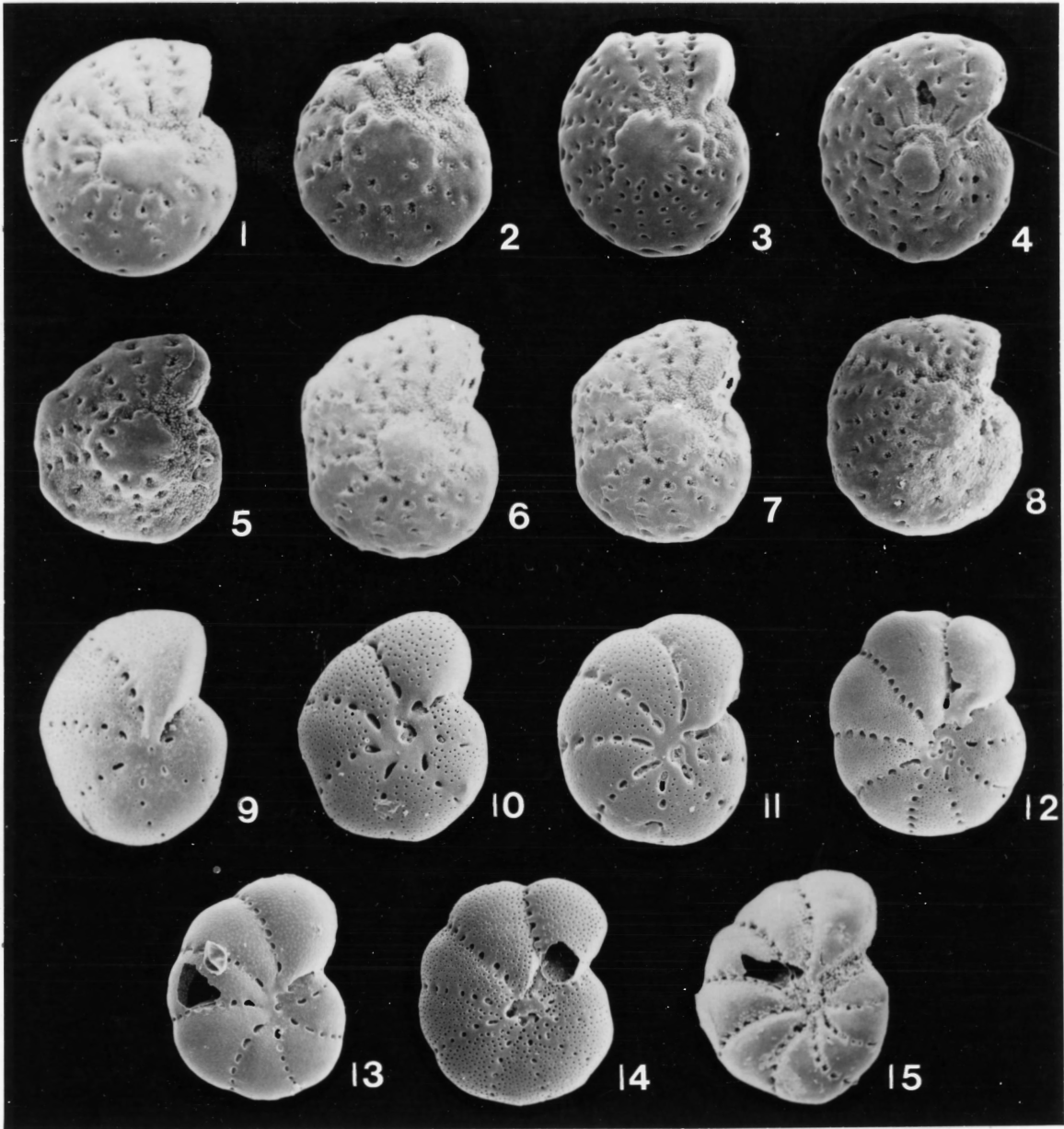


PLATE 17

Elphidium excavatum (Terquem), two formae observed in a Holocene assemblage from the Wadden Sea, the Netherlands. One forma (E. excavatum forma williamsoni) comprised over 95% of the population. There were no intermediate specimens observed.

1-12. E. excavatum forma williamsoni Haynes ab Williamson. 1. SN393 x 125. 2. SN392 x 125. 3. SN395 x 101. 4. SN397 x 130. 5. SN417 x 94. 6. SN408 x 86. 7. SN377 x 79. 8. SN396 x 82. 9. SN336 x 97. 10. SN402 x 80. 11. SN420 x 78. 12. SN375 x 93.

13-15. E. excavatum forma gunteri Cole. 13. Typical specimen, SN425 x 106. 14-15 Etched specimens. 14. SN424 x 110. 15. Intermediate specimen, approaching forma lidoensis, SN426 x 123.

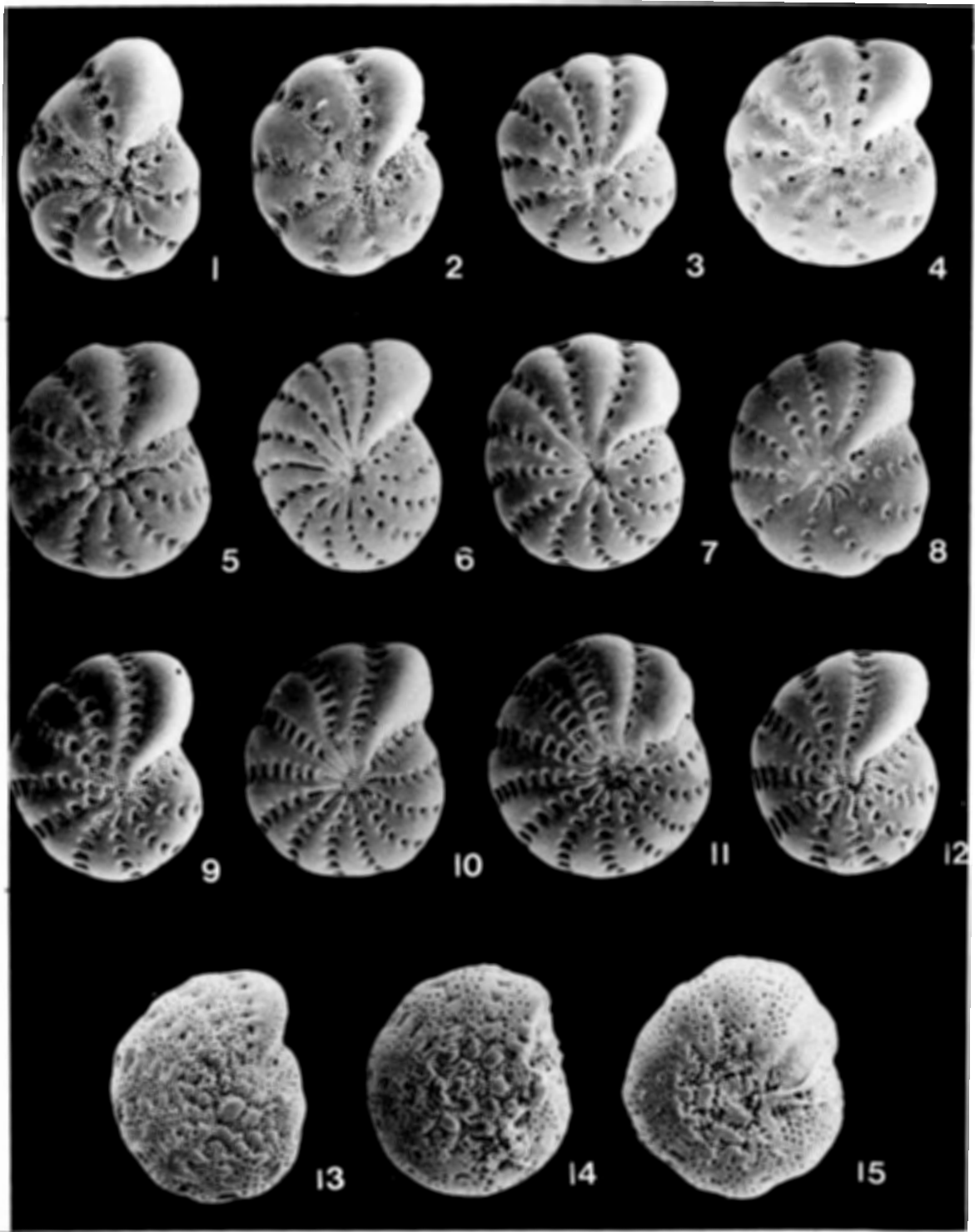


PLATE 18

Elophidium excavatum (Terquem), from a population from Venice Lagoon, Italy. Four formae are identified and one other is tentatively identified.

1-4. E. excavatum forma gunteri Cole. 1. Typical specimen, SN629 x 159. 2. SN622 x 131. 3. SN617 x 181. 4. Intermediate specimen approaching forma lidoensis, SN619 x 123.

5-7. E. excavatum forma lidoensis Cushman. 5. SN623 x 137. 6. SN369 x 83. 7. SN633 x 158.

8-15. E. excavatum forma cuvillieri Lévy. Note the variation in the umbilical regions, sutures, and ponticuli. No intermediate specimens were found linking this forma to other members of the group. 8. SN368 x 87. 9. SN630 x 144. 10. SN384 x 79. 11. SN627 x 116. 12. SN616 x 75. 13. SN635 x 95. 14. SN385 x 71. 15. SN380 x 51.

16,18. E. excavatum forma galvestonensis Kornfeld. Specimens tentatively identified by comparison with specimens from San Diego Bay and not included in the analysis. No intermediate specimens were found linking this forma to other members of the group. 16. SN387 x 125. 18. SN388 x 120.

17. E. excavatum forma williamsoni Haynes ab Williamson, SN662 x 106.

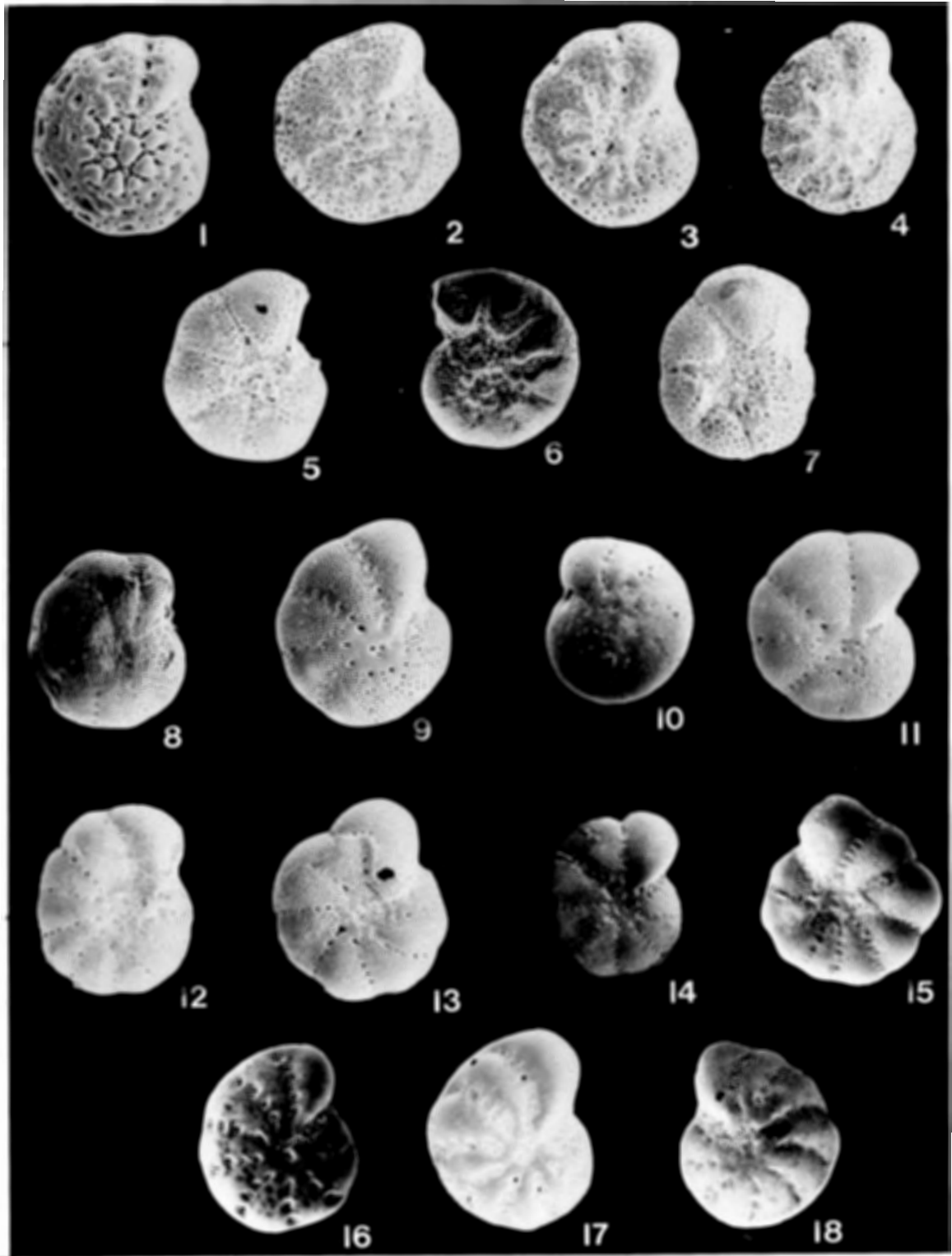


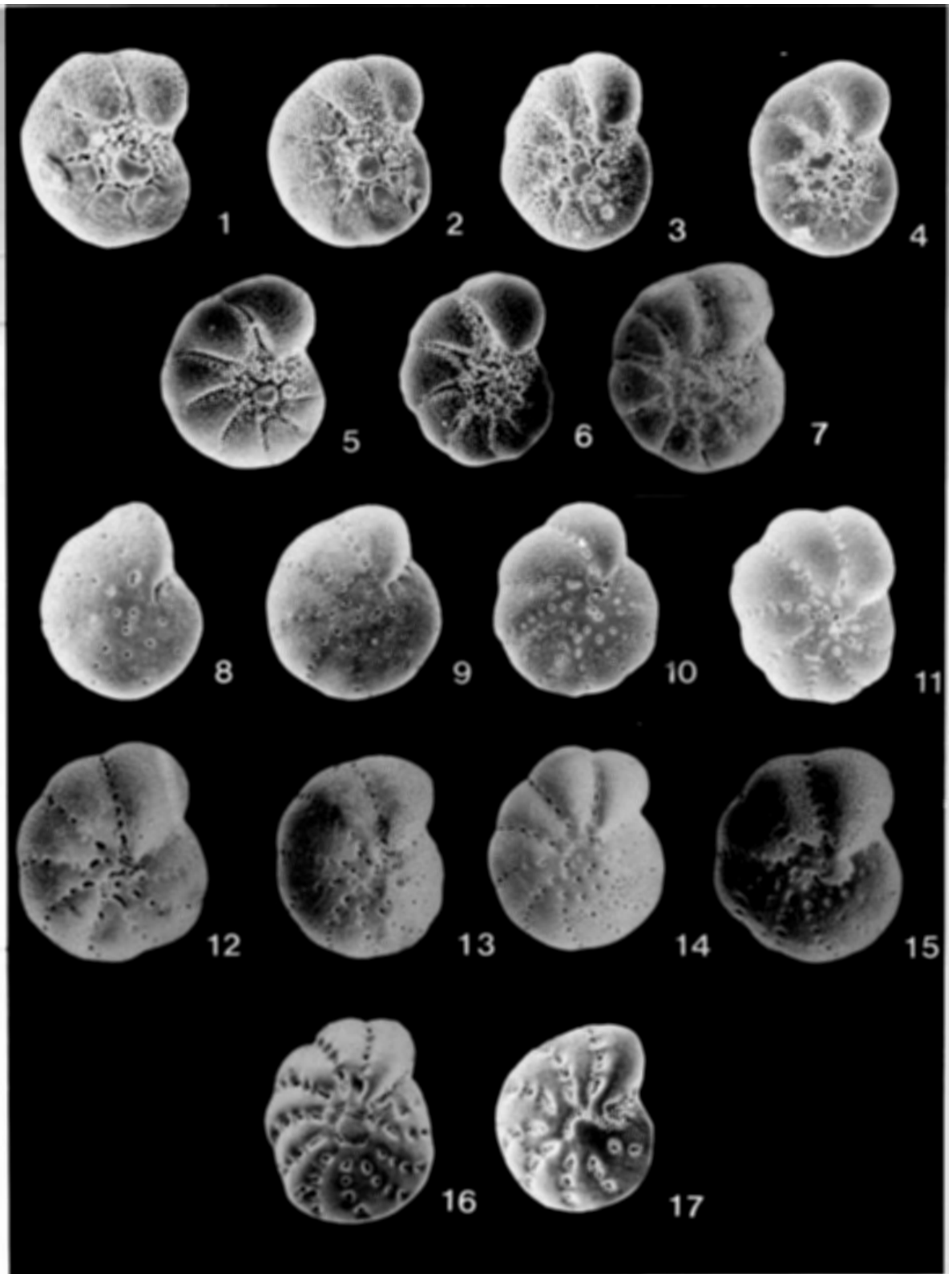
PLATE 19

E. excavatum (Terquem) from a late Pleistocene assemblage, from the Bay of Izmir, Turkey. Three formae were observed, but no intermediate specimens linking these three formae to one another or to other members of the group were observed.

1-7. E. excavatum forma lidoensis Cushman. 1. SN16 x 212. 2. SN75 x 124. 3. SN204 x 121. 4. SN247 x 87. 5. SN259 x 76. 6. SN265 x 102. 7. SN607 x 66.

8-15. E. excavatum forma cuvillieri Lévy. 8. SN299 x 186. 9. SN329 x 86. 10. SN330 x 84. 11. SN333 x 80. 12. SN581 x 81. 13. SN578 x 70. 14. SN580 x 79. 15. SN590 x 97.

16-17. E. excavatum forma williamsoni Haynes ab Williamson 16. SN661 x 96. 17. SN324 x 137.



PLATES 20-28

The following 9 plates illustrate the morphological features of, and variation within, each morphotype of E. excavatum.

The following code of letters has been used on these plates (plus plate 30), to designate these morphological features.

- a umbilicus
- b suture
- c umbilical boss
- d imperforate umbilical collar
- e papillae
- f ponticulus
- g retral process pit
- h umbilical aperture
- i sutural pore
- j apertural face
- k interiomarginal apertural arches
- l fossette
- m foramen
- n chamber

PLATE 20

Elphidium excavatum forma excavata Terquem. Note the lobate peripheral outline, straight intercameral sutures which extend to the umbilicus and contain papillae, and the absence of an imperforate umbilical collar.

1-2. Recent specimens from Miramichi estuary, equatorial views.

1. SN123 x 77. 2. SN120 x 81.

3. Recent specimen from Annapolis Basin, equatorial view, SN64 x 78.

4-7. Recent specimens from Long Island Sound. 4-6. Equatorial views. 4. SN537 x 96. 5. SN536 x 82. 6-7. SN540, an intermediate specimen approaching forma selseyensis. 6. x 71. 7. Enlargement showing the wall porosity, chambers, umbilicus, imperforate ponticuli, intercameral sutures, and papillae, x 358.

8-10. Pleistocene specimens from San Francisco Bay. 8-9. Equatorial views. 8. SN757 x 88. 9-10. SN768. 9. x 113. 10. Enlargement of the ultimate intercameral suture, showing ponticuli and sutural papillae, x 454.

11. Enlargment of a specimen (equatorial view illustrated by Miller 1979, Pl. 1:1; Pl. 2:1; Miller et al. 1982, Pl. 2:10, Pl. 3:1, Pl. 4:1) from the Labrador Shelf (Late Pleistocene in age). Note the excavated umbilicus, straight intercameral sutures extending to the umbilicus, and lack of imperforate umbilical collar, SN1 x 480.

12. Recent specimen from Chezzetcook Inlet, intermediate specimen approaching forma lidoensis, SN296 x 106.

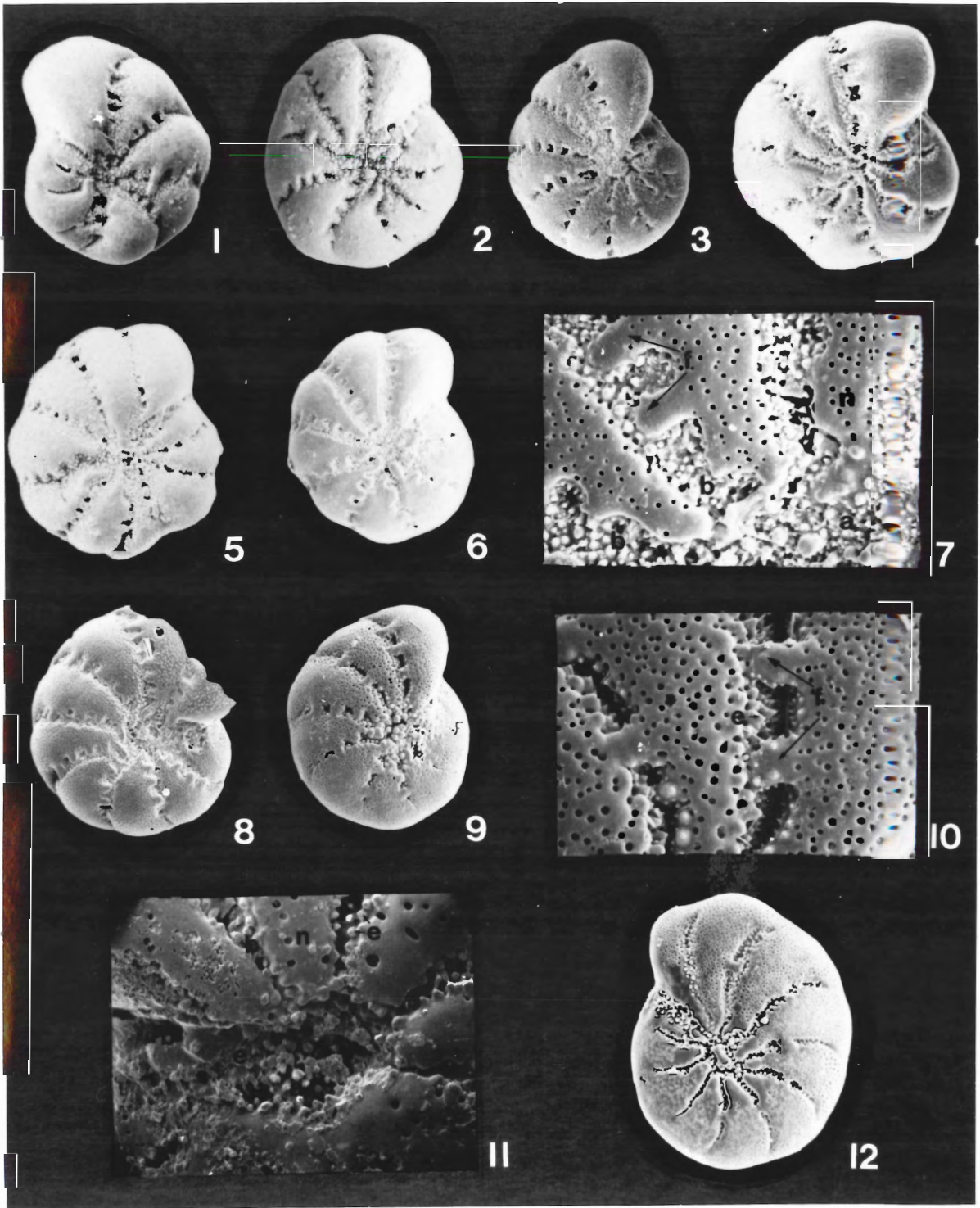


PLATE 21

Elphidium excavatum forma williamsoni Haynes ab Williamson. This form is characterized by the inflated chambers, smooth peripheral outline, and numerous large, well developed, regular ponticuli. The ponticuli extend across the periphery, the walls are very finely perforate, and an boss may be present.

1-3. Recent specimens from a Maine-New Brunswick estuary. 1-2. Representative plesiotype of the forma. 1. Equatorial view; note partially formed umbilical boss, SN471 x 69. 2. Detail of the ultimate intercameral suture of the same specimen showing finely perforate wall, suture, papillae in the suture and the regularity of the ponticulli, x 347. 3. Juvenile specimen (?) with a umbilical boss, SN438 x 101.

4. Late Pleistocene specimen from San Francisco Bay, juvenile specimen (?) with partially formed umbilical boss, SN774 x 143.

5-6. Recent specimens from Chezzetcook Inlet, Nova Scotia; equatorial views. 5. SN209 x 60. 6. SN297 x 54.

10-11. Recent specimens from Bay of Chaleur. Both have umbilical bosses. 10. SN279 x 83. 11. SN270 x 56.

7-9, 12-24. Holocene specimens from the Wadden Sea, Netherlands. 7, 12-13. SN897. 7. Peripheral view of the apertural face. Note the ponticuli extending across the periphery, x 70. 8-9. SN361. 8. Equatorial view, x 48. 9. Detail of the intercameral suture and regular ponticuli, x 274. 12. Detail of the apertural face showing papillae and the cribrate interiomarginal apertures, x 129. 13. Detail of the interiomarginal apertural arches, x 261.

14-15, 19. SN418. 14. Equatorial view, showing broken ultimate chamber and outer wall of the previous chamber underneath, x 54. 15,19. Interior of the broken ultimate chamber wall, showing the septal wall structure and previous interiomarginal apertual arches, x 467, and x 354, respectively.

16-18. SN375. 16. Equatorial view, x 57. 17-18. Detail of the intercameral suture, showing the finely perforate wall, regular ponticuli, and spinose sutural papillae. 17. x 371. 18. x 530.

20-21. SN378. 20. Equatorial view, x 61. 21. Enlargement of the umbilicus, and intercameral suture extentions into the umbilicus, x 308.

22-24. SN405. 22. Equatorial view, with broken ultimate chamber, x 50. 23. Interior of the ultimate chamber wall, viewed from the inside, illustrating the perforations extending through the wall, x 121. 24. Retral process pits (inside the hollow ponticuli) which house the cytoplasm of the retral processes, x 397.

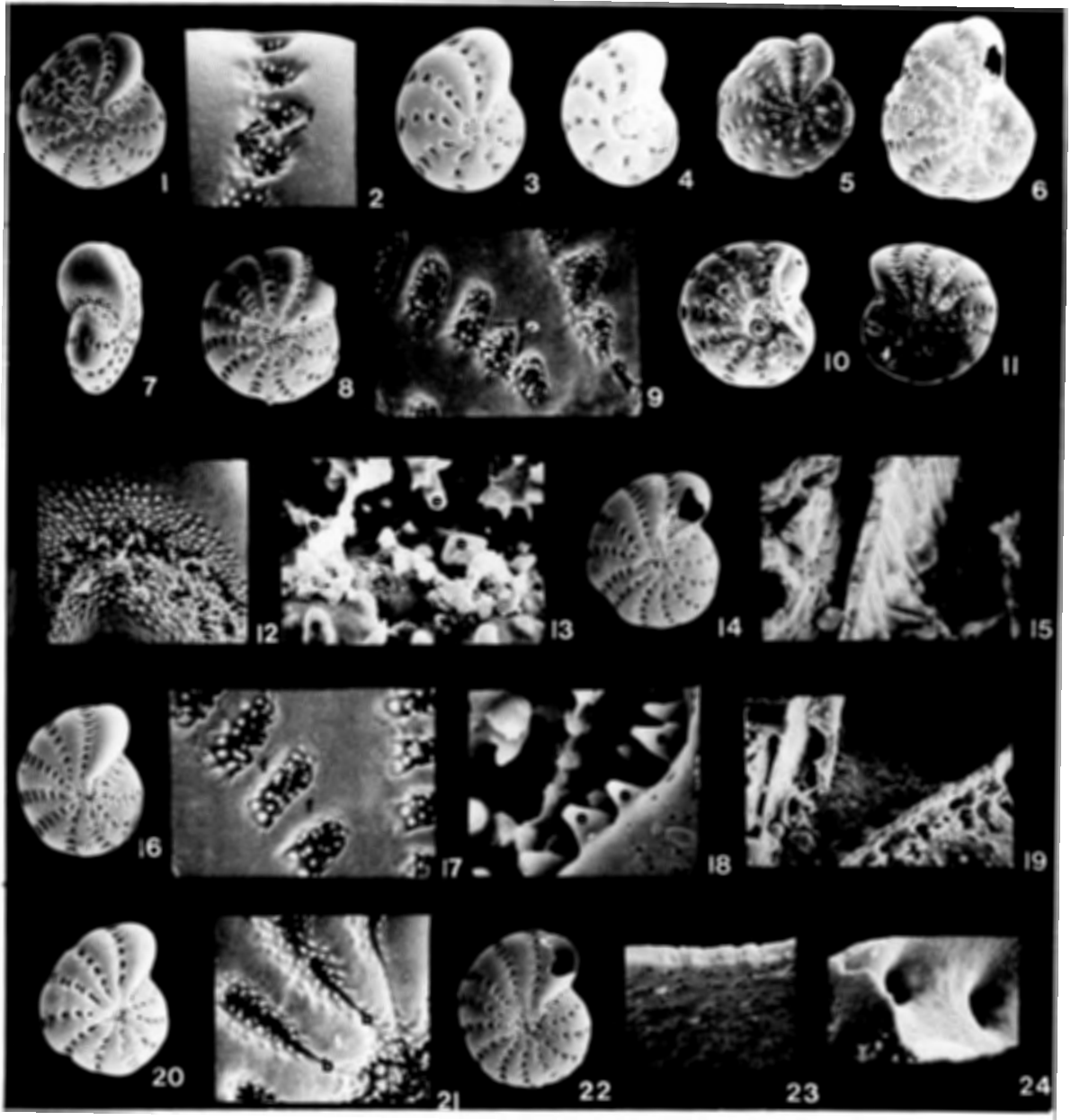


PLATE 22

Elphidium excavatum forma selsevensis (Heron-Allen and Earland).

1-3. Recent specimens from Long Island Sound. Note the development (particularly the thickness) and regularity of the ponticuli, and the fine papillae in the umbilici and intercameral sutures. 1-3. Equatorial views. 1. SN534 x 48. 2. SN560 x 50. 3-4. SN103. 3. x 73. 4. Detail of the umbilicus, umbilical papillae, chamber extentions into the umbilicus, and intercameral sutures extending into the umbilicus, x 427. 5-6. SN567. 7-8. SN98. 5,7. Equatorial views, x 58, and x 52, respectively. 6,8. Enlargements of intercameral sutures filled with thick imperforate ponticuli and papillae, so the suture is only visible in the horizontal slits (8); x 479, and x 261, respectively.

9-13. Equatorial views. 9. SN553 x 77. 10. SN562 x 73. 11. SN557 x 67. 12. SN558 x 69. 13. SN101 x 72.

14-18. Holocene specimens from Baie Verte, Northumberland Strait. 14-15. SN149. 14. Equatorial view, x 88. 15. Detail of the umbilical region, showing umbilical papillae and bosses, and the perforate chamber wall, x 226. 16. Equatorial view, SN647 x 68.

17-18. SN139, oblique-axial view, penultimate septal face (ultimate chamber removed) showing previous interiomarginal apertures, x 54, and x 128, respectively.

19. Recent specimen, from Chezzetcook Inlet, SN200 equatorial view, x 59.

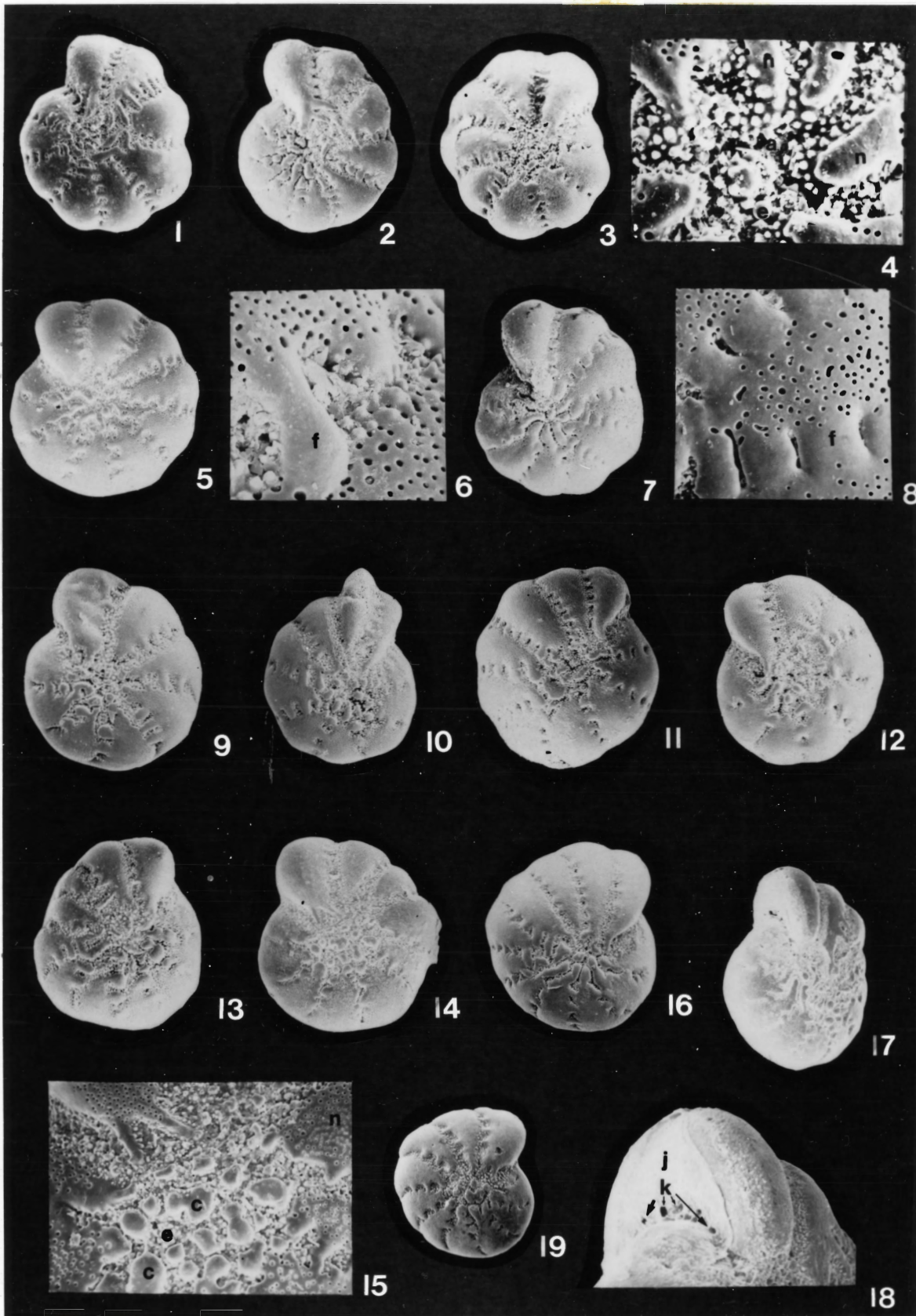


PLATE 23

Elphidium excavatum forma clavata Cushman. Note the smooth peripheral outline, curved depressed intercameral sutures, complete or incomplete umbilical boss(es) and imperforate umbilical collar. The ponticuli are irregular and often poorly developed.

1. Late Pleistocene specimen from Hirtshals, Denmark, equatorial view, SN89 x 76.

2-3. Late Pleistocene specimens from the Champlain Sea, equatorial views. 2. SN484 x 110. 3. SN492 x 108.

4-7. Recent specimens from Miramichi Estuary, New Brunswick. 4-5. SN515. 6-7. SN521. 4,6 Equatorial views. Note coarser porosity and large umbos, 108, and x 119, respectively. 5,7. Enlargement of the umbilical regions. 5. Note the pores in the umbilical boss, x 432. 7. Enlargement of the imperforate umbilical collar, showing fusion of the chamber ends and abrupt truncation of the intercameral sutures, x 475.

8-11,14-19. Holocene specimens from Baie Verte, Northumberland Strait. 8-10. Specimen beginning to approach forma galvestonensis, SN142. 8. Equatorial view, x 90. 9. Enlargement of the umbilical region, x 276. 10. Detail of the umbilicus, showing the smooth imperforate umbo and intercameral suture ending against the imperforate umbilical collar, x 422. 11. Equatorial view, SN229 x 149. 14. Equatorial view, SN246 x 84. 15-17. SN147. 15. Equatorial view, x 79. 16. Umbilical-apertural equatorial view, x 273. 17. Enlargement of the umbilical region with complete ring of papillae, x 259. 18-19. Equatorial views. 18. SN649 x 82. 19. SN658 x 90.

12. Recent specimen from the Annapolis Basin, equatorial view,
SN513 x 104.

13. Recent specimens from the Bay of Chaleur, equatorial view,
SN281 x 89.

20-21. Recent specimens from San Diego Bay, equatorial views.

20. SN368 x 108. 21. SN301 x 139.

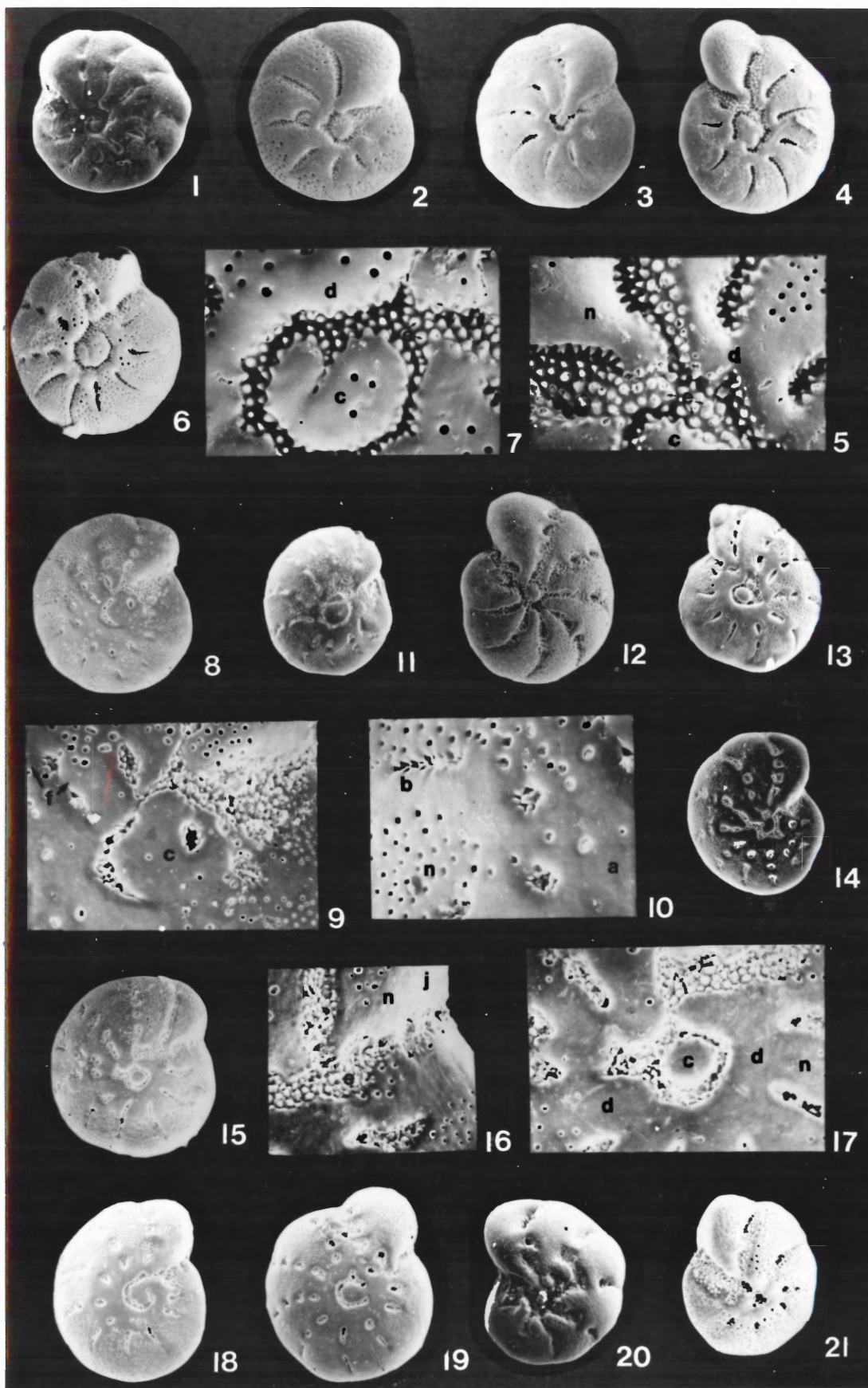


PLATE 24

Elphidium excavatum forma gunteri Cole.

A rotund, inflated form, small to medium sized, with a coarsely perforate wall. The sutures are straight, not depressed, and marked by many regular, raised, rectangular shaped ponticuli. The umbilicus contains papillae/irregular bosses.

1-12. Recent specimens from San Antonio Bay, Texas. 1-3. SN744.
 1. Equatorial view, x 96. 2-3. Detail of the chamber walls, illustrating the coarse porosity and umbilical bosses and papillae.
 2. "Poorly" developed ponticuli on the ultimate intercameral suture, x 251. 3. "Strongly" developed ponticuli on earlier intercameral sutures of the same specimen. The ponticuli are so closely spaced that the suture appears as a horizontal slits between ponticuli, x 274. 4,8. SN696. 4. Equatorial view, x 62. 8. Unusual ponticuli, appearing as bosses in the intercameral suture, x 419. 5-7. Representative plesiotype, SN745. 5. Equatorial view, specimen with raised umbilicus, x 91. 6. Oblique-axial view of the apertural face, showing the foramen and the ponticuli extending across the periphery, x 94. 7. Detail of the interiomarginal apertural arches, x 350.
 9-10. Equatorial views. Note the width of the ponticuli as a contrast to the width of the chamber wall, 9. SN730 x 57. 10. SN743 x 59. 11-12. SN715. 11. Equatorial view, x 82. 12. Peripheral-umbilical view showing detail of the intercameral sutures and umbilicus, x 250.

13. Recent specimen from San Diego Bay, equatorial view, x 131.

14-20. Holocene specimens from Baie Verte, Northumberland

Strait. 14-16. SN154. 14. Equatorial view, x 106. 15-16. Detail of (15) the chamber wall, x 370; and (16) the umbilicus, x 419. 17-18. SN153. 17. Equatorial view, x 98. 18. Enlargement of umbilical bosses and papillae, x 343. 19-20. Equatorial views. 19. SN670 x 85. 20. An intermediate specimen, providing a morphological link between forma clavata and forma gunteri, SN231 x 83.

21-24. Equatorial views. The ponticuli are not as well developed on these specimens. 21. Late Pleistocene specimen from San Francisco Bay, SN773 x 133.

22-24. Recent specimens from Venice Lagoon. 22. SN634 x 137. 23. SN366 x 97. 24. SN381 x 157.

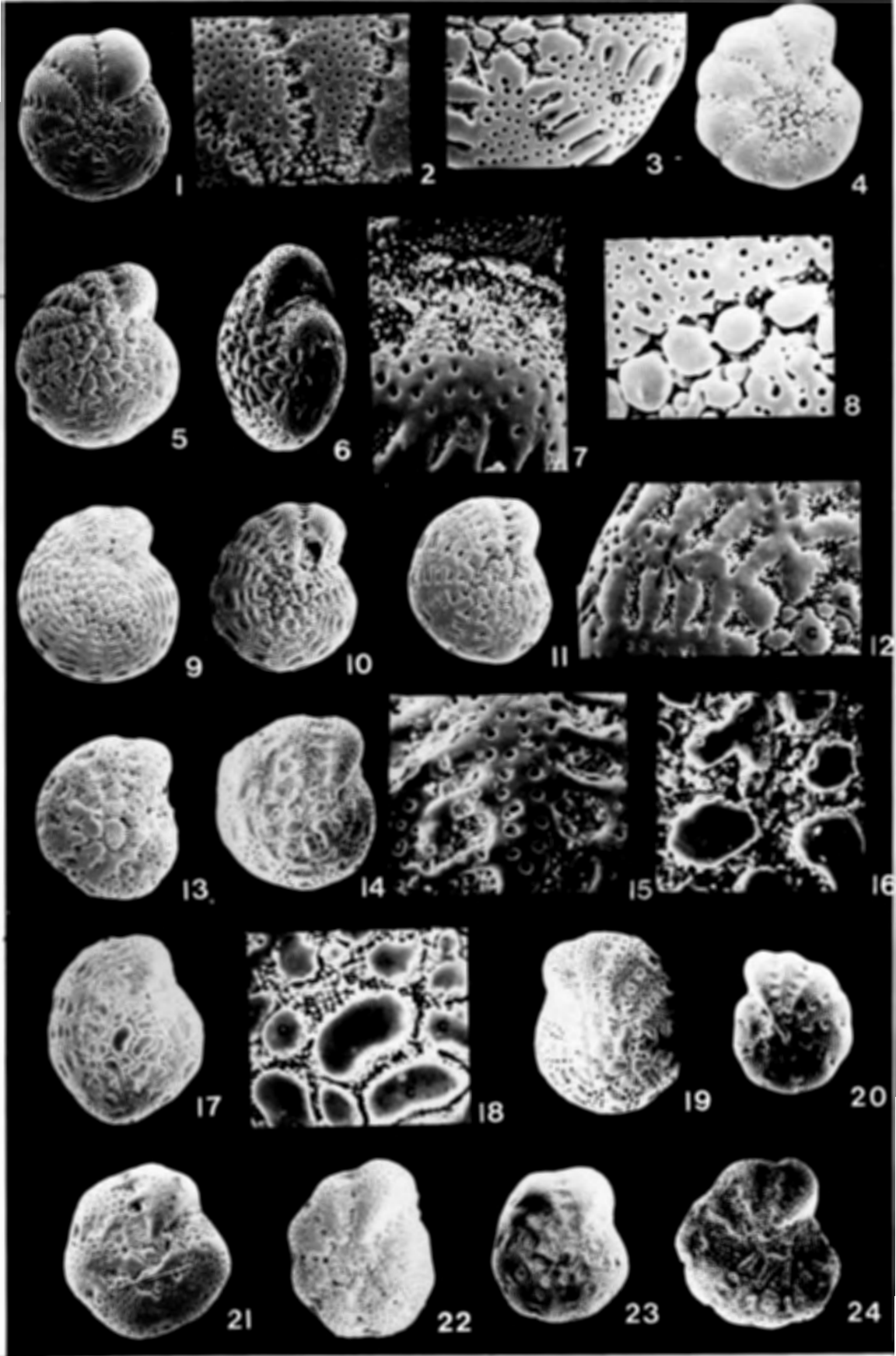


PLATE 25

Elphidium excavatum forma galvestonensis, Kornfeld.

This is a large, many chambered umbonate form, with many regular, distinct ponticuli. There may be a ring of papillae surrounding the boss or in the sutures. The wall is heavily calcified and very finely perforate.

1-10. Recent specimens from San Antonio Bay. 1-3.

Representative plesiotype, SN795. 1. Equatorial view, x 51. 2.

Detail of the intercameral suture, chamber wall, and ponticuli, x 325.

3. Enlargement of an etched ponticuli, showing fine porosity extending across the ponticuli, x 1298. 4-7. SN699. 4. Equatorial view, x 61.

5. Enlargement of the umbo, x 306. 6. Detail of papillae in a

intercameral suture, x 1088. 7. Detail of papillae on the chamber

wall, x 374. 8-9. SN731. 8. Equatorial view, x 151. 9. Intercameral

suture, papillae, and umbilical aperture. 10. Equatorial view, SN701

x 86.

11-14. Holocene specimens from Baie Verte, Northumberland Strait.

11. Etched specimen, equatorial view, SN669 x 225. 12,14. Detail of

fine wall porosity and imperforate ponticuli. 12. SN161 x 364. 14.

SN160 x 282. 13. Equatorial view, SN160 x 94.

15-16. Recent specimens, tentatively identified as this forma;

from San Diego Bay, equatorial views. 15. SN324 x 151. 16. SN302

x126.

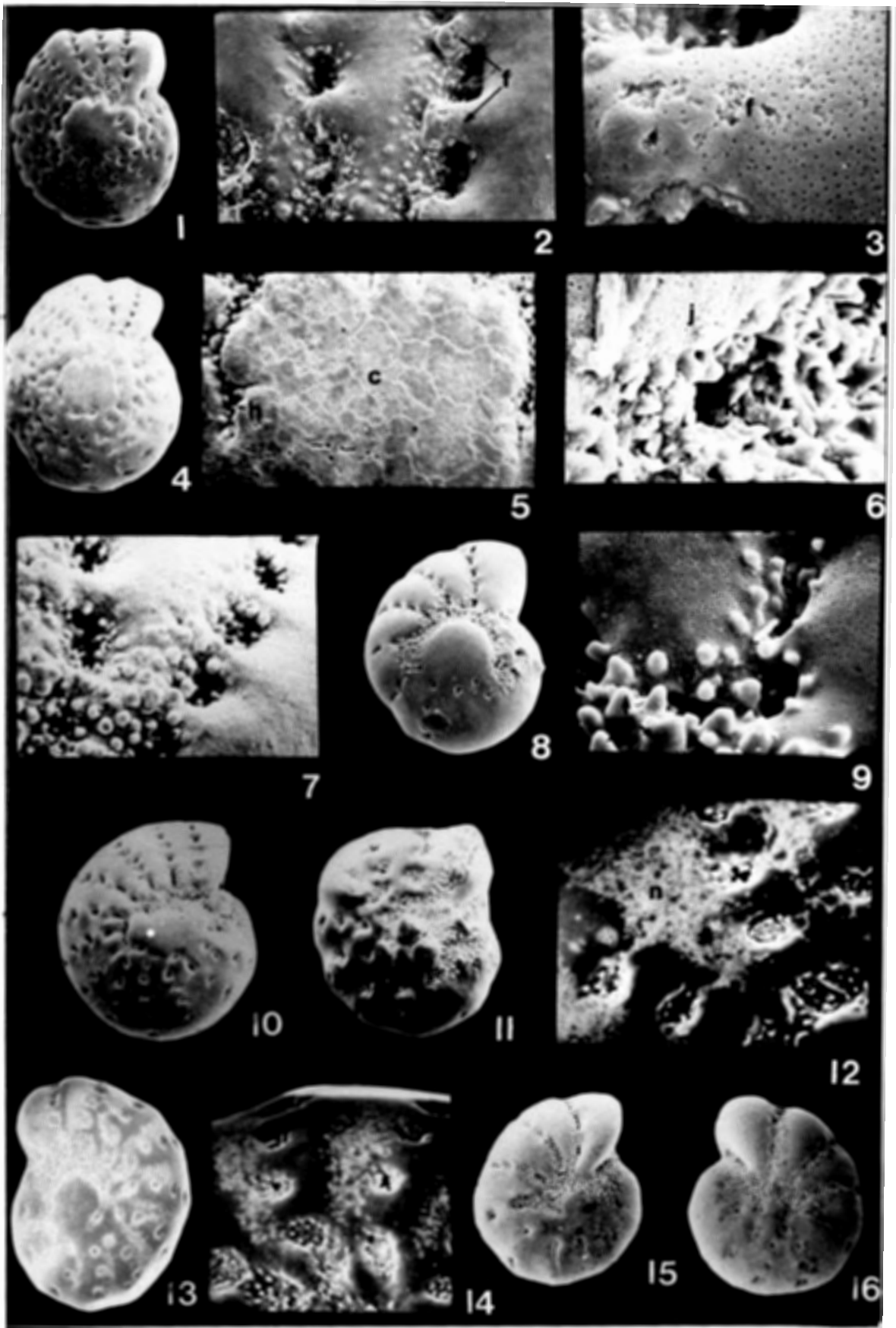


PLATE 26

Elphidium excavatum forma lidoensis Cushman. This is a smaller form, with a large open umbilicus filled with papillae/umbilical bosses. The sutures are backwards curved, distinctly broadening towards the umbilicus and also filled with papillae. The ponticuli are not generally well developed. Within this forma there are two "subforma".

1-2. E. excavatum forma lidoensis, "boreal" form. This form grades into forma excavata, the wall perforations are fine, and the papillae are small. The ponticuli are more strongly developed on this form.

1-3. Recent specimen from Annapolis Basin, SN509. 1. Equatorial view, x 68. 2. Detail of the umbilicus showing umbilical apertures, x 229.

3-5, 10-11. Recent specimens from Long Island Sound. Note the fine porosity, sutures broadening towards the umbilicus, and fine papillae. 3,5. Equatorial views. 3. SN104 x 64. 4. SN545, detail of the umbilical papillae, x 265. 5. SN545, equatorial view, x 71. 10-11. Oblique-axial view of penultimate septal face (ultimate chamber removed), SN546. 10. x 65. 11. Enlargement of the interiomarginal apertural arches and umbilical aperture, x 160.

6. Recent specimen from Chezzetcook Inlet, equatorial view, SN208 x 65.

7-9. Recent specimens from Miramichi Estuary. 7-8. Equatorial views. 7. SN517 x 80. 8. SN117 x 62. 9. Detail of umbilicus, SN117 x 217.

12-13. Holocene specimens from Baie Verte, equatorial views. 12. SN656 x 78. 13. SN639 x 88.

14-32. *E. excavatum* forma lidoensis, "Lusitanian" form. This form grades into forma gunteri. The periphery is rounded, porosity coarse, and ponticuli not as well developed as in the "boreal" form.

14-19. Recent specimens from San Antonio Bay. 14-16. Representative plesiotype of the "Lusitanian" form, SN717. 14. Equatorial view, x 87. 15. Detail of sutures broadening towards the umbilicus. Compare to figs. 4 and 9; there are less papillae in the sutures of this form, x 271. 16. Enlargement of the umbilicus, note papillae on the edges of the bosses, x 271. 17-18 SN674. 17. Equatorial view, x 155. 18. Umbilical-apertural view, x 421. 19. Equatorial view, x 75.

20-22, 27. Recent specimens from San Diego Bay. 20-21. Equatorial views. 20. Intermediate specimen, approaching forma tumidum. Note closed intercameral sutures and lack of papillae, SN359 x 97. 21-22, 27. SN306. 21. Note closed intercameral sutures, x 120. 22. Detail of suture, x 622. 27. Umbilical-apertural view, x 480.

23-25. Recent specimens from Venice Lagoon. 23-24. Equatorial views. 23. SN620 x 90. 24. SN364, note umbilical bosses, x 46. 25. Enlargement of same bosses, x 309.

26. Late Pleistocene specimen from San Francisco Bay, equatorial view, SN762 x 96.

28-32. Late Pleistocene specimens from Bay of Izmir, Turkey. 28-31. SN579. 28. Equatorial view, x 51. 29. Detail of intercameral suture, lacking papillae and broadening towards the umbilicus, x 368.

30. Detail of another suture with papillae, x 275. 31. Umbilical
bosses with papillae, x 342. 32. Equatorial view, SN612 x 64.

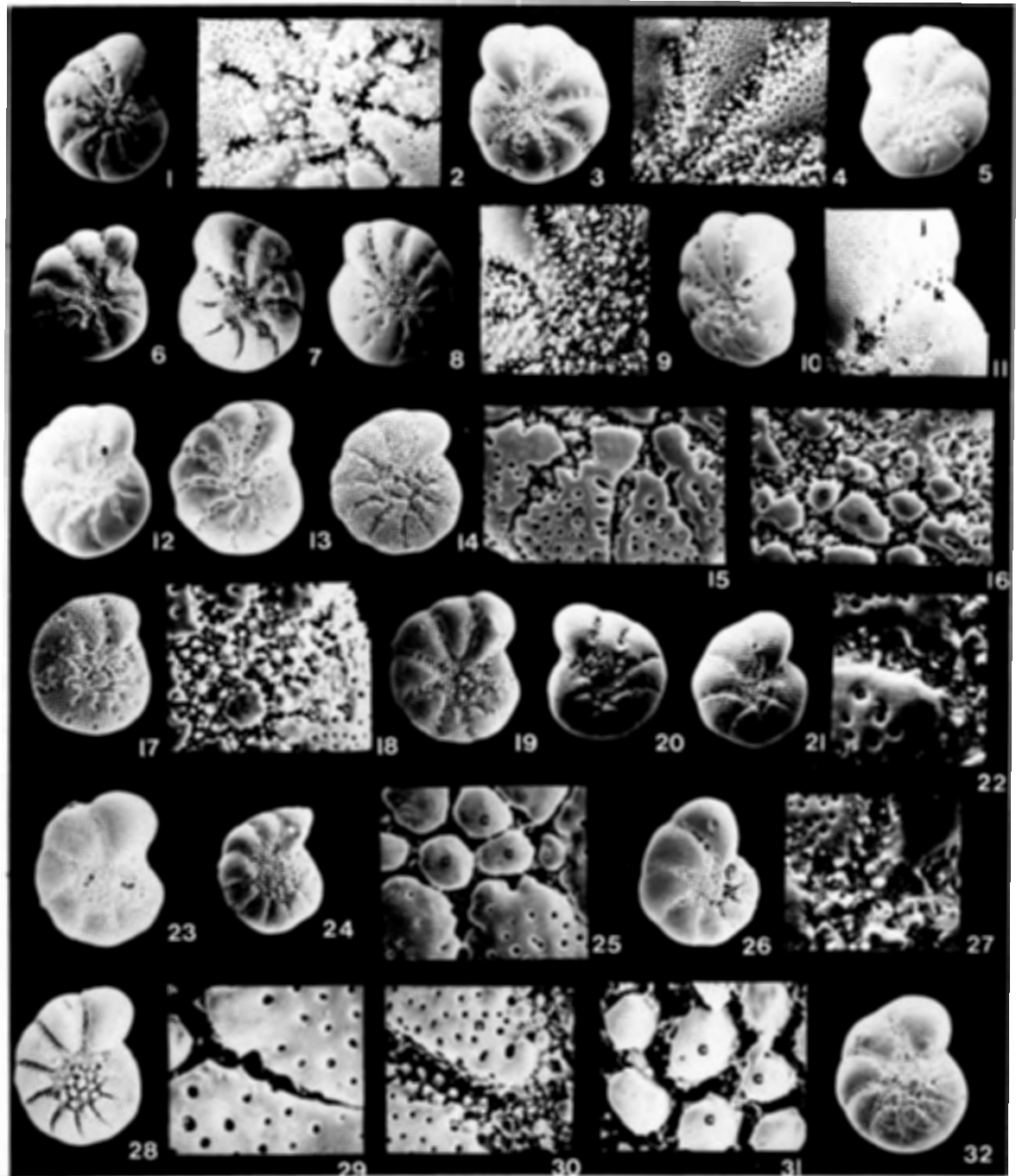


PLATE 27

1-11. Elphidium excavatum forma magna. This form is recognized by its larger size, smooth peripheral outline, subacute periphery, and strongly convex walls, which give the umbilicus a raised appearance. The umbilicus is usually large and filled with one knobby boss. The sutures are backwards curved, and some (or all) may be constricted before reaching the umbilicus.

1-4. Recent specimens from Chezzetcook Inlet. 1-3. Equatorial views. 1. SN214 x 95. 2. SN198 x 62. 3. SN899 x 78. 4. Axial view, illustrating the subacute periphery, SN898 x 84.

5-8,11. Recent specimens from the Scotian Shelf. 5,7,11. Equatorial views. 5-6. SN911. 5. x 70. 6. Enlargement of the umbilicus showing ring of papillae inside the imperforate collar, x 58. 7-8. SN906. 7. x 299. 8. Detail of the intercameral sutures ending in the umbilicus, x 198. 11. SN910 x 60.

9-10. Late Pleistocene specimens from the Champlain Sea. 9. Oblique-axial view of the penultimate septal face (ultimate chamber removed) showing an interiomarginal apertural arch, SN603 x 151. 10. Equatorial view, SN486 x 107.

12-19. Elphidium excavatum forma tumidum. This is a large, ornamented form resembling forma selseyensis, but the ornamentation and ponticuli are much more regular on forma tumidum. The umbilicus is large, circular, depressed, and filled with papillae/bosses. The chamber extensions into the umbilicus are truncated sharply. The periphery is broadly rounded and the chambers inflated. Recent specimens from San Diego Bay. 12-13. SN355. 12. Equatorial view, x

79. 13. Enlargment of the umbilicus, x 394. 14. Equatorial view, SN317 x 107. 15. Representative plesiotype of the forma, SN360 x 85. 16. Equatorial view, SN304 x 94. 17. Equatorial view, SN315 x 84. 18-19. Etched specimen, SN351. 18. Equatorial view, x 84. 19. Detail of etched umbilicus, x 420.

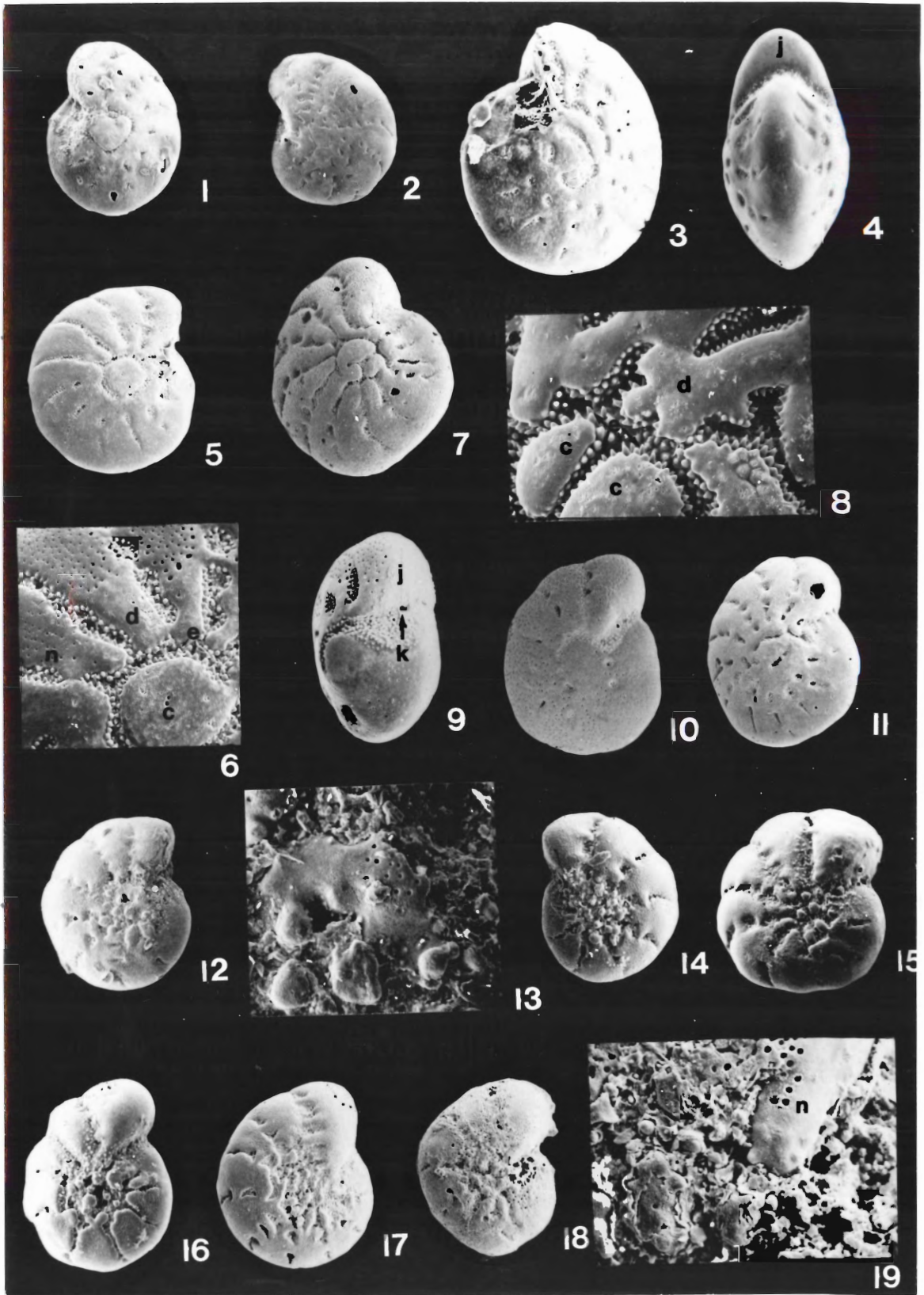


PLATE 28

Elphidium excavatum forma cuvillieri Lévy. This is a smooth, round disc shaped Elphidium about the same size as form clavata. The peripheral outline can range from smooth to very lobate. The sutures are straight or gently backwards curved, and characterized by very regular rows of sutural pores.

1-10,12,15. Late Pleistocene specimens from the Bay of Izmir, Turkey. 1,3,5,6,8,10,15 Equatorial views. 1-2. SN548. 1. x 70. 2. Enlargement of the umbilicus, x 220. 3-4. SN576. 3. x 81. 4. Umbilical-axial view (turned sideways); interiomarginal apertural arches visible, x 306. 5. SN577 x 71. 6-7. SN586. 6. x 75. 7. Enlargement of the intercameral suture, appearing as slits or pores between ponticuli. 8. Representative plesiotype, SN347 x 74. 9. SN584 x 98. 10. SN591 x 89. 12. SN580, (pl. 19:14), enlargement of the umbilicus showing apertural aperture, x 312. 15. SN344 x 62.

11,13-14,16-21. Recent specimens from Venice Lagoon. 11. Enlargement of a perforate umbilicus, SN368 (pl. 18:8), x . 13-14. SN376. 13. Peripheral (axial) view of the penultimate septal face, (ultimate chamber removed), x 64. 14. Detail showing the interiomarginal apertural arches and retral process pits, x 207. 16-18. SN380. 16. Equatorial view, x 411. 17. Enlargement of the umbilicus, x 155. 18. Detail of the intercameral suture, x 218. 19. Equatorial view, SN371 x 62. 20-21. SN379. 20. Equatorial view, x 62. 21. Ponticuli and an intercameral suture appearing as sutural pores, x 339.

22. Recent specimen from San Antonio Bay, equatorial view, SN742

x 89.

23. Recent specimen from Chezzetcook Inlet, equatorial view,
SN205 x 80.

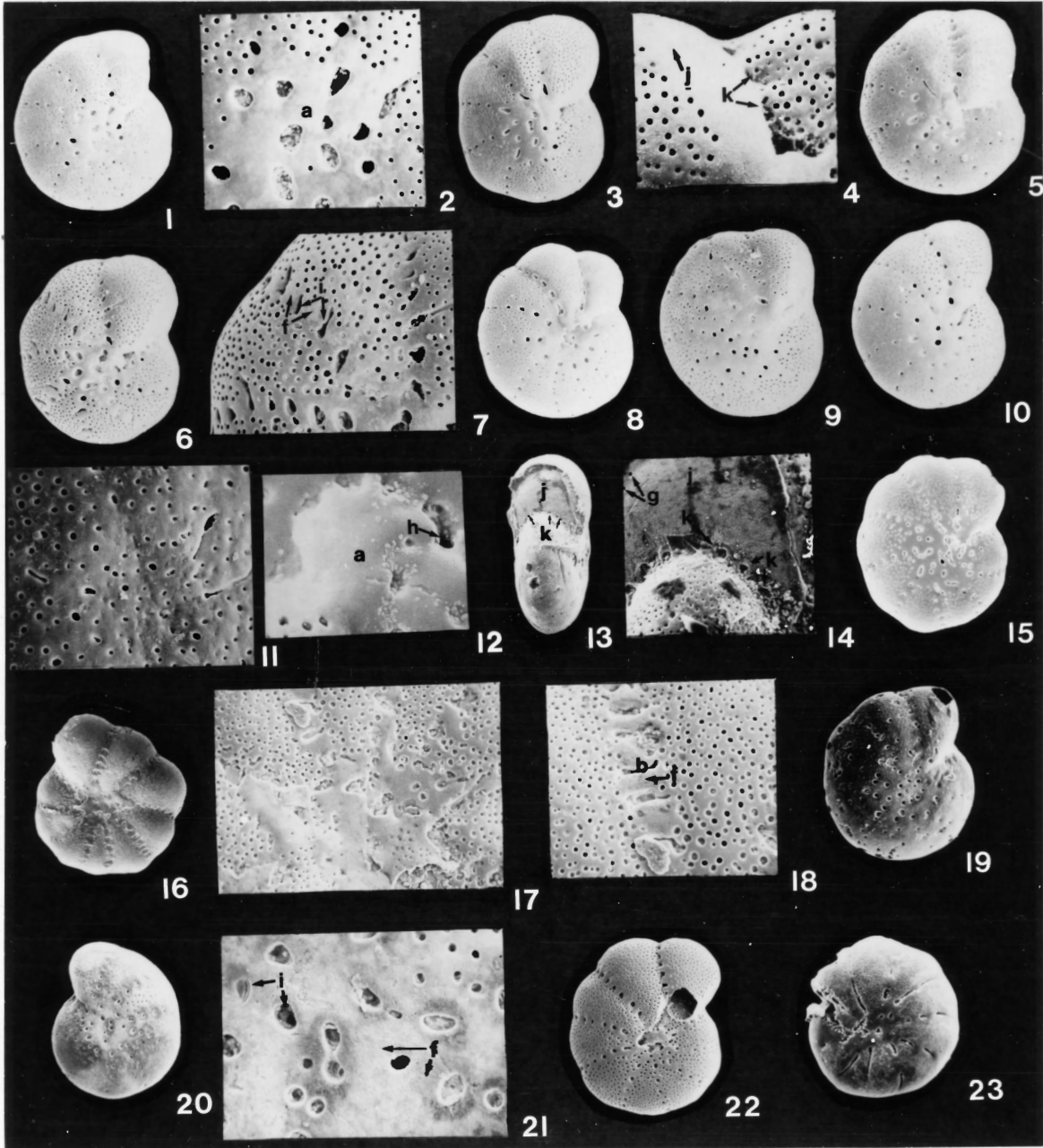


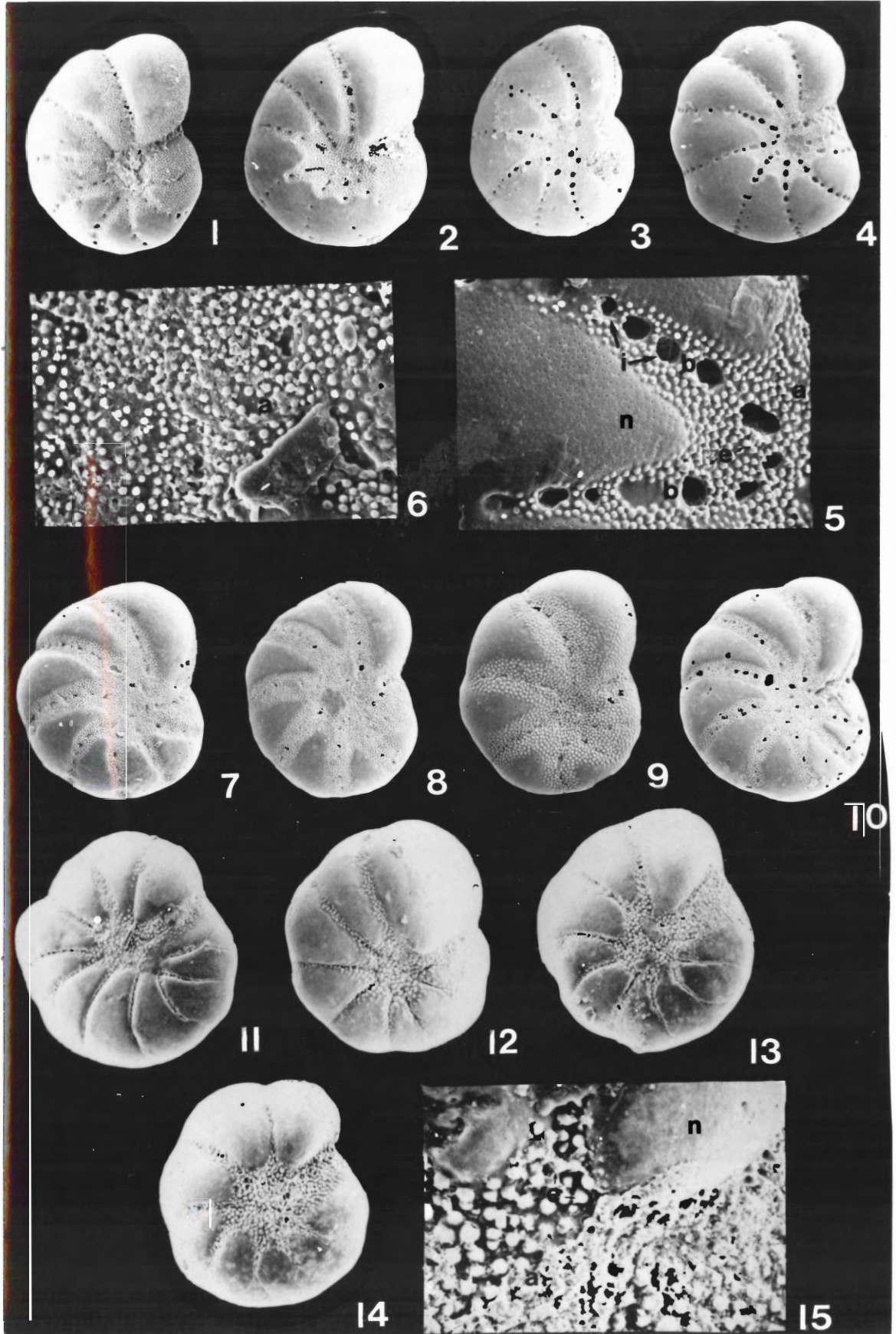
PLATE 29

Three other species of Elphidiidae tested in the statistical analysis.

1-5. Recent specimens of Elphidium bartletti Cushman from the Scotian Shelf. 1-4. Equatorial views. 1. SN865 x 78. 2. SN878 x 88. 3. SN863 x 58. 4-5. SN888. 4. x 59. 5. Enlargement of the sutural pores, x 294.

6-10. Recent specimens of Elphidium subarcticum Cushman from the Scotian Shelf. 6-7. SN844. 6. Detail of the umbilicus, x 464. 7. Equatorial view, x 69. 8-10. Equatorial views. 8. SN830 x 76. 9. SN857 x 113. 10. SN884 x 78.

11-15. Late Pleistocene specimens of Haynesina orbiculare (Brady) from the Champlain Sea. 11-14. Equatorial views. 11. SN811 x 84. 12. SN801 x 105. 13. SN812 x 94. 14. SN818 x 104. 15. Enlargement of the umbilicus, x 540.



APPENDIX A: TEST and Foraminiferal (raw) data, as measured from
specimen photographs.

Table A1: TEST foraminiferal data.

Tables A2 and A3: Foraminiferal (raw) data used in the statistical analysis. Columns numbered 1-18 represent the following variables:

Column	Variable
1	LOC
2	SPNO (SN)
3	FORM
4	PAP
5	UMCO
6	DEUM
7	POR
8	AOMA
9	NOBO
10	PERO
11	DEPO
12	SUT
13	CHAM
14	PONT
15	POSU
16	GSD
17	GS90
18	REPO

POSU and GSR were computed internally; POSU by taking the square root of PONT and GSR by dividing GS90 by GSD.

Table A2: Elphidium excavatum (Terquem) data used in the analysis.

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
18	719	6	1	0	0	2	1	1	1	3	2	3	3	3	277	244	3	3
18	720	7	1	1	0	0	1	1	1	1	1	1	1	1	554	443	3	3
18	721	0	1	0	0	0	1	1	1	1	1	1	1	1	540	453	3	3
18	722	4	1	1	0	0	1	1	1	1	1	1	1	1	245	273	1	1
18	723	6	1	1	0	0	1	1	1	1	1	1	1	1	253	207	3	3
18	724	6	1	1	0	0	1	1	1	1	1	1	1	1	35	250	2	2
18	725	8	1	1	0	0	1	1	1	1	1	1	1	1	273	224	4	4
18	726	8	1	1	0	0	1	1	1	1	1	1	1	1	283	225	3	3
18	727	8	1	1	0	0	1	1	1	1	1	1	1	1	240	215	5	5
18	728	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	729	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	730	0	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	731	0	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	732	6	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	733	6	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	734	7	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	735	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	736	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	737	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	738	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	739	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	740	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	741	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	742	0	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	743	6	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	744	6	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	745	6	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	746	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	747	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	748	8	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	750	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	795	7	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	796	7	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	797	7	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
18	900	7	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	771	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	901	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	902	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	903	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	907	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	908	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	909	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
19	911	1	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	751	2	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	752	2	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	753	3	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	754	3	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	756	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	759	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	760	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	761	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	762	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	763	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	764	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	765	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	766	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	767	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	768	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	770	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	773	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	774	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	775	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	776	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	777	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	778	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	779	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	780	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3
20	781	4	1	1	0	0	1	1	1	1	1	1	1	1	273	223	3	3

Table A3: Data of three additional species of Elphidiidae used in the analysis. FORM 11 is Haynesina orbiculare, 12 is Elphidium bartletti and 13 is Elphidium subarcticum.

APPENDIX B: DERIVATION OF FUNCTIONS

DERIVATION OF THE DISCRIMINANT FUNCTION

The statistical decision rule that forms the basis of discriminant analysis, is that a ratio of the between group to the within group variance, of a linear combination of variables, be formed, and maximized. The ratio is:

$$\lambda = \frac{W'BW}{W'W} \quad (\text{equation 4})$$

where

B is the between groups variance-covariance matrix and

W is the within groups variance-covariance matrix.

The latter can be expressed as:

$$W = \frac{A_1 + A_2 + \dots + A_g}{n_1 + n_2 \dots n_g - g} \quad (\text{equation 5})$$

where

g is the number of groups

n_i is the number of specimens in the i^{th} group and

A_i is the sum of squares and cross product matrix for the i^{th} group.

A_i can be represented as:

$$A_i = \begin{bmatrix} a_{11} & a_{21} \dots a_{1p} \\ a_{21} & a_{22} \dots a_{2p} \\ a_{p1} & a_{p2} \dots a_{pp} \end{bmatrix}$$

where

p is the number of variables

and

$$a_{1p} = \sum_{j=1}^{n_i} (x_{1j} - \bar{x}_1)(x_{pj} - \bar{x}_p)$$

where

\bar{x}_1 is the mean for the 1st variable in the i th group.

A_i can also be expressed as:

$$A_i = X_i'X_i - n_i\bar{x}_i\bar{x}_i' \quad (\text{equation 6})$$

where

\bar{x}_i is the mean vector for the i th group

X_i is the independent variables matrix ($n_i \times x_i$) and

n_i is the number of specimens in the i th group.

Equation 6 will be used to calculate A for each group and then these values substituted into equation 5 to calculate W . W will then be used in equation 4. The between groups variance-covariance matrix B can be calculated as (Gnanadesikan 1977):

$$B = \frac{1}{g-1} \sum_{i=1}^g n_i (x_i - \bar{x}_{..}) (x_i - \bar{x}_{..})' \quad (\text{equation 7})$$

where

$\bar{x}_{..}$ is the pooled variable means vector.

$$\text{To maximize } \lambda = \frac{W'BW}{W'W},$$

$\frac{d\lambda}{dW}$ is set to zero and the resulting equations are solved

for W .

The derivative is

$$\frac{d\lambda}{dW} = \frac{W'W W 2BW - W'BW 2W W}{(W'W W)^2}$$

$$= \frac{2B\underline{W}}{\underline{W}'\underline{W}\underline{W}} - \frac{\underline{W}'B\underline{W}}{\underline{W}'\underline{W}\underline{W}} \times \frac{2\underline{W}\underline{W}}{\underline{W}'\underline{W}\underline{W}}$$

$$= 2 \left(\frac{B\underline{W}}{\underline{W}'\underline{W}\underline{W}} - \frac{\underline{W}\underline{W}}{\underline{W}'\underline{W}\underline{W}} \right)$$

Equating this to zero gives

$$B\underline{W} - \lambda \underline{W}\underline{W} = 0$$

$$\text{or } (B - \lambda \underline{W})\underline{W} = 0$$

$$(W^{-1}B - \lambda \underline{I})\underline{W} = 0$$

$$W^{-1}B\underline{W} = \lambda \underline{W} \quad (\text{equation 8})$$

Therefore, λ is the largest eigenvalue of the matrix $W^{-1}B$, and the discriminant function coefficients are contained in the corresponding eigenvector \underline{W} . Now $W^{-1}B$ is not necessarily a symmetric matrix, and many programs including MINITAB cannot calculate the eigenvectors and values of a non-symmetric matrix. However, the problem can be re-expressed using two algebraic manipulations so that λ and \underline{W} can be obtained from eigenvectors and eigenvalues of a symmetric matrix. The first manipulation is to find a matrix R such that $W = R'R$. But W is symmetric so it can be written as the product:

$$W = TDT'$$

where

D is a diagonal matrix with the eigenvalues of W on the diagonal and T has the eigenvectors of W in its columns.

$$\text{Therefore } R'R = TD^{1/2}D^{1/2}T'$$

where

$D^{1/2}$ has the square root of the eigenvalues of W on its diagonal and R is chosen to be:

$$R = D1/2T' \quad (\text{equation 9})$$

The second is to let $\underline{\delta} = R\underline{w}$, so that $\underline{w} = R^{-1}\underline{\delta}$ and

$$\begin{aligned} \lambda &= \frac{\underline{\delta}'(R-1)'B(R-1)\underline{\delta}}{\underline{\delta}'(R-1)'R'R(R-1)\underline{\delta}} \\ &= \frac{\underline{\delta}'(R-1)'B(R-1)\underline{\delta}}{\underline{\delta}'\underline{\delta}} \quad (\text{equation 10}) \end{aligned}$$

To maximize this ratio, set $\frac{d\lambda}{d\underline{\delta}} = 0$ and solve.

The $\underline{\delta}$ which maximizes this ratio can be converted to the desired \underline{w} using the fact that $\underline{w} = R^{-1}\underline{\delta}$.

Now

$$\begin{aligned} \frac{d\underline{\delta}}{d\lambda} &= \frac{\underline{\delta}'\underline{\delta} \times 2(R-1)'B(R-1)\underline{\delta} - 2\underline{\delta}'(R-1)'B(R-1)\underline{\delta}}{(\underline{\delta}'\underline{\delta})^2} \\ &= \frac{2(R-1)'B(R-1)}{\underline{\delta}'\underline{\delta}} - \frac{2\underline{\delta}}{\underline{\delta}'\underline{\delta}} \times \frac{\underline{\delta}'(R-1)'B(R-1)}{\underline{\delta}'\underline{\delta}} \\ &= \frac{2(R-1)'B(R-1)}{\underline{\delta}'\underline{\delta}} - \frac{2\underline{\delta}\lambda}{\underline{\delta}'\underline{\delta}} \end{aligned}$$

Equating to zero gives

$$[(R-1)'B(R-1) - I]\underline{\delta} = 0 \quad (\text{equation 11})$$

$$\text{or } (R-1)'B(R-1)\underline{\delta} = \lambda\underline{\delta}.$$

Therefore λ is the largest eigenvalue of $(R-1)'BR^{-1}$ and $\underline{\delta}$ is the corresponding eigenvector. In general, $(R-1)'BR^{-1}$ has $g-1$ nonzero eigenvalues and corresponding eigenvectors. The second and subsequent largest eigenvectors gives the second and subsequent discriminant functions. Writing the eigenvectors of $(R-1)'BR^{-1}$ in the columns of a matrix Γ , the discriminant function coefficients are obtained by

pre-multiplying by R^{-1} .

To summarize:

$$\frac{\underline{W}'\underline{B}\underline{W}}{\underline{W}'\underline{W}\underline{W}} = \lambda = \frac{\underline{\gamma}'(R^{-1})\underline{B}R^{-1}\underline{\gamma}}{\underline{\gamma}'\underline{\gamma}}$$

$$\underline{W} = R'R$$

$$\underline{W}'\underline{W}\underline{W} = \underline{W}'R'R\underline{W} = \underline{\gamma}'\underline{\gamma}$$

$$\underline{\gamma} = R\underline{W}$$

$$\underline{W} = R^{-1}$$

and $\underline{Z} = \underline{X}'\underline{W}$ (equation 2)

For the two group case there is only one discriminant function, which also can be expressed in terms of the pooled within groups variance-covariance matrix (W) and a difference between the group mean vectors, ie:

$$\underline{Z} = \underline{X}'\underline{W}^{-1}(\bar{\underline{X}}_i - \bar{\underline{X}}_j) \quad (\text{equation 12})$$

where:

\underline{X}' is the transpose of the independent variable vector and

$\bar{\underline{X}}_i$ is the mean vector for the i th group.

The vector of coefficients $\underline{W} = W^{-1}(\bar{\underline{X}}_i - \bar{\underline{X}}_j)$ does maximize λ as is shown below. For the two group case, equation 7 simplifies to:

$$B = \frac{1/2 n_i^2 + n_j^2}{n_i + n_j} (\bar{\underline{X}}_i - \bar{\underline{X}}_j)(\bar{\underline{X}}_i - \bar{\underline{X}}_j)'$$

$$\text{or} = k\underline{d}\underline{d}'$$

where

$$\underline{d} = \bar{\underline{X}}_i - \bar{\underline{X}}_j \quad \text{and}$$

$$k = \frac{1 n_i^2 + n_j^2}{2 n_i + n_j}$$

$$\text{To maximize } \lambda = \frac{\underline{w}'B\underline{w}}{\underline{w}'W\underline{w}}$$

it has been shown that $\underline{w} = R^{-1}\underline{\zeta}$ where $\underline{\zeta}$ is the eigenvector associated with the largest eigenvalue of $(R^{-1})'BR^{-1}$.

$$\begin{aligned} \text{But } (R^{-1})'BR^{-1} &= k(R^{-1})'d d'R^{-1} \\ &= k\underline{h}\underline{h}' \end{aligned}$$

where

$$\underline{h} = (R^{-1})'d$$

so B has the single nonzero eigenvector

$$\underline{\zeta} = \frac{\underline{h}}{\|\underline{h}\|}$$

Therefore the discriminant function coefficients are given up to a scale factor by

$$\underline{w} = R^{-1}\underline{h} = R^{-1}(R^{-1})'d = W^{-1}(\bar{x}_i - \bar{x}_j)$$

For the two group case Z will be calculated from equation 12 rather than equations 4, 7, 8, 9, 10, and 11.

DERIVATION OF THE CLASSIFICATION FUNCTION

The a priori probability that a specimen with measurements \underline{x} will belong to group i is usually taken to be $1/g$ where g is the number of groups. The probability density for \underline{x} in group i is:

$$\frac{1}{(2\pi)^{p/2}} \left\{ |V|^{-1/2} \exp \left[-1/2[(\underline{x} - \underline{\mu}_i)'V^{-1}(\underline{x} - \underline{\mu}_i)] \right] \right\}$$

where:

$\underline{\mu}_i$ is the true mean vector (which is estimated by \bar{x}_i)

V is the true within group variance-covariance matrix
 (which is estimated by W) and
 p is the number of variables.

Using Bayes rule, the posterior probability for group i is the a priori probability times the likelihood (where the likelihood has the same form as the density for \underline{x}).

Therefore the estimated ln posterior probability is

$$\ln 1/g - p/2 \ln 2\pi - 1/2 \ln |W| - 1/2 (\underline{x} - \bar{\underline{x}}_i)' W^{-1} (\underline{x} - \bar{\underline{x}}_i). \quad (\text{equation 13})$$

Expanding the last term [$1/2 (\underline{x} - \bar{\underline{x}}_i)' W^{-1} (\underline{x} - \bar{\underline{x}}_i)$] gives:

$$\begin{aligned} & 1/2 \underline{x}' W^{-1} (\underline{x} - \bar{\underline{x}}_i) - 1/2 \bar{\underline{x}}_i' W^{-1} (\underline{x} - \bar{\underline{x}}_i) \\ & = 1/2 \underline{x}' W^{-1} (\underline{x}) - 1/2 \underline{x}' W^{-1} \bar{\underline{x}}_i - 1/2 \bar{\underline{x}}_i' W^{-1} (\underline{x}) + 1/2 \bar{\underline{x}}_i' W^{-1} \bar{\underline{x}}_i. \end{aligned} \quad (\text{equation 14})$$

Omitting terms constant for all groups, (the second and third terms of equation 13, the first term of equation 14), leaves:

$$\ln \frac{1}{g} + 1/2 \underline{x}' W^{-1} \bar{\underline{x}}_i + 1/2 \bar{\underline{x}}_i' W^{-1} \underline{x} - 1/2 \bar{\underline{x}}_i' W^{-1} \bar{\underline{x}}_i \quad (\text{equation 15}).$$

In equation 15, terms 2 and 3 are constants and the transpose of one another; they combine to: $\underline{x}' W^{-1} \bar{\underline{x}}_i$.

The result is the classification function:

$$\ln \frac{1}{g} + \frac{\underline{x}' W^{-1} \bar{\underline{x}}_i - \bar{\underline{x}}_i' W^{-1} \bar{\underline{x}}_i}{2}$$

or

$$\ln \frac{1}{g} + \frac{(\underline{x} - \bar{\underline{x}}_i)' W^{-1} \bar{\underline{x}}_i}{2}. \quad (\text{equation 3})$$

The classification rule is to evaluate these g functions for each specimen, and classify the specimen with independent variables \underline{x} into

the group which gives the largest function value.

For the two group case this is equivalent to calculating the discriminant score and classifying the specimen according to its proximity to the group centroids. Applying equation 3, the specimen is classified as belonging to group one instead of group three, if:

$$\ln \frac{1}{g} + \frac{(\mathbf{x} - \bar{\mathbf{x}}_1)' \mathbf{W}^{-1} \bar{\mathbf{x}}_1}{2} - \frac{(\mathbf{x} - \bar{\mathbf{x}}_3)' \mathbf{W}^{-1} \bar{\mathbf{x}}_3}{2} - \ln \frac{1}{g} > 0 \quad (\text{equation 16}).$$

$\ln 1/g$ cancels, and expanding the remaining terms gives:

$$\mathbf{x}' \mathbf{W}^{-1} \bar{\mathbf{x}}_1 - \frac{\bar{\mathbf{x}}_1' \mathbf{W}^{-1} \bar{\mathbf{x}}_1}{2} - \mathbf{x}' \mathbf{W}^{-1} \bar{\mathbf{x}}_3 + \frac{\bar{\mathbf{x}}_3' \mathbf{W}^{-1} \bar{\mathbf{x}}_3}{2} > 0$$

$$\mathbf{x}' \mathbf{W}^{-1} (\bar{\mathbf{x}}_1 - \bar{\mathbf{x}}_3) - \frac{\bar{\mathbf{x}}_1' \mathbf{W}^{-1} \bar{\mathbf{x}}_1}{2} + \frac{\bar{\mathbf{x}}_3' \mathbf{W}^{-1} \bar{\mathbf{x}}_3}{2} > 0$$

$$\text{or } \mathbf{x}' \mathbf{W}^{-1} (\bar{\mathbf{x}}_1 - \bar{\mathbf{x}}_3) > \frac{\bar{\mathbf{x}}_1' \mathbf{W}^{-1} \bar{\mathbf{x}}_1}{2} - \frac{\bar{\mathbf{x}}_3' \mathbf{W}^{-1} \bar{\mathbf{x}}_3}{2} \quad (\text{equation 17}).$$

The left hand side is simply the discriminant score. The right hand side is the average of the two group centroids:

$$\frac{\bar{\mathbf{x}}_1' \mathbf{W}^{-1} (\bar{\mathbf{x}}_1 - \bar{\mathbf{x}}_3)}{2} + \frac{\bar{\mathbf{x}}_3' \mathbf{W}^{-1} (\bar{\mathbf{x}}_1 - \bar{\mathbf{x}}_3)}{2}. \quad (\text{equation 18})$$

Therefore it is not necessary to evaluate the classification functions but simply use the discriminant scores and the midpoint between the group centroids, the critical Z value, $(Z_1 + Z_3)/2$ to determine if a specimen is correctly classified. For two groups of equal size, if $Z > (Z_1 + Z_3)/2$ the specimen is classified as belonging to group one, if $(Z_1 + Z_3)/2 > Z$, then the specimen is classified as belonging to group three. If the groups are not of equal size, then a weighed average of the group centroids provides the optimal cutting score (Hair et al. 1979). If the group size in the analysis is irrelevant, or unrelated to the actual group size, then

the unweighed method is employed.

APPENDIX C: ILLUSTRATIVE EXAMPLE - TEST

TEST ONE

The SPSS output of the options and statistics calculated in subprogram DISCRIMINANT gives the group means, group standard deviations, and the pooled within groups variance-covariance matrix.

GROUP MEANS:

FORM	NOBO	GSD
1	1.00000	.34148
3	3.24138	.45893
TOTAL	1.45455	.36530

GROUP STANDARD DEVIATIONS:

FORM	NOBO	GSD
1	.56443	.09522
3	.73946	.09229
TOTAL	1.08594	.10555

POOLED WITHIN-GROUPS COVARIANCE MATRIX:

	NOBO	GSD	
NOBO	.3639032		
GSD	-2731719E-03	.8957548E-02	(matrix 1)

The calculation of this covariance matrix can be illustrated with the aid of MINITAB. Recall that:

$$A_1 = X_1'X_1 - n_1\bar{X}_1\bar{X}_1' \quad (\text{equation 6})$$

$$\text{and } W = \frac{A_1 + A_3}{n_1 + n - 2} \quad (\text{equation 5})$$

X_1 for group one is the 2 x 114 matrix, containing the two variables

x_1 (NOBO) and x_2 (GSD) for the 114 group one specimens used in TEST (Appendix A, Table A1).

$$\text{Now } X_1'X_1 = \begin{bmatrix} 150.000 & 39.303 \\ 39.303 & 14.318 \end{bmatrix}$$

$$\text{and } \bar{x}_1 = \begin{bmatrix} 1.0000 \\ .34148 \end{bmatrix} \quad (\text{mean vector})$$

$$\text{so } 114\bar{x}_1\bar{x}_1' = \begin{bmatrix} 114.000 & 38.929 \\ 38.929 & 13.293 \end{bmatrix}$$

$$\text{and } A_1 = X_1'X_1 - 114\bar{x}_1\bar{x}_1' = \begin{bmatrix} 36.0000 & .3743 \\ .3743 & 1.0247 \end{bmatrix} \quad (\text{matrix 2})$$

Similarly for group three, X_3 is a 2 x 29 matrix (Appendix A, Table A1). Using the same procedure as above:

$$X_3'X_3 = \begin{bmatrix} 320.000 & 42.727 \\ 42.727 & 6.346 \end{bmatrix}$$

$$\text{and } A_3 = X_3'X_3 - 29\bar{x}_3\bar{x}_3' = \begin{bmatrix} 15.3065 & -.4127 \\ -.4127 & .2385 \end{bmatrix} \quad (\text{matrix 3})$$

The pooled variance - covariance matrix is therefore:

$$W = \frac{1}{114 + 29 - 2} \times \begin{bmatrix} 51.3065 & -.0384 \\ -.0384 & 1.2632 \end{bmatrix}$$

$$= \begin{bmatrix} .363876 & -.000272 \\ -.000272 & .008959 \end{bmatrix} \quad (\text{matrix 4})$$

which compares quite well to the SPSS output (matrix 1). Based on this pooled variance-covariance matrix the discriminant function can be calculated:

$$Z = \mathbf{x}'W^{-1}(\bar{x}_1 - \bar{x}_3) \quad \mathbf{x}' = (x_1, x_2) \quad (\text{from equation 12})$$

$$Z = -6.1698x_1 - 13.2972x_2$$

The group centroid for group one is -10.711, for group three it is -26.101, and the Z values can be represented on histograms (Figures C1 and C2).

The SPSS output gives the unstandardized discriminant function coefficients:

	FUNC 1	
NOBO	1.572599	(matrix 5)
GSD	3.390245	
(CONSTANT)	-3.525875	

which can be calculated from the MINITAB coefficients by standardizing the discriminant scores to have zero mean and unit standard deviation. The mean and standard deviation of the discriminant scores for the two groups are:

$$\bar{Z}_1 = -10.711 \quad (\text{s.d.} = 3.78)$$

$$\text{and } \bar{Z}_3 = -26.101 \quad (\text{s.d.} = 4.46)$$

which give the pooled mean:

$$\frac{n_1 Z_1 + n_3 Z_3}{n_1 + n_3}$$

$$= [114(-10.711) + 29(-26.101)]/143$$

$$= -13.832$$

The pooled variance is $[113(3.78)^2 + 28(4.46)^2]/141$

$$= 15.4011$$

so the standard deviation is 3.9244.

Standardizing the discriminant scores gives:

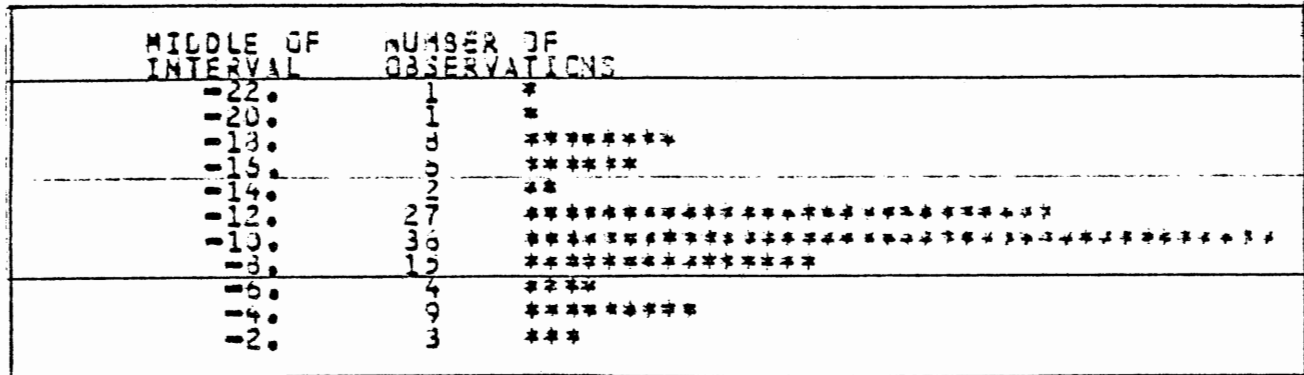


Figure C1: Histogram of group one Z values (discriminant scores).

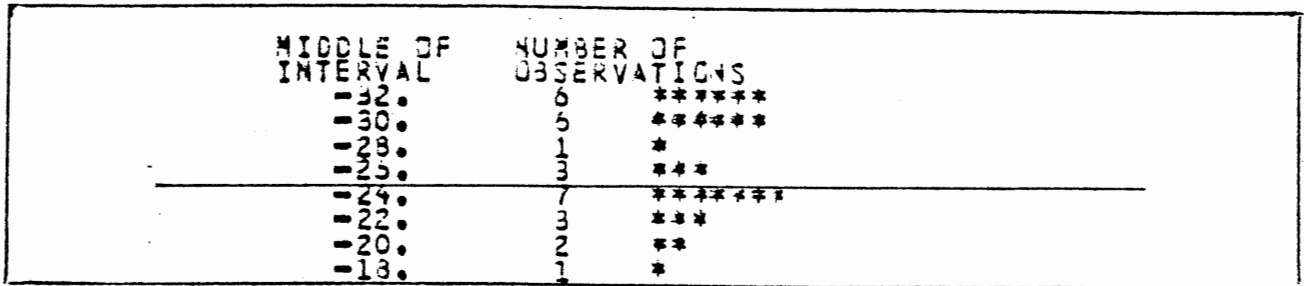


Figure C2: Histogram of group three Z values (discriminant scores).

$$\begin{aligned}
 Z_{sz} &= \frac{Z - (-13.832)}{3.9244} && \text{(equation 19)} \\
 &= \frac{-6.1698x_1 - 13.2972x_2 + 13.832}{3.9244} \\
 &= -1.5722x_1 - 3.5246x_2 + 3.3883
 \end{aligned}$$

by substituting \bar{z}_1 and \bar{z}_3 in turn into equation 19 the result is the standardized group centroids which, apart from the change in sign, is equivalent to the SPSS result.

GROUP	FUNC	1	
1	-.79557		(matrix 6)
3	3.12740		

SPSS also gives the standardized function coefficients:

	FUNC	1	
NOBO	.94866		(matrix 7)
GSD	.32087		

which can be calculated from the unstandardized functions (matrix 5)

by standardizing the independent variables as well:

$$Z_{sz} = +1.5725x_1 + 3.390245x_2 - 3.525875$$

If $x_1 = s_1y_1 + \bar{x}_1$

and $x_2 = s_2y_2 + \bar{x}_2$

where:

\bar{x} is the pooled variable mean

s is the pooled standard deviation

and

$$\bar{x}_1 = 1.45455$$

$$\bar{x}_2 = .36530$$

$$s_1 = .5999$$

$$s_2 = .0946,$$

the result is:

$$Z_S = +1.5722(.5999y_1 + 1.45455) + 3.39024(.0946y_2 + .3653) - 3.525875$$

$$Z_S = .9431y_1 + 2.2868 + .320716y_2 + 1.23845 - 3.525$$

$$\begin{aligned} Z_S &= .9431y_1 + .320716y_2 + 3.5213 - 3.525 \\ &= .9431y_1 + .320716y_2 \end{aligned}$$

which compares well with matrix 7 (above).

The SPSS output gives the classification function coefficients:

(FISHER'S LINEAR DISCRIMINANT FUNCTIONS)

FORM =	1	3	
NOBO	2.776665	8.945923	(matrix 8)
GSD	38.20699	51.50682	
(CONSTANT)	-8.604988	-27.01075	

The calculation of these functions is easily illustrated on

MINITAB. Recall the general equation for the classification function:

$$\ln \frac{1}{g} + (\mathbf{x} - \bar{\mathbf{x}}_1)' \mathbf{W}^{-1} \bar{\mathbf{x}}_1 \quad (\text{equation 3})$$

Then:

$$\mathbf{W} = \begin{bmatrix} .3639032 & -.0002731719 \\ -.0002731719 & .00895748 \end{bmatrix} \quad (\text{matrices 1 and 4})$$

$$\mathbf{W}^{-1} = \begin{bmatrix} 2.748 & .084 \\ .084 & 111.640 \end{bmatrix} \quad (\text{matrix 9})$$

$$\mathbf{W}^{-1} \bar{\mathbf{x}}_1 = \begin{bmatrix} 2.7767 \\ 38.2067 \end{bmatrix} \quad (\text{matrix 10})$$

(Classification function coefficients for group one, compare to matrix 8).

$$\frac{\bar{X}_1'W^{-1}\bar{X}_1}{2} = 7.9117$$

$$\ln .5 - \frac{\bar{X}_1'W^{-1}\bar{X}_1}{2} = -.6931471 - 7.9117 = -8.60484 \quad (\text{constant 1})$$

Similarly, for group three:

$$W^{-1}\bar{X}_3 = \begin{matrix} 8.9459 \\ 51.5067 \end{matrix} \quad (\text{matrix 11})$$

$$\frac{\bar{X}_3'W^{-1}\bar{X}_3}{2} = 26.3176$$

$$\ln .5 - \frac{\bar{X}_3'W^{-1}\bar{X}_3}{2} = -.6931471 - 26.3176 = -27.010747 \quad (\text{constant 2})$$

Combining matrices 10 and 11, and constants 1 and 2 the classification function coefficients from MINITAB are:

	GROUP 1	GROUP 3	
NOBO	2.7767	8.9459	
GSD	38.2067	51.5067	(matrix 12)
CONSTANT	-8.60484	-27.010747	

which compare very well with matrix 8.

As illustrated earlier, for a two group case, the classification function scores are closely related to the discriminant scores, and will not be evaluated separately.

Looking at the discriminant scores from SPSS (Table C1), there are two group one specimens and one group three specimen misclassified.

From the MINITAB discriminant (unstandardized) scores (Table C2) the critical \bar{Z} value is:

-10.711

-26.101

-36.812 / 2 = 18.406

There are two group one and one group three Z values lying outside

CASE	MIS	SEL	ACTUAL	HIGHEST	PROBABILITY	2ND HIGHEST	DISCRIMINANT
SUBFILE	SEQNUM	VAL	GROUP	GROUP	P(X/G)	P(G/X)	SCORES
SAMPLE1	1		1.	1	.0824	1.0000	3 .0000 -2.5325
SAMPLE1	2		1.	1	.0562	1.0000	3 .0000 -2.7054
SAMPLE1	3		1.	1	.7467	.9999	3 .0001 -1.1159
SAMPLE1	4		1.	1	.8801	.9997	3 .0003 -.9464
SAMPLE1	5		1.	1	.9960	.9996	3 .0004 -.8006
SAMPLE1	6		1.	1	.5864	.9999	3 .0001 -1.3396
SAMPLE1	7		1.	1	.8217	.9998	3 .0002 -1.0210
SAMPLE1	8		1.	1	.9716	.9995	3 .0005 -.7599
SAMPLE1	9		1.	1	.7385	.9999	3 .0001 -1.1294
SAMPLE1	10		1.	1	.8508	.9998	3 .0002 -.9837
SAMPLE1	11		1.	1	.9015	.9993	3 .0007 -.6716
SAMPLE1	12		1.	1	.8138	.9998	3 .0002 -1.0311
SAMPLE1	13		1.	1	.0842	1.0000	3 .0000 -2.4952
SAMPLE1	14		1.	1	.9526	.9994	3 .0006 -.7362
SAMPLE1	15		1.	1	.1013	1.0000	3 .0000 -2.4342
SAMPLE1	16		1.	1	.3102	1.0000	3 .0000 -1.8104
SAMPLE1	17		3.	3	.9678	.9995	3 .0005 3.0871
SAMPLE1	18		3.	3	.6561	.9974	3 .0026 -.3531
SAMPLE1	19		1.	3	.0741	.6658	3 .3342 1.3416
SAMPLE1	20		1.	1	.6729	.9976	3 .0024 -.3734
SAMPLE1	21		1.	1	.0760	.6757	3 .3243 .9788
SAMPLE1	22		1.	1	.8374	.9990	3 .0010 -.5904
SAMPLE1	23		1.	1	.9689	.9995	3 .0005 -.7585
SAMPLE1	24		1.	1	.6508	.9973	3 .0027 -.3429
SAMPLE1	25		1.	1	.8349	.9996	3 .0002 -1.0045
SAMPLE1	26		1.	1	.5501	.9996	3 .0004 -.8481
SAMPLE1	27		1.	1	.8640	.9991	3 .0009 -.6243
SAMPLE1	28		1.	1	.8256	.9998	3 .0002 -1.0108
SAMPLE1	29		1.	1	.5365	.9994	3 .0006 -.7158
SAMPLE1	30		1.	1	.0766	1.0000	3 .0000 -2.5664
SAMPLE1	31		1.	1	.8934	.9992	3 .0008 -.6616
SAMPLE1	32		1.	1	.9013	.9993	3 .0007 -.6716
SAMPLE1	33		1.	1	.9770	.9995	3 .0005 -.7667
SAMPLE1	34		1.	1	.9608	.9996	3 .0004 -.6447
SAMPLE1	35		1.	1	.8507	.9992	3 .0008 -.6582
SAMPLE1	36		1.	1	.7694	.9999	3 .0001 -1.0888
SAMPLE1	37		1.	1	.9231	.9997	3 .0003 -.8921
SAMPLE1	38		1.	1	.0349	1.0000	3 .0000 -2.9055
SAMPLE1	39		1.	1	.5911	.9999	3 .0001 -1.3329
SAMPLE1	40		1.	1	.0443	1.0000	3 .0000 -2.8071
SAMPLE1	41		1.	1	.6315	.9999	3 .0001 -1.2755
SAMPLE1	42		1.	1	.9122	.9993	3 .0007 -.6855
SAMPLE1	43		1.	1	.9068	.9993	3 .0007 -.6785
SAMPLE1	44		1.	1	.8828	.9997	3 .0003 -.9430
SAMPLE1	45		1.	1	.8775	.9998	3 .0002 -.9498
SAMPLE1	46		1.	1	.8256	.9998	3 .0002 -1.0108

Table C1: SPSS discriminant scores for TEST One. The symbol *** indicates those specimens misclassified.

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES		
SUBFILE	SEQNUM	VAL	SEL	GROUP	P(X/G) P(G/X)	GROUP P(G/X)	
SAMPLE1	47		1.	1	.8344 .9998	3 .0002	-1.0040
SAMPLE1	48		1.	1	.9825 .9996	3 .0004	-1.6175
SAMPLE1	49		1.	1	.7206 .9999	3 .0001	-1.1532
SAMPLE1	50		1.	1	.0766 1.0000	3 .0000	-2.5664
SAMPLE1	51		1.	1	.0896 1.0000	3 .0000	-2.5258
SAMPLE1	52		1.	1	.0635 1.0000	3 .0000	-2.6512
SAMPLE1	53		1.	1	.1041 1.0000	3 .0000	-2.4207
SAMPLE1	54		1.	1	.0755 1.0000	3 .0000	-2.5733
SAMPLE1	55		1.	1	.0743 .9995	3 .0000	-1.7633
SAMPLE1	56		1.	1	.1835 1.0000	3 .0000	-2.1257
SAMPLE1	57		1.	1	.6780 .9999	3 .0001	-1.2108
SAMPLE1	58		1.	1	.8270 .9998	3 .0002	-1.0142
SAMPLE1	59		1.	1	.0866 .7231	3 .2769	.9212
SAMPLE1	60		1.	1	.1812 .9205	3 .0795	.5415
SAMPLE1	61		1.	1	.1725 .9124	3 .0876	.5086
SAMPLE1	62		1.	1	.7308 .9999	3 .0001	-1.1396
SAMPLE1	63		1.	1	.5162 .9942	3 .0058	-1.1463
SAMPLE1	64		1.	1	.9906 .9996	3 .0004	-1.6074
SAMPLE1	65		1.	1	.1154 1.0000	3 .0000	-2.3598
SAMPLE1	66		1.	1	.7979 .9986	3 .0012	-1.5395
SAMPLE1	67		1.	1	.7564 .9985	3 .0015	-1.4853
SAMPLE1	68		1.	3	.5110 .9340	3 .0660	2.4700
SAMPLE1	69		1.	3	.8790 .9992	3 .0003	2.9752
SAMPLE1	70		1.	3	.2288 1.0000	3 .0000	4.3308
SAMPLE1	71		1.	1	.9264 .9994	3 .0006	-1.7057
SAMPLE1	72		1.	1	.6189 .9989	3 .0011	-1.5667
SAMPLE1	73		1.	1	.1561 .8983	3 .1037	.6161
SAMPLE1	74		1.	1	.8934 .9992	3 .0008	-1.6616
SAMPLE1	75		1.	3	.3583 1.0000	3 .0000	4.0460
SAMPLE1	76		1.	1	.7876 .9998	3 .0002	-1.0650
SAMPLE1	77		1.	1	.2642 1.0000	3 .0000	-1.9121
SAMPLE1	78		1.	1	.7772 .9999	3 .0001	-1.0786
SAMPLE1	79		1.	1	.1026 .7643	3 .2157	.8369
SAMPLE1	80		1.	1	.9320 1.0000	3 .0000	-1.4205
SAMPLE1	81		1.	1	.7901 .9987	3 .0013	-1.5294
SAMPLE1	82		1.	3	.6597 .9974	3 .0026	2.6870
SAMPLE1	83		1.	1	.0577 .5622	3 .4378	1.2297
SAMPLE1	84		1.	1	.3675 .9867	3 .0133	2.2632
SAMPLE1	85		1.	3	.3728 .9852	3 .0148	2.2361
SAMPLE1	86		1.	1	.6305 .9970	3 .0030	2.6463
SAMPLE1	87		1.	1	.1972 .9331	3 .0669	.4940
SAMPLE1	88		1.	1	.8265 .9989	3 .0011	2.9108
SAMPLE1	89		1.	1	.0771 .6815	3 .3185	.9720
SAMPLE1	90		1.	1	.1468 1.0000	3 .0000	4.5783
SAMPLE1	91		1.	1	.8005 .9988	3 .0012	-1.5529
SAMPLE1	92		1.	1	.9634 .9995	3 .0005	-1.7497
SAMPLE1	93		1.	1	.6401 .9972	3 .0028	2.6559
SAMPLE1	94		1.	1	.0766 .6766	3 .3214	.9754
SAMPLE1	95		1.	1	.9689 .9995	3 .0005	-1.7365
SAMPLE1	96		1.	1	.0914 1.0000	3 .0000	4.8156

Table C1: continued.

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP P(X/G)	P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES
AM	67			.	78	.0000	3	-1.48
AM	68			.	78	.0000	3	-1.48
AM	69			.	78	.0000	3	-1.48
AM	70			.	78	.0000	3	-1.48
AM	71			.	78	.0000	3	-1.48
AM	72			.	78	.0000	3	-1.48
AM	73			.	78	.0000	3	-1.48
AM	74			.	78	.0000	3	-1.48
AM	75			.	78	.0000	3	-1.48
AM	76			.	78	.0000	3	-1.48
AM	77			.	78	.0000	3	-1.48
AM	78			.	78	.0000	3	-1.48
AM	79			.	78	.0000	3	-1.48
AM	80			.	78	.0000	3	-1.48
AM	81			.	78	.0000	3	-1.48
AM	82			.	78	.0000	3	-1.48
AM	83			.	78	.0000	3	-1.48
AM	84			.	78	.0000	3	-1.48
AM	85			.	78	.0000	3	-1.48
AM	86			.	78	.0000	3	-1.48
AM	87			.	78	.0000	3	-1.48
AM	88			.	78	.0000	3	-1.48
AM	89			.	78	.0000	3	-1.48
AM	90			.	78	.0000	3	-1.48
AM	91			.	78	.0000	3	-1.48
AM	92			.	78	.0000	3	-1.48
AM	93			.	78	.0000	3	-1.48
AM	94			.	78	.0000	3	-1.48
AM	95			.	78	.0000	3	-1.48
AM	96			.	78	.0000	3	-1.48
AM	97			.	78	.0000	3	-1.48
AM	98			.	78	.0000	3	-1.48
AM	99			.	78	.0000	3	-1.48
AM	100			.	78	.0000	3	-1.48
AM	101			.	78	.0000	3	-1.48
AM	102			.	78	.0000	3	-1.48
AM	103			.	78	.0000	3	-1.48
AM	104			.	78	.0000	3	-1.48
AM	105			.	78	.0000	3	-1.48
AM	106			.	78	.0000	3	-1.48
AM	107			.	78	.0000	3	-1.48
AM	108			.	78	.0000	3	-1.48
AM	109			.	78	.0000	3	-1.48
AM	110			.	78	.0000	3	-1.48
AM	111			.	78	.0000	3	-1.48
AM	112			.	78	.0000	3	-1.48
AM	113			.	78	.0000	3	-1.48
AM	114			.	78	.0000	3	-1.48
AM	115			.	78	.0000	3	-1.48
AM	116			.	78	.0000	3	-1.48
AM	117			.	78	.0000	3	-1.48
AM	118			.	78	.0000	3	-1.48
AM	119			.	78	.0000	3	-1.48
AM	120			.	78	.0000	3	-1.48
AM	121			.	78	.0000	3	-1.48
AM	122			.	78	.0000	3	-1.48
AM	123			.	78	.0000	3	-1.48
AM	124			.	78	.0000	3	-1.48
AM	125			.	78	.0000	3	-1.48
AM	126			.	78	.0000	3	-1.48
AM	127			.	78	.0000	3	-1.48
AM	128			.	78	.0000	3	-1.48
AM	129			.	78	.0000	3	-1.48
AM	130			.	78	.0000	3	-1.48
AM	131			.	78	.0000	3	-1.48
AM	132			.	78	.0000	3	-1.48
AM	133			.	78	.0000	3	-1.48
AM	134			.	78	.0000	3	-1.48
AM	135			.	78	.0000	3	-1.48
AM	136			.	78	.0000	3	-1.48
AM	137			.	78	.0000	3	-1.48
AM	138			.	78	.0000	3	-1.48
AM	139			.	78	.0000	3	-1.48
AM	140			.	78	.0000	3	-1.48
AM	141			.	78	.0000	3	-1.48
AM	142			.	78	.0000	3	-1.48
AM	143			.	78	.0000	3	-1.48
AM	144			.	78	.0000	3	-1.48
AM	145			.	78	.0000	3	-1.48
AM	146			.	78	.0000	3	-1.48
AM	147			.	78	.0000	3	-1.48
AM	148			.	78	.0000	3	-1.48
AM	149			.	78	.0000	3	-1.48
AM	150			.	78	.0000	3	-1.48
AM	151			.	78	.0000	3	-1.48
AM	152			.	78	.0000	3	-1.48
AM	153			.	78	.0000	3	-1.48
AM	154			.	78	.0000	3	-1.48
AM	155			.	78	.0000	3	-1.48
AM	156			.	78	.0000	3	-1.48
AM	157			.	78	.0000	3	-1.48
AM	158			.	78	.0000	3	-1.48
AM	159			.	78	.0000	3	-1.48
AM	160			.	78	.0000	3	-1.48
AM	161			.	78	.0000	3	-1.48
AM	162			.	78	.0000	3	-1.48
AM	163			.	78	.0000	3	-1.48
AM	164			.	78	.0000	3	-1.48
AM	165			.	78	.0000	3	-1.48
AM	166			.	78	.0000	3	-1.48
AM	167			.	78	.0000	3	-1.48
AM	168			.	78	.0000	3	-1.48
AM	169			.	78	.0000	3	-1.48
AM	170			.	78	.0000	3	-1.48
AM	171			.	78	.0000	3	-1.48
AM	172			.	78	.0000	3	-1.48
AM	173			.	78	.0000	3	-1.48
AM	174			.	78	.0000	3	-1.48
AM	175			.	78	.0000	3	-1.48
AM	176			.	78	.0000	3	-1.48
AM	177			.	78	.0000	3	-1.48
AM	178			.	78	.0000	3	-1.48
AM	179			.	78	.0000	3	-1.48
AM	180			.	78	.0000	3	-1.48
AM	181			.	78	.0000	3	-1.48
AM	182			.	78	.0000	3	-1.48
AM	183			.	78	.0000	3	-1.48
AM	184			.	78	.0000	3	-1.48
AM	185			.	78	.0000	3	-1.48
AM	186			.	78	.0000	3	-1.48
AM	187			.	78	.0000	3	-1.48
AM	188			.	78	.0000	3	-1.48
AM	189			.	78	.0000	3	-1.48
AM	190			.	78	.0000	3	-1.48
AM	191			.	78	.0000	3	-1.48
AM	192			.	78	.0000	3	-1.48
AM	193			.	78	.0000	3	-1.48
AM	194			.	78	.0000	3	-1.48
AM	195			.	78	.0000	3	-1.48
AM	196			.	78	.0000	3	-1.48
AM	197			.	78	.0000	3	-1.48
AM	198			.	78	.0000	3	-1.48
AM	199			.	78	.0000	3	-1.48
AM	200			.	78	.0000	3	-1.48

Table C1: continued.

Table C2: MINITAB (unstandardized) discriminant scores for TEST One. Group 1 in column C8, group 3 in C9. The scores circled are those specimens misclassified.

COLUMN COUNT	C8 114	C9 29
1	-3.6961	-25.9424
2	-11.1961	-23.5223
3	-12.2179	-25.5036
4	-10.4542	-30.5223
5	-10.1190	-29.7054
6	-10.6903	-24.3733
7	-8.5763	-16.6337
8	-10.2265	-22.7112
9	-10.8504	-24.2136
10	-9.4310	-25.2503
11	-9.9728	-31.7930
12	-9.7866	-24.2669
13	-4.0423	-32.7238
14	-10.9434	-30.9154
15	-4.2817	-23.2627
16	-6.7234	-13.4528
17	-12.6460	-27.7507
18	-10.0942	-30.7957
19	-12.3662	-27.0195
20	-17.6717	-24.5452
21	-11.5152	-20.1849
22	-10.8637	-19.6929
23	-12.4839	-31.7698
24	-9.9930	-29.7313
25	-10.5046	-31.8535
26	-11.3823	-29.8813
27	-9.8664	-29.9713
28	-11.0232	-29.2001
29	-3.7631	-18.3099
30	-11.2360	
31	-11.1961	
32	-10.8233	
33	-10.5179	
34	-11.2493	
35	-9.5603	
36	-10.3315	
37	-2.4334	
38	-8.6031	
39	-12.8190	
40	-11.8292	
41	-11.1429	
42	-11.1691	
43	-10.1323	
44	-10.1037	
45	-9.8664	
46	-9.8930	
47	-10.8243	
48	-9.3079	
49	-9.7631	
50	-9.9227	
51	-9.4907	
52	-14.3349	
53	-9.7363	
54	-10.8371	
55	-9.4917	
56	-9.0818	
57	-9.8531	
58	-17.4456	
59	-11.9363	
60	-16.0627	
61	-9.3611	
62	-13.2572	
63	-10.6642	
64	-9.5343	
65	-11.7147	
66	-11.9274	
67	-11.0631	
68	-11.6033	
69	-16.2459	
70	-11.2360	
71	-9.6326	
72	-8.3295	
73	-8.6504	
74	-17.1133	
75	-8.2576	

C8 continued

76	-11.7546
77	-22.5043
78	-15.7702
79	-17.5431
80	-11.7014
81	-10.8903
82	-17.6564
83	-10.8657
84	-9.6536
85	-17.7116
86	-11.5019
87	-10.0934
88	-12.5391
89	-17.1398
90	-17.8844
91	-11.7014
92	-12.5258
93	-11.4385
94	-12.2732
95	-12.2599
96	-17.3260
97	-10.5578
98	-11.4620
99	-10.4382
100	-11.2360
101	-10.7573
102	-16.7542
103	-11.1961
104	-9.7066
105	-9.8132
106	-10.1589
107	-16.4882
108	-9.0951
109	-8.6962
110	-8.7893
111	-10.2735
112	-9.7467
113	-8.8425
114	-11.8073

their respective territories, with Z scores -19.0945, -22.6048 (group one), and -18.3099 (group three), see Table C3 and Figure C3.

Regardless of whether or not the data is standardized, the classification results are the same, the same specimens are misclassified.

The overall classification results are shown in the following classification matrix:

ACTUAL GROUP	NO. OF CASES	PREDICTED GROUP MEMBERSHIP	
		1	3
GROUP 1	114	112 98.2	2 1.8
GROUP 3	29	1 3.4	28 96.6
PERCENT OF GROUPED CASES CORRECTLY CLASSIFIED -		97.90	

Table C3: Classification results, TEST One.

TEST FOUR

Adding a third group and a third variable to TEST One results in a case where there are two discriminant functions and three classification functions. The SPSS output is:

GROUP MEANS:

FORM	NOBO	GSD	POSQ
1	1.00000	.34148	2.78762
3	3.24137	.45893	4.83609
4	1.38298	.31909	1.94113

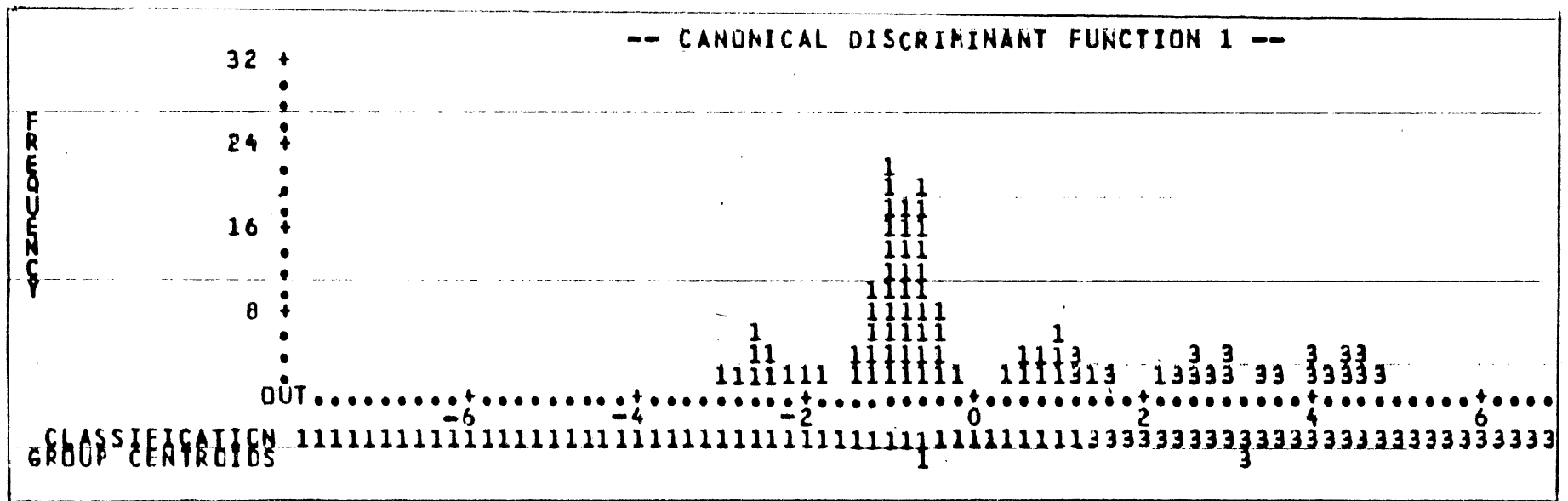


Figure C3: All groups stacked histogram.

TOTAL	1.43684	.35387	2.89089
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GROUP STANDARD DEVIATIONS:

FORM	NOBO	GSD	POSQ
1	.56443	.09522	1.44555
3	.73946	.09229	.95904
4	1.36020	.06650	1.39022
TOTAL	1.15640	.09923	1.63320

POOLED WITHIN-GROUPS COVARIANCE MATRIX:

	NOBO	GSD	POSQ
NOBO	.7295012		
GSD	.2898133E-02	.7841989E-02	(matrix 13)
POSQ	-.1217422	.6908217E-01	1.875848

A₁ and A₃ are calculated as they were for TEST One, except a third variable (POSQ), has been added.

$$A_1 = \begin{bmatrix} 36.000 & .374 & 7.063 \\ .374 & 1.025 & 9.427 \\ 7.063 & 9.427 & 236.164 \end{bmatrix} \quad (\text{matrix 14})$$

$$A_3 = \begin{bmatrix} 15.3065 & -.4127 & -2.9847 \\ -.4127 & .2385 & 1.1850 \\ -2.9847 & 1.1850 & 25.7478 \end{bmatrix} \quad (\text{matrix 15})$$

A₄ = X₄'X₄ - 47 $\bar{x}_4\bar{x}_4'$ (again, X₄ is similar to X₁, it is the 3 x 47 matrix, containing the three variables for the 47 group four specimens used in the analysis).

$$\text{Now } \underline{X}_4 = \begin{bmatrix} 1.38298 \\ .31909 \\ 1.94113 \end{bmatrix}$$

$$\text{and } A_4 = \begin{bmatrix} 85.1036 & .5798 & -26.8409 \\ .5798 & .2033 & 2.3093 \\ -26.8409 & 2.3093 & 88.9096 \end{bmatrix} \quad (\text{matrix 16})$$

Using equation 6:

$$A_1 + A_3 + A_4 = (\text{matrix 14} + \text{matrix 15} + \text{matrix 16})$$

	190	190	
Therefore W =	.72947	.00289	-.12173
	.00289	.00784	.06910 (matrix 17)
	-.12173	.06910	1.87605

which compares well with matrix 13.

The SPSS unstandardized discriminant function coefficients are:

	Z_{sz} FUNC 1	Z_{sz} FUNC 2	
NOBO	-.9862163	.5983976	
GSD	-.9753932	1.409323	(matrix 18)
POSQ	-.4136835	-.6320159	
(CONSTANT)	2.958110	.4685691	

There are now 2 discriminant functions, which cannot be calculated from equation 12, but must be calculated from equations 4 (λ), 5(W, above), and 7(B, below):

$$B = \frac{1}{g-1} \sum_{i=1}^g n_i (\bar{x}_i - \bar{x}_{..})(\bar{x}_i - \bar{x}_{..})'$$

B is calculated on MINITAB to be:

$$B = \begin{bmatrix} 58.1627 & 3.1015 & 54.6713 \\ 3.1015 & .1972 & 3.8125 \\ 54.6713 & 3.8125 & 76.6711 \end{bmatrix} \quad (\text{matrix 19})$$

and W as calculated for matrices 13 and 17.

Employing the two algebraic manipulations, the function coefficients can be calculated:

$$D1/2 = \begin{bmatrix} 1.37518 & 0 & 0 \\ 0 & .84667 & 0 \\ 0 & 0 & .07221 \end{bmatrix} \quad (\text{matrix 20})$$

$$D1/2T' = R \begin{bmatrix} -.14312 & .04992 & 1.36680 \\ .84203 & .01199 & .08773 \\ -.00074 & .07216 & -.00271 \end{bmatrix} \quad (\text{matrix 21})$$

$$(R-1)' = \begin{bmatrix} -.0757 & .0264 & .7227 \\ 1.1746 & .0167 & .1224 \\ -.1428 & 13.8380 & -.5203 \end{bmatrix} \quad (\text{matrix 22})$$

$$(R-1)'BR^{-1} = \begin{bmatrix} 34.5356 & 47.6692 & 3.1945 \\ 47.6692 & 97.2539 & 7.8640 \\ 3.1945 & 7.8640 & .6702 \end{bmatrix} \quad (\text{matrix 23})$$

$$\text{Now} = \begin{bmatrix} 123.533 & 8.931 & -.005 \end{bmatrix} \quad (\text{matrix 24})$$

$$\text{and} = \begin{bmatrix} -.472958 & .879135 & -.058589 \\ -.878416 & -.465311 & .108952 \\ -.068521 & -.102996 & -.992319 \end{bmatrix} \quad (\text{matrix 25})$$

$$\text{giving} \quad \begin{matrix} Z_{sz1} & Z_{sz2} \\ R^{-1} \begin{bmatrix} -.9862 & -.5984 & .2741 \\ -.9754 & -1.4098 & -13.7314 \\ -.4137 & .6320 & .4873 \end{bmatrix} \end{matrix} \quad (\text{matrix 26})$$

The eigenvalues (matrix 24) do not compare at all with those from the SPSS output:

AFTER FUNCTION	EIGENVALUE	PERCENT OF VARIANCE	CUMULATIVE PERCENT	CANONICAL CORRELATION
1	1.32120	93.26	93.26	.7544463
2	.09552	6.74	100.00	.2952766

This is due to the fact that SPSS calculates B differently. SPSS uses the equation:

$$B = \frac{1}{n_1+n_2+\dots+n_{g-1}} \sum_{i=1}^g n_i (\bar{x}_i - \bar{x})(\bar{x}_i - \bar{x})' \quad (\text{equation 20})$$

rather than equation 7:

$$B = \frac{1}{g-1} \sum_{i=1}^g n_i (\bar{x}_i - \bar{x})(\bar{x}_i - \bar{x})'$$

The B calculated from equation 7 can easily be calculated from B (equation 20) by the following:

$$B(\text{eq. 7}) \times \frac{g-1}{n_1+n_2+\dots+n_{g-1}} \quad . \text{ (equation 21)}$$

The same conversion factor can be applied to B (MINITAB) to result in B (SPSS):

$$\begin{bmatrix} 123.533 \\ 8.931 \end{bmatrix} \times \frac{2}{187} = \begin{bmatrix} 1.3212086 \\ .095518 \end{bmatrix}$$

which agrees reasonably well with those given on the SPSS output (above).

It should be noted that no reference was found to a B matrix calculated from equation 21; though two different equations were found: equation 7 (from Gnanadesikan 1977) and also the following:

$$B = \frac{1}{g} \sum_{i=1}^g (x_i - \bar{x})(x_i - \bar{x})'$$

from Lachenbruch (1975).

However it should be remembered that W is calculated using equation 5:

$$\frac{A_1 + A_2 + \dots + A_g}{n_1 + n_2 + \dots + n_{g-1}}$$

so it is not unreasonable that the same scaling factor is applied by SPSS to both the within groups (W) and between groups (B) pooled variance-covariance matrices.

To arrive at the constants for the functions (matrix 18), the (\bar{Z}) means of each group for each function are calculated (on MINITAB) and the means pooled; arriving at a pooled (\bar{Z}) mean for each function. These means are then subtracted from matrix 26 to arrive at matrix 18. For function 1 the group (\bar{Z}_{sz1}) means are:

$$\text{group one } \bar{z}_{sz11} \quad -2.4725$$

$$\text{group three } \bar{z}_{sz13} \quad -5.6450$$

$$\text{group four } \bar{z}_{sz14} \quad -2.4781$$

the pooled (\bar{z}_{sz1}) mean:

$$= 114(-2.4725) + 29(-5.6450) + 47(-2.4781)$$

190

$$= -281.867 - 163.705 - 116.4707$$

190

$$= -562.0427$$

190

$$= -2.9581195$$

For function 2 the group (\bar{z}_{sz1}) means are:

$$\text{group one } \bar{z}_{sz21} \quad = -.68217$$

$$\text{group three } \bar{z}_{sz23} \quad = -.47012$$

$$\text{group four } \bar{z}_{sz24} \quad = .050495$$

the pooled (\bar{z}_{sz2}) mean:

$$= 114(-.68217) + 29(-.47012) + 47(.050495)$$

190

$$= -77.7638 - 13.63348 + 2.373265$$

190

$$= -89.024015$$

190

$$= -.4685478$$

Therefore the constants are the pooled \bar{z} means for the 2 functions,

for function 1: -2.9581195

for function 2: $-.4685478$

which when subtracted from matrix 26 give matrix 18.

If the pooled \bar{Z} means (\bar{Z}_{sz1} , \bar{Z}_{sz2}) are each subtracted from the individual group means for the two functions the result is the functions evaluated at the group centroids (from the SPSS output below).

GROUP	FUNC 1	FUNC 2
1	.48562	-.21360
3	-2.68684	-.00150 (matrix 27)
4	.47995	.51901

For function 1:

$$\text{group one } -2.4725 + 2.958110 = 0.48561$$

$$\text{group three } -5.6450 + 2.958110 = -2.68689$$

$$\text{group four } -2.4781 + 2.958110 = 0.48001$$

and for function 2:

$$\text{group 1 } -.68217 + .4685478 = -.2136222$$

$$\text{group 3 } -.47012 + .4685478 = -.0015722$$

$$\text{group 4 } .050495 + .4685478 = .4180528$$

The SPSS output also gives the standardized canonical discriminant function coefficients:

	FUNC 1	FUNC 2
NOBO	-.84234	.51110
GSD	-.08638	.12480 (matrix 28)
POSQ	-.56659	-.86562

which can be calculated from the unstandardized functions (matrix 17)

by standardizing the independent variables as well:

$$Z_{1sz} = -.9862163x_1 - .9753932x_2 - .4136835x_3 + 2.958110$$

$$Z_{2sz} = .5983976x_1 + 1.409323x_2 - .6320159x_3 + .4685691$$

$$\begin{aligned}
 x_1 &= s_1 y_1 + \bar{x}_1 & \bar{x}_1 &= 1.43684 & s_1 &= .8541 \\
 x_2 &= s_2 y_2 + \bar{x}_2 & \bar{x}_2 &= .35387 & s_2 &= .08855 \\
 x_3 &= s_3 y_3 + \bar{x}_3 & \bar{x}_3 &= 2.89089 & s_3 &= 1.3683
 \end{aligned}$$

$$\begin{aligned}
 Z_{1s} &= -.9862163 (.8541y_1 + 1.43684) - .9753932 (.08855y_2 + .35387) - \\
 &.4136835 (1.3683y_3 + 2.89089) + 2.95110 \\
 &= -.8423273y_1 - 1.417037 - .086371068y_2 - .3451623 - .5660431y_3 \\
 &- 1.1959135 + 2.958110 \\
 &= -.8423273y_1 - .086371068y_2 - .5660431y_3 - 2.9581128 + 2.958110 \\
 &= -.84233y_1 - .08637y_2 - .56604y_3
 \end{aligned}$$

$$\begin{aligned}
 Z_{2s} &= .5983976 (.8541y_1 + 1.43684) + 1.409323 (.08855y_2 + .35387) - \\
 &.6320159 (1.3683y_3 + 2.89089) + .4685691 \\
 &= .5110828y_1 + .8598016 + .1247955y_2 + .4987171 - .8647873y_3 - \\
 &1.8270878 + .4685691 \\
 &= .5110828y_1 + .1247955y_2 - .8647873y_3 + 1.8270878 - 1.8270878 \\
 &= .5110828y_1 + .1247955y_2 - .8647873y_3
 \end{aligned}$$

Z_{1s} and Z_{2s} compare very well with matrix 28.

The classification function coefficients from SPSS:

(FISHER'S LINEAR DISCRIMINANT FUNCTIONS)

FORM =	1	3	4
NOBO	1.192389	4.448038	1.636374
GSD	43.41741	46.81071	44.45542
POSQ	-.3549533e-01	1.142855	-.4961663 (matrix 29)
(CONSTANT)	-9.058475	-21.81242	-8.841117

were calculated on MINITAB. Recall:

$$W = \begin{bmatrix} .7295012 & .002898133 & -.1217422 \\ .002898133 & .007841989 & .06908217 \\ -.1217422 & .06908217 & 1.875848 \end{bmatrix} \text{ (matrix 13)}$$

$$\text{so } W^{-1} = \begin{bmatrix} 1.406 & -1.959 & .163 \\ -1.959 & 191.484 & -7.179 \\ .163 & -7.179 & .808 \end{bmatrix} \text{ (matrix 30)}$$

For group one:

$$W^{-1}\bar{X}_1 = \begin{bmatrix} 1.1924 \\ 43.4169 \\ -.0355 \end{bmatrix} \text{ (matrix 31)}$$

$$\frac{\bar{X}_1 W^{-1} \bar{X}_1}{2} = 7.9598$$

$$\ln .33333 - \frac{\bar{X}_1 W^{-1} \bar{X}_1}{2} = -1.0986124 - 7.9598 = -9.0584 \text{ (constant 3)}$$

Similarly for group three:

$$W^{-1}\bar{X}_3 = \begin{bmatrix} 4.4480 \\ 46.8105 \\ 1.1429 \end{bmatrix} \text{ (matrix 32)}$$

$$\frac{\bar{X}_3 W^{-1} \bar{X}_3}{2} = 20.7138$$

$$\ln .33333 - \frac{\bar{X}_3 W^{-1} \bar{X}_3}{2} = -1.0986124 - 20.7138 = -21.8124 \text{ (constant 4)}$$

and for group four:

$$W^{-1}\bar{X}_4 = \begin{bmatrix} 1.6364 \\ 44.4563 \\ -.4962 \end{bmatrix} \text{ (matrix 33)}$$

$$\frac{\bar{X}_4 W^{-1} \bar{X}_4}{2} = 7.7427$$

$$\ln .33333 - \frac{\bar{X}_4 W^{-1} \bar{X}_4}{2} = -1.0986124 - 7.7427 = -8.8413 \text{ (constant 5)}$$

Combining matrices 31, 32, 33 and constants 3, 4, and 5 results in the classification functions:

	GROUP 1	GROUP 3	GROUP 4
NOBO	1.1924	4.4480	1.6364
GSD	43.4169	46.8105	44.4563 (matrix 34)

POSQ	-.0355	1.1429	-.4962
CONSTANT	-9.0584	-21.8124	-8.8413

which compares very well with matrix 29.

Because there are now two discriminant functions and three groups, it is impossible to determine if a specimen is correctly classified solely from the discriminant scores and critical (\bar{Z}) values $(\bar{Z}_1 + \bar{Z}_3)/2$, $(\bar{Z}_3 + \bar{Z}_4)/2$, $(\bar{Z}_1 + \bar{Z}_4)/2$.

The discriminant scores will not be calculated on MINITAB; instead, the classification scores for all three groups must be evaluated on MINITAB for each specimen and the highest function score taken as the theoretical classification. The theoretical classification and the actual classification are then compared; if they differ, the specimen is misclassified, if they agree the specimen is correctly classified.

The classification functions have been evaluated for all the groups in turn and the three sets of classification scores are listed on Table C4 (group one), Table C5 (group three), and Table C6 (group four).

For the group one specimens, 74 specimens were correctly classified, and the 40 remaining specimens misclassified; three into group three and 37 into group four (Table C4). All of the group three specimens were correctly classified (Table C5). Of the group four specimens, 21 were correctly classified and 26 were misclassified; six into group three and 20 into group four (Table C6).

Looking at the discriminant scores from SPSS, (Table C7) there are 40 group one specimens misclassified, zero group three specimens misclassified, and 26 group four specimens misclassified. The data is

Table C4: MINITAB classification scores, group one, TEST Four. Those specimens circled are specimens misclassified into the group at the head of the column.

Count	Group 1	Group 3	Group 4
1	3.6628	-8.0969	4.1844
2	8.4587	3.1289	8.3844
3	1.4485	-10.4842	1.9171
4	2.8225	-4.6593	3.2796
5	4.9933	-2.3186	5.5024
6	6.8343	.5307	7.0508
7	-.0075	-8.8917	.8417
8	4.0737	-4.4915	5.0206
9	7.4168	-.8870	8.4437
10	2.6843	-5.9894	3.5980
11	4.4898	-1.9971	4.6502
12	3.9434	-4.6319	4.8872
13	4.0465	-4.5580	3.3605
14	7.6705	1.0566	8.0533
15	4.8042	-2.9485	3.8277
16	12.7559	6.8555	11.4907
17	12.4763	9.5794	11.6731
18	15.1911	17.0177	14.3432
19	12.2289	8.8756	11.5899
20	10.5992	10.2810	10.3366
21	9.5166	3.7392	9.6741
22	7.4247	.3026	7.9920
23	12.6065	9.7198	11.8065
24	4.2406	-2.6414	4.5412
25	6.2524	-.9613	6.7916
26	9.0825	3.2711	9.2296
27	4.2039	-4.3510	5.1539
28	7.8690	3.3352	7.4526
29	3.1576	-6.2793	2.7474
30	8.5637	4.0842	8.1639
31	8.4663	2.8855	8.4901
32	7.2798	.6353	7.6532
33	6.2604	.2284	6.3399
34	8.6691	2.1333	9.0758
35	3.2053	-5.4277	4.1314
36	5.6733	-1.0967	6.0083
37	-1.2308	-9.4552	-2.3517
38	-.0076	-5.9992	-.2846
39	.0958	-10.2726	-.1182
40	.7559	-6.0228	.8269
41	8.2345	4.5691	7.5000
42	8.3523	3.6650	8.0221
43	4.9784	-.3908	4.7301
44	4.9145	-1.2227	4.9617
45	4.1170	-1.5522	3.9387
46	4.2907	-4.2574	5.2428
47	6.5722	1.7458	6.1993
48	2.3449	-5.1742	2.7906
49	3.1347	-5.5410	2.4269
50	3.6627	-5.2044	3.0581
51	2.0817	-7.7558	1.7690
52	4.9891	-3.1236	4.1629
53	3.0189	-4.6998	1.9321

Count	Group 1	Group 3	Group 4
54	7.2865	1.8650	7.1840
55	8.6919	3.3478	6.9890
56	1.5484	-4.0888	1.2181
57	4.0483	-.7841	3.5405
58	9.9046	8.0829	10.1897
59	5.1003	.9590	6.0274
60	5.4122	2.4764	5.8868
61	2.5186	-4.9870	2.9684
62	15.0624	14.4428	13.5131
63	6.7150	1.4816	6.5084
64	5.6094	-1.4237	4.3965
65	10.0883	7.0048	9.2280
66	10.7709	8.1423	9.7706
67	7.9568	4.8436	6.9921
68	9.7329	6.8920	8.7587
69	5.9845	4.2746	6.0130
70	8.5384	4.8968	7.8112
71	3.4223	-2.3011	3.2274
72	11.4379	5.9506	9.9401
73	3.1582	.4271	1.7838
74	27.7035	26.3984	27.1086
75	17.7756	11.3781	16.9768
76	10.1990	7.7772	9.0870
77	7.7909	9.1814	8.3571
78	4.4665	1.1402	5.0418
79	10.5218	9.8822	10.3802
80	10.0253	7.5900	8.9091
81	7.4531	2.2774	7.2641
82	10.5268	11.1679	9.8868
83	7.3096	4.0089	6.3828
84	3.4383	-2.8143	3.4502
85	10.7294	10.4214	10.4700
86	9.4439	4.6387	9.2188
87	4.7542	2.4928	3.2838
88	12.7933	9.4842	12.1679
89	8.8770	7.9412	8.7613
90	11.2985	10.8803	11.1129
91	10.0627	6.3854	9.4321
92	12.7247	10.2488	11.7711
93	9.3589	5.9313	8.5928
94	11.9077	9.1034	11.0376
95	11.8526	9.4349	10.8289
96	9.4658	9.2091	9.1178
97	6.3439	1.8740	5.8198
98	9.2720	5.8377	8.5039
99	5.9770	.6858	5.7526
100	8.5820	3.4945	8.4200
101	6.9952	2.5761	6.4866
102	7.6179	6.5836	7.4721
103	8.4037	4.9015	7.6148
104	3.5890	-1.8887	3.3075
105	3.9509	-1.9828	3.8666
106	5.0362	.6377	4.4131

Count	Group 1	Group 3	Group 4
107	6.7786	4.7126	6.9889
108	1.5854	-3.8340	1.1722
109	.3477	-7.3275	.7456
110	.6078	-5.5872	.4435
111	5.3952	2.0818	4.3691
112	3.7128	-1.5403	3.3506
113	.7815	-5.4000	.6213
114	10.3654	8.1976	9.1636

Count	Group 1	Group 3	Group 4
1	18.6045	23.6371	18.3406
2	10.7542	13.4559	10.9711
3	17.1859	21.6352	17.0721
4	15.5960	23.2051	15.8121
5	11.9564	19.0342	12.1815
6	13.5237	16.7466	13.6882
7	13.7868	14.5564	13.2741
8	8.1447	9.3479	8.8031
9	12.9745	17.0946	12.7598
10	16.3893	19.8361	16.6224
11	18.7515	27.0714	18.8624
12	13.1519	17.1607	12.9902
13	21.7777	30.7653	21.7932
14	15.9191	22.9156	16.3913
15	10.2538	12.2313	10.7254
16	13.2112	14.3090	12.5394
17	5.5247	13.7450	4.9552
18	15.5243	22.6235	15.9351
19	22.0757	28.8973	21.3040
20	14.0563	18.3826	13.8201
21	18.7180	21.9305	17.5222
22	17.0876	20.9699	15.5424
23	18.5713	27.0956	18.5928
24	12.0287	19.5952	12.0675
25	19.0189	25.6838	19.7888
26	11.8966	15.6809	11.7541
27	12.8034	20.6584	12.7719
28	10.3355	16.3216	10.8975
29	12.6295	14.2559	11.7203

Table C5: MINITAB classification scores, group three specimens, TEST Four. All specimens correctly classified.

Count	Group 1	Group 3	Group 4
1	11.0247	11.9688	11.9406
2	5.9504	-2.2903	5.2263
3	9.2971	5.9728	10.1342
4	6.0806	-2.1499	5.3596
5	4.1404	-7.5820	4.6734
6	7.5470	2.1459	7.4508
7	7.2547	.5002	6.0229
8	8.6687	5.0372	7.9445
9	2.9246	-6.0002	2.3024
10	8.3163	-1.8985	8.4895
11	10.6858	11.3246	11.7021
12	10.2866	11.1730	11.1849
13	15.1095	19.1449	15.0438
14	9.4542	10.5270	10.2345
15	12.7670	17.4939	13.9515
16	5.6095	-2.8728	4.9609
17	7.9684	.6366	7.0001
18	8.7721	10.8169	9.1370
19	8.6122	.1320	8.1262
20	4.7758	-1.0934	4.7113
21	6.6450	-1.5414	5.9376
22	5.3741	-2.5171	4.4826
23	4.8391	1.2036	3.9083
24	2.5577	-5.7450	1.6733
25	7.2410	-.5043	6.3942
26	6.1122	-1.7214	5.2383
27	4.2654	-.5501	3.7628
28	6.5811	5.2922	6.4781
29	4.9285	-.2296	4.5953
30	4.3378	-3.8257	3.4960
31	1.4252	-7.3469	2.3087
32	-.3983	-9.3129	.4416
33	1.5374	-5.1802	1.6271
34	1.2394	-9.5286	1.2431
35	2.0638	-7.1798	1.5189
36	-1.7442	-10.7641	-.9366
37	3.8333	-1.5882	5.1899
38	6.5220	7.6356	8.7738
39	9.5845	7.7750	11.4942
40	11.2857	10.7903	12.7762
41	6.0476	.7992	7.4571
42	7.6741	5.7153	9.5381
43	7.5873	5.6217	9.4492
44	2.2501	-6.4575	3.1534
45	4.0235	-2.8755	4.3189
46	11.0887	13.7404	12.9900
47	7.5004	5.5281	9.3602

Table C6. MINITAB classification scores, group four specimens, TEST Four. Those scores circled are specimens misclassified into the group at the head of the column.

CASE	MIS	SEL	ACTUAL	HIGHEST	PROBABILITY	2ND HIGHEST	DISCRIMINANT	SCORES
SUBFILE	SEQNUM	VAL	GROUP	GROUP	P(X/G) P(G/X)	GROUP P(G/X)		
SAMPL4A	1		1.***	4	.0847 .6475	1 .3725	2.6723	.6018
SAMPL4A	2		1.***	4	.0776 .6150	1 .3850	2.7221	.6096
SAMPL4A	3		1.***	4	.6862 .6122	1 .3875	1.3173	.7631
SAMPL4A	4		1.***	4	.6929 .6244	1 .3753	1.2682	.6535
SAMPL4A	5		1.***	4	.9042 .5534	1 .4458	1.4237	.4515
SAMPL4A	6		1.***	4	.3050 .7004	1 .2996	1.7753	1.3221
SAMPL4A	7		1.***	4	.3053 .7204	1 .2795	1.7053	1.4545
SAMPL4A	8		1.***	4	.2598 .7382	1 .2657	1.6286	1.5030
SAMPL4A	9		1.***	4	.3061 .7137	1 .2863	1.7349	1.4074
SAMPL4A	10		1.***	4	.6750 .5346	1 .4597	1.9766	.3753
SAMPL4A	11		1.***	1	.9662 .5174	4 .4801	.7899	.6218
SAMPL4A	12		1.***	4	.3054 .7156	1 .2801	1.7060	1.4503
SAMPL4A	13		1.***	4	.4759 .6650	1 .3349	1.5671	.6775
SAMPL4A	14		1.***	4	.8455 .5942	1 .4053	1.6357	.6751
SAMPL4A	15		1.***	4	.4629 .7261	1 .2755	1.2720	1.5738
SAMPL4A	16		1.***	4	.3507 .7783	1 .2196	1.6614	1.5732
SAMPL4A	17		1.***	3	.9736 .9886	1 .0064	-2.6953	-1.5325
SAMPL4A	18		1.***	4	.5648 .6633	1 .2960	-1.2436	-1.5492
SAMPL4A	19		1.***	3	.3785 .1331	1 .1369	-1.7356	1.6222
SAMPL4A	20		1.***	1	.7460 .6390	4 .3377	-1.0847	1.7241
SAMPL4A	21		1.***	4	.3279 .4007	1 .3061	-1.0087	1.2175
SAMPL4A	22		1.***	4	.9528 .5385	1 .4601	1.7524	.3695
SAMPL4A	23		1.***	4	.7013 .6379	1 .3518	1.2139	.5324
SAMPL4A	24		1.***	4	.5654 .6644	1 .2965	-1.2456	1.5450
SAMPL4A	25		1.***	4	.6171 .5743	1 .4253	1.1133	.5276
SAMPL4A	26		1.***	4	.6960 .6314	1 .3663	1.1246	.8544
SAMPL4A	27		1.***	4	.9482 .5355	1 .4627	1.7622	.3554
SAMPL4A	28		1.***	4	.3052 .7211	1 .2769	1.7007	1.4560
SAMPL4A	29		1.***	1	.9841 .5986	4 .3948	1.3077	1.4172
SAMPL4A	30		1.***	4	.3652 .6011	1 .3968	1.8547	1.3966
SAMPL4A	31		1.***	1	.9655 .5946	4 .3980	1.2921	1.3547
SAMPL4A	32		1.***	4	.9278 .5050	1 .4932	1.6782	1.6654
SAMPL4A	33		1.***	4	.8430 .5919	1 .4076	1.8455	.6664
SAMPL4A	34		1.***	4	.9118 .5192	1 .4796	1.8256	.6556
SAMPL4A	35		1.***	4	.8511 .5499	1 .3995	1.6145	.6599
SAMPL4A	36		1.***	4	.3059 .7152	1 .2837	1.7232	1.4263
SAMPL4A	37		1.***	4	.6307 .5926	1 .4169	1.6316	.6143
SAMPL4A	38		1.***	1	.3351 .7534	4 .2459	1.4576	1.5697
SAMPL4A	39		1.***	4	.5175 .5680	1 .4301	1.7001	.4232
SAMPL4A	40		1.***	4	.2427 .5533	1 .4407	1.1863	1.4265
SAMPL4A	41		1.***	4	.6159 .5175	1 .4620	1.0009	1.2541
SAMPL4A	42		1.***	4	.7256 .6643	1 .3187	1.0049	1.2541
SAMPL4A	43		1.***	4	.4850 .5787	1 .4159	.3647	1.2492
SAMPL4A	44		1.***	4	.9545 .5603	1 .4371	.5382	1.1852
SAMPL4A	45		1.***	4	.8911 .5113	1 .4877	.0926	.4201
SAMPL4A	46		1.***	4	.9723 .5435	1 .4546	.6874	1.0644
SAMPL4A	47		1.***	4	.3051 .7215	1 .2785	1.6286	1.4516

Table C7: SPSS discriminant scores for TEST Four. The symbol *** indicates those specimens misclassified.

SUBFILE	CASE ID	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST GROUP	PROBABILITY P(X/G)	PROBABILITY P(G/X)	2ND HIGHEST GROUP	PROBABILITY P(G/X)	DISCRIMINANT	SCORES
SAMPL4A	48	48			1.	1	.9865	.5894	4	.4059	.4041	-.3576
SAMPL4A	49	49			1.	4	.6767	.6095	1	.3903	1.3280	.7676
SAMPL4A	50	50			1.	1	.4575	.6696	4	.3301	1.5876	-.8047
SAMPL4A	51	51			1.	1	.4550	.6467	4	.3532	1.6571	-.6035
SAMPL4A	52	52			1.	1	.3222	.5775	4	.4224	1.4899	-.2825
SAMPL4A	53	53			1.	1	.4957	.6954	4	.3044	1.3951	-.9680
SAMPL4A	54	54			1.	1	.4024	.7475	4	.2551	1.2510	-1.3656
SAMPL4A	55	55			1.	1	.9663	.5244	4	.4732	.6162	.0135
SAMPL4A	56	56			1.	1	.1468	.6421	4	.1135	.4459	-2.1726
SAMPL4A	57	57			1.	1	.1468	.6421	4	.1135	.4459	-2.1726
SAMPL4A	58	58			1.	1	.9809	.5806	4	.4173	.6638	.2565
SAMPL4A	59	59			1.	1	.9437	.6212	4	.3733	.6635	-.2541
SAMPL4A	60	60			1.	4	.6267	.5336	1	.4013	-.4834	.3344
SAMPL4A	61	61			1.	4	.6564	.7132	1	.2823	.3067	1.4467
SAMPL4A	62	62			1.	4	.8064	.6042	1	.3759	-.7655	.6555
SAMPL4A	63	63			4.	3	.1450	.4235	4	.4117	-.1982	1.2689
SAMPL4A	64	64			4.	1	.5266	.6735	4	.3264	1.4486	-.6686
SAMPL4A	65	65			4.	4	.6737	.6903	1	.2969	1.2491	1.6166
SAMPL4A	66	66			4.	4	.5234	.5728	4	.3271	1.4457	-.6244
SAMPL4A	67	67			4.	4	.0862	.6302	1	.3698	2.6616	.8970
SAMPL4A	68	68			4.	1	.9851	.5229	4	.4743	.6104	.0220
SAMPL4A	69	69			4.	4	.3826	.7735	4	.2256	.9337	-1.3354
SAMPL4A	70	70			4.	1	.6760	.6105	4	.3853	1.3241	1.7735
SAMPL4A	71	71			4.	1	.7289	.6617	4	.3207	-.6043	-.8356
SAMPL4A	72	72			4.	1	.4413	.5507	4	.3492	1.6733	-.4677
SAMPL4A	73	73			4.	4	.2449	.3431	1	.4569	2.1535	1.4017
SAMPL4A	74	74			1.	1	.6673	.5712	3	.4074	-1.6301	-1.9740
SAMPL4A	75	75			1.	1	.9945	.5499	4	.4472	-.5477	-1.2666
SAMPL4A	76	76			1.	1	.3790	.7703	4	.2260	1.6233	1.4966
SAMPL4A	77	77			1.	1	.5719	.5806	4	.2880	-.1908	-1.0660
SAMPL4A	78	78			4.	4	.4266	.6945	4	.2554	-.3461	-1.2161
SAMPL4A	79	79			4.	3	.1485	.4004	3	.3349	-1.1427	1.5268
SAMPL4A	80	80			3.	3	.6749	.8642	4	.0725	-1.9161	1.5001
SAMPL4A	81	81			3.	3	.9820	.9784	1	.0114	-2.4975	-.0260
SAMPL4A	82	82			3.	3	.6773	.5969	4	.0866	-3.4640	-.4170
SAMPL4A	83	83			1.	1	.5040	.7015	4	.2673	-.1905	-1.1591
SAMPL4A	84	84			1.	4	.4671	.5965	4	.2629	-.2771	-1.1851
SAMPL4A	85	85			1.	4	.5607	.4656	1	.4525	-.5421	1.1857
SAMPL4A	86	86			1.	3	.7286	.6625	4	.3201	-.6019	-.8435
SAMPL4A	87	87			1.	1	.7566	.5981	4	.0611	-3.2936	.5505
SAMPL4A	88	88			1.	1	.9716	.5476	4	.4506	-.7630	1.1113
SAMPL4A	89	89			4.	4	.1462	.4174	3	.4126	-1.2016	1.3046
SAMPL4A	90	90			1.	1	.2446	.8145	4	.1821	1.5099	1.8916
SAMPL4A	91	91			1.	1	.2299	.7586	4	.1920	-.5482	-1.7255
SAMPL4A	92	92			1.	1	.4323	.5488	4	.3025	-.7266	-.6650
SAMPL4A	93	93			1.	1	.7165	.6890	4	.3099	-.8608	-.4350
SAMPL4A	94	94			1.	1	.3367	.7054	4	.2220	-.4217	-1.3726
SAMPL4A	95	95			3.	3	.7046	.5201	4	.0432	-2.0854	-.5571
SAMPL4A	95	95			3.	3	.7046	.5201	1	.2862	-1.3733	-.5561

Table C7: continued.

SUBFILE	CASE SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST GROUP	PROBABILITY P(X/G)	PROBABILITY P(G/X)	2ND HIGHEST GROUP	HIGHEST P(G/X)	DISCRIMINANT SCORES	
AMPL4A	96			3.	4	.2559	.5316	4	.3065	-1.4033	1.0370
AMPL4A	97			1.	3	.3151	.5927	4	.2590	-1.4710	.5162
AMPL4A	98			3.	3	.9461	.9715	1	.0158	-2.4029	-.1627
AMPL4A	99			4.	4	.9473	.9669	1	.0171	-2.3625	-.0412
AMPL4A	100			1.	4	.8244	.6318	1	.3524	-.0175	.5343
AMPL4A	101			3.	3	.7817	.4328	4	.0375	-2.1498	.4502
AMPL4A	102			1.	4	.3805	.4175	4	.3624	-.0951	-.0511
AMPL4A	103			3.	3	.2760	.2992	4	.0003	-.8976	-.2711
AMPL4A	104			1.	1	.3576	.7067	4	.2312	-.4174	-1.5778
AMPL4A	105			1.	4	.9931	.5454	4	.4512	-.5311	-.1049
AMPL4A	106			3.	3	.9432	.9075	1	.0176	-2.3632	-.0504
AMPL4A	107			1.	1	.5510	.5543	1	.2918	-1.3448	-.7323
AMPL4A	108			3.	3	.4646	.6960	4	.2763	-1.1275	-1.1170
AMPL4A	109			3.	4	.4646	.6997	4	.0001	-3.9170	.1396
AMPL4A	110			1.	4	.6661	.5022	1	.4965	-.0849	.1722
AMPL4A	111			4.	4	.1984	.4790	4	.3572	-1.2033	1.2033
AMPL4A	112			3.	3	.6355	.4976	4	.0015	-3.2474	1.7002
AMPL4A	113			1.	1	.3263	.3990	4	.3078	-1.0106	-.2131
AMPL4A	114			1.	4	.9953	.5536	4	.4419	-.1325	-.1325
AMPL4A	115			1.	1	.1575	.7496	4	.1723	-.5047	-1.0616
AMPL4A	116			1.	4	.7474	.6363	4	.3404	-.0975	-.7058
AMPL4A	117			1.	1	.4294	.4380	4	.3901	-.0995	-.1122
AMPL4A	118			1.	1	.3454	.4016	4	.3333	-.0648	-.1177
AMPL4A	119			3.	3	.4372	.7353	4	.1650	-1.0847	1.7798
AMPL4A	120			4.	4	.2117	.9636	4	.0279	-2.4665	1.7467
AMPL4A	121			4.	1	.5077	.6566	4	.3433	1.5320	-.7244
AMPL4A	122			4.	4	.5139	.7245	4	.2751	1.1398	-1.1641
AMPL4A	123			4.	1	.7920	.6420	4	.3413	-.0183	-.7115
AMPL4A	124			1.	1	.4373	.3805	4	.2623	-.3901	-1.1526
AMPL4A	125			1.	4	.6772	.5679	4	.3104	-.0730	-.6974
AMPL4A	126			1.	1	.5301	.5760	4	.2631	-.2790	-1.0510
AMPL4A	127			1.	1	.3887	.6905	4	.2480	-.4152	-1.2226
AMPL4A	128			3.	3	.3616	.6049	1	.2213	-1.4916	1.7796
AMPL4A	129			3.	3	.4630	.9946	1	.0003	-3.7284	-.0558
AMPL4A	130			3.	3	.6609	.9979	1	.0012	-3.2855	-.6640
AMPL4A	131			3.	3	.5365	.9964	1	.0011	-3.3050	-.5262
AMPL4A	132			1.	1	.3125	.4033	3	.3120	-1.0348	-.3542
AMPL4A	133			1.	4	.9204	.6236	4	.3693	-.0278	-.5541
AMPL4A	134			4.	4	.4999	.7599	4	.1417	-1.5954	-.0341
AMPL4A	135			3.	3	.9551	.9769	1	.0129	-2.4700	-.1526
AMPL4A	136			4.	4	.5466	.6192	4	.3907	1.5459	-.0501
AMPL4A	137			4.	1	.9257	.5134	4	.4831	1.7611	-.0667
AMPL4A	138			4.	4	.5357	.6698	4	.3500	1.4330	-.0600
AMPL4A	139			4.	4	.4930	.7090	4	.2907	1.3232	-1.0579
AMPL4A	140			1.	1	.6766	.6684	4	.3100	-.0710	-.9062
AMPL4A	141			4.	4	.5819	.7039	4	.2775	-.0225	-1.1217
AMPL4A	142			4.	1	.4423	.7076	4	.2922	1.4537	1.0470
AMPL4A	143			1.	1	.9551	.5344	4	.4429	1.2645	-.1528

Table C7: continued.

represented graphically on Figure C4 and the territorial map on Figure C5.

Comparing the SPSS and MINITAB outputs it can be seen that not only are the same number of specimens from each group misclassified but in each case the same specimens are misclassified and misclassified into the same (incorrect) groups.

The overall classification results (64.74 %, Table C8) are not nearly as good as those obtained for TEST One (97.90 %, Table C3). Though these three variables (NOBO, GSD, POSQ) are good discriminators for two groups, they are poor discriminators for these three groups, particularly for groups one and four.

The overall classification results are given on the classification matrix below:

ACTUAL GROUP	NO. OF CASES	PREDICTED GROUP MEMBERSHIP		
		1	3	4
GROUP 1	114	74 64.9	3 2.6	37 32.5
GROUP 3	29	0 0	29 100.0	0 0
GROUP 4	47	21 44.7	6 12.8	20 42.6
PERCENT OF GROUPED CASES CORRECTLY CLASSIFIED -		64.74		

Table C8: Classification results, TEST Four.

SUMMARY OF TEST RESULTS

There are three major stages to discriminant analysis; derivation, validation, and interpretation. To understand the derivation stage an illustrative example, TEST, was devised and the

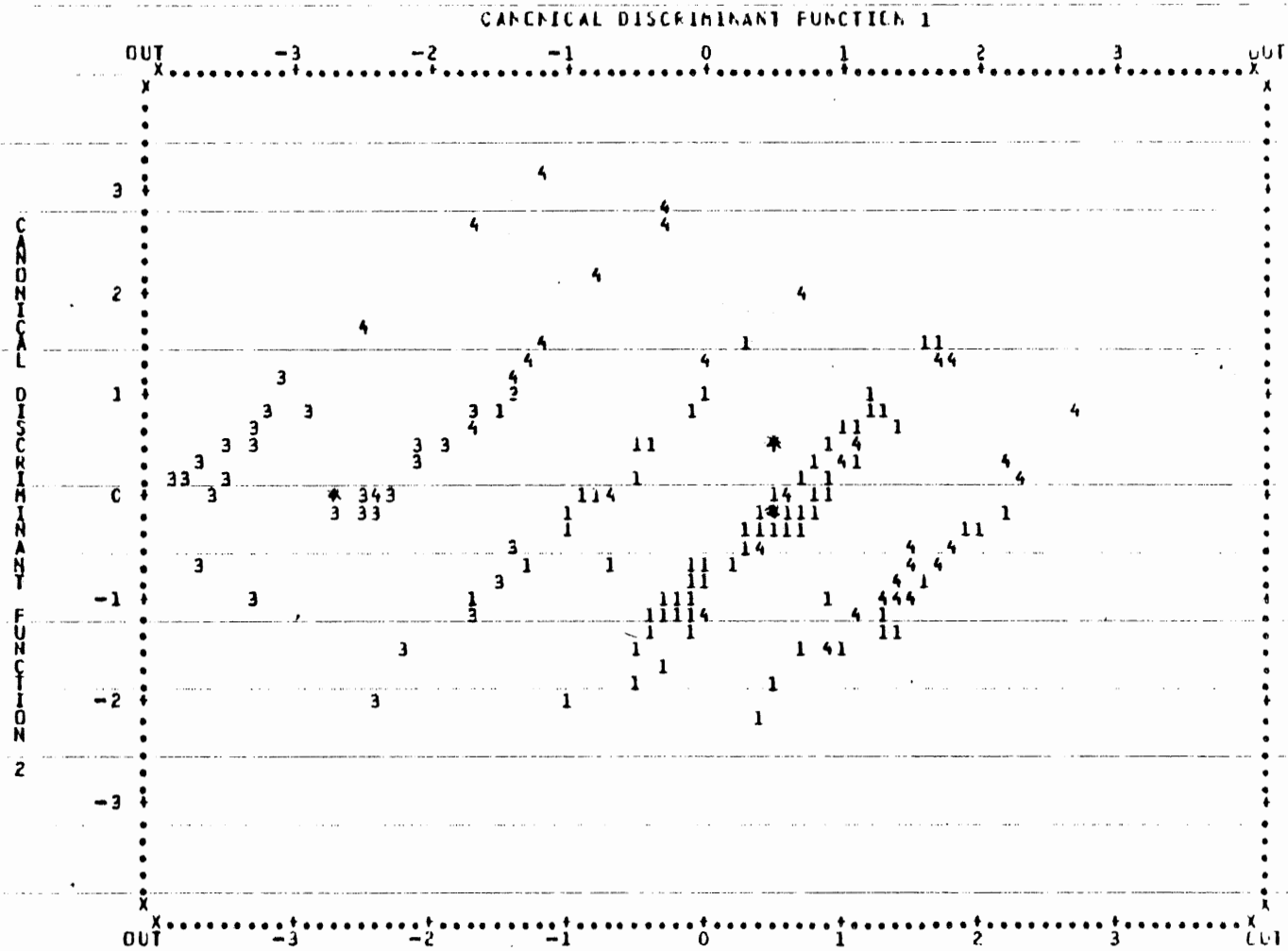


Figure C4: Scatter plot, TEST Four.

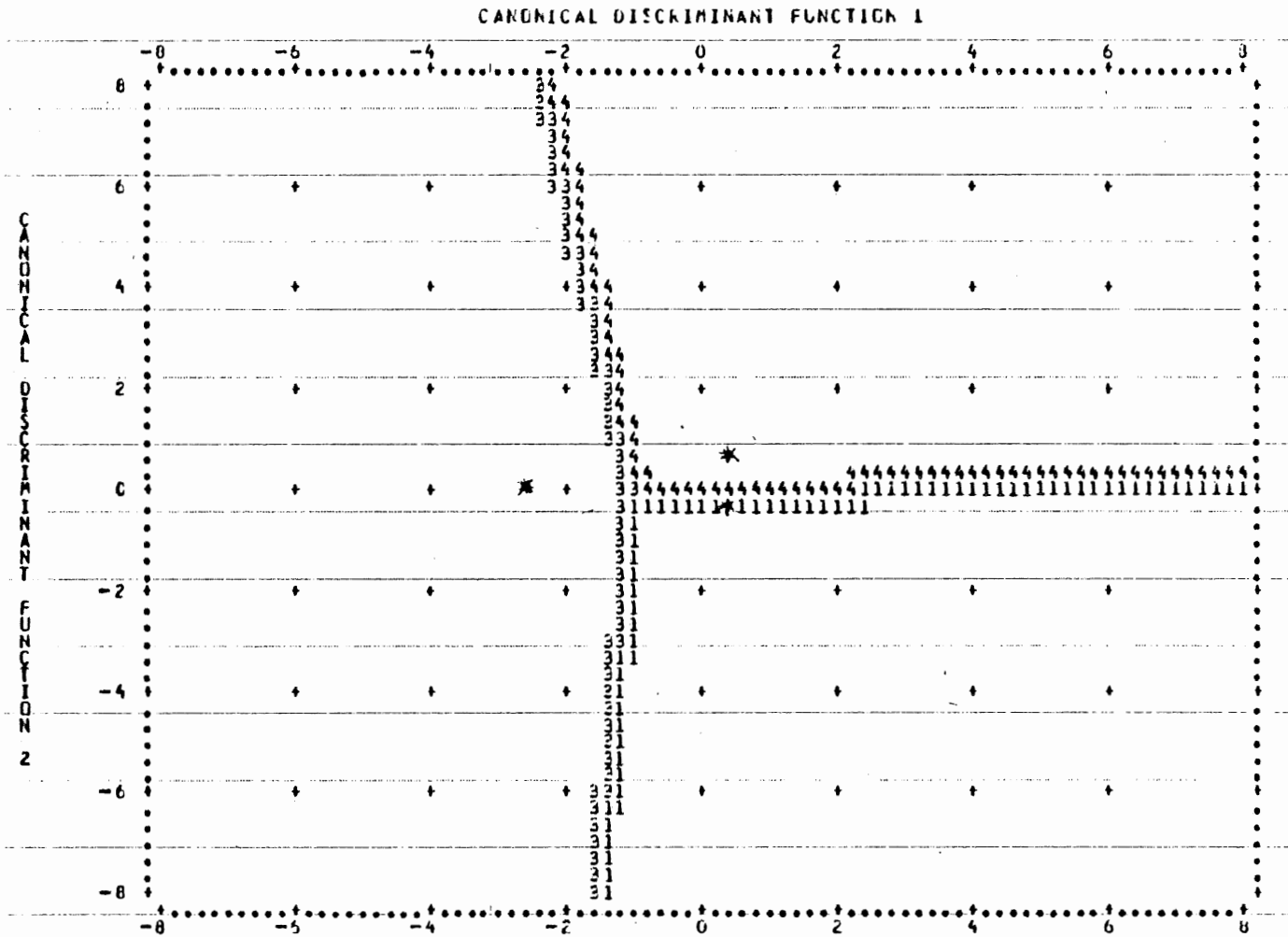


Figure C5: Territorial map, TEST Four.

following derivations / calculations are understood / duplicated with the aid of MINITAB.

1. Calculation of the pooled within group variance matrix (W).
2. Calculation of the pooled within groups variance-covariance matrix (B) for the three group case (TEST Four), and the corresponding eigenvalues (λ).
3. Derivation of the unstandardized (Z only; Z_{SZ}) discriminant functions.
4. Derivation of the standardized (Z and variable, Z_S) discriminant functions.
5. Calculation of \bar{Z} values of the group centroids (= group means).
6. Calculation of critical Z values.
7. Calculation of the discriminant (Z) scores, two group case (TEST One).
8. Derivation of the classification functions.
9. Calculation of the classification scores and classification matrices, three group case (TEST Four).

APPENDIX D: SPSS Output of analysis A-1.

----- DISCRIMINANT ANALYSIS -----

ON GROUPS DEFINED BY FORM

721 (UNWEIGHTED) CASES WERE PROCESSED.
 0 OF THESE WERE EXCLUDED FROM THE ANALYSIS.
 721 (UNWEIGHTED) CASES WILL BE USED IN THE ANALYSIS.

NUMBER OF CASES BY GROUP

FORM	NUMBER OF CASES		LABEL
	UNWEIGHTED	WEIGHTED	
1	163	163.0	
2	61	61.0	
3	51	51.0	
4	134	134.0	
5	46	46.0	
6	69	69.0	
7	20	20.0	
8	67	67.0	
9	100	100.0	
10	10	10.0	
TOTAL	721	721.0	

GROUP MEANS

FORM	PAP	UNCU	DEUH	POR	AOHA	NOBJ	PERO	REPO
1	.85890	.199387	1.76687	2.07975	1.00613	1.02454	1.70552	1.49080
2	.96721	.13115	1.16393	2.73770	1.01639	.80328	2.26230	2.26230
3	.96078	.18608	2.37255	3.01961	1.01373	3.00000	2.15686	2.84314
4	.97015	.19403	1.53731	2.20896	1.02239	1.67164	2.01493	.87313
5	.58696	.193478	2.64793	2.28261	1.197826	1.08696	1.52174	1.58696
6	.97101	.01449	2.72484	1.05797	1.88406	3.07246	1.95652	3.43478
7	.95000	.140000	3.00000	3.00000	2.00000	1.20000	1.87000	3.65000
8	.01493	.163582	1.61194	2.19403	1.00000	.01493	1.91045	3.79104
9	.97000	.070000	1.39000	3.98000	1.00000	.62000	1.89000	3.98000
10	1.00000	0	1.50000	2.40000	2.00000	2.90000	1.80000	2.40000
TOTAL	.83079	.144521	1.84050	2.46741	1.121775	1.34674	1.90430	2.36061

FORM	SUT	CHAM	POSU	DEPO	POSQ	GSR
1	1.93865	9.142945	1.05521	1.94479	2.66123	.85221
2	1.55738	9.21311	1.59016	2.01639	4.07776	.83462
3	1.68627	10.49020	1.82353	2.23529	4.81855	.85410
4	1.82090	9.02239	.67910	.99254	1.68398	.84089
5	1.95652	10.184783	1.06522	1.97826	3.05525	.84931
6	1.17391	10.42029	2.26037	2.85507	3.56213	.86507
7	1.30000	13.90000	2.40000	2.90000	6.17333	.85082
8	1.79104	9.165672	2.79104	3.00000	6.66865	.85074

FORM	SUT	CHAM	POSU	DEPD	POSQ	GSR
9	1.95000	12.23000	2.94000	2.98000	7.076124	.83866
10	1.50000	9.50000	1.40000	2.60000	3.65683	.82683
TOTAL	1.75967	10.13037	1.66574	2.16089	4.27089	.85016

GROUP STANDARD DEVIATIONS

FORM	PAP	UMCO	DEUM	POR	ADNA	NOBO	PERO	REPO
1	.34920	.03594	.49154	.33299	.07833	.60808	.55481	.86330
2	.17956	.034036	.37327	.65577	.12804	1.06175	.62985	.79376
3	.19604	.40098	.56430	.42380	.46862	.93808	.57871	.80926
4	.17081	.39694	.64470	.69450	.14850	1.53543	.64871	.83551
5	.49732	.24964	.36316	.50169	.14744	.55080	.58648	1.06617
6	.16839	.12039	.44937	.29123	.82250	.92861	.71609	.77608
7	.22361	.50262	0	.61559	0	.69585	.58714	.74516
8	.12217	.37323	.49037	.46836	0	.12217	.71205	.56508
9	.17145	.25643	.49021	.14071	0	.67838	.54855	.14071
10	0	0	.84934	.51640	0	.73786	.63246	.69921
TOTAL	.37920	.50012	.73848	.92907	.41301	1.29707	.64161	1.38756

FORM	SUT	CHAM	POSU	DEPD	POSQ	GSR
1	.24071	1.04804	.54713	.97659	1.49197	.05366
2	.50092	1.03465	.58013	.69503	1.00820	.05434
3	.46862	1.23891	.65440	.70960	1.13895	.05322
4	.38488	1.28301	.55658	.88049	1.49688	.05378
5	.20618	1.22868	.80006	1.20165	1.91839	.05104
6	.38181	1.47933	.65640	.39390	1.88734	.07880
7	.47016	2.26878	.68056	.30779	1.57602	.05372
8	.40963	1.06086	.59123	.17408	1.07627	.04220
9	.21904	1.48973	.31203	.14071	1.12508	.03891
10	.52705	.52705	.51640	.51640	.95894	.03159
TOTAL	.42819	1.075900	1.00445	1.02153	2.54039	.05396

POOLED WITHIN-GROUPS COVARIANCE MATRIX WITH 711 DEGREES OF FREEDOM

	PAP	UMCO	DEUM	POR	AOMA	NOBO	PERO	REPO
PAP	.6389658E-01							
UMCO	-.3018988E-02	.8893532E-01						
DEUM	-.4252759E-02	-.5216680E-02	.2596907					
POR	.4523600E-02	-.2512821E-02	.1980124E-03	.2250744				
AOMA	-.1060525E-02	.2902498E-03	.2551533E-01	-.1422889E-02	.3357280E-01			
NOBO	-.1026763E-01	-.3448370E-02	.1592617	-.4282257E-01	.1522352E-01	.8672382		
PERO	.6414417E-02	-.1512785E-01	-.3808780E-01	.2107687E-01	-.3303071E-02	.7826102E-02	.3799354	
REPO	-.2698672E-01	-.5712867E-02	.2230493E-01	.3696527E-01	-.2528093E-02	-.1452810E-01	-.8947199E-03	.5825896
SUT	-.1068162E-02	.1636005E-02	.4174982E-02	-.1783317E-02	.3387179E-02	-.1129869E-01	-.2437879E-01	-.2696679E-01
CHAM	-.2354988E-01	.1320359E-01	.1459734	.9919795E-01	.1358269E-01	.3098667	-.4314957E-01	.26968095
POSU	-.1546519E-01	-.7425925E-02	.4183047E-02	.5693599E-01	-.1083965E-01	.5472888E-02	.1345274E-01	.3049801
DEPO	-.2483729E-01	.1501327E-01	-.1170347E-01	.6607042E-01	-.4057181E-02	-.7012543E-01	-.1830249E-01	.3357316
POSQ	-.4983730E-01	-.5663800E-02	.4389289E-01	.1938389	-.1020108E-01	.6499169E-01	.2871122E-01	.8978083
GSR	-.5228996E-03	.1370901E-02	.1743561E-02	.17154997E-03	.1528599E-03	.3440857E-02	-.6371782E-03	.2947607E-02

	SUT	CHAM	POSU	DEPO	POSQ	GSR
SUT	.1258344					
CHAM	-.1702283E-01	1.1619841				
POSU	-.2203069E-01	.1956435	.3289645			
DEPO	-.4617028E-03	.1373847	.2118951	.5588333		
POSQ	-.4102440E-01	.9737417	.6256405	.5722890	1.940760	
GSR	-.2024894E-02	.9016332E-02	.3163508E-02	.2114171E-02	.8856217E-02	.2670106E-02

WILKS LAMBDA (U-STATISTIC) AND UNIVARIATE F-RATIO WITH 9 AND 711 DEGREES OF FREEDOM

VARIABLE	WILKS	LAMBDA	F	SIGNIFICANCE
PAP	.44822		97.25	0
UMCO	.35113		146.0	0
DEUM	.47023		89.00	0
POR	.25749		227.8	0
AOMA	.19494		326.2	0
NOBO	.51021		75.84	0
PERO	.91139		7.680	0
REPO	.29881		185.4	0
SUT	.67775		37.56	0
CHAM	.51698		73.61	0
POSU	.32197		166.4	0
DEPO	.52885		70.38	0
POSQ	.29697		187.0	0
GSR	.97339		2.160	.0233

ON GRUUPS DEFINED BY FORM

ANALYSIS NUMBER 1

DIRECT METHOD- ALL VARIABLES PASSING THE TOLERANCE TEST ARE ENTERED.

MINIMUM TOLERANCE LEVEL..... .0010

CANONICAL DISCRIMINANT FUNCTIONS

MAXIMUM NUMBER OF FUNCTIONS..... 9
 MINIMUM CUMULATIVE PERCENT OF VARIANCE... 100.00
 MAXIMUM SIGNIFICANCE OF WILKS LAMBDA... 1.0000

CANONICAL DISCRIMINANT FUNCTIONS

FUNCTION	EIGENVALUE	PERCENT OF VARIANCE	CUMULATIVE PERCENT	CANONICAL CORRELATION	AFTER FUNCTION	WILKS	LAMBDA	CHI-SQUARED	D.F.	SIGNICANCE
					0	.0012348		4741.4	126	0
1*	5.45895	36.07	36.87	.9193346	1	.0079755		3420.6	104	0
2*	3.85768	26.05	62.93	.8911656	2	.0387424		2301.6	84	0
3*	2.70047	18.24	81.17	.8542621	3	.1433649		1375.2	66	0
4*	1.68465	11.33	92.54	.7921565	4	.3848345		676.01	50	0
5*	.49746	3.35	95.90	.5763688	5	.5763679		390.14	36	0
6*	.29001	1.95	97.85	.4741444	6	.7434752		207.85	24	0
7*	.16390	1.11	98.97	.3752631	7	.8653568		102.39	14	.0000
8*	.12908	.97	99.64	.3301142	8	.9770549		16.434	6	.0116
9*	.02348	.15	100.00	.1514764						

* MARKS THE 9 FUNCTION(S) TO BE USED IN THE REMAINING ANALYSIS.

STANDARDIZED CANONICAL DISCRIMINANT FUNCTION COEFFICIENTS

	FUNC 1	FUNC 2	FUNC 3	FUNC 4	FUNC 5	FUNC 6	FUNC 7	FUNC 8	FUNC 9
PAP	.10875	.21445	.44611	-.07312	-.07202	.34058	.04500	-.02603	.28446
UMCO	-.10883	-.08591	-.43012	-.35430	-.41336	-.13722	.128362	-.16337	-.02708
DEUM	.12750	-.21327	-.19225	-.13903	-.26797	-.43212	-.07205	-.19888	.45401
POR	-.38491	.34149	.05227	.57332	.18303	.50200	.25863	-.19074	.04805
ADMA	.81192	.13563	-.08650	-.31759	.132137	.15932	.13102	.44334	.06930
NUBO	.13039	.03424	.47273	.21695	-.17949	.68089	.40597	.24500	-.31533
PERO	.01578	-.04345	-.00141	.06531	.05582	-.10518	-.11324	.31457	.34885
REPO	.08835	.27773	-.23996	.27475	-.23990	-.34148	.08777	.32216	.64746
SUT	-.25064	-.00599	-.08179	-.10203	.04731	-.13111	.03243	.82316	.28316
CHAM	.03659	.25785	-.01468	.33732	-.13612	.40571	-.48731	-.03553	-.04315
POSU	.12543	.11519	-.33919	.11541	-.02632	.05421	-.17600	.21191	-.42081
DEPO	.17135	-.09173	.09479	-.06526	.35347	.11558	.46103	-.06342	-.01099
POS 9	-.07270	-.05714	-.12734	.34948	.05655	.17004	.02273	.35822	1.23119
GSR	-.03125	-.00885	.05172	.03532	-.08303	.00175	-.09408	.21419	.09655

UNSTANDARDIZED CANONICAL DISCRIMINANT FUNCTION COEFFICIENTS

	FUNC 1	FUNC 2	FUNC 3	FUNC 4	FUNC 5	FUNC 6	FUNC 7	FUNC 8
PAP	.4302008	.10483675	-1.923072	-1.2892740	-2.658532	1.3472254	.1780140	-.1029888
UMCO	-.3647865	-1.6322723	-1.442304	-1.188329	-1.375033	-.3663323	.0510538	-.5478210
DEUM	.2502030	-.4185455	-.3772490	-.3905583	-.5257843	-.7891505	-1.358022	-.3932681
POR	-.8113287	1.147476	-.1101682	-1.208463	.3357981	-.8473452	.5431542	-.4020423
ADMA	4.424587	.7790613	-.4386692	-1.730149	1.751318	.8694874	1.801436	.7811518
NOBO	.1398495	.3572877E-01	-.5070933	.2348470	-1.925131	-.7302554	-.4354575	.2627799
PERO	-.2559348E-01	-.17375851E-01	-.2281612E-02	.1059579	-.1903537E-01	-.2583032	-.1674949	-.5102633
REPO	.1157577	.3639315	-.3143818	.3599600	-.3143011	-.4473899	.1149944	-.4655678
SUT	-.7065684	-.1970576E-01	-.2305570	-.2876139	-.1333793	-.3696172	.1000294	2.321026
CHAM	.2874916E-01	.20254939	-.1153112E-01	-.3121763	-.1069524	.3195545	-.3829291	-.2791576E-01
POSU	.2186899	.2008272	-.5913856	-.2012230	-.4536588E-01	.9451889E-01	-.3103379	-.3694748
DEPO	.2292115	-.01227756	-.1268729	-.8729524E-01	-.4728337	.1546104	.0167113	-.8483535E-01
POSQ	-.5219228E-01	.4819327E-01	-.9140444E-01	.2508654	.4052143E-01	.1220563	.1635225E-01	.2373604
GSR	-.3832731	-.11554572	.9654178	.6593004	-1.549840	.3266939E-01	-1.756130	3.998117
(CONSTANT)	-3.741379	-5.932150	.8679790	7.098891	5.092652	.4308970	2.161113	-5.924132

FUNC 9

PAP	1.125334
UMCO	-.9080966E-01
DEUM	.8909217
POR	.1012862
ADMA	.3776765
NOBO	-.3382166
PERO	.5659603
REPO	-.6219086
SUT	.7982353
CHAM	-.6594910
POSU	-.7336324
DEPO	-.1469506E-01
POSQ	.8837699
GSR	1.802171
(CONSTANT)	-1.173932

CANONICAL DISCRIMINANT FUNCTIONS EVALUATED AT GROUP MEANS (GROUP CENTROIDS)

GROUP	FUNC 1	FUNC 2	FUNC 3	FUNC 4	FUNC 5	FUNC 6	FUNC 7	FUNC 8	FUNC 9
1	-1.22679	-2.108632	-.09948	-.54599	-.88073	.13919	.13931	-.03849	-.02762
2	-1.09906	-.79276	.92920	.77753	-.55744	.00117	.41067	-.69274	.28129
3	.63231	1.134502	.99582	-.12025	-.40378	-1.79374	-.12463	-.12008	-.09794
4	-1.12185	-.73854	2.09544	.13583	.72438	-.00331	-.33314	.12142	-.09417
5	3.11921	-1.31022	-1.40823	-3.76645	.92709	.07626	-.01411	.43582	.25177
6	5.52688	-.01316	.40176	1.76955	-.56232	.31546	-.21657	-.10366	.02294
7	3.49874	3.25975	-1.32173	-3.31690	.01174	.21541	-.58962	-1.30069	-.40544
8	-1.03644	-1.11089	-3.88559	1.68462	.76742	-.32319	-.03006	-.06769	-.08246
9	-1.81660	3.73843	-.66557	-1.10325	-.35238	.29314	-.04881	.37024	.01201
10	4.13784	1.134414	1.93422	-.28093	1.28208	.13436	2.73960	.47108	-.57972

PRIOR PROBABILITY FOR EACH GROUP IS .10000

CLASSIFICATION FUNCTION COEFFICIENTS
(FISHER'S LINEAR DISCRIMINANT FUNCTIONS)

FORM	1	2	3	4	5	6	7	8
PAP	16.80354	18.02542	18.77399	17.37733	12.89328	21.06864	19.13090	4.70221
UMCO	3.415272	-3.914106	-2.883572	-3.533101	4.549295	-5.072266	-1.209308	4.342923
DEUM	1.912073	-1.945973	1.799617	1.0506688E-01	3.524596	1.732824	3.015640	1.806201
POR	6.523835	3.930137	10.42047	7.778815	7.299190	.1721819	12.39101	5.345884
AOA	25.03579	28.10658	33.91866	28.32273	53.15531	32.21828	54.75522	26.55139
NOBO	-1.506989	-1.250458	.7837305	-5.221079	-2.465622	.3735250E-01	-2.659938	-2.837859
PERO	7.526853	7.951832	8.040744	7.645329	7.438773	7.910618	7.574920	7.987877
REPO	5.774709	5.320834	7.721449	5.308975	5.112214	7.744424	8.078690	7.856330
SUT	19.61322	17.27771	18.41703	18.91060	18.64437	14.19003	13.79578	13.51891
CHAM	6.069159	5.904320	5.929414	6.113423	6.802439	6.5912699	8.660727	5.404785
POSU	5.460443	5.141631	5.672727	4.712715	6.763777	7.591883	7.737360	8.346511
DEPO	3.415373	2.699577	2.492834	4.393498	3.366239	4.193729	3.181631	1.713855
POSQ	-7.814407	-7.232874	-7.866323	-7.757264	-8.084998	-7.372502	-9.011923	-6.923091
GSR	301.1314	297.6939	299.9413	302.3213	295.4073	299.7886	288.6562	296.1737
(CONSTANT)	-214.9818	-209.4777	-236.2932	-210.2677	-257.6783	-249.2740	-289.6322	-207.9839

CLASSIFICATION FUNCTION COEFFICIENTS
(FISHER'S LINEAR DISCRIMINANT FUNCTIONS)

FORM	9	10
PAP	19.09032	19.73179
UMCO	-4.864065	-6.044951
DEUM	-1.9219218	-1.386767
POR	12.95869	3.043713
AOA	27.67763	58.91623
NOBO	-1.758962	1.453514
PERO	6.978407	5.920817
REPO	7.711052	5.374471
SUT	20.75124	15.94750
CHAM	7.138652	5.943509
POSU	7.106769	5.328894
DEPO	2.070630	4.385229
POSQ	-7.160445	-3.264663
GSR	301.4712	292.5937
(CONSTANT)	-250.8330	-250.4430

TEST OF EQUALITY OF GROUP COVARIANCE MATRICES USING BOX'S M

THE RANKS AND NATURAL LOGARITHMS OF DETERMINANTS PRINTED ARE THOSE OF THE GROUP COVARIANCE MATRICES.

GROUP LABEL	RANK	LOG DETERMINANT
1	14	-30.450691
2	14	-27.198444
3	14	-25.222006
4	14	-26.050209
5	14	-27.674586
6	14	-29.326704
7	12	(SINGULAR)
8	13	(SINGULAR)
9	13	(SINGULAR)
10	< 10	(TOO FEW CASES TO BE NON-SINGULAR)
POOLED WITHIN-GROUPS COVARIANCE MATRIX	14	-25.042928

SINCE SOME COVARIANCE MATRICES ARE SINGULAR, THE USUAL PROCEDURE WILL NOT WORK. THE NON-SINGULAR GROUPS WILL BE TESTED AGAINST THEIR OWN POOLED WITHIN-GROUPS COVARIANCE MATRIX. THE LOG OF ITS DETERMINANT IS -23.809609

BOX'S M	APPROXIMATE F	DEGREES OF FREEDOM	SIGNIFICANCE
2196.9	3.8253	525,	150258.1 .0000

SYMBOLS USED IN TERRITORIAL MAP

SYMBOL GROUP LABEL

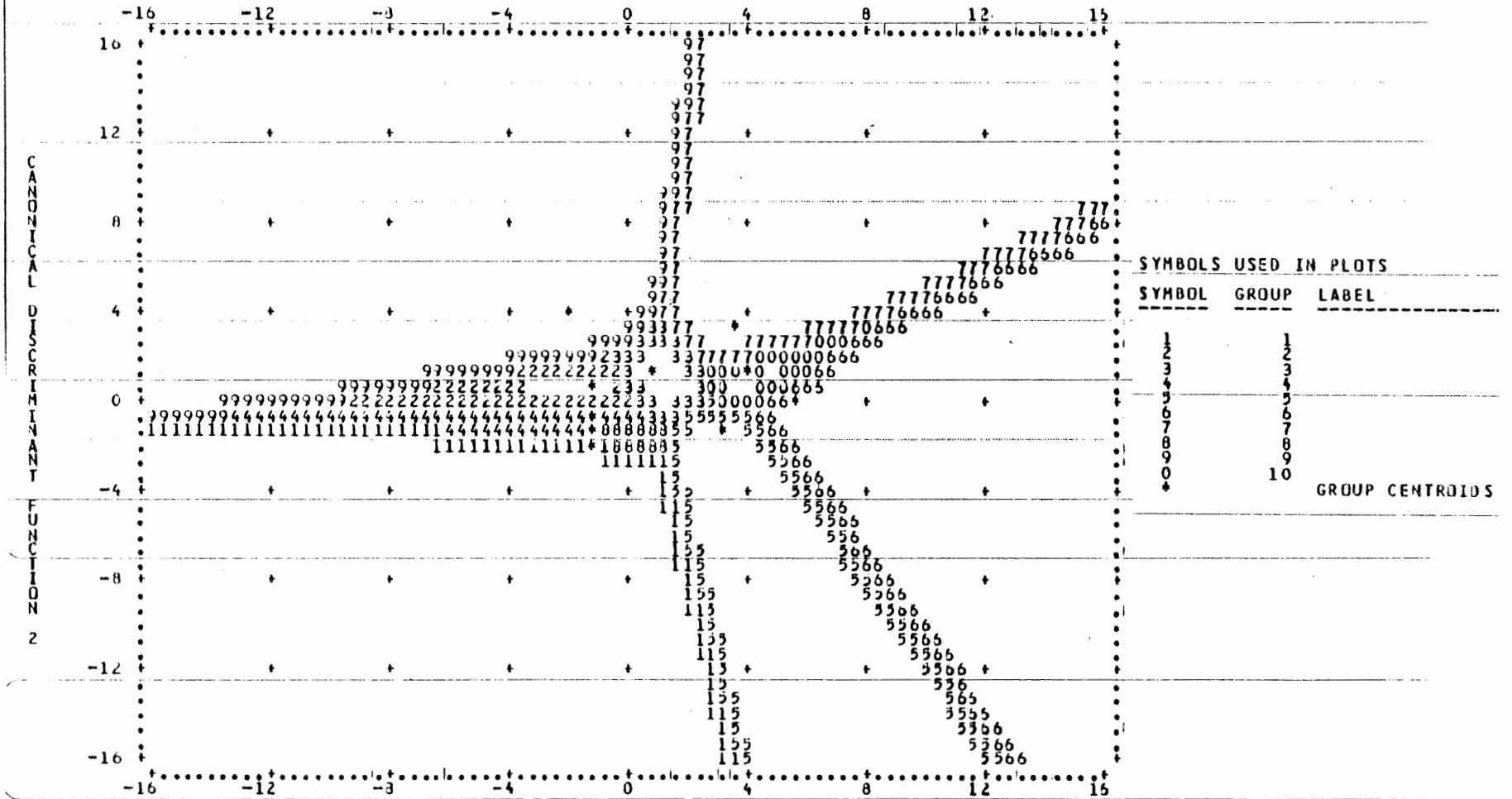
1
2
3
4
5
6
8
9
10

1
2
3
4
5
6
8
9
10

GROUP CENTROIDS

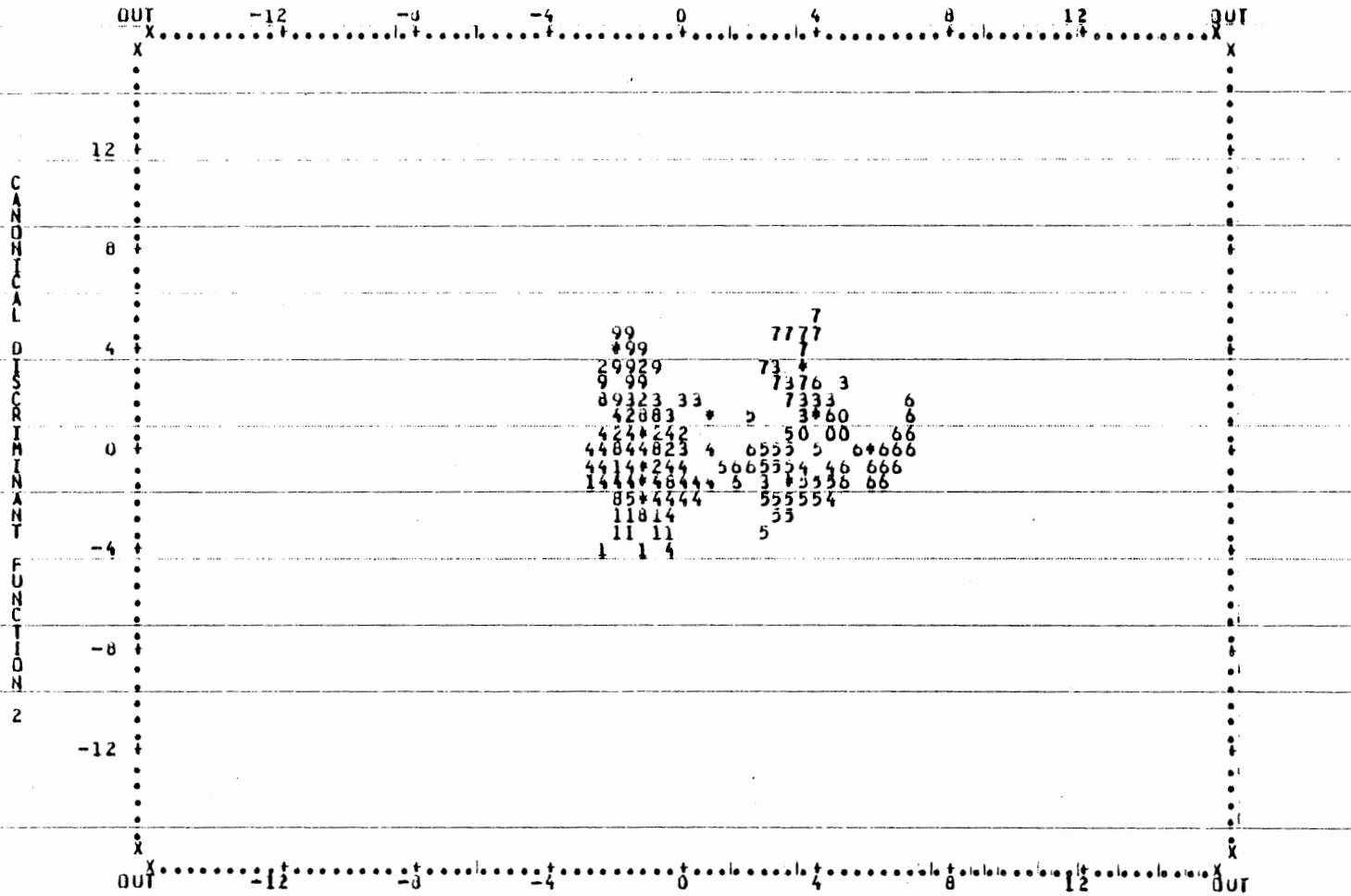
TERRITORIAL MAP ASSUMING ALL FUNCTIONS BUT THE FIRST TWO ARE ZERO * INDICATES A GROUP CENTROID

CANONICAL DISCRIMINANT FUNCTION 1



ALL-GROUPS SCATTERPLOT - * INDICATES A GROUP CENTROID

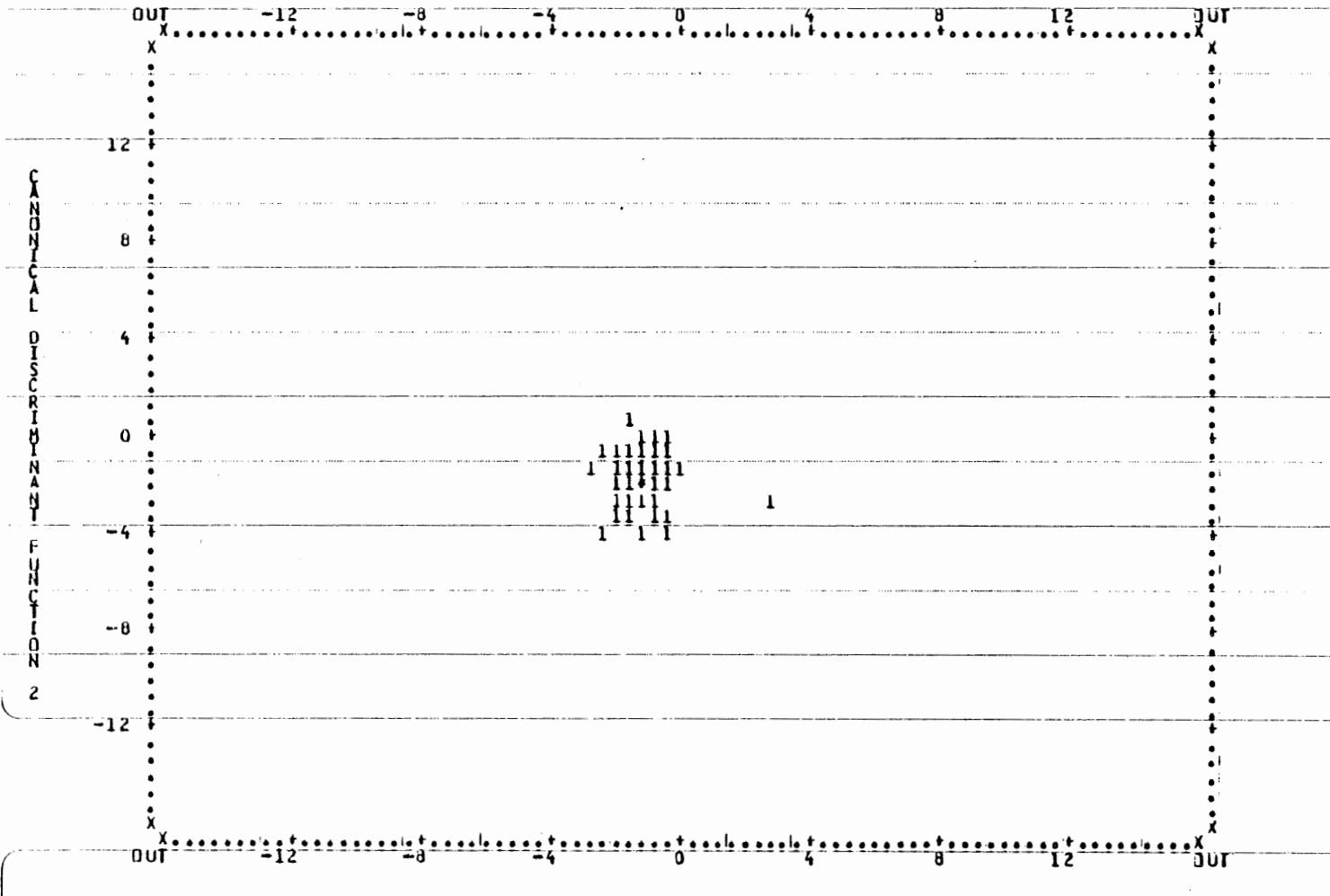
CANONICAL DISCRIMINANT FUNCTION 1



GROUP 1

* INDICATES A GROUP CENTROID

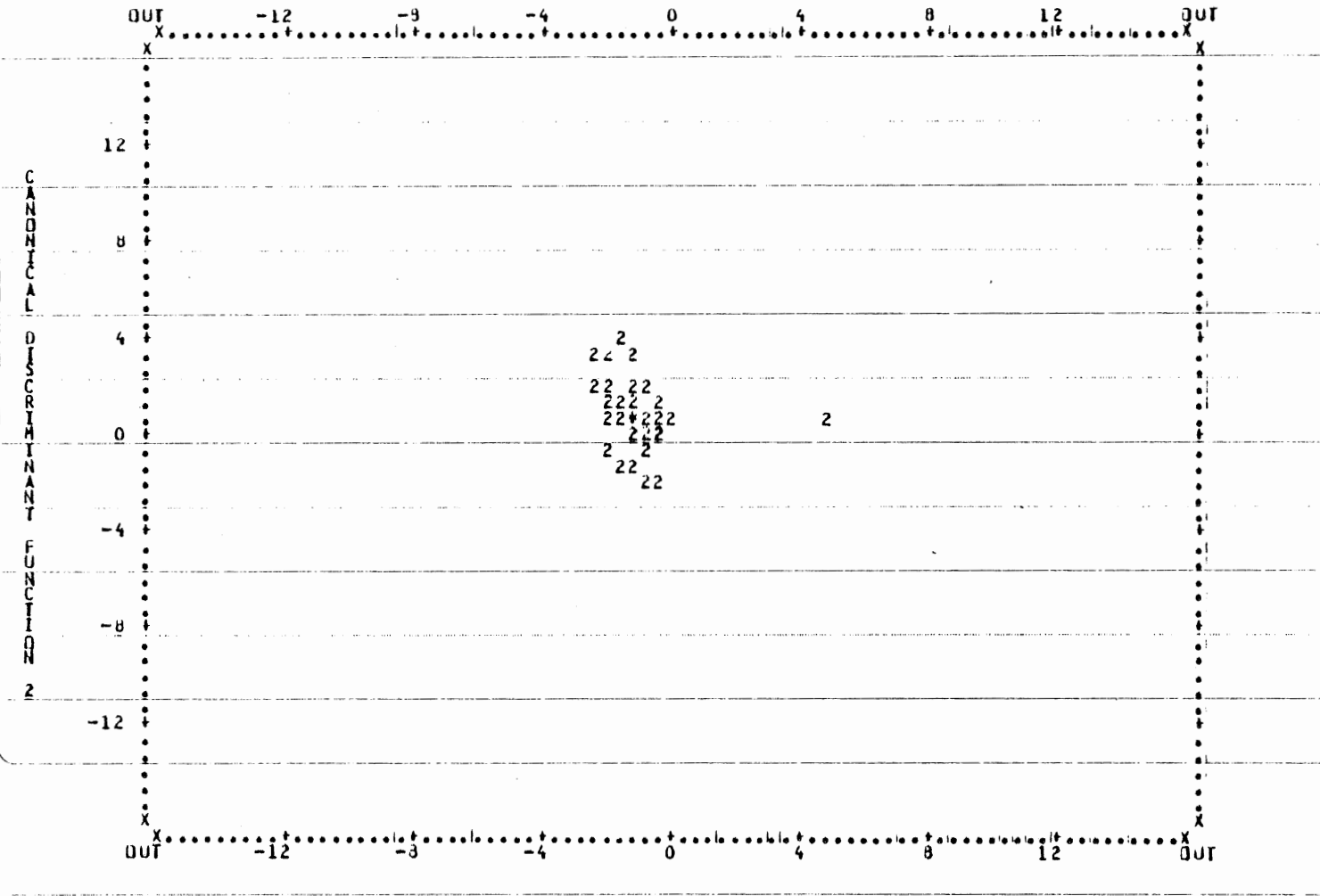
CANONICAL DISCRIMINANT FUNCTION 1



GROUP 2

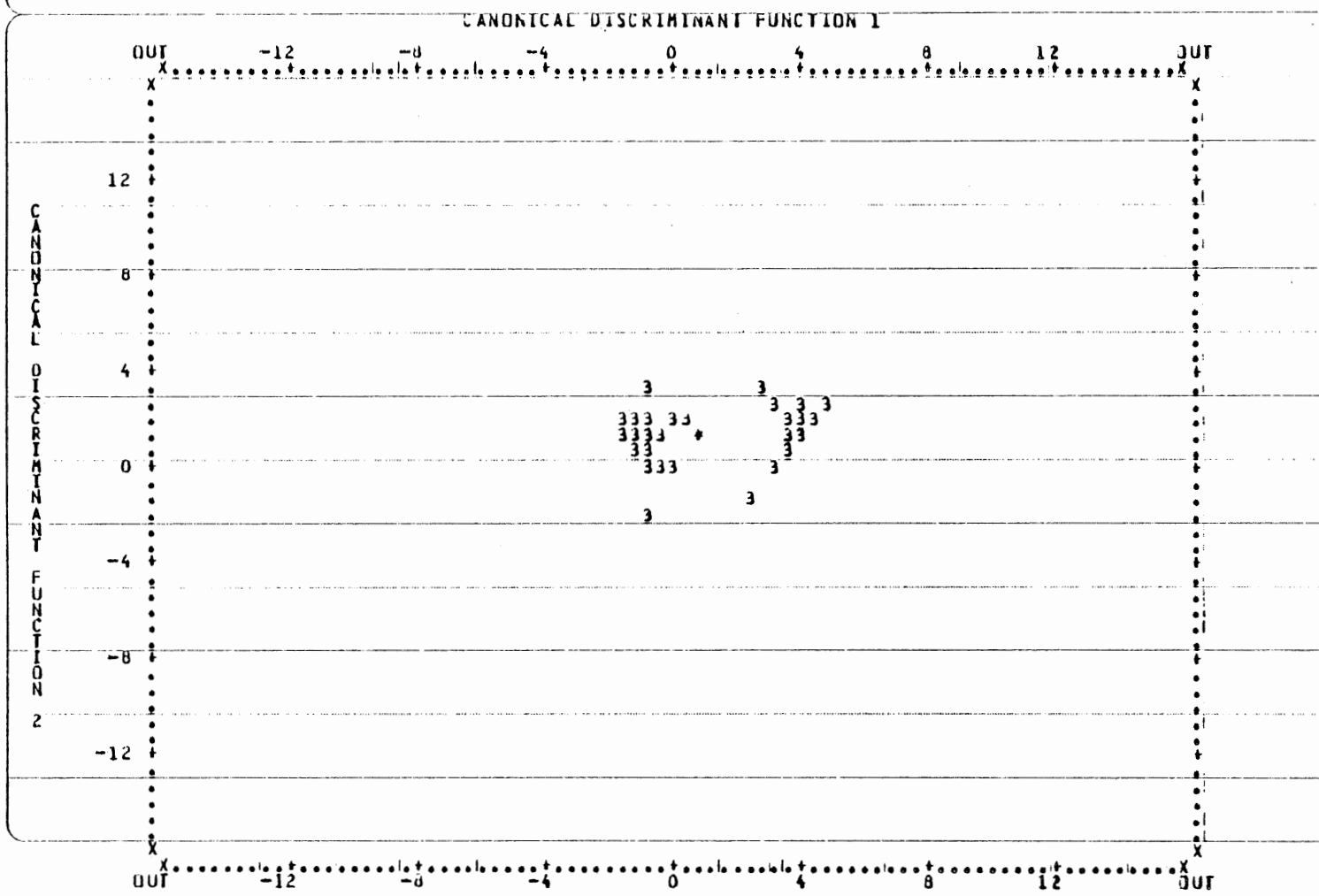
* INDICATES A GROUP CENTROID

CANONICAL DISCRIMINANT FUNCTION 1



GROUP 3

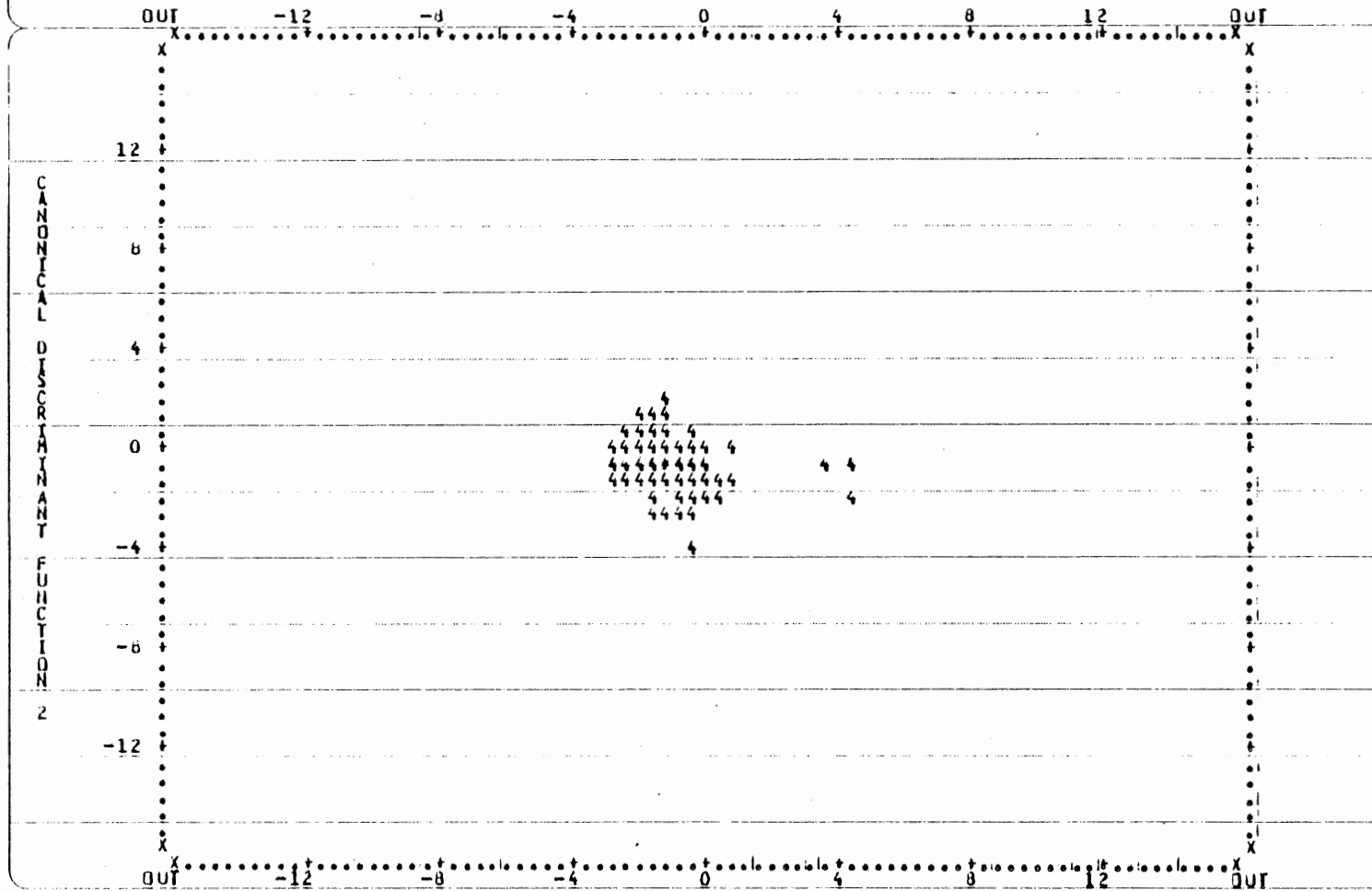
+ INDICATES A GROUP CENTROID

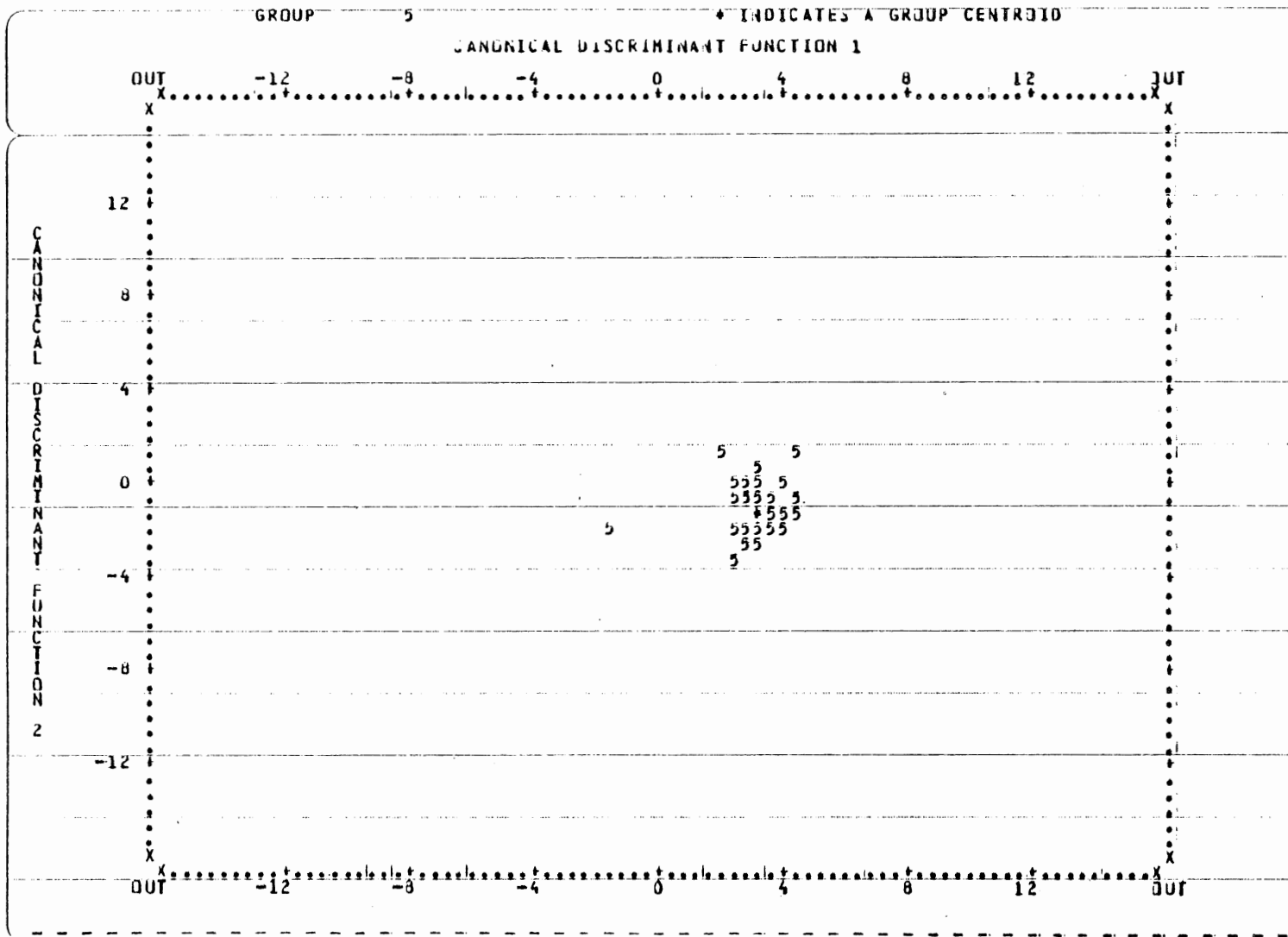


GROUP 4

* INDICATES A GROUP CENTROID

CANONICAL DISCRIMINANT FUNCTION 1

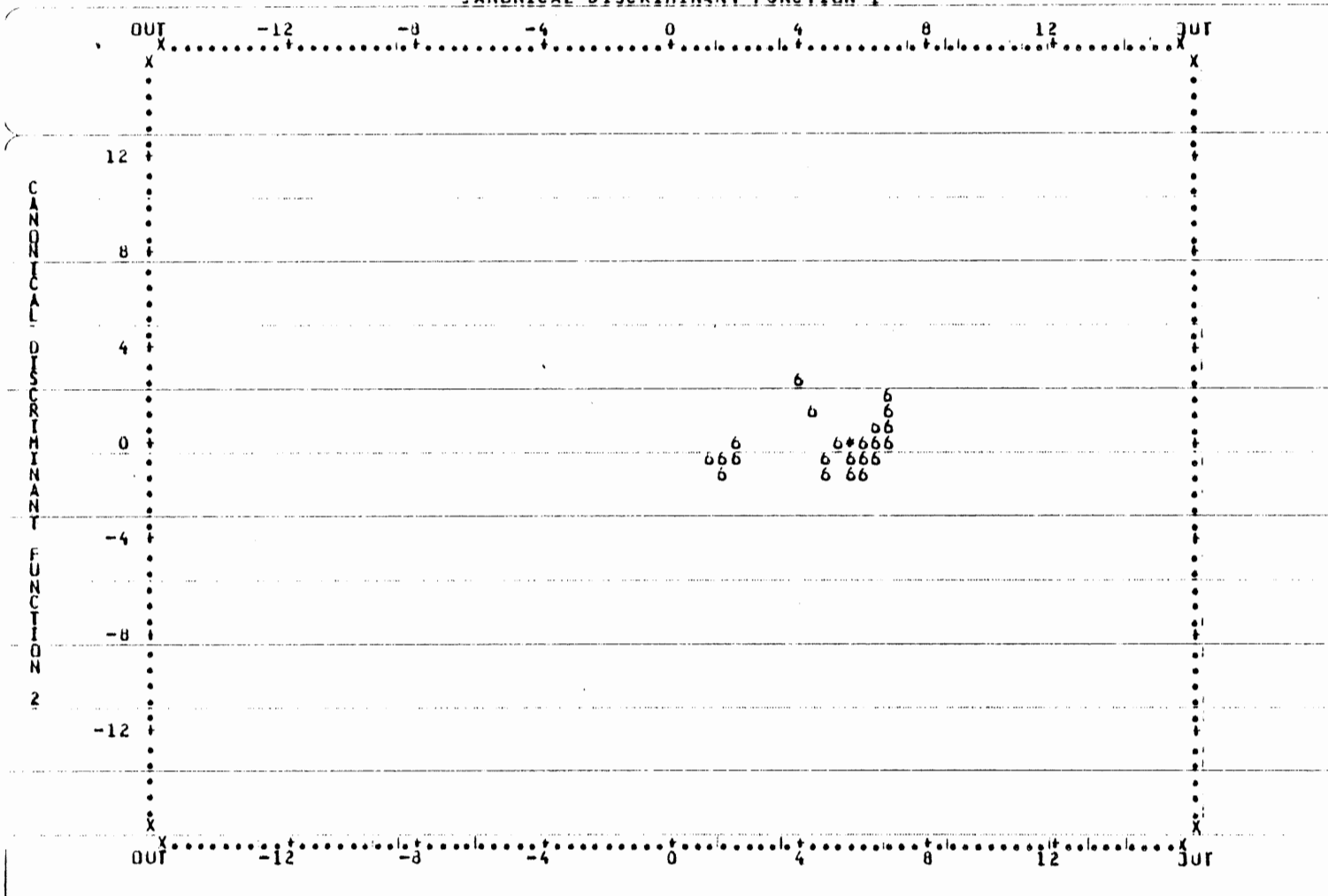




GROUP 6

* INDICATES A GROUP CENTROID

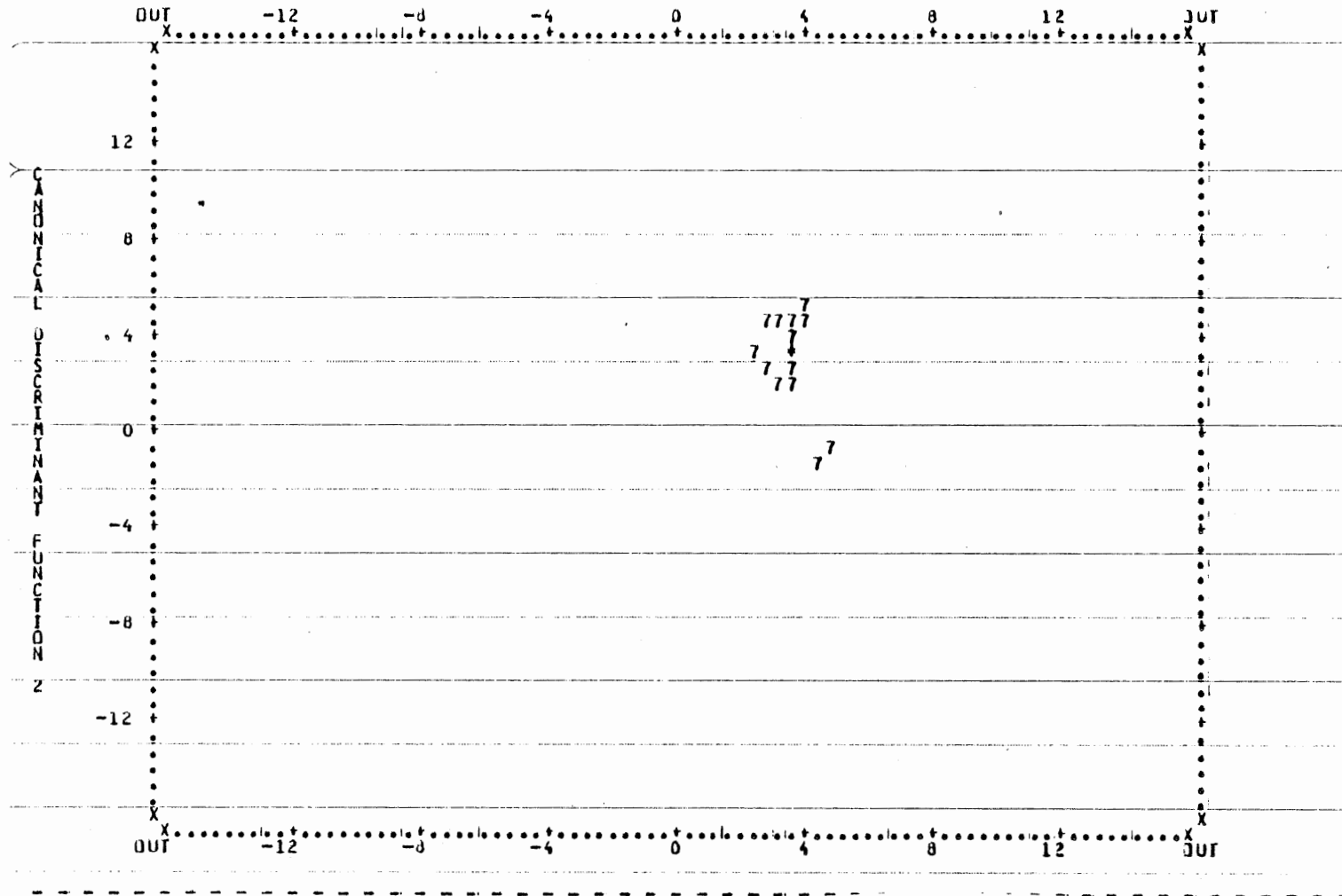
CANONICAL DISCRIMINANT FUNCTION 1



GROUP 7

* INDICATES A GROUP CENTROID

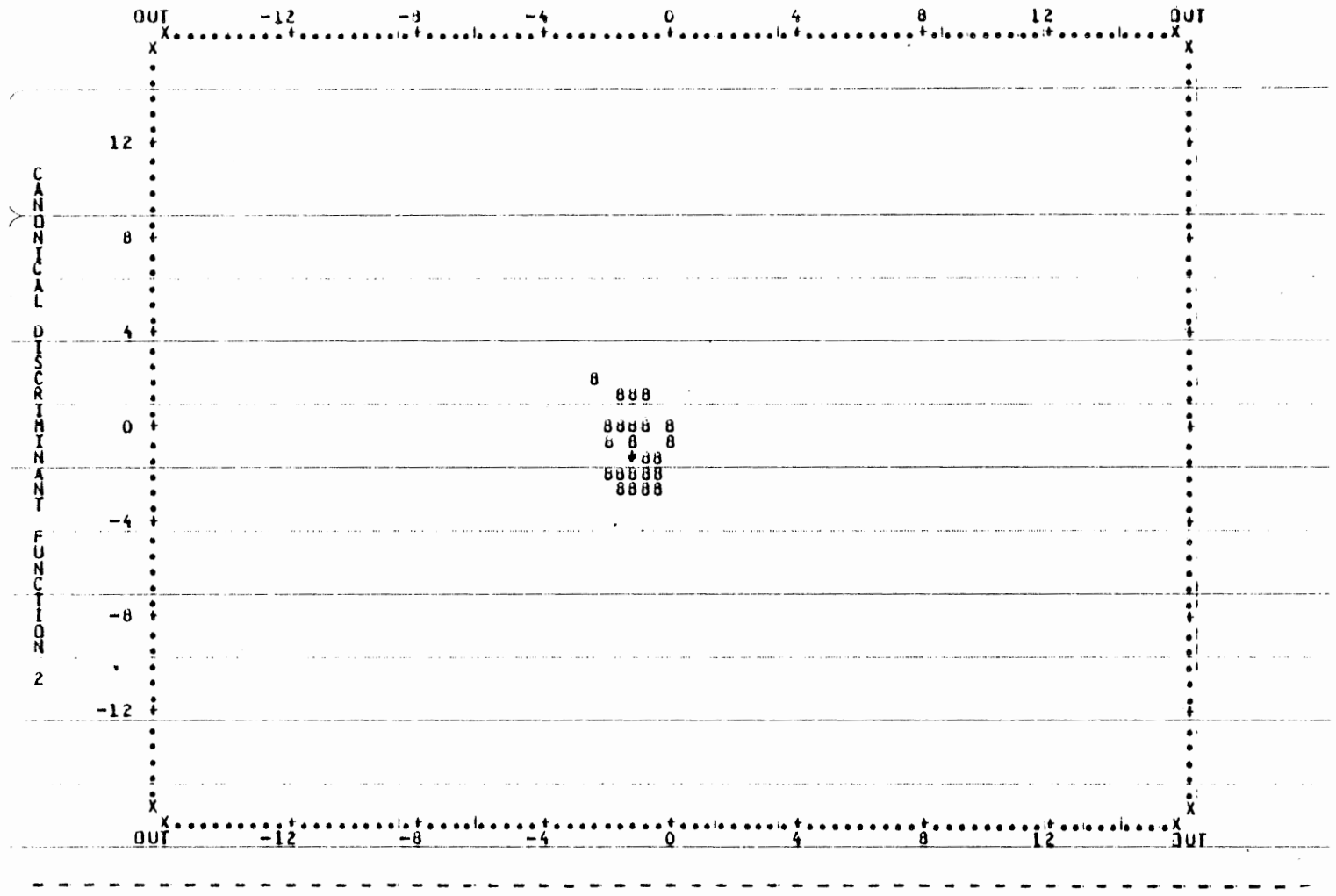
CANONICAL DISCRIMINANT FUNCTION 1



GROUP 9

* INDICATES A GROUP CENTROID

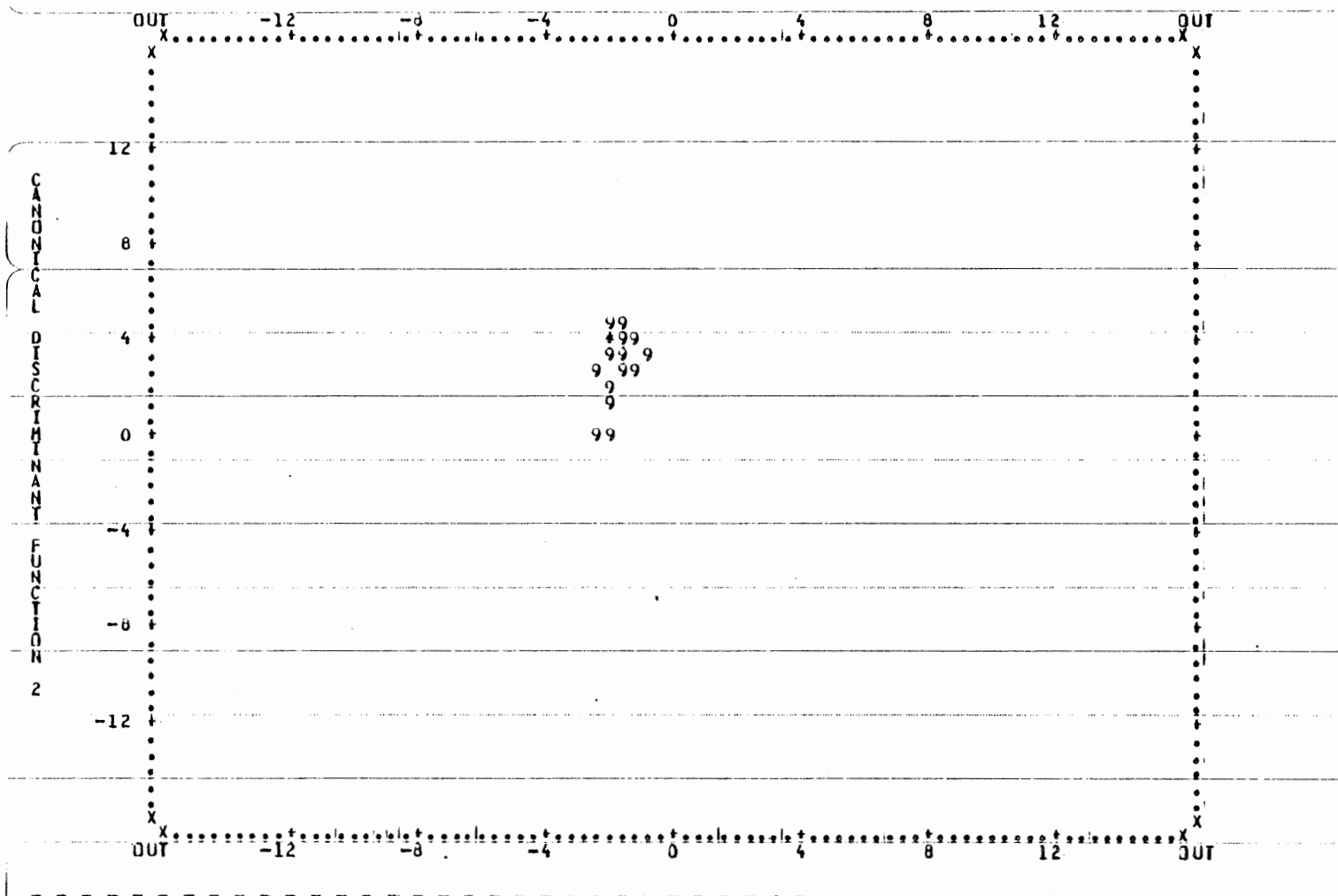
CANONICAL DISCRIMINANT FUNCTION 1



GROUP 7

* INDICATES A GROUP CENTROID

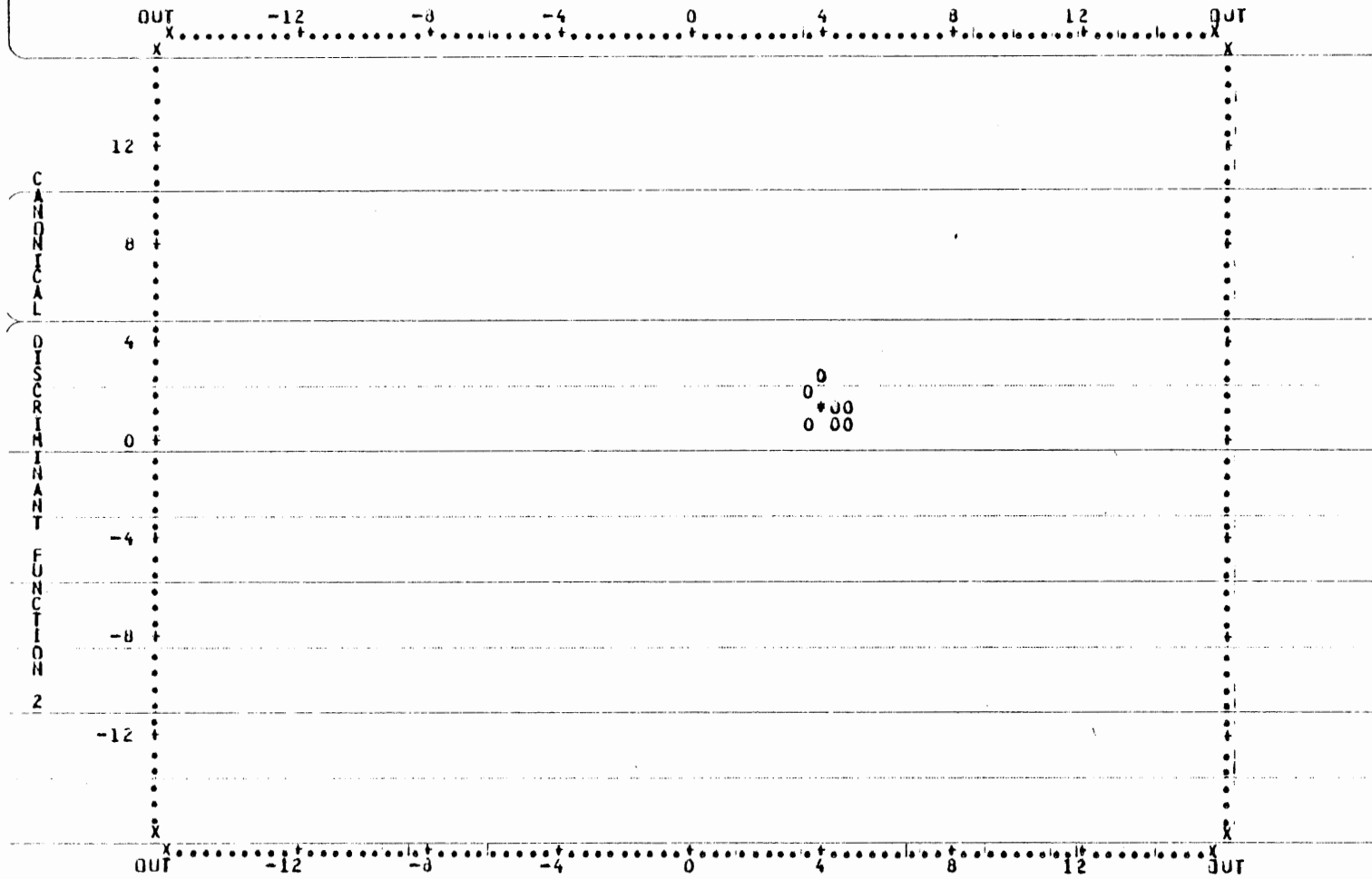
CANONICAL DISCRIMINANT FUNCTION 1



GROUP 10

* INDICATES A GROUP CENTROID

CANONICAL DISCRIMINANT FUNCTION 1



CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	1			1.	1 .4536 .9563	4 .0335	-1.9409	-2.9547	.5742	-1.9555	.2497
ELPHEXA	2			1.	1 .6547 .9691	4 .0308	.6544	-1.5119	-.8287	.4488	
ELPHEXA	3			1.	1 .9984 .9889	4 .0107	-2.0327	-3.0942	-.6501	-1.7069	.1665
ELPHEXA	4			1.	1 .7878 .9172	4 .0825	.6053	-1.0744	-.0336	.6582	
ELPHEXA	5			1.	1 .9975 .9954	4 .0044	-1.3267	-2.6328	-.2363	-.5742	-.6503
ELPHEXA	6			1.	1 .4707 .9141	4 .0957	-.4711	-.2813	-.4037	.3097	
ELPHEXA	7			1.	1 .6628 .9175	4 .0823	-1.8742	-2.8004	1.0176	-1.5584	-.0336
ELPHEXA	8			1.	1 .6880 .9112	4 .0887	.3154	-.9307	.3367	.4758	
ELPHEXA	9			1.	1 .6757 .9367	4 .0931	-1.0709	-2.3130	-.2608	-1.2105	-1.2877
ELPHEXA	10			1.	1 .9944 .9950	4 .0037	.4116	-.3962	-.4065	-.3999	
ELPHEXA	11			1.	1 .2423 .9185	8 .0791	-1.9071	-3.3369	1.1847	-1.0414	.1421
ELPHEXA	12			1.	1 .0053 .9446	4 .0502	-.7122	-.4568	-.1774	1.5795	
ELPHEXA	13	***		8	8 .3069 .7534	1 .2464	-1.8347	-3.1220	1.1009	-1.4029	.1511
ELPHEXA	14			1.	1 .1645 .9306	8 .0473	-.3951	-.7083	-.5044	.7852	
ELPHEXA	15			1.	1 .6032 .9815	2 .0127	-1.8244	-2.9246	1.1199	-1.6943	-.0049
ELPHEXA	16			2.	2 .6870 .9356	4 .0605	-.0745	-1.1467	-.4060	.1827	
ELPHEXA	17			2.	2 .3003 .7549	4 .1389	-1.8509	-2.9321	1.1638	-1.6643	-.0752
ELPHEXA	18			1.	1 .7724 .9730	2 .0184	-.0730	-1.2265	-.2244	.2645	
ELPHEXA	19			1.	1 .0565 .9343	2 .0426	-1.0917	-2.7078	-.2232	-.6276	-.9765
ELPHEXA	20			1.	1 .6820 .9582	4 .0328	-.2296	.4799	-.6018	.8050	
ELPHEXA	21			1.	1 .7722 .9932	4 .0039	-1.5272	-3.1236	-1.7465	-.7540	1.4298
ELPHEXA	22			1.	1 .9975 .9870	4 .0126	-.8488	-.5286	-.1960	-.9259	
ELPHEXA	23			1.	1 .9989 .9937	4 .0042	-2.2893	-3.9773	-.7816	-1.0860	2.7446
ELPHEXA	24			1.	1 .9919 .9952	4 .0033	-1.7410	-.9600	-.2337	-.2645	
ELPHEXA	25			1.	1 .0134 .7921	8 .1784	-1.6477	-2.9248	-2.5195	-.3375	1.2479
ELPHEXA	26			1.	1 .7064 .5922	4 .4045	-.5914	-.3911	-.1179	-.8445	
ELPHEXA	27			1.	1 .9680 .9210	4 .0714	-1.6144	-3.5214	-1.5303	-.6296	1.3567
ELPHEXA	28			1.	1 .4872 .6410	4 .9576	-1.3427	.2128	.5021	-.9082	
ELPHEXA	29			1.	1 .6684 .7377	2 .1436	-1.5711	-2.0338	-.2394	.5687	-.6727
							.9884	1.8889	.4726	1.3355	
							-.9188	-.0017	.3888	1.9359	.8653
							1.6239	.3009	.6492	1.3081	
							-.3813	.9146	1.5153	2.5993	-.2374
							-.2723	.9351	1.2113	-.8716	
							-1.1630	-1.3350	-.4982	.5672	-.9798
							1.1025	1.3848	1.0109	-.3616	
							-.5807	-1.0942	-.7687	2.0467	-1.9162
							-2.2711	2.2997	1.3046	.3455	
							-1.5348	-2.0147	.5752	.1122	-.7630
							1.1466	1.1590	1.0551	1.4873	
							-.7765	-2.0166	-.2928	.7591	-1.5678
							-1.0488	.6124	.4405	1.1506	
							-1.3799	-2.3904	.1826	-.7019	-.7957
							-.0278	-.7374	.0295	.6258	
							-1.0823	-2.5568	.3738	-1.0584	-1.2659
							.0038	-.0881	-.4239	-.3228	
							-1.0591	-1.6250	-.2918	-.5310	-1.6118
							.5903	-.6283	-.2421	.5428	
							-2.0220	-2.7553	-1.3626	-.3126	2.3629
							-.3910	.7009	.6540	-2.5771	
							-2.1384	-2.4668	1.4382	-1.0170	.4773
							.8385	1.322	.4587	.2734	
							-1.5580	-1.7598	.4931	-.6612	-.2883
							1.0790	.3380	.1636	-1.0275	
							-2.1186	-2.4375	1.5300	-1.3880	.3814
							.9849	.3232	.6212	-1.2171	
							-.9536	-1.8875	-.6335	-.2333	-.0081
							1.2711	.6318	-1.8118	-.1307	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP PIX/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES					
ELPHEXA	30			1.	1 .9577 .9558	4 .0257	-1.7566	-2.2021	-.0351	.0111	-.2703	.1148
ELPHEXA	31			2.	2 .5824 .9752	4 .0045	1.1702	.6548	-.0351	.6334	-.2703	.5942
ELPHEXA	32			2.	2 .4060 .8032	4 .1944	1.5753	.2071	-1.8239	.3563	2.6818	1.7211
ELPHEXA	33			2.	2 .4805 .9558	4 .0430	1.6930	-.5588	-2.1747	1.6474	1.4290	1.3710
ELPHEXA	34			1.	1 .1696 .7267	8 .2620	1.7057	-.7803	-2.4334	1.2007	1.7563	1.3367
ELPHEXA	35			2.	2 .4421 .7488	4 .2507	-1.7554	-2.1284	-1.6192	-.0403	1.3367	1.6117
ELPHEXA	36			2.	2 .8390 .8319	9 .1437	-.0257	.3165	-.4684	-2.2050	1.6117	1.0578
ELPHEXA	37			1.	1 .9785 .9734	4 .0186	1.6953	-.6828	-1.8924	1.0291	1.0578	1.0578
ELPHEXA	38			1.	1 .9943 .9955	4 .0044	1.0403	-.5806	-.3899	-.3539	1.0578	1.0578
ELPHEXA	39			1.	1 .9689 .9951	4 .0047	1.4988	1.2788	.1064	-.6137	1.0578	1.0578
ELPHEXA	40			1.	1 .9991 .9881	4 .0115	-1.2020	-2.4836	-.5582	-.9785	1.0578	1.0578
ELPHEXA	41			1.	1 .8699 .9054	4 .0945	-.3267	-.1321	.6899	-.2887	1.0578	1.0578
ELPHEXA	42			1.	1 .9973 .9959	4 .0038	-1.8108	-2.7247	.8726	-.4416	1.0578	1.0578
ELPHEXA	43			1.	1 .9580 .9805	2 .0121	-1.810	.1880	.1873	1.3673	1.0578	1.0578
ELPHEXA	44			1.	1 .8199 .9727	4 .0228	-1.3222	-.25110	-.3073	.0242	1.0578	1.0578
ELPHEXA	45			1.	1 .9017 .9898	4 .0085	-1.008	-.6652	-.3073	.0242	1.0578	1.0578
ELPHEXA	46			1.	1 .9420 .9704	4 .0255	-1.8587	-2.9343	-1.1768	-.16555	1.0578	1.0578
ELPHEXA	47			1.	1 .6092 .9880	4 .0076	-.0726	-1.2501	-1.706	-.2888	1.0578	1.0578
ELPHEXA	48			1.	1 .9901 .9985	4 .0012	-1.1016	-2.7306	-.2960	-.6774	1.0578	1.0578
ELPHEXA	49			1.	1 .6018 .9866	4 .0104	-.2667	.3922	-.5025	-.6067	1.0578	1.0578
ELPHEXA	50			2.	2 .7352 .9419	4 .0576	-1.5112	-1.7090	-.2263	-.0766	1.0578	1.0578
ELPHEXA	51			1.	1 .9095 .9875	4 .0120	1.3603	.8857	.0653	.6542	1.0578	1.0578
ELPHEXA	52			1.	1 .9863 .9862	4 .0132	-1.4484	-.21932	.6091	-.1139	1.0578	1.0578
ELPHEXA	53			1.	1 .4854 .9157	4 .0832	-.9179	1.8864	.7812	-.1173	1.0578	1.0578
ELPHEXA	54			1.	1 .9788 .9582	4 .0188	-1.5522	-2.2850	.2264	-.5881	1.0578	1.0578
ELPHEXA	55			1.	1 .9856 .9223	4 .0742	1.5228	1.3798	.3754	-.4037	1.0578	1.0578
ELPHEXA	56			1.	1 .7916 .9873	4 .0091	-1.4036	-2.4068	.7025	-.1673	1.0578	1.0578
ELPHEXA	57			1.	1 .9435 .9953	4 .0104	-.5630	1.5522	.1212	.1056	1.0578	1.0578
ELPHEXA	58			1.	1 .9326 .9709	4 .0258	-1.42209	-1.5069	.4348	-.0053	1.0578	1.0578

CASE SUBFILE SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	59		1.	1 .9215 .9705	2 .0070	-1.3223	-2.2520	-.0488	1.5335	-1.1277
ELPHEXA	60		1.	1 .8255 .9914	2 .0058	.8373	2.2651	-.0495	1.6766	-1.3781
ELPHEXA	61		1.	1 .8260 .9889	4 .0072	-1.3098	-1.8304	-.0798	.0491	-1.6800
ELPHEXA	62		1.	1 .2840 .9227	2 .0726	1.5342	1.4442	.1568	.8321	-1.6800
ELPHEXA	63		1.	1 .9224 .9988	4 .0010	-1.0737	-1.6358	.4844	-.2993	-1.0737
ELPHEXA	64		1.	1 .9995 .9957	4 .0027	.9989	1.5240	.0307	-1.1073	-1.8639
ELPHEXA	65		1.	1 .9995 .9580	4 .0309	-.9077	-.1606	-1.1881	-.7142	-2.1512
ELPHEXA	66		1.	1 .1422 .9369	4 .0595	.7016	.1605	-2.1799	-.1238	-1.6184
ELPHEXA	67		1.	1 .9410 .8232	4 .1669	-.9457	-2.3845	.1623	-.0896	-1.8732
ELPHEXA	68		4. ***	1 .2776 .9939	4 .0154	-1.355	1.0156	.4549	.7993	-1.9105
ELPHEXA	69		4. ***	1 .6550 .9910	4 .0063	-1.1182	-2.0187	-.1140	-.2470	-1.8732
ELPHEXA	70		4. ***	1 .1309 .9887	4 .0072	.0894	.1898	.2682	.4412	-1.9105
ELPHEXA	71		4. ***	1 .5167 .9911	4 .0068	-1.1713	-2.0991	.4972	-.7005	-1.4016
ELPHEXA	72		4. ***	1 .0961 .9059	4 .0907	-.3638	-.4525	-.0744	-.0288	-1.8732
ELPHEXA	73		4. ***	1 .8308 .9305	4 .0162	-1.929	-2.1698	-.9429	-.7815	-1.8732
ELPHEXA	74		4. ***	2 .9632 .9347	4 .0452	-1.929	-.6972	-2.5207	-2.1606	-1.8732
ELPHEXA	75		2.	2 .8104 .9832	4 .0163	-1.4674	-1.9287	1.0290	-.1024	-2.1046
ELPHEXA	76		2.	2 .9817 .9279	4 .0680	.0298	1.1251	.5257	-.6741	-1.8732
ELPHEXA	77		2.	2 .8557 .9753	4 .0218	-.8693	-2.2362	.8414	-1.2499	-1.8732
ELPHEXA	78		1.	1 .7093 .9956	4 .0042	-1.3391	-1.9359	.9463	-.7160	-1.8732
ELPHEXA	79		4. ***	1 .6138 .7750	2 .1635	-2.295	-.3323	-.6442	1.0740	-1.8732
ELPHEXA	80		4. ***	2 .8592 .7386	4 .2597	-1.4827	-1.3702	-.5778	-.3926	-1.8732
ELPHEXA	81		4.	4 .0177 .6280	2 .3608	-1.6724	-1.2699	-.7396	-.3733	-1.8732
ELPHEXA	82		5.	5 .6873 1.0000	7 .0000	-2.0817	-.9557	-.1963	-.2655	-1.8732
ELPHEXA	83		1.	1 .6965 .9006	4 .0993	.7496	-.7455	-.6033	.4455	-1.8732
ELPHEXA	84		4.	4 .4884 .5095	2 .4481	-2.6391	-1.5802	1.4324	-2.9753	-1.8732
ELPHEXA	85		1.	1 .9503 .9961	4 .0037	1.1777	1.5827	-.0231	-.7213	-1.8732
ELPHEXA	86		1.	1 .9603 .9985	4 .0113	-1.0351	-1.1353	.1309	-.3216	-1.8732
ELPHEXA	87		4. ***	2 .7429 .7108	4 .2722	-.3474	.0142	-.5181	.3832	-1.8732
						-1.7411	1.3145	.6169	.3913	.9744
						1.0555	.2614	.5273	.6131	1.0385
						1.3834	-1.699	-2.4483	.5692	1.0274
						.9452	.0845	-.6365	.7264	.7912
						-1.5132	.6877	-.8090	.8969	.7912
						.2372	1.4955	-.1677	2.0018	-1.3837
						-.9886	-2.1414	-.2908	-1.7236	-1.3837
						.6409	-.7451	-.7277	-1.7536	-1.3837
						-2.1238	-.0841	-.2666	-1.4162	-1.3837
						.4882	1.2012	-.1350	-.5040	-1.3837
						-1.9428	.6897	1.5787	-.6174	1.4087
						1.2163	.6072	-1.1304	.4385	.9380
						-1.8077	1.2700	1.7209	-1.8873	.9380
						1.9995	-.6459	-1.4036	-2.7382	1.8391
						-3.3734	-2.6366	-.3067	-3.3104	1.8391
						-.1111	.1720	.7090	-1.3957	-1.2526
						-1.4648	-2.6057	.9939	-1.3942	-1.2526
						-.4434	-.7741	.5544	-1.5373	.6246
						-1.9226	1.0349	.8514	-.7379	.6246
						.4852	-1.7328	.3460	.2643	-1.3668
						-1.0707	-2.2520	.2769	-1.4066	-1.3668
						.6406	-.2063	-.0409	-1.2750	-1.6629
						-1.0093	-2.2006	.8098	-.9706	-1.6629
						-.0155	.0944	.7011	-.9475	1.1254
						-1.8435	.5528	-.7863	-1.2387	1.1254
						-.4940	-.8957	-.8140	1.4086	

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES					
SUBFILE	VAL	GROUP	GROUP P (X7G) P (G7X)	GROUP P (G7X)						
SEQNUM	SEL									
ELPHXA	88	1.	1 .9679 .9984	4 .0016	-.8290	-2.4522	.4231	-1.3731	-1.7840	
ELPHXA	89	2.	2 .9903 .9760	9 .0094	.5270	.2034	-.5460	-.6832		
ELPHXA	90	1.	1 .9915 .9986	4 .0012	-1.4051	1.3424	.4540	.9042	.3003	
ELPHXA	91	1.	1 .5935 .9956	2 .0022	.4113	1.4106	-.1045	.3884		
ELPHXA	92	1.	1 .9870 .9980	4 .0018	-.8231	-2.0419	.0715	-.6527	-2.1694	
ELPHXA	93	1.	1 .8348 .9895	2 .0077	.2331	.2014	-.3825	-.0760		
ELPHXA	94	8.	8 .2348 .9999	2 .0000	-.5754	-.9016	-1.2148	.0909	-2.5254	
ELPHXA	95	8.	8 .3650 1.0000	2 .0000	.9270	-.0125	.1183	.1189		
ELPHXA	96	1.	1 .5983 .8589	8 .0304	-.9404	-2.4062	.3885	-1.0109	-1.8592	
ELPHXA	97	1.	1 .8621 .9932	2 .0026	.6806	.0817	.0938	.5507		
ELPHXA	98	2.	2 .6766 .5334	9 .3329	-1.2573	-1.5381	-.1868	-.2002	-1.3753	
ELPHXA	99	4. ***	2 .1152 .5001	4 .6728	1.9017	1.1736	-.0064	.4255		
ELPHXA	100	3.	3 .0413 .5326	4 .4429	-1.2558	-.0533	-3.7396	.8484	1.1124	
ELPHXA	101	3.	3 .1005 .5974	4 .3510	-.3682	1.0203	-2.1244	-2.1619		
ELPHXA	102	3.	3 .1436 .9545	4 .0316	-1.2558	-.0316	-3.7782	.8483	1.1315	
ELPHXA	103	2. ***	6 .0157 .6685	10 .2835	-.3190	1.0301	-2.0280	-1.8090		
ELPHXA	104	7. ***	5 .2270 .9993	7 .0004	-1.3372	-2.5395	-2.0502	-.84106	.4261	
ELPHXA	105	1.	1 .1340 .8837	2 .0815	-.3329	-.0869	.4339	-1.0966		
ELPHXA	106	1.	1 .8695 .9976	4 .0015	-.8002	-1.2257	-1.1889	.4603	-1.9281	
ELPHXA	107	1.	1 .8317 .9673	4 .0292	-.0391	-.5227	-.0234	.2370		
ELPHXA	108	1.	1 .6717 .9998	4 .0002	-1.2677	1.7750	.2795	.3377	-.2303	
ELPHXA	109	3.	3 .1593 .9851	4 .0131	-.0803	-1.2910	.9378	.6357		
ELPHXA	110	1.	1 .6192 .9987	4 .0013	-1.0093	.9245	-3.7231	.3444	1.4543	
ELPHXA	111	4.	4 .3702 .9329	3 .0524	-.8151	1.2243	-1.7845	-1.7978		
ELPHXA	112	1.	1 .6887 .9962	8 .0019	-.6596	-2.2677	-1.2486	-.0869	.1534	
ELPHXA	113	6.	6 .9985 1.0000	10 .0000	-2.1232	-2.3526	-1.8721	1.7827		
ELPHXA	114	6.	6 .9235 1.0000	10 .0000	-.0377	-.3233	1.2143	1.8278	-1.1555	
ELPHXA	115	1.	1 .7433 .9974	2 .0013	-1.4487	-2.0890	1.2179	1.3697		
ELPHXA	116	7. ***	5 .0043 .6843	6 .2258	-2.2110	-2.7961	.9331	-.8179	-.7390	
					4.6847	.6835	-.0450	-.4722	1.4252	
					2.9105	.3582	-1.0121	.8839	-.9503	
					1.4802	.3440	-1.9590	.6966		
					-.6726	-.1434	-2.4955	-.7289	-1.9360	
					-.1221	.7543	-2.4955	-1.2909		
					-.8876	-1.3431	-.8858	-.3672	-1.7642	
					.9816	-.7966	.7631	-.5240		
					-.3733	-2.2189	1.0193	-.2765	-1.5151	
					.2103	-.2889	-1.5719	-.1603		
					-.5924	-2.1132	-.4956	-1.2003	-2.7330	
					.1793	-1.2275	-.2580	.5463		
					-.8979	.9070	1.6999	.6125	-.3833	
					-3.4624	-1.8719	.1571	1.5973		
					-.3484	-.36349	.1340	.5157	-1.5977	
					1.0282	-.6149	.3633	.8624		
					-1.5571	.9450	2.7567	-.8894	.2052	
					-1.0314	-.9575	1.4226	-1.4418		
					-1.0405	-1.6386	-1.4248	-.1106	-1.3425	
					1.1631	-.9902	.0533	1.3041		
					6.2283	-.1563	.4237	2.2569	-.8196	
					-.0678	.3333	.1016	.4311		
					6.4715	.2985	.8319	1.8156	-1.1244	
					-.1615	.1407	-.0053	-1.3676		
					-.6232	-1.3670	-1.0301	.3123	-2.4647	
					.1682	.0093	.2376	.6852		
					4.9734	-.5941	-2.0394	-1.1326	-1.2114	
					.1283	.7134	-2.8387	.0027		

CASE	SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST GROUP	PROBABILITY P(X/G)	P(G/X)	2ND HIGHEST GROUP	P(G/X)	DISCRIMINANT SCORES				
ELPHEXA		117			6.	6	.9486	1.0000	10	.0000	6.4380	.5609	.6623	1.7099	-1.0840
ELPHEXA		118			1.	1	.8401	.9971	2	.0015	.2783	-.1050	-.0500	-1.2841	
ELPHEXA		119			1.	1	.4134	.9999	4	.0001	-.3409	-1.3699	-1.0921	.3392	-2.3209
ELPHEXA		120			7.	7	.4129	1.0000	5	.0000	.1139	-.1846	-.3145	.1294	
ELPHEXA		121			7.	7	.6585	1.0000	5	.0000	-.8384	-2.3828	-.8552	-1.1206	-2.6075
ELPHEXA		122			7.	7	.8062	1.0000	5	.0000	.5603	-1.4692	-.1410	1.4973	
ELPHEXA		123			7.	7	.4867	1.0000	5	.0000	3.3343	2.2486	-1.9926	-3.7739	-.6381
ELPHEXA		124			1.	1	.4140	.9785	2	.0168	-.5002	1.2869	-2.9883	-.5841	
ELPHEXA		125			8.	8	.2147	.9958	1	.0042	3.5541	2.8366	-.9828	-3.8689	-1.4560
ELPHEXA		126		3. ***	5	5	.2775	.9947	3	.0041	-1.0920	.5701	-2.0217	-1.1191	
ELPHEXA		127		3. ***	7	7	.1571	.8929	3	.0616	3.4785	2.6394	-2.6603	-3.5361	-.7641
ELPHEXA		128		3. ***	5	5	.1640	.9932	3	.0051	-.1626	.3075	-2.4304	-.1062	
ELPHEXA		129			6.	6	.7992	1.0000	10	.0000	-1.6774	2.6685	-1.5128	-3.1015	-1.3683
ELPHEXA		130			8. ***	1	.0194	.9094	2	.0189	-.6263	.9443	-1.5128	-.7611	
ELPHEXA		131			1.	1	.9260	.9502	4	.0323	-.7413	-.2491	-.3050	-.3679	1.4053
ELPHEXA		132			5.	5	.2490	.9999	7	.0001	.1327	-1.3033	-2.2090	-.9749	
ELPHEXA		133			6.	6	.9845	1.0000	10	.0000	3.0740	-.0427	.0765	-3.4256	-.4558
ELPHEXA		134			3.	3	.0991	.8117	1	.1359	-1.5696	1.1588	.3627	1.0921	
ELPHEXA		135			6.	6	.9520	1.0000	10	.0000	3.1407	2.3394	.5376	-2.2768	1.0492
ELPHEXA		136			8.	8	.9794	.9998	1	.0002	-.5111	-.9889	1.0242	.2583	
ELPHEXA		137		4. ***	2	2	.2932	.5019	4	.2839	3.0573	-.2461	.2687	-3.5218	-.5819
ELPHEXA		138			6.	6	.2351	1.0000	5	.0000	-1.8803	1.5877	.7816	.0908	
ELPHEXA		139			1.	1	.8767	.9520	4	.0376	6.5175	2.284	.7289	1.1518	-1.1220
ELPHEXA		140			3.	3	.6666	.5575	4	.2932	.1932	-.1288	-.1871	-1.8248	
ELPHEXA		141			1.	1	.4482	.9508	8	.0375	.0643	-.7131	-1.7181	-.5964	-2.5603
ELPHEXA		142			5.	5	.1174	1.0000	7	.0000	1.4623	-.5384	-2.4198	-1.2830	
ELPHEXA		143			6.	6	.0109	.9788	5	.0012	-.8474	-2.1864	-.4732	-.4668	-1.6258
ELPHEXA		144			2.	2	.8110	.7563	4	.2389	-.9152	.4954	.3948	-.1102	
ELPHEXA		145			3.	3	.0474	.5796	10	.4188	2.5838	-.0696	-3.8268	-3.0639	1.9654
											-1.1457	-.4158	.9989	-.5726	
											8.4738	2.2405	-.8053	2.0158	-1.0323
											-.3864	-.0355	-.3916	-.4741	
											-.8216	-.3883	-.2141	-2.1349	-2.1126
											-1.3098	-.0976	-1.9292	-2.1349	
											6.3891	.2702	.2298	1.5159	-.7792
											.2655	-.1218	-.6618	-1.2068	
											-1.0470	-1.8671	-3.8600	.7299	.1881
											-.8745	-.3634	-.6464	-.6763	
											-1.3147	1.8522	1.2095	-1.1840	.1525
											-.4239	.7702	1.5491	-1.1428	
											5.9218	-1.6533	-1.0552	.9751	-2.2040
											-.3723	.7619	-1.0214	.2996	
											-1.1978	-1.9697	.7395	-1.1255	-1.2907
											-.1500	-1.1602	.3581	-1.1778	
											-1.1295	1.4341	2.1949	-.5164	-.2073
											-1.0644	-.3853	-.0365	-.9846	
											-1.1703	-2.7281	-1.4511	-0.130	.0293
											-1.4600	.6987	.6114	-1.3153	
											4.3307	-1.5866	-3.4124	-1.5945	1.2183
											-.2314	-.1735	-1.0582	-1.7411	
											5.4776	-1.1380	-2.0192	1.3987	2.4389
											-.3859	-.7022	-2.0066	-.2514	
											-.7744	.2001	1.0108	1.7927	.4144
											1.1566	.1962	1.0788	.0341	
											3.8339	2.0475	1.0355	-.7901	.4912
											-2.5436	.8120	1.5312	1.3309	

CASE SUBFILE	SEJNUM	MIS VAL	SEL	ACTUAL SPJOP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	146			1. ***	2 .6549 .6484	4 .2700	-.4761	-.1333	.9985	1.3245	-1.1013
ELPHEXA	147			1.	1 .3548 .9737	4 .0239	-.20129	-.5430	-.3147	-2.2339	-.8360
ELPHEXA	148			5.	5 .7228 1.0000	7 .0000	3.5028	-1.7650	-3.2536	-2.1493	1.4302
ELPHEXA	149			5.	5 .2666 1.0000	7 .0000	3.7214	-1.6188	-3.6907	-2.1143	1.2293
ELPHEXA	150			3.	3 .1000 .8187	10 .0883	-.6824	-.1090	.6598	-1.7038	.6290
ELPHEXA	151			5.	5 .1331 1.0000	7 .0000	-1.8351	-.1995	.6115	1.0609	
ELPHEXA	152			5.	5 .7041 1.0000	8 .0000	-2.4502	.4309	-.34257	-2.4607	2.1456
ELPHEXA	153			2. ***	1 .8997 .9533	2 .0419	3.3844	-2.4398	-.3008	-2.2580	1.7080
ELPHEXA	154			1.	1 .6098 .9944	3 .0031	-.3174	.5215	-.2597	-.8735	
ELPHEXA	155			1.	1 .9223 .9977	2 .0013	-1.3506	-1.3141	-.6662	-.4413	-1.0012
ELPHEXA	156			3. ***	5 .0251 .7542	7 .1524	.7450	1.1857	-.7362	.1026	-2.7481
ELPHEXA	157			1.	1 .6063 .9983	4 .0016	-.4966	-1.3535	-.4459	.7028	
ELPHEXA	158			2.	2 .7680 .9118	4 .0875	-.2982	.3510	-.5926	.1644	-2.4543
ELPHEXA	159			4. ***	1 .3462 .6936	4 .2565	.7349	-1.3907	-.4450	-.2348	
ELPHEXA	160			5.	5 .9353 1.0000	7 .0000	.2986	.0408	-.6498	-.0295	-1.7131
ELPHEXA	161			5.	5 .4287 1.0000	1 .0000	3.7163	.9120	-.3167	-2.8892	-1.5777
ELPHEXA	162			7.	7 .0942 1.0000	5 .0000	-1.7510	1.01799	1.0387	-.3716	
ELPHEXA	163			1.	1 .9482 .9988	4 .0005	-.9979	-3.0466	-.4903	-.2307	-1.5777
ELPHEXA	164			4. ***	1 .6806 .6593	2 .2079	-.3566	1.2049	.0454	1.9739	.6241
ELPHEXA	165			6.	6 .9788 1.0000	10 .0000	1.3803	.4337	-1.8191	-.4471	-1.0229
ELPHEXA	166			1.	1 .9249 .9840	4 .0156	-1.6410	-.1164	-.12629	-.8336	
ELPHEXA	167			7.	7 .8903 1.0000	5 .0000	3.5976	-1.2279	.9531	-2.9438	-.6229
ELPHEXA	168			1.	1 .3506 .9727	8 .0266	.3282	-.1355	.7008	-.5134	-1.045
ELPHEXA	169			4. ***	1 .3032 .7023	4 .2477	3.4451	-2.1029	-.5232	-2.6616	
ELPHEXA	170			3.	3 .1153 .9928	1 .0796	.3102	.2286	.7941	2.7224	-1.7886
ELPHEXA	171			1.	1 .1289 .9744	2 .0224	3.5082	2.1000	-1.3688	-3.6081	
ELPHEXA	172			1.	1 .9752 .9988	4 .0010	-1.8284	1.2451	-3.5762	-.1152	-2.1511
ELPHEXA	173			5.	5 .0142 1.0000	8 .0000	-.6881	-1.7642	-.6652	-.0504	-2.2536
ELPHEXA	174			4. ***	1 .4082 .9551	4 .0426	.5225	-.0641	.4008	.6029	
							-2.3558	-.2014	.2349	-1.1824	
							-.0721	.9222	-.1887	-.4235	
							6.2607	-.2514	-.6452	1.4399	-.4533
							.4797	-.7638	-.7795	.7441	
							-1.0446	-2.5840	-.9599	-.4327	-1.5761
							-.9004	-.0778	.1308	.8286	
							3.4952	-.9891	-1.0141	-2.9425	.7382
							.7475	-.8713	-2.5073	.6830	
							-1.2856	-2.6124	-1.8346	-.7064	.2301
							-.5025	-.4672	.4278	-2.1852	
							-1.8624	-.7518	1.12557	-2.0103	-.9887
							-1.6689	-.0713	.1307	-1.0720	
							-.8614	.7044	-.9979	-.4148	-2.8234
							-1.9745	.9203	.5216	-.5093	
							.1064	-1.1973	-.8463	.7207	-2.3455
							.6477	-.6528	-2.4847	.2351	
							-.8385	-1.9652	.0569	-.7702	-2.2328
							.5003	.3796	.0379	-.6735	
							3.6128	-.6921	-.41215	-1.3046	1.7048
							.7816	.8261	1.19026	-2.1153	
							-2.1718	-.7675	.2925	-2.6266	-.7474
							.0329	-.8280	.1672	-1.1194	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GRJUP	HIGHEST GROUP	PROBABILITY P(X/G) P(G/X)	2ND HIGHEST GRJUP	P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	175			1.	1	.9936 .9956	4	.0043	-1.1585	-2.2251	.2762	-1.1496	-1.4530
ELPHEXA	176			1.	1	.9844 .9987	4	.0010	.7434	-.3283	.4788	-.4098	
ELPHEXA	177		***	1.	8	.2598 .9971	1	.0027	-.8943	-2.1532	.0503	-.3394	-2.1345
ELPHEXA	178			4.	4	.5119 .6964	2	.2958	.2250	.7623	.2362	.3258	
ELPHEXA	179			1.	1	.3306 .7502	8	.2461	-1.3046	-1.0344	-3.2563	-.1872	1.0368
ELPHEXA	180			1.	1	.9235 .9974	4	.0016	-.3272	-.0348	-2.7133	-.1780	
ELPHEXA	181			1.	1	.9665 .9986	4	.0010	-1.9804	.3773	1.1717	-.8545	1.2003
ELPHEXA	182			1.	1	.9998 .9914	4	.0071	.2758	-1.5437	-.0347	1.5250	
ELPHEXA	183			1.	1	.9585 .9975	4	.0004	-1.54305	-2.5816	-1.6285	2.0666	.7782
ELPHEXA	184			5.	5	.7154 1.0000	8	.0000	-1.5513	.0785	.8569	-.8310	
ELPHEXA	185			1.	1	.7451 .9987	2	.0005	-.9484	-1.5478	-.8409	.0024	-1.7225
ELPHEXA	186			1.	1	.9255 .9992	2	.0005	.6770	-.4908	.9994	.3141	
ELPHEXA	187			3.	3	.6874 .6473	2	.1927	-.9124	-2.1202	-.0274	-.2227	-2.0740
ELPHEXA	188		***	3.	4	.0412 .6111	9	.1500	.2981	.8129	.2994	.8192	
ELPHEXA	189			6.	6	.9839 1.0000	10	.0000	-1.3218	-1.8886	-.2832	-.1887	-1.0774
ELPHEXA	190			7.	7	.9781 1.0000	5	.0000	-.0666	-.3497	.1771	.3766	
ELPHEXA	191			6.	6	.7477 1.0000	10	.0000	-1.0005	-2.0082	-.4135	-.9850	-2.0092
ELPHEXA	192			1.	1	.9387 .9982	4	.0017	1.2323	-.1026	-.0530	-.2668	
ELPHEXA	193			8.	8	.4103 .5787	1	.8211	3.2843	-2.4424	-3.2821	-2.7889	1.9552
ELPHEXA	194		***	4.	2	.2604 .6412	4	.2816	.0053	.3454	.1296	.0081	
ELPHEXA	195			3.	3	.0045 .8351	10	.2016	-.6208	-1.3330	-.9194	.2295	-2.6168
ELPHEXA	196			5.	5	.6878 .9999	7	.0001	.3447	.1295	.6024	-.5143	
ELPHEXA	197			5.	5	.2537 .9999	10	.0001	-.9397	-1.5909	-.8317	-.3476	-2.3632
ELPHEXA	198			1.	1	.2608 .9999	4	.0001	.9186	.0370	-.2517	.0730	
ELPHEXA	199			1.	1	.9740 .9951	4	.0047	-1.1181	1.7455	.9652	.1873	-.5931
ELPHEXA	200			1.	1	.3973 .9382	2	.0322	-.9417	-1.0161	.6434	-.5244	
ELPHEXA	201			1.	1	.6664 .9911	2	.0055	-1.5026	1.8916	1.6859	-1.2583	.0344
ELPHEXA	202			1.	1	.5371 .9214	3	.0369	1.230	-2.4334	.1775	-1.9472	
ELPHEXA	203			1.	1	.0764 .9358	3	.0505	-.1583	-.3057	.2098	-.2609	-1.0136
									-.1583	-1.8802	.34870	-2.9415	.8133
									-.2436	-.8184	-2.0238	.0306	
									6.3791	.6066	-.8010	1.6725	-.9981
									.7531	-1.1710	-.2973	-1.5221	
									-.9817	-2.3827	.4896	-1.1777	-2.0650
									.9009	.1308	.7530	-.2627	
									-1.5266	-2.8591	-2.5215	-.13250	.8127
									-.2358	-.2999	-.4361	.1270	
									-1.5341	1.2754	.7978	-1.3985	.2599
									1.0257	-1.7920	-.8432	-.6214	
									2.8030	3.0014	2.0495	-3.4618	1.0879
									-2.5149	.8880	.6641	.5250	
									4.0429	-1.1281	-.5128	-2.8724	-1.1020
									.3986	.5521	.1095	.6219	
									1.5393	-1.12149	-1.1203	-3.3398	-.1777
									2.1067	1.2996	.2137	-1.1593	
									-2.6030	-1.5644	-.7541	-2.2543	-1.9357
									-.7610	1.8961	-.2247	-.8415	
									-1.2559	-2.4127	.3125	-.6349	-1.4487
									.4988	-.0792	.9859	.9205	
									-1.3844	-.2679	-1.0146	-.2620	-1.8710
									-.9141	.5113	-.9020	1.4873	
									-.9386	-1.1739	-.8802	.4470	-1.8907
									-.8106	.1311	-1.3758	1.4257	
									-1.7445	-.8836	.4149	-1.0788	-1.2284
									-1.7216	1.1142	-.5459	.9278	
									-1.7359	-1.1634	-.1179	-1.3149	-1.2645
									-2.6316	-.8446	-.7845	2.0722	

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES									
SUBFILE	SEQNUM	VAL	SEL	GROUP	GROUP	P(17/G)	P(17/X)	GROUP	P(17/X)					
ELPHEXA	204			3. ***	5	.0097	.9999	3	.0001	2.3900	-1.2169	-1.8557	-2.6226	2.8269
ELPHEXA	205			4.	4	.0001	.4792	1	.4568	-3.9082	1.0836	.4679	.5935	1.8403
ELPHEXA	206			4. ***	2	.0154	.6804	4	.3177	-2.2227	-1.0446	.2797	-1.4896	1.2872
ELPHEXA	207			4. ***	2	.8857	.9121	4	.0833	-3.1925	3.0236	.7580	-1.8526	1.5581
ELPHEXA	208			2.	2	.8913	.7563	4	.2118	-2.0018	.6853	1.6414	-.5131	.6090
ELPHEXA	209			2.	2	.8714	.9853	9	.0371	1.2429	.4638	.2540	.7625	.4856
ELPHEXA	210			2.	2	.7717	.9892	4	.0105	-2.1366	1.1365	1.1303	2.503	1.2902
ELPHEXA	211			4. ***	2	.7580	.6333	4	.3609	.4728	-.2049	-.3371	1.4577	.6729
ELPHEXA	212			4. ***	3	.5893	.5765	4	.3259	-1.6292	1.3150	2.2144	.4575	-.1856
ELPHEXA	213			4.	4	.8384	.9062	2	.0559	-.4481	.9017	.1691	.5877	.7901
ELPHEXA	214			4.	4	.6242	.5061	2	.3065	-.8104	2.1437	-.9520	1.2196	.5456
ELPHEXA	215			1. ***	8	.2470	.6302	1	.3697	.8326	-.2440	-1.9520	-.2836	.9700
ELPHEXA	216			1.	1	.9685	.9486	2	.0337	-1.1750	1.1307	1.8396	.3136	-.9013
ELPHEXA	217			4.	4	.9254	.8830	2	.0657	1.2793	.6587	-2.6596	.0220	.1679
ELPHEXA	218			1.	1	.9996	.9766	4	.0025	-1.6259	.6127	2.2313	-.3382	.1679
ELPHEXA	219			5.	5	.0840	1.0000	8	.0000	.6887	1.5683	.3665	.5204	1.4256
ELPHEXA	220			5.	5	.0691	1.0000	8	.0000	-1.2838	.8543	2.3972	.2593	2.3193
ELPHEXA	221			1.	1	.6154	.8851	8	.0132	-1.8162	.7105	.9017	.1716	.4411
ELPHEXA	222			1. ***	8	.1394	.9999	1	.0001	-1.7563	.3081	2.2397	-.2181	1.0164
ELPHEXA	223			1.	1	.5455	.7286	8	.2708	-1.2613	-.1632	1.0591	.7918	.6129
ELPHEXA	224			1.	1	.8581	.9776	2	.0124	-1.6461	.8702	1.7672	.2661	-1.2097
ELPHEXA	225			1.	1	.8215	.9672	2	.0229	-1.4910	-.3414	.3285	1.2509	.0028
ELPHEXA	226			3. ***	7	.1459	.9061	10	.0652	-1.4226	-3.3242	-2.3820	-.0056	1.0913
ELPHEXA	227			3. ***	7	.0578	.8025	3	.1061	-1.0245	.4381	-.6784	.3693	1.0913
ELPHEXA	228			3.	3	.6779	.9873	4	.0113	-1.3307	-1.4070	.0564	.1066	1.0913
ELPHEXA	229			3.	3	.0641	.7039	2	.2464	1.0432	1.2044	-.0680	.1096	1.0913
ELPHEXA	230			1.	1	.6338	.9457	4	.0492	-1.4378	.5037	2.3728	-.3760	1.0913
ELPHEXA	231			2. ***	1	.3135	.7435	4	.0888	-.7644	-.5023	.3898	-.1655	1.0913
ELPHEXA	232			2.	2	.8499	.9272	4	.0449	-1.0898	-1.9916	-.2147	-.2212	1.0913

CASE SUBFILE	SEJNUM	MIS VAL	SEL	ACTUAL GRJUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHXA	233			1.	1 .9729 .9956	4 .0019	-1.5143	-1.5123	-1.4924	-1.0792	-1.8849
ELPHXA	234			4.	4 .4825 .8483	3 .0976	-1.6278	-.1774	-1.8454	-.2052	.6002
ELPHXA	235			3.	3 .5873 .9128	2 .0436	-1.8079	-.2886	1.1257	1.3438	-.8142
ELPHXA	236			5.	5 .8226 1.0000	1 .0000	-1.5189	-1.1797	1.14918	-.4320	1.9683
ELPHXA	237			5.	5 .9104 1.0000	7 .0000	-1.3429	-.5589	-.9392	-.0942	-.1629
ELPHXA	238			4.	4 .7809 .6220	2 .3704	-.4461	-1.16811	-.19731	1.2226	.8848
ELPHXA	239			8. ***	1 .0963 .8879	8 .1102	-1.1835	-1.1726	1.1546	-.5699	.9154
ELPHXA	240			4. ***	1 .2248 .6331	2 .2188	2.0665	-.0945	-.6280	-.4979	-.4258
ELPHXA	241			4. ***	1 .4426 .9282	4 .0434	-1.5287	-1.7247	1.1846	-1.0594	-1.3057
ELPHXA	242			4. ***	1 .2161 .6128	2 .3343	-.7883	1.1846	-2.0848	-1.3057	-.4585
ELPHXA	243			9. ***	8 .0182 .9708	9 .0272	1.5415	-.4659	-.1209	-2.2410	-.1171
ELPHXA	244			2.	2 .8631 .9446	3 .0298	-1.6873	-1.7084	-.4187	-1.5638	1.3498
ELPHXA	245			5.	5 .5063 .9397	7 .0003	-1.3243	-.8129	-2.0051	-1.5361	.2714
ELPHXA	246			5.	5 .1310 .9887	6 .0093	-1.5260	1.12037	-2.0291	1.1966	-.4123
ELPHXA	247			1.	1 .3623 .8411	4 .0817	-1.476	1.4232	-1.6851	-2.299	-1.7666
ELPHXA	248			9.	9 .1449 .8929	3 .0739	-1.1198	1.5889	-.0489	1.5825	-.4302
ELPHXA	249			5. ***	7 .2368 .5532	5 .4348	4.4175	-1.4124	-1.9601	-1.9114	-1.8726
ELPHXA	250			4. ***	1 .6591 .7773	2 .1561	-2.958	-.9060	1.1961	-1.7039	-.8552
ELPHXA	251			5.	5 .0031 .9989	6 .0007	1.2170	-1.3358	-.4996	-1.8995	-.8527
ELPHXA	252			9.	9 .5376 1.0000	2 .0000	-1.2282	1.3870	-.1548	-1.4705	3.1771
ELPHXA	253			4. ***	2 .3701 .7592	4 .2302	-1.2771	2.0144	-1.5378	-1.1414	.1228
ELPHXA	254			2. ***	1 .5932 .8765	4 .0657	-.7289	2.0216	-1.2994	-1.3007	2.0377
ELPHXA	255			2. ***	3 .0768 .6529	4 .2754	1.38531	-.8118	-2.1798	-1.1084	-.1539
ELPHXA	256			2.	2 .6500 .9365	4 .0632	-2.1699	-.0815	-.0945	-1.1323	-.8527
ELPHXA	257			2.	2 .5924 .9356	4 .0348	.5206	1.1056	.0962	-1.808	-.1539
ELPHXA	258			3. ***	7 .1792 .9196	6 .0708	4.0865	-.0310	-3.16382	-.0035	3.1771
ELPHXA	259			3.	3 .2675 .6814	2 .3131	2.103	-1.1848	1.1102	1.1808	.1228
ELPHXA	260			3. ***	7 .1792 .9196	6 .0708	-1.17050	4.3926	-1.5966	-.9163	2.0377
ELPHXA	261			3. ***	10 .0237 .6154	3 .2490	1.4132	.0230	1.4184	1.7735	2.0377
							-2.2284	-.4253	1.7747	1.7655	2.0377
							-.4914	.6746	.3586	2.3519	
							-1.5426	-1.3124	-.4282	-1.2886	-.1539
							-.4797	.3595	-2.1957	.5030	
							-2.1852	1.7493	2.0817	-1.5112	.3953
							-2.1809	.4781	.6389	.7835	
							-.6253	-.0879	1.2215	1.6188	1.2658
							1.9370	-.6292	-1.9364	-.0657	
							-.3784	-.4936	-.2763	-1.7743	-.2300
							1.8923	.5988	.7813	-.6448	
							-1.5334	1.1733	.7574	-.3520	.4663
							1.2260	.8545	.5779	.7540	
							-.1066	2.0353	1.1668	1.9703	-.8756
							-1.1351	-.2857	-2.2359	.0021	
							4.7377	-2.7217	-1.7010	-.3861	.1492
							-.3838	-.3814	-1.1099	.6778	
							4.8282	2.4257	1.1666	-1.3882	-.3624
							-2.3888	.6779	-1.3378	.1888	

CASE SUBFILE	SEJNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST GROUP	PROBABILITY P(X/G)	HIGHEST PROBABILITY P(G/X)	2ND HIGHEST GROUP	2ND HIGHEST PROBABILITY P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	262			3.	3	.0138	.9652	7	.0238	2.8123	3.5139	1.0083	-2.2832	1.2620
ELPHEXA	263			2.	2	.0758	.4986	9	.2535	-3.2501	.3849	1.0671	.5859	
ELPHEXA	264		***	4.	2	.7629	.9911	4	.0074	-1.6856	1.3065	-2.0364	-1.1733	.5435
ELPHEXA	265			3.	3	.2717	.9739	2	.0185	-1.1146	1.4510	1.2357	.5511	.7492
ELPHEXA	266		***	4.	2	.9319	.9502	4	.0492	.7822	.5300	-2.7009	-0.7490	
ELPHEXA	267			2.	2	.0295	.9581	8	.0117	-1.3828	1.2021	2.8891	1.4242	-.6258
ELPHEXA	268			1.	1	.9978	.9803	4	.0113	-2.8390	.8031	-1.5786	-0.4573	
ELPHEXA	269		***	4.	2	.1894	.6956	1	.2566	-1.5127	1.1948	-1.1512	.4385	1.3953
ELPHEXA	270			3.	3	.0661	.9435	2	.0303	1.0779	-.3615	-1.7867	.0518	
ELPHEXA	271			3.	3	.1537	.9919	2	.0379	-1.1137	1.1009	-1.2313	.21828	3.1500
ELPHEXA	272		***	4.	2	.4404	.8750	4	.1086	-1.6391	1.0371	-1.2777	-1.2460	
ELPHEXA	273			2.	2	.8591	.9377	3	.0511	-1.4255	-2.2817	.4032	-.6613	-.8238
ELPHEXA	274			1.	1	.0140	.9722	4	.0122	.3189	-.5182	.6124	.5883	
ELPHEXA	275		***	4.	2	.6622	.5127	2	.0387	-1.4895	-.7940	.6750	-.2094	-.3883
ELPHEXA	276			2.	2	.8509	.9629	4	.0345	-0.4532	1.9243	-2.7918	.7644	
ELPHEXA	277		***	4.	1	.8357	.9881	4	.0097	-1.1321	.6463	.7340	-1.06485	-1.5614
ELPHEXA	278			4.	4	.0016	.9982	4	.0016	-2.2227	1.8461	-2.2536	-.4339	
ELPHEXA	279			2.	2	.7138	.9414	4	.0579	-2.099	1.19361	1.18459	1.18578	-1.3104
ELPHEXA	280		***	2.	2	.8591	.9377	3	.0511	-2.5407	1.0883	-1.9315	-1.4629	
ELPHEXA	281			1.	1	.0140	.9722	4	.0122	-1.6265	1.4002	.8192	-.0109	.7980
ELPHEXA	282		***	4.	2	.6622	.5127	2	.0387	-.3022	-.4100	-1.2369	2.7032	
ELPHEXA	283			2.	2	.8591	.9377	3	.0511	-1.7628	1.7184	1.16145	1.1753	.2812
ELPHEXA	284			1.	1	.0140	.9722	4	.0122	-.6236	.7235	-1.3203	.2867	
ELPHEXA	285		***	4.	2	.6622	.5127	2	.0387	-.9147	-2.9202	-.4504	.9479	.0996
ELPHEXA	286			2.	2	.8509	.9629	4	.0345	-3.4754	1.7682	.7720	-.5083	
ELPHEXA	287		***	4.	1	.8357	.9881	4	.0097	-1.6340	-.4338	1.10035	.1096	-.6343
ELPHEXA	288			2.	2	.8509	.9629	4	.0345	1.3739	-1.2324	.1145	.7205	
ELPHEXA	289		***	4.	1	.8357	.9881	4	.0097	-1.0799	.9221	2.0323	.17625	1.0536
ELPHEXA	290			3.	3	.1024	.9746	2	.0218	.1582	1.3238	-2.1784	-.0448	
ELPHEXA	291		***	2.	2	.8591	.9377	3	.0511	-1.6391	-2.2578	.2241	-.3322	-.6084
ELPHEXA	292			4.	4	.0016	.9982	4	.0016	1.6255	1.2609	.8882	.4584	
ELPHEXA	293		***	4.	1	.8357	.9881	4	.0097	-1.4154	1.5805	-.1719	-.1641	-.0175
ELPHEXA	294			2.	2	.7138	.9414	4	.0579	.5164	-1.1518	.3612	.4404	
ELPHEXA	295			1.	1	.0140	.9722	4	.0122	-.4029	-.2713	1.4364	1.5970	.8442
ELPHEXA	296		***	2.	2	.8591	.9377	3	.0511	1.7151	.3301	-2.0798	.0511	
ELPHEXA	297			1.	1	.0140	.9722	4	.0122	-.9751	-2.8050	.3738	-.1123	-1.5848
ELPHEXA	298		***	2.	2	.8591	.9377	3	.0511	.0402	.7161	-.0064	1.7994	
ELPHEXA	299			4.	4	.0016	.9982	4	.0016	-1.7908	-2.3307	.4063	-.2276	-.2233
ELPHEXA	300		***	4.	1	.8357	.9881	4	.0097	1.6224	1.2003	-2.6564	-.7254	
ELPHEXA	301			2.	2	.8591	.9377	3	.0511	-1.52839	-2.2288	.0948	-.3336	-.4236
ELPHEXA	302		***	4.	2	.6622	.5127	2	.0387	1.5477	1.4639	.4888	.4422	
ELPHEXA	303			1.	1	.0140	.9722	4	.0122	-1.5374	-2.3100	.0958	-.3123	-.3157
ELPHEXA	304			2.	2	.8591	.9377	3	.0511	1.3530	1.3225	-.1437	.7892	
ELPHEXA	305		***	4.	1	.8357	.9881	4	.0097	-1.1178	-2.3190	-.0513	1.1031	-1.4712
ELPHEXA	306			3.	3	.1024	.9746	2	.0218	-.5384	.3581	-.2053	1.4144	
ELPHEXA	307		***	4.	2	.6622	.5127	2	.0387	-2.09412	1.0923	-2.1604	1.0821	-.5430
ELPHEXA	308			2.	2	.8591	.9377	3	.0511	-.9412	1.0923	-2.1604	1.0821	
ELPHEXA	309		***	4.	2	.6622	.5127	2	.0387	-1.5214	.3945	1.0984	-.2573	-.0773
ELPHEXA	310			2.	2	.8591	.9377	3	.0511	-.1747	.2956	.2138	1.6702	
ELPHEXA	311		***	4.	2	.6622	.5127	2	.0387	-1.3746	1.0260	-.4168	1.7972	1.2748
ELPHEXA	312			2.	2	.8591	.9377	3	.0511	1.2011	-1.1793	-1.5716	.7500	
ELPHEXA	313		***	4.	2	.6622	.5127	2	.0387	-1.4228	1.5633	.4303	.7407	.1596
ELPHEXA	314			2.	2	.8591	.9377	3	.0511	.7932	.9746	.1359	.2345	
ELPHEXA	315		***	4.	2	.6622	.5127	2	.0387	-1.5224	.5078	1.3453	1.1294	.4880
ELPHEXA	316			2.	2	.8591	.9377	3	.0511	-.9036	.1625	-.3945	1.8303	
ELPHEXA	317		***	4.	2	.6622	.5127	2	.0387	-1.3952	1.2166	-.1240	.5508	.2270
ELPHEXA	318			2.	2	.8591	.9377	3	.0511	-.3645	-1.1471	-.5945	2.0883	

CASE SUBFILF	SEQNUM	MIS VAL	SEL	ACTUAL GRJUP	HIGHEST GROUP	PROBABILITY P(X/G) P(G/X)	2ND HIGHEST GRJUP	P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	291			2.	2	.8555 .9788	9	.0194	-.8159	1.9705	-.1611	1.5890	.7371
ELPHEXA	292			1.	1	.8920 .9764	4	.0171	1.2762	-.0754	-1.2618	.0145	
ELPHEXA	293			2.	2	.6129 .9259	4	.0014	-.813742	-1.6343	-.3647	.0511	-1.2006
ELPHEXA	294			1.	1	.2250 .9737	2	.0214	-.1173	1.4407	.7501	-.8049	
ELPHEXA	295			4. ***	2	.8571 .7778	4	.2165	-1.1403	-1.7987	-.2740	1.4249	1.4702
ELPHEXA	296			2.	2	.9691 .8970	4	.1344	.6629	-.3860	-2.6517	.4787	
ELPHEXA	297			3.	3	.3301 .9358	2	.0135	-2.0118	-1.0088	-1.5322	-1.3492	-.7147
ELPHEXA	298			2. ***	1	.3626 .8249	2	.1554	-.4860	.1070	-1.5322	2.5065	
ELPHEXA	299			3.	3	.0067 .7555	1	.1103	-2.0488	1.0260	1.7011	.6308	1.3341
ELPHEXA	300			3.	3	.6413 .8880	2	.1085	-.3448	.7356	.4182	.9877	
ELPHEXA	301			1.	1	.5357 .9446	8	.5547	-1.7749	1.3576	1.6767	.0942	.7638
ELPHEXA	302			3.	3	.3345 .9164	2	.0823	.5494	.6936	.2804	.2985	
ELPHEXA	303			3.	3	.7690 .9788	2	.0198	-2.4147	2.3191	1.0783	2.6026	-.9696
ELPHEXA	304			4. ***	2	.9839 .9374	3	.0251	-.6613	-.0168	-1.5874	-.6250	-.7519
ELPHEXA	305			3.	3	.0704 .9740	1	.0170	.6601	1.6744	-2.1704	-.0339	
ELPHEXA	306			3.	3	.4612 .5146	4	.4445	-1.2689	.9420	-.0509	1.1604	-2.3352
ELPHEXA	307			3.	3	.5378 .9897	2	.0367	-1.1651	2.8016	.5814	-2.4032	
ELPHEXA	308			3.	3	.8399 .6220	2	.3324	-.7925	1.4183	1.6438	1.5830	-.8237
ELPHEXA	309			3.	3	.5436 .9718	2	.0047	-2.1583	1.1219	-.3141	.8447	
ELPHEXA	310			3. ***	2	.8194 .6311	3	.3550	-2.1583	1.1219	-.3141	.8447	.3229
ELPHEXA	311			2.	2	.3051 .9431	4	.0520	-.8007	.0946	-2.8090	-1.8256	-1.3495
ELPHEXA	312			2. ***	3	.2206 .4767	2	.4729	-.1739	2.1031	1.6487	2.4438	-1.0143
ELPHEXA	313			9. ***	8	.6842 1.0000	1	.0000	-1.7640	.0732	-1.5582	.4781	
ELPHEXA	314			2. ***	3	.0873 .5632	9	.2186	-.6196	1.8151	1.0873	1.4867	-1.1505
ELPHEXA	315			2.	2	.3010 .5386	3	.4138	-2.4338	.5915	-.3427	4.005	
ELPHEXA	316			5.	5	.0608 .3566	6	.2559	-1.4426	1.6474	1.8183	1.1756	.5147
ELPHEXA	317			9.	9	.6867 1.0000	2	.0000	-.3597	.6080	.0141	.4954	
ELPHEXA	318			2. ***	8	.2281 .9494	2	.0471	-.2511	-.0699	1.7360	-.2615	-2.3797
ELPHEXA	319			2.	2	.7508 .9455	1	.0336	-2.4911	1.9204	-2.0614	-1.1010	
									-1.4362	1.0350	2.4378	-.1207	.1105
									-2.0671	-.6020	.4175	-.8640	
									-.6507	1.5356	2.0100	1.2352	-1.2483
									-2.4802	.7416	1.1271	.0066	
									-.9142	1.4844	.9190	.8011	-.7697
									-.7871	.1683	.4020	.8360	
									-.6323	1.5090	2.0694	1.1173	-1.2899
									-2.5414	.9093	1.0515	-.4157	
									-.2863	1.5095	1.2018	1.2619	-.7669
									-.5603	-.0927	-1.3634	.5981	
									-.9623	.4675	.2712	1.7082	.6467
									2.4440	-.5138	1.1382	1.3648	
									-1.2447	1.8211	1.0765	1.6323	.1028
									-1.4982	1.7023	1.0476	-.4043	
									-2.0471	1.2487	-3.6638	.8575	1.2643
									-1.4648	.3920	.2070	-1.1464	
									-2.0229	3.1466	1.7410	.4099	.5730
									-2.1886	1.7883	.1649	.3030	
									-1.2256	1.7773	1.6841	1.8427	.2666
									-1.7059	1.6058	.5315	.5474	
									4.2647	1.0224	.6147	-1.4646	.2120
									.3330	-.8600	.8813	-.9238	
									-1.7565	4.6455	1.2104	.0428	-.6992
									.5459	.4580	1.8302	-1.5395	
									-1.1368	-.1816	-1.6451	2.3205	2.2338
									.7776	-.2744	1.3288	-.3116	
									-1.4764	.6449	-.6448	.2957	-.5176
									.3753	-.0389	-1.9734	.2236	

CASE SUBFILE	SEJNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST GROUP	PROBABILITY P(X%G) P(Y%G)	2ND HIGHEST GROUP	PROBABILITY P(X%G) P(Y%G)	DISCRIMINANT SCORES				
ELPHEXA	320			2. ***	3	.0224 .4965	9	.4925	-1.5042	4.0893	1.8997	.2589	-.6623
ELPHEXA	321			9. ***	8	.2035 .9369	1	.0106	-2.1142	1.8469	.0379	-1.6645	1.0922
ELPHEXA	322			1.	1	.1199 .5790	8	.4119	-.2540	1.2302	-.3192	-1.4138	
ELPHEXA	323			2. ***	9	.3711 .8156	2	.1325	-1.6796	-2.1102	-1.7390	-.0739	1.5125
ELPHEXA	324			3.	3	.1805 .9175	9	.0531	-.0804	.5111	.0707	-2.2657	
ELPHEXA	325			9.	9	.3075 .8625	2	.1299	-2.2378	1.3715	.7514	-.2281	.5435
ELPHEXA	326			2. ***	9	.3774 .5581	2	.4301	-1.6946	-1.2473	-.3039	1.7459	-.4272
ELPHEXA	327			1.	1	.4577 .9646	8	.0335	-1.3926	1.3298	-1.5348	-1.5818	-.1963
ELPHEXA	328			1.	1	.8540 .9790	2	.0142	-.2625	1.2476	-.5330	-.0311	1.3835
ELPHEXA	329			1.	1	.8445 .9760	4	.0167	-.4338	-.3737	-1.7898	-.1395	
ELPHEXA	330			4. ***	1	.6121 .8518	4	.0914	-1.4274	-2.2205	-1.5029	-1.6104	.3386
ELPHEXA	331			8.	8	.1068 .7001	1	.2864	-1.0832	-1.5408	.2244	.0089	-1.4273
ELPHEXA	332			2. ***	1	.9780 .9623	4	.0209	1.1841	1.7645	-.0601	.0422	
ELPHEXA	333			5. ***	7	.0078 .6972	5	.2922	-1.5106	-1.8391	-.9888	-.3371	-1.1139
ELPHEXA	334			1.	1	.2173 .9732	8	.0184	-.4982	1.8007	.8302	-.1219	
ELPHEXA	335			1.	1	.3367 .9679	4	.0235	-2.2960	-.9102	.7384	-1.3873	-.3514
ELPHEXA	336			1.	1	.9948 .9639	4	.0295	1.703	1.7643	.02185	.5862	
ELPHEXA	337			4. ***	1	.4344 .9218	4	.0409	-2.1667	-.8070	-2.2032	-1.7476	1.3881
ELPHEXA	338			1.	1	.9835 .9858	4	.0124	-.4843	1.3250	-.5053	-1.4491	
ELPHEXA	339			1.	1	.9982 .9959	4	.0039	-1.3606	-1.4480	.2235	.1012	-1.1289
ELPHEXA	340			10.	10	.3636 1.0000	3	.0000	1.0014	.3577	.3813	.0193	
ELPHEXA	341			4.	4	.5351 .9547	2	.0352	-2.1674	1.4796	-.1970	-4.8341	.1426
ELPHEXA	342			4.	4	.4920 .9333	2	.0643	-.3191	2.6817	-.5828	.3112	
ELPHEXA	343			2. ***	1	.2211 .9645	2	.0187	-1.6124	-3.0539	-1.1344	-1.1394	1.4572
ELPHEXA	344			1.	1	.3834 .9124	2	.0799	-2.2480	1.6852	.4018	-1.2167	
ELPHEXA	345			1.	1	.1705 .9223	2	.0551	-1.2675	-.8558	.2269	-.8964	-1.4813
ELPHEXA	346			1.	1	.9992 .9952	4	.0046	1.9266	-.2186	.2784	-2.1809	
ELPHEXA	347			3. ***	1	.8849 .9508	4	.0366	-1.1281	-2.2438	.2784	-.4311	-1.0052
ELPHEXA	348			10.	10	.7428 .9996	6	.0004	-.4120	-.0702	.8466	.2372	
									-2.3586	-.5339	.1947	-2.2020	-.3693
									1.5607	.5618	-.0470	-.2389	
									-1.3904	-2.3412	.0527	-.5311	-.6751
									.0715	-.6252	.0183	1.2568	
									-1.1001	-2.7563	.9672	.7586	-1.1298
									-.3163	.3278	-.4711	.3028	
									3.7700	1.8427	3.4361	-1.4663	2.3610
									-1.1712	3.6111	-.5814	-2.1286	
									-1.0717	-.9865	3.2076	-.3945	2.0077
									1.4001	-.3325	-1.1525	-1.2778	
									-1.0466	1.1738	3.1299	-.6459	2.2578
									1.0776	.2126	-1.4339	-.7848	
									-.5124	-2.3097	1.0038	.0452	-1.0047
									1.4165	2.3668	-1.6050	-.8506	
									-.2496	-1.3237	-.0484	-.6703	-1.5187
									1.6461	1.1402	-1.4190	-.9470	
									-.3849	-1.7917	.6410	-.3608	-1.3426
									1.0935	1.9056	-2.3756	-1.0061	
									-1.1082	-2.2918	.2329	-1.0717	-1.2965
									.4737	-.4211	-.2014	.0796	
									-.8320	-2.2523	1.4573	-.3164	-1.5316
									-1.1625	.3270	-.0628	.6112	
									4.2050	.3346	1.9032	-.1631	.8658
									-.1643	2.4811	1.4619	1.3006	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	349			3. ***	10 .9823 1.0000	6 .0000	3.8303	1.0467	2.1926	-.1857	1.7521
ELPHEXA	350			10.	10 .3204 .9975	6 .0025	.7025	2.8544	1.6658	-.8905	
ELPHEXA	351			10.	10 .8546 1.0000	6 .0000	4.6355	1.4760	3.356	.4211	1.3784
ELPHEXA	352			10.	10 .7288 1.0000	6 .0000	2.4682	2.2053	-.8169	-1.3075	
ELPHEXA	353			10.	10 .9693 1.0000	6 .0000	4.0444	1.3931	1.9829	.2268	.8245
ELPHEXA	354			10.	10 .6519 1.0000	6 .0000	.7821	3.6723	2.0577	-1.2551	
ELPHEXA	355			10.	10 .8407 1.0000	3 .0000	3.9137	1.2858	1.5834	.3418	1.6207
ELPHEXA	356			4.	4 .9588 .7310	2 .2647	.9317	2.4146	2.6482	-.5184	
ELPHEXA	357			1.	1 .5849 .9695	2 .0270	3.7592	.8110	2.2998	.0299	1.7604
ELPHEXA	358			4.	4 .9632 .7313	2 .2453	.3216	3.1409	1.7512	-.5971	
ELPHEXA	359			4. ***	2 .8612 .5106	4 .4834	4.7049	.7765	2.6520	.0578	1.3630
ELPHEXA	360			10.	10 .1272 .7846	3 .2120	1.0893	3.8678	-.2895	-2.1966	
ELPHEXA	361			10.	10 .2952 .9958	3 .0035	3.9154	2.7993	1.4788	.1276	2.0112
ELPHEXA	362			3.	3 .1388 .6052	10 .9881	-1.1348	2.8840	-.9165	-.4055	
ELPHEXA	363			3.	3 .7607 .8808	2 .0602	-.9848	-.6588	2.0557	.9602	.9035
ELPHEXA	364			3.	3 .7429 .9351	2 .0424	.9263	.8633	.3701	.0301	
ELPHEXA	365			1.	1 .8922 .9985	4 .0010	-.8518	-.9451	-.6721	.8897	-1.9081
ELPHEXA	366			4.	4 .7597 .9311	2 .0680	.9567	1.4023	.5182	-.2416	
ELPHEXA	367			4.	4 .5286 .9142	2 .0855	-1.3465	-.6978	1.5801	.7938	1.4797
ELPHEXA	368			3.	3 .0192 .7228	10 .2661	1.1475	-.5384	-.0994	.4765	
ELPHEXA	369			4.	4 .6572 .9823	2 .0172	-1.0991	-.5431	1.4929	.6483	1.0935
ELPHEXA	370			6.	6 .9395 1.0000	10 .0000	1.7695	-.1218	-.2643	.7278	
ELPHEXA	371			4.	4 .4428 .9913	2 .0081	3.9143	1.6259	-.7319	-1.0806	.2409
ELPHEXA	372			6. ***	10 .1133 .8713	6 .1287	-2.1203	1.9235	.9972	1.3157	
ELPHEXA	373			4.	4 .7158 .9919	2 .0078	4.4458	1.4966	1.6187	-1.3048	.3949
ELPHEXA	374			4.	4 .7071 .9737	2 .0257	-1.6592	1.7954	-1.6012	-.0047	
ELPHEXA	375			1.	1 .0465 .6517	8 .3339	3.2440	1.7317	1.2562	-1.6612	.6892
ELPHEXA	376			4.	4 .7033 .9694	2 .0300	-2.1142	.0916	.3918	.8797	
ELPHEXA	377			4.	4 .6693 .9836	2 .0158	-1.1708	1.4457	2.0282	.4453	-.5946
							-1.8785	.4406	.5705	-.6252	
							-.9827	1.4511	1.4296	.9847	-.5160
							-2.0887	.5288	.9687	-.6046	
							-.8861	2.4373	-.9375	-1.0673	-1.8639
							-1.4080	1.0709	-.5527	1.1365	
							-1.1855	-.1978	2.4540	1.1689	.8458
							.7446	.2804	1.9130	-.5050	
							-2.4829	.2322	1.9827	-1.0316	2.5038
							.7395	-.5939	.3476	-.2148	
							3.4336	1.0756	1.6171	-1.6681	1.1718
							-2.2896	.7674	.3958	2.4921	
							-1.8548	-.9116	2.4413	-.3442	2.0743
							1.8115	-1.1914	.3061	-.2626	
							6.1531	-.5816	1.0459	.7849	-.2810
							1.0999	-.7463	.0441	.1274	
							-1.9378	-.8541	2.5385	-.3882	1.8312
							2.0831	-1.1967	1.2099	-.6512	
							4.9614	-1.2369	2.2250	.3812	.7359
							.9807	1.7911	2.0361	.7467	
							-1.27103	-.8725	2.9007	-.2494	1.8714
							1.3479	-.4975	.8122	-1.2801	
							-1.9133	-1.3170	2.4660	.2813	2.2855
							1.1725	-.4285	.3688	1.0594	
							-1.8000	-3.4573	-1.6891	-.2567	1.9680
							-1.7658	-.5172	-.0469	-.3833	
							-1.8308	-1.8859	-.24387	.0659	2.2907
							1.2233	-.7546	-.2259	.9409	
							-1.9080	-1.0401	2.4532	-.1393	2.0937
							1.7602	-.6375	.8364	-.1727	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	378			4.	4 .5536 .9025	2 .0369	-1.8790	-1.3884	2.4494	.3775	2.3991
ELPHEXA	379			4.	4 .7392 .9742	2 .0251	-1.9033	-1.2332	2.4093	1.5987	2.2896
ELPHEXA	380			4.	4 .4372 .9929	2 .0066	1.4388	-1.539	.6351	.3835	1.8755
ELPHEXA	381			4.	4 .2467 .9986	2 .0010	1.2860	-1.476	.5996	.4243	1.2400
ELPHXA	382			4.	4 .1094 .9995	2 .0004	1.4601	-1.8665	1.6720	-1.3216	1.0508
ELPHEXA	383			4.	4 .2191 .9979	2 .0020	-1.4343	-1.8980	1.4277	-1.7474	1.4780
ELPHEXA	384			4.	4 .1880 .9261	2 .0682	-1.2649	-1.553	1.4178	-1.8707	.5839
ELPHEXA	385			4.	4 .9125 .9910	2 .0089	-1.3648	-1.5060	3.8419	-1.1338	1.9789
ELPHXA	386			2.	2 .0198 .9230	4 .0747	.2049	-1.021	1.1157	-2.7106	2.2466
ELPHEXA	387			8.	8 .0997 .9999	1 .0001	-1.5436	-1.1046	3.4073	.5045	1.0429
ELPHEXA	388			8.	8 .8797 1.0000	1 .0000	-1.2308	-1.0006	.1116	-1.792	.6487
ELPHEXA	389			8.	8 .9996 1.0000	1 .0000	-1.426	-1.8162	2.0388	-1.6295	.8085
ELPHEXA	390			8.	8 .9906 1.0000	1 .0000	-1.5985	-1.0337	-3.1723	-1.7425	1.0318
ELPHEXA	391			4.	4 .3789 .7984	1 .2000	-1.4574	1.6645	-2.7772	-1.8046	.2438
ELPHEXA	392			4.	4 .4056 .8076	1 .1910	-1.2226	-1.3310	-4.0124	2.0458	.1690
ELPHEXA	393			4.	4 .5195 .9941	2 .0050	.0974	1.1603	1.1533	-1.1149	1.7476
ELPHEXA	394			4.	4 .2008 .9392	2 .0979	-1.1157	-1.2633	-4.1870	-1.3302	1.1226
ELPHEXA	395			4.	4 .1688 .9770	2 .0141	-1.118	-1.5370	-1.0874	-1.6888	.6160
ELPHEXA	396			4.	4 .1725 .9987	2 .0011	-1.1312	-1.4336	-4.2779	-2.3142	1.1303
ELPHEXA	397			8.	8 .1102 .9937	1 .0056	-1.3782	1.0385	-1.1961	.2766	1.0507
ELPHEXA	398			8.	8 .5726 1.0000	1 .0000	-1.7543	-2.4211	2.3442	-1.8799	1.5109
ELPHEXA	399			8.	8 .9997 1.0000	1 .0000	-1.7481	1.2044	.2317	-1.5100	.7557
ELPHEXA	400			8.	8 .9776 1.0000	1 .0000	-1.7824	-2.4291	2.4908	-1.8481	.6193
ELPHEXA	401			8.	8 .9201 .9998	1 .0002	-1.7466	1.1196	.4248	-1.4230	.7221
ELPHEXA	402			8.	8 .8496 1.0000	1 .0000	-1.8779	-2.2095	2.7596	-1.3498	.6647
ELPHEXA	403			8.	8 .8045 1.0000	1 .0000	1.0060	-1.542	.6256	-2.109	.6739
ELPHEXA	404			8.	8 .8987 1.0000	1 .0000	.6448	-2.1864	2.5096	-2.4348	-.0966
ELPHEXA	405			8.	8 .9268 1.0000	1 .0000	.9000	.1110	.1760	1.7356	-.0404
ELPHEXA	406			8.	8 .7738 1.0000	1 .0000	-1.7449	-2.634	3.5345	1.2758	-.0851
							-1.1879	1.1058	1.1659	-2.2227	
							-1.3809	-1.7411	3.2207	1.7425	
							.2613	-1.6672	1.6990	-2.1875	
							-1.3113	-2.0617	-2.4110	1.6805	
							-1.6960	1.8455	-1.1100	2.3364	
							-1.4015	-1.8410	-3.7472	1.3945	
							1.2128	.6211	1.4160	1.0680	
							-1.1483	-1.0017	-4.3125	1.9201	
							.3555	.1797	.1837	-2.2624	
							-1.1777	-1.9416	-4.2584	1.7802	
							.6004	.8011	.7401	-1.9508	
							-1.2741	-1.4577	-3.4678	1.3170	
							-1.0855	1.0039	-2.2772	-1.5767	
							-1.2212	-1.0674	-4.2727	1.9741	
							.5485	.8644	1.2035	-1.9010	
							-1.1546	-1.1074	-4.2165	1.7193	
							.4333	.9224	.8412	-1.7825	
							-1.9573	-1.3122	-4.5585	1.2335	
							.0392	-1.0921	.9921	-1.8414	
							-1.9356	-1.3000	-4.6028	1.2471	
							.0552	-1.9325	.6977	-1.7764	
							-1.9395	-1.0673	-4.6886	1.0518	
							.4392	-1.2606	.7026	-1.0104	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GRJUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GRJUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	407			8.	8 .9335 1.0330	1 .0000	-1.0071	-1.3229	-4.6829	1.5619	.0523
ELPHEXA	408			8.	8 .9848 1.0330	1 .0000	-.0711	-1.1317	.8000	.8627	
ELPHEXA	409			8.	8 .7955 1.0000	1 .0000	-.9715	-1.5123	-4.5751	1.5042	.0629
ELPHEXA	410			8.	8 .9878 1.0330	1 .0000	-.2902	-.5475	.8386	-.3092	
ELPHEXA	411			8.	8 .6024 1.0000	1 .0000	-.9047	-1.8214	-4.5331	1.5830	.2966
ELPHEXA	412			8.	8 .9970 .9999	1 .0001	-.7815	-.2663	-.0195	.0964	
ELPHEXA	413			4.	4 .0192 .9917	3 .0076	-.9665	-1.3964	-4.5305	1.5197	.0859
ELPHEXA	414			4.	4 .1992 .9933	2 .0059	-.5659	-.7894	.4767	.2635	
ELPHEXA	415			8.	8 .9924 1.0330	1 .0000	-.8562	-1.0998	-4.6682	.7868	-.0232
ELPHEXA	416			8.	8 .9973 1.0330	1 .0000	.3285	-1.1642	-.4207	-1.9112	
ELPHEXA	417			8.	8 .9689 1.0330	1 .0000	-1.1076	-1.8188	-3.8428	1.32019	.2098
ELPHEXA	418			8.	8 .9579 1.0330	1 .0000	-.7461	-.3699	-.3226	.2749	
ELPHEXA	419			4.	4 .4119 .9987	3 .0009	-.0428	-1.3747	3.9062	-.3226	.3240
ELPHEXA	420			8.	8 .9675 1.0000	1 .0000	-1.9995	-2.5976	-1.8382	-1.3437	
ELPHEXA	421			8.	8 .9627 1.0330	1 .0000	-.4713	-1.4737	3.8080	.6639	1.3292
ELPHEXA	422			4.	4 .2370 .9526	2 .0374	-1.3872	-1.0966	-2.1697	.0070	
ELPHEXA	423			4.	4 .1948 .9973	3 .0014	-.9483	-1.5912	-4.5606	1.5991	.1342
ELPHEXA	424			8.	8 .9388 1.0000	1 .0000	-.5659	-.7346	.3720	.2123	
ELPHEXA	425			8.	8 .9932 1.0330	1 .0000	-.8385	-1.3817	-4.6302	1.5289	.2536
ELPHEXA	426			8.	8 .9388 1.0000	1 .0000	-.5781	-.5972	.0378	.0060	
ELPHEXA	427			8.	8 .7983 1.0000	1 .0000	-.9723	-1.5588	-4.5864	1.7554	.1167
ELPHEXA	428			8.	8 .9932 1.0330	1 .0000	-.4983	-.7709	.6180	.7494	
ELPHEXA	429			8.	8 .9910 1.0330	1 .0000	-.9747	-1.3368	-4.6714	1.5350	.0727
ELPHEXA	430			8.	8 .9787 1.0330	1 .0000	-.1166	-1.0977	.6147	.4977	
ELPHEXA	431			8.	8 .9931 1.0330	1 .0000	-.9894	-.9729	4.0360	-.1381	.7435
ELPHEXA	432			8.	8 .3282 1.0000	1 .0000	-1.5806	-1.1090	-.7534	-1.4678	
ELPHEXA	433			8.	8 .9935 1.0000	1 .0000	-.9855	-1.5243	-4.6330	1.5856	.1704
ELPHEXA	434			4.	4 .7366 .9892	2 .0064	-.4751	-.7159	.5512	.8712	
ELPHEXA	435			4.	4 .7366 .9892	2 .0064	-1.2396	-1.3480	-4.7181	2.4095	.8216
							-.0383	1.0538	.8259	.3717	
							-.8033	-.8809	3.6181	-.4743	2.0185
							.4501	-.4686	-2.2714	-1.2038	
							-.3620	-1.1701	4.3616	.5187	.8498
							-1.5277	-.9779	-1.1940	-1.4507	
							-1.0166	-1.4822	-4.6215	1.6859	.0650
							-.2063	-.2652	1.0975	.3224	
							-1.3096	-1.2523	3.0528	-.2824	1.2042
							-.4390	-1.6324	.3364	-.0955	
							-.9868	-1.7701	-4.6077	2.0029	.2906
							-.8359	-.3081	.4244	1.2055	
							-.9403	-1.5711	-4.6242	1.6444	.2062
							-.5337	-.6514	.2886	.3920	
							-.9791	-1.6135	-4.6502	1.6744	.2671
							-.7044	-.8859	.1404	1.7268	
							-.9297	-1.4956	-4.6321	1.5314	.1321
							-.2657	-.4753	.7565	-.1931	
							-.9437	-1.3522	-4.6345	1.4461	.0864
							-.1641	-1.0708	.4403	.1241	
							-.9414	-1.4993	-4.6376	1.4838	.1557
							-.2834	-.4444	.6450	.3509	
							-.4492	-1.3015	-4.6313	1.7986	.0404
							.8100	.4457	1.3712	.9350	
							-.9245	-1.8071	-4.5566	1.7660	.3008
							-.9427	-.2710	.0842	.3860	
							-1.0316	-1.4581	-4.6313	1.7986	.0404
							-.1592	-.6043	1.3010	.7058	
							-1.6291	-1.4505	2.0285	-.5622	1.6441
							.9695	-1.9093	.2498	.6298	

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES						
SUBFILE	VAL	GROUP	GROUP P(X7G) P(G7X)	GROUP P(G7X)							
SEQNUM	SEL										
ELPHEXA	436	8.	8 .9766 1.0000	1 .0000	-1.0116	-1.7136	-4.5574	1.8429	.01525		
ELPHEXA	437	4.	4 .0632 .9959	3 .0024	-.6007	-.1863	.9269	.4366			
ELPHEXA	438	4.	4 .5718 .9984	2 .0012	-.2212	-1.6168	3.4268	-.1412	.5974		
ELPHEXA	439	4.	4 .0750 .9951	3 .0044	-1.5683	-2.0797	-2.0557	-.3156			
ELPHEXA	440	8.	8 .9727 1.0000	1 .0000	-1.6144	-.9260	3.0272	-.7804	1.0331		
ELPHEXA	441	8.	8 .9745 1.0000	1 .0000	-.0672	-2.6198	-.1065	-.7930			
ELPHEXA	442	4.	4 .3701 .9987	3 .0010	-.6897	-1.1830	3.6130	-.8643	.1657		
ELPHEXA	443	4.	4 .8313 .9989	2 .0007	-2.0514	-2.7876	.1933	-1.3003			
ELPHEXA	444	4.	4 .6576 .9990	2 .0005	-.9492	-1.3706	-4.5978	1.4011	.0237		
ELPHEXA	445	1.	1 .9923 .9553	4 .0206	-.1956	-1.1340	2.052	-.0519			
ELPHEXA	446	5.	5 .5145 1.0000	7 .0000	-.9841	-1.5664	-4.6000	1.7461	.1385		
ELPHEXA	447	1.	1 .9943 .9961	4 .0038	-.4988	-.7462	.5618	.7241			
ELPHEXA	448	1.	1 .4282 .9761	4 .0238	-1.0343	-1.0667	4.1504	.0475	.6467		
ELPHEXA	449	1.	1 .6346 .9369	4 .0929	-1.8450	-1.4887	.17263	-.6840	.9888		
ELPHEXA	450	1. ***	5 .6451 1.0000	1 .0000	-1.1783	-1.2180	3.5741	-.0378			
ELPHEXA	451	5.	5 .3752 .9999	7 .0001	-1.1688	-1.2228	.6581	-.4071	.9572		
ELPHEXA	452	1.	1 .5047 .9482	4 .0457	-1.2592	-1.4354	3.6720	.3333			
ELPHEXA	453	4. ***	1 .9917 .9661	4 .0331	-1.4854	-.9969	1.0434	.4135	-1.0367		
ELPHEXA	454	5.	5 .6574 1.0000	1 .0000	-1.2473	-1.5783	-.3057	-.1400			
ELPHEXA	455	5.	5 .7603 1.0000	1 .0000	.5743	.8303	-.6008	-.6195	.9980		
ELPHEXA	456	1.	1 .7460 .9988	4 .0012	2.9046	-.21398	-.2864	-.44356			
ELPHEXA	457	1.	1 .5367 .9992	4 .0008	.6451	-1.4791	-.0483	.1425	-1.3972		
ELPHEXA	458	5.	5 .8255 1.0000	7 .0000	-1.1232	-2.4874	.4014	-.11470			
ELPHEXA	459	2. ***	4 .9772 .9638	2 .0333	-2.730	.0333	.1913	-.0415	.4698		
ELPHEXA	460	5.	5 .3004 1.0000	1 .0900	-1.0343	-3.9971	1.0049	-.5966			
ELPHEXA	461	1.	1 .8730 .9977	4 .0022	1.0409	-1.5115	.4770	-.4978	-.0222		
ELPHEXA	462	5. ***	1 .7705 .9279	4 .0713	-1.8799	-3.1405	1.2089	-1.3232			
ELPHEXA	463	5.	5 .9806 1.0000	7 .0000	-.3914	-.9047	-.0572	.9868	1.1053		
ELPHEXA	464	1.	1 .9250 .9536	4 .0100	2.8760	-2.3424	-.2979	-.41235	.5982		
					.3256	-1.0959	-.0210	.8017			
					4.0533	-1.7528	-.5325	-2.7870			
					.7939	-.6294	-2.1320	.6931			
					-2.0789	-1.1346	.2871	-2.2943			
					-.8970	-.5696	-1.2494	.0311			
					-1.2472	-2.3409	-.6596	-.4136			
					-.7395	-.2825	.3178	.4803			
					2.8158	-2.2784	.3573	-.11904			
					.5958	-1.0325	.7262	.3425			
					3.5010	-2.1393	-.4684	-2.8837			
					.2084	.2531	.4941	1.9511			
					-.2741	-3.6848	.2283	.2557			
					.9018	-.0312	.0954	-.0532			
					-.8657	-2.7643	.0146	-1.6112			
					-.4123	-1.5749	-.4563	-.4005			
					3.5698	-1.7836	-.3500	-3.2997			
					.6336	-.6453	-.0191	1.5670			
					-.9369	-.9890	2.5827	.0060			
					1.2159	.0458	.1535	-.8453			
					2.8111	-2.0698	-.1892	-2.8097			
					-.1411	-.6303	.3365	3.0783			
					-.9109	-3.0901	-.8958	-1.7630			
					.4834	.4144	-.3396	-.6170			
					-1.7588	-2.0629	.6217	-1.4419			
					.3621	-1.8050	-.3214	.0505			
					3.3300	-1.8224	-.6077	-3.1933			
					.3398	-.7043	.3761	1.0218			
					-1.3079	-2.2753	.9281	-.7527			
					.4056	.9260	-.4369	-1.3673			

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES									
SUBFILE	VAL	SEL	GROUP	GROUP	P(X/G)	P(G/X)	GROUP	P(G/X)						
SEQNUM														
ELPHEXA	465		5.	5	.7324	1.0000	1	.0000	2.8081	-2.4751	.3339	-3.9021	1.0858	
ELPHEXA	466		1.	1	.5269	.9060	4	.0939	.2751	-.5861	.6096	-.9369		
ELPHEXA	467		1.	1	.6852	.9397	4	.0901	-1.8048	-2.7246	1.1236	-1.7951	-.1361	
ELPHIXA	468		5.	5	.7003	1.0000	1	.0000	.2458	-1.5573	-.3710	-.5484		
ELPHEXA	469		1.	1	.9759	.9993	4	.0006	-1.8331	-2.9271	1.1343	-1.6844	-.0279	
ELPHEXA	470		5.	5	.4443	1.0000	1	.0000	-.0740	-1.1730	-.3463	.2096		
ELPHEXA	471		5.	5	.2176	.9982	10	.0018	2.7884	-2.4807	.1366	-3.8797	1.0333	
ELPHEXA	472		5.	5	.6915	1.0000	1	.0000	2.762	-.6456	.7450	.9979		
ELPHEXA	473		5.	5	.0157	.9352	10	.0148	-1.8356	-2.5623	-.0662	-1.1919	-1.5284	
ELPHEXA	474		5.	5	.0870	1.0000	1	.0000	.9783	.0917	-.5528	.6814		
ELPHEXA	475		5.	5	.2658	1.0000	7	.0000	2.5578	-1.8593	.7309	-3.8023	1.4544	
ELPHEXA	476		1.	1	.9370	.9983	4	.0015	1.3848	.3321	1.1015	-.5551		
ELPHEXA	477		9.	9	.5253	.9336	2	.0515	3.4805	-1.7623	-.0054	-2.7159	-.0727	
ELPHEXA	478		9.	9	.9862	.9976	2	.0022	1.4190	2.4739	1.1310	.3989		
ELPHEXA	479		9.	9	.6669	.9993	2	.0004	2.7836	-2.4820	.3745	-3.8743	1.0206	
ELPHEXA	480		9.	9	.9070	.9986	2	.0009	.2764	-.6600	.1778	1.0127		
ELPHEXA	481		9.	9	.5571	.9547	2	.0452	3.1831	-.8412	-.6499	-1.1024	4.4681	
ELPHEXA	482		9.	9	.9996	.9996	2	.0004	.3113	.7601	1.8634	-.4411		
ELPHEXA	483		9.	9	.9938	.9999	2	.0001	2.4067	-3.1208	-1.6007	-3.7250	3.16375	
ELPHEXA	484		9.	9	.9949	.9994	2	.0005	-.7528	-1.1470	.16845	-.8480		
ELPHEXA	485		9.	9	.9393	1.0000	2	.0009	2.6966	-2.1149	-.1479	-4.7282	.9868	
ELPHEXA	486		9.	9	.9543	1.0000	2	.0000	1.6461	-1.8754	.4914	.0465		
ELPHEXA	487		9.	9	.9959	.9999	2	.0001	-.8466	-2.4833	.5257	-1.4327	-1.9048	
ELPHIXA	488		9.	9	.9854	.9895	2	.0103	.4792	.0787	-.3843	-.9283		
ELPHEXA	489		9.	9	.7006	.8743	2	.1222	-.9065	3.4467	-.2518	.1120	-.5466	
ELPHEXA	490		9.	9	.9489	.9889	2	.0108	-.5802	-.4702	-2.0402	-.0948		
ELPHEXA	491		9.	9	.9914	.9999	2	.0001	-1.8717	3.8820	-.2007	.5426	-.0661	
ELPHEXA	492		9.	9	.9983	.9988	2	.0012	-.0869	1.0695	.6667	.2833		
ELPHEXA	493		9.	9	.7265	.9850	3	.0090	-1.7348	4.1765	.3548	.3149	-.5202	
									-2.2508	1.2248	1.4804	-1.4547	-.3925	
									-1.7598	4.1487	.2659	.6619		
									-.4121	1.0711	1.2210	-.0400		
									-1.2943	3.8721	-.5065	.5958	.2999	
									1.0202	.5775	-1.9969	-.1525		
									-2.0013	4.0588	-.7345	.0661	.0163	
									.9947	.2399	.4775	.2060		
									-2.0288	4.2914	-.7824	-.0428	-.1096	
									1.4042	-.1860	.7521	.2466		
									-1.3714	3.8795	-.6330	-.0099	.0081	
									.8098	.7947	.6843	-.6880		
									-2.0180	4.3296	-.7110	-.3297	-.2337	
									1.5837	-.0350	1.0953	-.9504		
									-2.0195	4.3488	-.7638	-.2682	-.1820	
									1.6209	.0143	1.0748	-.7285		
									-1.9635	4.2640	-.7610	-.2563	-.0665	
									1.3138	-.1157	.3874	-.4815		
									-2.1513	3.5650	.0402	-.2928	.0920	
									.3520	.9018	-.3446	-.0562		
									-2.1639	3.1338	-.0984	.1629	.3149	
									-.3629	1.5959	-.5343	.6801		
									-2.1827	3.4777	-.6700	.5297	.7849	
									-.3797	.8064	-.0596	.2462		
									-2.0133	4.1161	-.6971	-.1175	-.0960	
									1.2237	.3923	.9267	-.6363		
									-2.0314	1.8155	-.6631	.3175	.0617	
									.6368	.5627	.5469	.6353		
									-1.86014	3.3922	-.5827	.2767	-.5043	
									-1.1362	-.4613	-.2025	1.8045		

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST GROUP	PROBABILITY P(X%G)	PROBABILITY P(G%X)	2ND HIGHEST GROUP	PROBABILITY P(G%X)	DISCRIMINANT SCORES				
ELPHEXA	494			9.	9	.9971	.9952	2	.0046	-1.9729	3.5600	-.5205	-.0923	.1229
ELPHEXA	495			9.	9	.9845	.9998	2	.0002	-.1276	.9395	.1321	-.0946	
ELPHEXA	496			9.	9	.9496	.9962	2	.0038	1.1892	.4732	.6530	-.9623	-.0316
ELPHEXA	497			9.	9	.9965	.9999	2	.0001	-1.9905	3.7904	-.7396	-.4480	.2624
ELPHEXA	498			9.	9	.9988	.9988	2	.0012	.4136	.5085	-.1071	1.4339	
ELPHEXA	499			9.	9	.9887	.9999	2	.0001	-1.9773	4.2641	-.7494	-.2286	-.0918
ELPHEXA	500			9.	9	.9961	.9986	2	.0014	1.1221	-.1429	.4753	-.3012	
ELPHEXA	501			9.	9	.9964	.9994	2	.0006	-2.0201	3.8345	-.6701	-.2919	.0800
ELPHEXA	502			9.	9	.9591	1.0000	2	.0000	.6281	.9853	.4757	.5522	
ELPHEXA	503			9.	9	.7340	.9524	2	.0463	-2.0187	4.3193	-.8699	.0221	-.0113
ELPHEXA	504			9.	9	.9787	.9999	2	.0001	1.4510	-.0064	.6525	.5025	
ELPHEXA	505			9.	9	.9481	1.0000	2	.0000	-2.0073	3.8547	-.7379	.3279	.1609
ELPHEXA	506			9.	9	.9954	.9998	2	.0002	.5587	.5693	.3020	.7026	
ELPHEXA	507			9.	9	.9912	.9939	2	.0060	-2.0036	3.9030	-.5889	-.0863	.0523
ELPHEXA	508			9.	9	.9978	.9990	2	.0010	.8016	.0321	.7254	-.3402	
ELPHEXA	509			9.	9	.9978	.9987	2	.0013	-2.0394	4.3730	-.8152	-.1546	-.1498
ELPHEXA	510			9.	9	.9627	.9950	2	.0050	1.6786	.0367	1.1636	-.3239	
ELPHEXA	511			9.	9	.9993	.9995	2	.0005	-1.9681	3.3063	-.5821	-.7052	.3996
ELPHEXA	512			9.	9	.9827	.9950	2	.0050	-1.4221	1.2351	-.4370	1.3409	
ELPHEXA	513			9.	9	.9935	.9987	2	.0013	-1.9265	4.2284	-.6921	-.4388	-.0993
ELPHEXA	514			9.	9	.7650	.9930	2	.0036	1.2244	-.1312	.2067	-1.1233	
ELPHEXA	515			9.	9	.9993	.9998	2	.0002	-1.9418	4.5031	-.8696	-.4433	-.0825
ELPHEXA	516			9.	9	.9997	.9996	2	.0004	1.1055	-.4136	.3460	-.6777	
ELPHEXA	517			9.	9	.9993	.9995	2	.0005	-1.8004	4.4993	-.2697	-.7724	-.2200
ELPHEXA	518			9.	9	.9993	.9998	2	.0002	-.2670	3.678	.5032	-.9842	
ELPHEXA	519			9.	9	.9993	.9998	2	.0002	-1.9407	3.5795	-.6031	-.2288	.2380
ELPHEXA	520			9.	9	.7319	1.0000	2	.0000	1.453	1.0651	-.1053	-.0754	
ELPHEXA	521			6.	6	.0025	.9734	3	.0247	-1.9774	3.9886	-.7453	1.860	.1137
ELPHEXA	522			6.	6	.0002	.9383	3	.0407	.7341	.3785	-.0200	.8227	
										-2.0217	3.8424	-.6908	.3193	.0992
										.6440	.6032	.4741	.6532	
										-1.9877	3.7995	-.5990	.6320	.0402
										.428	.5610	.3246	-.0541	
										-1.9599	4.0718	-.7978	.0427	.1113
										1.0005	.3456	.2555	.1545	
										-2.0173	3.6323	-.6868	.5426	.2466
										.2907	1.0398	.3011	1.0168	
										-2.0315	3.8558	-.7206	.3739	.1195
										.5753	.6186	.5151	.8539	
										-1.7232	3.9000	-.3518	.7032	-.2958
										-.8576	1.4727	.9039	-.3241	
										-1.8523	4.3237	-.2942	.1053	-.2467
										.6438	.3192	.7967	-.3755	
										-1.9669	4.0136	-.6362	-.1406	-.0486
										.3887	.4722	.3288	-.5320	
										-2.1511	4.4258	-.8687	.2287	-.2064
										1.8444	-.0689	1.7943	.9890	
										-2.4430	2.9001	-.2285	-.1283	-1.6123
										1.4730	.3309	.5789	.3927	
										-2.2924	2.5998	-.2145	-.1612	-1.4644
										.8665	.8291	-.3414	-1.1628	
										-2.2282	2.4419	-.6091	-.3476	-1.6140
										-.1404	1.3649	.2370	-.4518	
										-2.1254	-.8888	1.0652	.3621	-2.3560
										-.8540	-1.7557	.0047	-1.4842	
										4.0518	-.0090	1.4112	.3879	-2.3745
										-.3090	-3.0680	-.7118	-1.6570	

CASE SUBFILL	SEQNUM	MIS VAL	SEL	ACTUAL GRUPO	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GRUPO P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	523			6.	6 .0043 .6330	2 .2042	1.5556	-.6435	1.5112	3.3660	-1.07495
ELPHEXA	524			4.	4 .0226 .9928	3 .0067	.3897	-1.2845	-1.0216	-.5281	
ELPHEXA	525			6.	6 .5030 1.0000	5 .0000	-1.9977	-2.5984	-1.6287	-1.2402	
ELPHEXA	526			8.	8 .2992 .9973	2 .0026	1.5463	-.6708	-.8034	1.9749	
ELPHEXA	527			8.	8 .2687 .9999	2 .0001	-.7400	1.1361	-2.9245	2.2394	2.2092
ELPHEXA	528			4.	4 .2876 .9992	3 .0006	-.6246	-.5300	-1.3679	.3297	2.2473
ELPHEXA	529			8.	8 .0999 .9999	2 .0001	-.8615	.0350	-2.6874	3.6831	2.2473
ELPHEXA	530			8.	8 .0737 .7263	2 .2702	-.1053	.4496	1.2081	.3165	
ELPHEXA	531			4.	4 .7881 .9971	2 .0028	-1.1017	-1.1182	4.1438	1.336	.6058
ELPHEXA	532			6.	6 .7678 .9997	10 .0003	-1.6286	-.7332	1.6891	-1.1950	
ELPHEXA	533			8.	8 .1987 .9765	2 .0034	-.1830	-.4663	-2.8082	3.4747	1.9785
ELPHEXA	534			8.	8 .0815 .9980	2 .0019	-.8845	-1.1163	-2.3650	.2515	
ELPHEXA	535			6.	6 .2546 .9963	10 .0037	-1.0344	1.2734	-2.1313	1.8888	2.9336
ELPHEXA	536			8. ***	9 .0113 .5414	8 .4313	.5657	.0741	-1.0875	-.6717	
ELPHEXA	537			8.	8 .0583 .8693	2 .1302	-1.4040	-.8736	3.9510	.4603	1.6022
ELPHEXA	538			6.	6 .1998 .8071	10 .1929	-.5473	-.0356	.5457	-.7287	
ELPHEXA	539			6.	6 .8391 .9999	10 .0001	3.2295	-.3894	1.9913	1.3305	1.0031
ELPHEXA	540			8.	8 .7700 1.0000	1 .0000	1.8094	-.3268	-.1613	.4084	
ELPHEXA	541			6.	6 .6984 .9722	10 .0578	-.6530	1.0914	-2.8727	1.9295	2.2430
ELPHEXA	542			4.	4 .9105 .9927	2 .0372	-.7608	-.4591	-1.8560	-.7355	
ELPHEXA	543			6.	6 .2639 .8810	10 .1190	-.2503	-.0036	-1.1506	.1991	2.7112
ELPHEXA	544			4.	4 .7693 .9759	2 .0225	5.4461	-.4611	1.0257	1.0131	1.4123
ELPHEXA	545			6.	6 .7714 .9981	10 .0019	2.7193	-.2927	-.2166	-.2969	
ELPHEXA	546			6.	6 .2213 .9401	10 .0599	-2.2362	1.8338	-2.2235	1.0196	2.6087
ELPHEXA	547			4.	4 .4796 .9329	2 .0670	-1.2281	-.1566	1.0471	2.6235	2.3299
ELPHEXA	548			6.	6 .9391 1.0000	10 .0000	-.2973	1.0011	-2.0347	.0852	
ELPHEXA	549			6.	6 .4032 .9793	10 .0307	4.8586	-.9097	1.4707	.6632	.4686
ELPHEXA	550			8.	8 .5445 1.0000	1 .0000	1.3103	1.2952	1.8113	.9620	
ELPHEXA	551			6.	6 .6978 .9992	10 .0008	5.5586	-.6787	1.5802	1.2639	-.5181
							-.4565	.2549	1.6682	.8657	.6389
							-1.6918	-.2761	-4.9853	.2757	
							-1.2976	-.6030	-.7164	-.5775	
							3.1892	-.0733	.3866	1.1876	.1379
							1.4269	.8342	1.7865	-.0543	
							-1.5712	-1.1125	3.4531	.5357	1.9054
							-.2372	-.0840	.0013	.2647	
							5.8429	-.7008	2.0564	1.8851	.6149
							.9963	1.3316	-1.3574	-.5259	
							-1.8753	-1.1118	2.4391	-.0413	2.2032
							1.4915	-.7839	.2773	.3713	
							6.1066	.1240	-1.0621	1.3072	.1275
							1.1304	.6895	-.9360	-1.2519	
							3.2518	.0253	1.9649	1.3633	-.2558
							.9673	1.3113	2.8449	-.8298	
							-1.0346	-1.2571	3.1833	.7899	2.2590
							.8112	-.0582	-1.7736	-.1149	
							6.3039	.7786	.0201	2.6158	-.2305
							1.0288	.4357	-.2965	-.7062	
							5.7943	-.2525	-.0500	1.5418	.8101
							2.2439	.9428	-1.2638	.1833	
							-.4907	-1.6505	-3.8796	2.8543	1.0778
							-.3891	1.1163	-2.1600	-.1305	
							5.4888	.0007	1.9595	1.1924	.7584
							1.6132	-.1427	-1.2775	-.0724	

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES								
SUBFILE	VAL	GROUP	GROUP P(%)	GROUP P(%)									
SEQNUM	SEL		P(%)	P(%)									
ELPHEXA	552	6.	6 .9548 1.0000	10 .0000	6.4969	.1767	.7771	2.1570	-.9180				
ELPHEXA	553	8.	8 .9684 1.0000	1 .0000	-.6344	-.1811	-.9018	.2217					
ELPHEXA	554	6.	6 .2502 .5126	10 .4874	-.8766	-1.9048	-4.5102	1.7546	.3643				
ELPHEXA	555	6.	6 .1567 .5755	10 .4239	-1.2696	-.4549	-.5303	.5358					
ELPHEXA	556	4.	4 .0999 .9737	2 .0259	4.8368	-.7111	1.6831	.9698	.9851				
ELPHEXA	557	6.	6 .2851 .9999	10 .0001	.4594	1.3722	1.6643	.6586					
ELPHEXA	558	8.	8 .8845 1.0000	1 .0000	5.1330	-.0893	1.8517	-.2849	.3491				
ELPHEXA	559	4.	4 .2306 .9283	2 .0715	-.6141	.8901	-1.0253	1.2890					
ELPHEXA	560	9.	9 .9669 .9996	2 .0003	-.2233	-2.3990	3.0400	1.9757	1.9264				
ELPHEXA	561	9.	9 .9959 .9997	2 .0003	1.6574	-.5411	-1.5086	-.2780					
ELPHEXA	562	9.	9 .9884 .9999	2 .0001	5.7225	-.5349	.0349	2.0784	.8635				
ELPHEXA	563	9.	9 .9925 .9999	2 .0001	1.6853	.7886	-1.2972	1.7748					
ELPHEXA	564	9.	9 .9761 .9997	2 .0003	-1.1931	-1.6555	-4.1820	2.6379	1.0189				
ELPHEXA	565	9.	9 .7877 1.0000	2 .0000	-.7134	1.2900	.0877	.9382					
ELPHEXA	566	9.	9 .4694 1.0000	2 .0000	-2.7477	-.2571	-2.6543	-.7662	2.6327				
ELPHEXA	567	9.	9 .9989 .9998	2 .0002	.0595	-1.973	-.2101	1.8769					
ELPHEXA	568	9.	9 .9996 .9999	2 .0001	-1.7382	4.3788	1.831	4.477	-.4347				
ELPHEXA	569	9.	9 .9985 .9978	2 .0002	-.0363	.7477	1.1925	-.3388					
ELPHEXA	570	9.	9 .9968 .9996	3 .0002	-2.0389	4.0830	-.7705	2.145	.0156				
ELPHEXA	571	9.	9 .6913 .7977	3 .1757	1.0626	.2227	.6794	.7209					
ELPHEXA	572	9.	9 .9625 .9966	2 .0028	-2.0807	4.2426	-.6005	-.0877	-.3442				
ELPHEXA	573	9.	9 .9862 .9979	2 .0001	1.3428	-.4359	1.1718	.0166					
ELPHEXA	574	9.	9 .8731 .9979	2 .0011	-2.0292	4.1279	-.7170	-.0495	-.0917				
ELPHEXA	575	9.	9 .8954 .9998	2 .0001	1.2550	.3895	1.0099	-.3989					
ELPHEXA	576	9.	9 .9211 .9655	3 .0276	-2.0393	4.0871	-.7389	3.274	-.0661				
ELPHEXA	577	9.	9 .8230 1.0000	2 .0000	1.1002	.1216	.9255	1.0874					
ELPHEXA	578	1.	1 .9917 .9986	4 .0010	-2.0681	4.5877	-.8118	-.3155	-.3287				
ELPHEXA	579	9.	9 .9953 .9985	3 .0008	2.0557	-.4410	1.4842	-.4807					
ELPHEXA	580	9.	9 .9942 .9991	3 .0006	-1.7949	4.3736	-1.2008	-1.0012	-.9669				
					1.5922	-2.1897	1.0998	-.1920					
					-1.8787	4.3162	-.2504	1.1352	-.3170				
					.6453	.2396	.9779	-.2938					
					-1.8394	4.4592	-.2916	-.0023	-.3206				
					.7324	-.3051	.5131	-.1288					
					-1.8091	4.3496	-.3358	-.2579	-.2412				
					.7147	.3083	.9915	.1559					
					-1.5858	3.9060	-.6877	-.3152	-.7399				
					-.1534	-1.0002	.3007	.4218					
					-1.5249	2.9495	-.3797	-.1380	-.4022				
					-1.8120	.5214	-.4808	.2690					
					-1.7815	3.8262	-.1204	1.1929	-.0579				
					-.2457	1.1170	.1782	-.2282					
					-1.5813	4.1453	-.7824	-.4668	-.7872				
					.2501	-1.3359	.3697	.3383					
					-1.6853	4.1095	.5130	-.3939	-.3663				
					-.3304	1.1290	.8068	-.9623					
					-1.5824	4.0880	-.8082	-.2277	-.6992				
					.0287	-1.5177	.0040	1.2618					
					-1.5864	3.1926	-.4283	-.1290	-.5610				
					-1.3622	.0487	.0045	.6461					
					-1.7519	4.0893	-1.2086	-.6992	-.7066				
					.9621	-1.8932	.3347	.6500					
					-.8362	-1.9041	-.1175	-.5885	-2.0549				
					.6150	.5728	-.0538	.0131					
					-1.5831	3.6969	-.6848	-.0854	-.5932				
					-.5038	-.5650	.1376	.8075					
					-1.5530	3.7058	-.5472	-.4999	-.7651				
					-.3911	-.4518	.3946	-.8553					

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GRUP	HIGHEST PROBABILITY GRUP P(X/G) P(G/X)	2ND HIGHEST GRUP P(G/X)	DISCRIMINANT SCORES							
ELPHEXA	581			9.	9 .9934 .9990	3 .0002	-1.6091	-1.38761	-.5834	-.3596	-.8681			
ELPHEXA	582			9.	9 .9761 .9999	3 .0000	-1.1961	-1.1345	.4978	.2270	-.9062			
ELPHEXA	583			9.	9 .9957 .9992	3 .0006	.1066	-1.4573	.05116	.0522	-.8036			
ELPHEXA	584			9.	9 .9969 .9999	3 .0001	-1.5724	-3.7049	-.5280	-.4639	-.8746			
ELPHEXA	585			9.	9 .9995 .9994	3 .0004	-.3813	-.4976	.5204	-.7423	-.7902			
ELPHEXA	586			9.	9 .9995 .9994	3 .0004	-1.6115	-3.9643	-.6418	-.31679	-.7902			
ELPHEXA	586			9.	9 .9197 .9589	3 .0329	.0859	-.8754	.8363	-.3204	-.7902			
ELPHEXA	587			9.	9 .9692 .9999	3 .0001	-1.6142	3.7368	-.5823	-.2835	-.4984			
ELPHEXA	589			9.	9 .9957 .9996	3 .0002	-1.2963	-.5030	.7371	-.1116	-.4984			
ELPHEXA	589			9.	9 .9835 .9929	3 .0054	-1.5519	3.1922	-.4566	-.0589	-.9107			
ELPHEXA	589			9.	9 .9835 .9929	3 .0054	-1.3837	.1245	-.2158	.4223	-.9107			
ELPHEXA	590			9.	9 .9838 .9999	2 .0001	-1.5928	-1.4501	.3714	-.3335	-.8134			
ELPHEXA	591			9.	9 .9685 .9999	2 .0000	-1.1895	-1.0744	.13794	.2218	-.7181			
ELPHEXA	592			9.	9 .9813 .9999	2 .0000	-1.6128	-3.4134	-.4442	-.0178	-.7181			
ELPHEXA	593			9.	9 .9194 .9573	3 .0265	-.9758	-.4069	.3004	.5451	-.1117			
ELPHEXA	594			9.	9 .8697 .9145	3 .0644	-1.9917	-4.0965	-.6604	-.2203	-.1117			
ELPHEXA	595			9.	9 .9882 .9997	2 .0002	1.1738	.3866	.8194	-.9983	-.9282			
ELPHEXA	596			9.	9 .9340 .9983	3 .0015	-1.6230	-4.1229	-.6833	-.4519	-.9282			
ELPHEXA	597			9.	9 .9533 .9947	3 .0038	-.2324	-1.4908	.6727	.3457	-.8411			
ELPHEXA	597			9.	9 .9407 .9998	3 .0001	-1.5978	4.1371	-.7451	-.4588	-.8411			
ELPHEXA	597			9.	9 .9397 .9999	3 .0001	.2444	-1.3954	.4886	.3491	-.6038			
ELPHEXA	600			9.	9 .9778 .9999	3 .0001	-1.6025	3.1880	-.4017	.1472	-.6038			
ELPHEXA	601			9.	9 .9280 .9999	2 .0001	-1.3616	.0003	.0148	.6959	-.6038			
ELPHEXA	602			9.	9 .8868 1.0000	2 .0000	-1.7755	3.0385	.1944	-.3705	-.5809			
ELPHEXA	603			9.	9 .9279 .9999	2 .0000	-1.2724	.5752	-.1649	.1541	-.7148			
ELPHEXA	604			9.	9 .9706 .9956	3 .0031	-1.6193	-3.9368	-.7456	-.1549	-.7148			
ELPHEXA	605			9.	9 .5856 1.0000	2 .0000	-.0755	-.9906	.4668	.9862	-.10323			
ELPHEXA	606			9.	9 .9931 .9996	3 .0002	-1.4429	4.0756	-.1620	-.1717	-.10323			
ELPHEXA	607			9.	9 .9743 .9998	3 .0001	-.7943	-1.2185	.3477	.3623	-.6305			
ELPHEXA	609			9.	9 .9509 .9999	3 .0001	-1.5482	-3.5722	-.5512	-.1533	-.6305			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.8740	-.8521	-.3510	.7298	-.7715			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.5510	4.0487	-.7155	-.4201	-.7715			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-.0584	-1.5783	-.0703	.6751	-.10141			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.5885	4.0689	-.5623	-.6893	-.10141			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	1.087	-1.5579	.5244	-.5054	-.8678			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.5649	4.1053	-.6862	-.6184	-.8678			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	.1666	-1.4069	.3269	-.2134	-.8678			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.7050	3.8693	-1.0514	-.4712	-.6201			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	.6004	-1.5233	.2901	1.0108	-.6201			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.5915	4.3473	-.7587	-.7032	-.9681			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	.5915	-1.8077	.5925	-.0814	-.9681			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.6502	4.1602	-.7569	-.2641	-.8908			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	.3250	-1.4709	.8511	1.0096	-.8908			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.4408	3.9169	-.1122	-.1691	-.9963			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-.9355	-.6243	.5717	-.2449	-.9963			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.7238	4.2982	-1.2357	-.9868	-.8004			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	1.2952	-2.2646	.3137	.0800	-.8004			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.5995	3.8668	-.5654	-.4072	-.8768			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-.2194	-1.1388	.4510	.0591	-.8768			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.6005	3.9107	-.6371	-.1517	-.8649			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.1002	-1.1540	.7720	.9499	-.8649			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.5745	4.0764	-.5935	-.6906	-.9707			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	.1150	-1.5107	.4358	-.4964	-.9707			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-1.1841	-2.4008	.7748	-.7222	-.10186			
ELPHEXA	609			4. ***	1 .9811 .9585	4 .0310	-.8803	-.3068	.0089	-.6079	-.10186			

CASE	MIS	ACTUAL	HIGHEST PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES									
SUBFILE	SEL	GRUP	GROUP P(X/G) P(G/X)	GROUP P(G/X)										
ELPHEXA	610	6. ***	2 .0015 .6167	6 .1897	1.2554	-.3754	.5008	2.8247	-1.5243					
ELPHEXA	611	4.	4 .1865 .9817	1 .0128	2.3632	-1.7907	-.6497	-1.1842						
ELPHEXA	612	4.	4 .1550 .9946	3 .0024	-.0899	-.8126	1.6414	2.6727	2.1630	-.7170				
ELPHEXA	613	4.	4 .1233 .9898	2 .0084	-.5782	-1.9115	1.4501	3.0016	2.8017	-.5406				
ELPHEXA	614	6.	6 .5721 1.0000	10 .0000	1.1131	-2.5487	2.4143	1.6375	1.6372	1.8927				
ELPHEXA	615	4.	4 .0936 .9950	1 .0020	1.7536	-1.2517	.5122	1.6375	2.3463	-1.1271				
ELPHEXA	616	4. ***	3 .3355 .7566	4 .02135	5.7186	.0071	2.1026	.9827	2.3463	-1.1271				
ELPHEXA	617	8.	8 .9755 1.0000	1 .0000	-1.0847	.3725	2.1026	.9827	2.3463	-1.1271				
ELPHEXA	618	6.	6 .5514 1.0000	10 .0000	-.3391	-1.4832	2.3122	-1.1010	2.1261	-.5150				
ELPHEXA	619	4.	4 .3263 .9976	1 .0016	-2.2329	-.4331	-.6478	-1.7731	-.6020					
ELPHEXA	620	6.	6 .9439 1.0000	10 .0000	-.8790	-2.0462	-.44883	1.8588	.4040					
ELPHEXA	621	4. ***	1 .5188 .9956	4 .0034	-1.3635	-.1322	-.1755	.2828	2.0839	-.8458				
ELPHEXA	622	4.	4 .1511 .9930	2 .0069	-.6954	-.2419	.1521	2.0839	-.8458					
ELPHEXA	623	4.	4 .0474 .9738	3 .0193	-.5030	-.3046	1.8506	1.8370	1.3460	1.0117				
ELPHEXA	624	6.	6 .9671 1.0000	10 .0000	-.1794	-1.9669	.2670	1.0336	1.2281	-.7317				
ELPHEXA	625	8.	8 .9338 1.0000	1 .0000	5.5014	-.0145	1.5576	.2828	1.2281	-.7317				
ELPHEXA	626	6. ***	10 .3639 .8586	6 .1410	.5504	-.1946	1.5576	.2828	1.2281	-.7317				
ELPHEXA	627	6.	6 .9660 1.0000	10 .0000	-.5876	-3.7229	.0258	1.8530	-1.0063					
ELPHEXA	628	4.	4 .0810 .8405	2 .1593	.5275	-.8293	.3690	1.3245	1.4913					
ELPHEXA	629	4. ***	5 .0000 .4730	10 .3069	-.1057	-2.1741	3.6201	1.9561	1.4913					
ELPHEXA	630	4.	4 .0190 .9907	3 .0041	1.2495	-.6422	-.9240	-1.0181	2.21923	-.8980				
ELPHEXA	631	6. ***	3 .0167 .9269	10 .0526	-.7845	-1.5689	1.2951	-2.1370	2.27710	-.5910				
ELPHEXA	632	6.	6 .9072 1.0000	10 .0000	-.3508	-.3475	-.8496	.7276	1.9449	.4726				
ELPHEXA	633	6.	6 .9043 1.0000	10 .0000	-1.4607	-2.1855	-4.4828	1.9449	.4726					
ELPHEXA	634	7.	7 .0000 1.0000	5 .0000	4.4527	.7525	.0949	-.0244	1.6765	.4185				
ELPHEXA	635	7.	7 .5931 1.0000	9 .0000	-.2561	1.6045	1.6249	1.5120	2.6138	-.8586				
ELPHEXA	636	7.	7 .5138 1.0000	9 .0000	6.6726	.2724	-.1230	-2.6138	-.8586					
ELPHEXA	637	6.	6 .9779 1.0000	10 .0000	-.5531	.1495	-.1045	-.2191	3.526	2.9712				
ELPHEXA	638	6.	6 .9658 1.0000	10 .0000	-2.6893	-.4350	2.5216	-.3526	2.9712					
					-2.649	.4580	-.17797	2.2671						
					3.6693	-.9100	-.0374	.2993	3.6798					
					-3.7632	.3531	1.7806	.9122						
					-1.5133	-2.0435	3.0050	2.9924	-.1905					
					4.1019	.4279	1.5296	1.4320						
					-3.0477	2.5809	1.2543	1.4218	.2282					
					6.0306	-.6393	1.2543	1.4218	-.2069					
					.8062	-.3352	-.3991	1.6543						
					6.5582	-.8154	-.3991	1.6543	-1.2516					
					.5621	-.5987	1.1845	-.4415						
					4.7737	4.7370	-3.4933	-1.0327	2.3799					
					.8023	-2.0099	1.13270	-3.6454						
					3.1252	-.85245	-1.6329	-1.8065	.2401					
					4.225	-.8822	1.0239	-.1938						
					3.1026	-.82097	-1.13718	-1.0366	.1562					
					4.081	-.8954	1.1920	-.2106						
					6.5874	-.5028	.1302	2.6138	-1.0972					
					-.1133	-.4108	.5528	.1639						
					6.5991	.5872	.0709	2.4658	-1.0888					
					.1529	-.1312	.8078	-.5171						

CASE	SUBFILE	SEQNUM	VAR	SEL	ACTUAL	HIGHEST	PROBABILITY	2ND HIGHEST	DISCRIMINANT SCORES					
					GROUP	GROUP	P(1 2) P(2 1)	GROUP						
ELPHEXA		639			6.	6	.1627 .9963	10 .0037	4.9134	-.6918	1.7461	.5635	.3860	
ELPHEXA		640			4.	4	.0002 .9934	1 .0283	-.5712	-.5719	2.0605	.5777		
ELPHEXA		641			6.	6	.9617 1.0000	10 .0000	.1336	-1.9377	2.2339	2.0127	1.1015	
ELPHEXA		642			6.	6	.9365 1.0000	10 .0000	-2.1661	-3.2881	1.8202	-2.3967		
ELPHEXA		643			6.	6	.9912 1.0000	10 .0000	5.4362	.0884	.8706	2.1175	-1.0512	
ELPHEXA		644	***		4.	3	.1179 .5599	4 .4349	-.4917	.4881	.0764	-.7343		
ELPHEXA		645			6.	6	.9997 1.0000	10 .0000	6.5143	.6838	-.4634	2.0370	-.8985	
ELPHEXA		646	***		4.	5	.0004 .8005	5 .0556	.9425	-1.1030	-.0050	-.2252		
ELPHEXA		647			6.	6	.7440 1.0000	10 .0000	6.4625	.2583	2.3153	2.0925	-1.0025	
ELPHEXA		648			6.	6	.9795 1.0000	10 .0000	-.3460	-.0116	-.0377	-.1983		
ELPHEXA		649			6.	6	.9831 1.0000	10 .0000	-2.2704	-.2294	2.3153	.9154	-1.0212	
ELPHEXA		650			6.	6	.9232 1.0000	10 .0000	6.2875	.0562	.9340	-.0211		
ELPHEXA		651			4.	4	.0174 .9894	2 .0057	.2714	.0731	-.0530	-.1926	-.8335	
ELPHEXA		652			4.	4	.0492 .9994	2 .0004	4.3587	-.6632	3.5458	-.1798	2.0365	
ELPHEXA		653			6.	6	.0072 .9538	2 .0302	-1.3957	-1.1104	-1.2934	-.2475		
ELPHEXA		654			7.	7	.5647 .9998	5 .0002	6.5068	.8592	-.5559	2.0960	-.8964	
ELPHEXA		655			6.	6	.6911 1.0000	10 .0000	1.1185	-1.6630	-.0215	.6079		
ELPHEXA		656			4.	4	.5640 .9585	3 .0364	6.6450	.4784	1.498	2.4246	-1.0599	
ELPHEXA		657			6.	6	.9468 1.0000	10 .0000	-.1936	-.3495	.0226	-.4815		
ELPHEXA		658	***		6.	6	.0006 .9992	6 .0003	6.6368	.4901	1.239	2.4766	-1.0420	
ELPHEXA		659			8.	8	.9718 1.0000	1 .0000	-.1667	-.3338	.2569	-.2953		
ELPHEXA		660			8.	8	.0892 .9961	2 .0037	6.3715	-.2025	2.2432	2.2123	-.4532	
ELPHEXA		661	***		4.	2	.3706 .9506	4 .0334	-.3537	.5037	-1.2052	.5673		
ELPHEXA		662			4.	4	.8365 .9107	2 .0789	1.0000	-1.4624	3.2982	2.1236	-.4370	
ELPHEXA		663			6.	6	.9751 1.0000	10 .0000	-.2961	-2.0241	-1.0538	-2.1267		
ELPHEXA		664			6.	6	.0043 1.0000	7 .0000	4.4552	2.3160	4.2418	1.7206	.4794	
ELPHEXA		665			7.	7	.2888 .9998	5 .0002	-.6807	-1.5050	-.8318	-1.7699		
ELPHEXA		666			6.	6	.9558 1.0000	10 .0000	1.6939	-.7427	.8628	4.3957	-1.5957	
ELPHEXA		667			6.	6	.0011 .9553	3 .0283	.1771	-.0913	-.0094	-.1026		
									2.7744	2.8148	-.2.8588	-4.1060	-1.0745	
									-.2112	-.0548	.0435	.3264		
									6.6461	1.0146	-.0061	-1.9303	-1.2588	
									.8404	-.8591	.7037	-1.3138		
									-.5911	-.5180	3.1733	1.0867	-.1054	
									-1.7701	-1.0423	1.0471	-.0663		
									6.5555	.3279	.9357	2.6151	-1.1518	
									-.2709	.0831	-.0357	-.4784		
									1.1598	-.8815	-1.5537	4.9201	1.1367	
									-1.4029	-1.6409	-.3346	-2.1403		
									-1.2421	-1.7907	-3.7985	1.5839	1.1216	
									.1929	.9745	.5916	-.6276		
									-1.6140	1.1325	-2.6749	2.5867	2.8527	
									-1.1959	.9667	.0453	1.0455		
									.1591	2.297	4.781	2.5034	-.3476	
									.4880	-1.2569	-1.3547	1.0626		
									-.0935	-.3963	2.7527	1.3183	.2345	
									-.2532	-1.3282	-.7516	-.0706		
									6.6324	.7394	.0090	2.3727	-1.0776	
									.2608	-.6841	.3569	-.2091		
									6.6219	1.9026	-.1523	1.1954	-1.7233	
									2.3478	-2.4874	1.4392	-2.0379		
									2.4250	3.0500	2.6295	-1.6376	1.4574	
									-.0951	-.8936	-.1641	2.363		
									6.3117	-.2569	-.7968	2.1011	-.7120	
									1.1780	-.7215	.2616	.0633		
									1.7489	-.8187	2.2945	3.8219	-1.6971	
									-.4178	-2.8900	-.4838	.3066		

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GROUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GRJUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	668			7.	7 .2320 .9995	5 .0005	2.5507	3.3424	-.1407	-3.9721	1.5004
ELPHEXA	669			4.	4 .0466 .9959	1 .0019	.2276	-1.1591	.6548	1.2023	-.6473
ELPHEXA	670			1.	1 .0027 .7207	4 .2712	.2124	-1.1537	3.0855	1.9375	2.0950
ELPHEXA	671			8.	8 .7536 .9995	1 .0005	-2.0317	-3.6346	2.8230	-1.0059	.7412
ELPHEXA	672			4. ***	10 .0005 .9725	6 .3373	-2.0122	-.3806	.9473	-2.0884	1.4914
ELPHEXA	673			8.	8 .7502 1.0000	1 .0000	-1.2444	-2.7817	-3.4875	1.2896	1.2660
ELPHEXA	674			4.	4 .5927 .6653	2 .3322	-1.1669	.8394	.2484	-.9756	.9928
ELPHEXA	675			6.	6 .9650 1.0000	10 .0000	-1.4017	-1.8803	2.3337	-.0811	-.7576
ELPHEXA	676			8.	8 .1127 .9999	2 .0001	-.7019	-.7829	2.3337	-.0811	1.8868
ELPHEXA	677			6.	6 .2396 1.0000	10 .0000	-1.9761	-.2286	-4.1127	1.9662	-1.9342
ELPHEXA	678			6.	6 .9611 1.0000	10 .0000	-.9533	1.6882	-.0107	-.3240	-.9420
ELPHEXA	679			6.	6 .9752 1.0000	10 .0000	-.3336	-1.2503	2.3117	2.1121	-.9831
ELPHEXA	680			6.	6 .8809 1.0000	10 .0000	-.0526	.2850	-.9340	.2232	-.7844
ELPHEXA	681			8.	8 .2080 .9966	2 .0033	6.5433	.2617	-.3812	2.6369	2.1179
ELPHEXA	682			7.	7 .6910 1.0000	5 .0000	.2793	-.4138	.1543	.9859	.1817
ELPHEXA	683			4. ***	3 .1650 .5196	2 .4309	-.4110	-1.5120	-1.5779	3.4926	-1.1960
ELPHEXA	684			7.	7 .1671 1.0000	9 .0000	6.3361	-.8703	4.4518	2.4307	-.1349
ELPHEXA	685			7.	7 .3922 1.0000	5 .0000	6.197	-1.9021	2.3421	.1812	1.502
ELPHEXA	686			7.	7 .9586 1.0000	3 .0000	6.6006	-.2900	1.350	2.8111	.4688
ELPHEXA	687			7.	7 .9492 1.0000	3 .0000	-.4772	.0372	3.2837	.4397	.2650
ELPHEXA	688			1.	1 .7311 .9653	2 .0201	6.5038	-.4082	1.7026	1.9764	-1.2155
ELPHEXA	689			1.	1 .3346 .5557	8 .4431	-.2472	-.5331	-.8359	.0324	-.9197
ELPHEXA	690			5.	5 .8165 1.0000	7 .0000	6.3219	-.6300	-1.3506	2.2395	1.9187
ELPHEXA	691			5.	5 .7602 1.0000	7 .0000	.7226	-1.2556	-.4122	.6853	1.9418
ELPHEXA	692			5.	5 .6691 .9941	7 .0058	-.9375	-.8919	-1.5537	-.7403	-.4607
ELPHEXA	693			5.	5 .4501 .9894	7 .0105	3.8649	4.8853	-1.2108	-3.1343	-.7518
ELPHEXA	694			5.	5 .3192 1.0000	7 .0000	1.2123	-1.9266	-1.5466	-1.2948	1.8883
ELPHEXA	695			5.	5 .1854 1.0000	7 .0000	-.8294	.2441	-1.6191	2.8571	2.6373
ELPHEXA	696			5.	5 .5117 .9999	7 .0001	3.8578	-.2441	-1.4610	-3.3308	-.4250
							2.0765	-2.5981	-1.3600	-1.0760	
							3.8792	5.1220	-1.3094	-3.3159	
							1.8043	-2.2420	-1.5494	-1.4772	
							3.6852	4.2251	-1.6274	-3.0726	
							3.3178	-.7803	-2.4039	-.4884	
							3.5385	4.4427	-1.5168	-3.1887	
							.7104	-1.2500	-1.0729	-1.4374	
							-1.9038	-.4531	-1.2247	-1.5956	
							-.1893	-.0875	-.4711	1.1469	
							-1.5536	-2.7361	-.0061	-1.0387	
							-1.1673	-.0654	-.2105	-.3757	
							2.5643	-.8785	-2.07137	-3.8713	
							-1.0322	.2318	1.907	4.070	
							2.6057	-.8973	-2.6960	-4.0114	
							-1.0924	.2724	-.0448	-.0723	
							2.9374	1.040	-.0448	-3.5682	
							-.0295	.2416	2.827	.7578	
							3.2350	-.0046	-1.3639	-4.1328	
							-.4787	1.0642	-.2911	.5318	
							2.6303	-.6452	-3.2068	-2.9503	
							-.5543	.8943	-.2060	1.8641	
							2.3913	-.9967	-2.4352	-4.9844	
							-.6534	-.7828	-.0771	-1.4381	
							2.8787	-.5638	-.3934	-4.5475	
							.6016	.8237	.8086	1.4333	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GRJUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	697			2.	2 .2084 .7949	4 .1937	-1.9129	-1.57362	-1.7242	-1.0319	1.0353
ELPHEXA	698		***	2.	4 .5214 .4261	2 .2780	-2.2562	-1.5717	-1.6469	2.7258	-1.2656
ELPHEXA	699			4.	4 .2424 .9620	1 .0167	-2.2491	-1.6154	1.5961	-1.1093	1.8764
ELPHEXA	700			3.	3 .4138 .5257	4 .4234	-2.0365	-1.7376	.3345	1.3520	-1.2213
ELPHEXA	701		***	4.	2 .7556 .7836	4 .2179	-1.3620	.1735	1.4775	-1.479	1.2695
ELPHEXA	702			6.	6 .8601 1.0030	10 .0000	-1.3374	-1.2279	1.0578	1.16941	-2.2347
ELPHEXA	703			4.	4 .6596 .5724	2 .4196	.9637	-.3693	-.0512	1.7463	1.0891
ELPHEXA	704		***	2.	2 .6610 .9755	3 .0184	6.2309	-.6800	1.7104	1.8795	-1.1390
ELPHEXA	705			4.	4 .7860 .6028	2 .3946	-1.2431	.0275	-.0825	1.4233	1.5775
ELPHEXA	706		***	2.	2 .0016 .6217	6 .2918	-1.2143	-.0985	1.6134	1.1578	-0.9922
ELPHEXA	707			4.	4 .7703 .9937	2 .0062	1.3724	.0738	.5460	1.4036	1.9063
ELPHEXA	708		***	1.	4 .8373 .7167	2 .1811	-.4386	1.1796	-.2175	1.6291	.3153
ELPHEXA	709			2.	2 .8557 .9724	4 .0243	.3829	-.3805	-2.7753	.3263	1.1879
ELPHEXA	710			2.	2 .9491 .9464	4 .0391	-1.3771	-.6435	1.45330	1.0252	1.4351
ELPHEXA	711			2.	2 .8931 .8883	4 .0724	1.2683	-.4571	.0100	1.2936	.0444
ELPHEXA	712			4.	4 .6570 .8189	2 .8776	-1.5710	-.2774	-2.0432	.9554	1.1819
ELPHEXA	713			6.	6 .4390 1.0030	10 .0000	-1.6314	-1.2430	3.4765	1.7485	.1866
ELPHEXA	714		***	3.	3 .2331 .5771	2 .3715	-2.2882	.4490	.8793	.3762	.0103
ELPHEXA	715			4.	4 .8909 .7414	2 .2490	-1.4398	-.5958	2.1558	1.4718	.8329
ELPHEXA	716		***	3.	3 .7777 .6296	4 .2691	-.3531	-.3868	.6399	1.2844	-1.1578
ELPHEXA	717			4.	4 .6339 .5145	2 .4797	-1.7245	1.0505	1.6001	1.8676	1.0598
ELPHEXA	718			4.	4 .8929 .6768	2 .1003	.4972	.5078	.0914	2.0458	1.0339
ELPHEXA	719			4.	4 .8528 .7323	2 .2440	-1.5352	1.1900	.17768	1.3653	.6603
ELPHEXA	720		***	2.	2 .3760 .5000	4 .4934	1.2267	.8191	.0585	.7904	1.1102
ELPHEXA	721			4.	4 .6198 .6336	2 .1923	-1.2522	-.7911	-.3075	-0.0507	.0515
							.4494	-.3693	-.0371	1.6334	
							-1.3015	-.1860	1.5007	1.2619	
							2.1447	-1.1509	.6029	-.7348	
							5.9047	-1.4866	.7429	1.6094	
							.4380	.2043	-.0562	2.3980	
							-2.0205	1.9844	1.2421	-.8409	
							-1.8002	1.6144	-.7687	.9815	
							-1.1332	-.5235	1.6555	.3944	
							1.9404	-.1227	.3073	-.4054	
							-1.2128	.9879	2.3558	1.0343	
							-1.7659	.2101	.2192	.0267	
							-1.1951	-.7513	1.5779	1.0776	
							1.4791	.0955	.2036	1.7495	
							-1.7329	1.3533	1.7584	-1.0194	
							-1.5332	-1.0358	-.3640	-.1930	
							-1.6106	.0813	1.8193	-.4600	
							-.5461	-.0809	.0451	1.6109	
							-1.2499	-.8679	1.5575	1.3219	
							1.4214	.6707	.7220	1.9861	
							-.7570	-1.3502	1.3173	-.0153	
							1.0340	.1315	.5334	1.0797	

CASE SUBFILE	SEQNUM	MIS VAL	SEL	ACTUAL GRUP	HIGHEST PROBABILITY GROUP P(X/G) P(G/X)	2ND HIGHEST GROUP P(G/X)	DISCRIMINANT SCORES				
ELPHEXA	697			2.	2 .2084 .7949	4 .1937	-1.9129	-1.577362	-0.677242	-2.0319	1.0353
ELPHEXA	698			2. ***	4 .5214 .4261	2 .2780	-2.2562	-1.5717	-0.6468	2.7258	
ELPHEXA	699			4.	4 .2424 .9020	1 .0167	-0.2491	-0.0959	1.0961	1.1093	-1.2656
ELPHEXA	700			3.	3 .4138 .5207	4 .4234	-2.0365	-1.7376	0.3345	-1.1771	1.8764
ELPHEXA	701			4. ***	2 .7556 .7836	4 .2179	-1.3620	-0.0210	1.4775	-1.479	-1.2213
ELPHEXA	702			6.	6 .8601 1.0030	10 .0000	-1.53374	-0.2279	1.5195	0.0877	1.2695
ELPHEXA	703			4.	4 .6596 .5724	2 .4196	-0.9637	-0.3893	-1.0578	1.1463	
ELPHEXA	704			3. ***	2 .6010 .9755	3 .0184	6.2509	-0.6800	1.7104	1.8795	-0.2347
ELPHEXA	705			4.	4 .7860 .6328	2 .3946	-2.431	0.0275	-0.825	1.4233	
ELPHEXA	706			6. ***	2 .0016 .6277	6 .2918	-1.2143	-0.8985	1.6134	1.1578	1.0891
ELPHEXA	707			4.	4 .7703 .9337	2 .0062	1.3724	0.6738	0.5460	1.4036	1.0891
ELPHEXA	708			1. ***	4 .0373 .7167	2 .1011	-0.4380	1.1796	0.2175	1.6291	-0.1390
ELPHEXA	709			2.	2 .6557 .9724	4 .0243	0.3829	-0.3805	-2.7753	0.3263	
ELPHEXA	710			2.	2 .9491 .9404	4 .0391	-1.33771	-1.4535	1.4530	1.0252	1.5775
ELPHEXA	711			2.	2 .8931 .8883	4 .0724	1.2683	-0.4571	0.0100	1.2436	
ELPHEXA	712			4.	4 .6570 .8189	2 .0776	1.15710	-1.2878	-2.0432	0.0059	-0.9922
ELPHEXA	713			6.	6 .4390 1.0030	10 .0000	0.0837	-0.2774	0.9257	0.554	
ELPHEXA	714			9. ***	3 .2331 .5771	2 .3715	-1.6314	-1.2430	3.4765	0.7485	1.9063
ELPHEXA	715			4.	4 .6909 .7414	2 .2490	-2.882	0.4490	0.8733	0.3762	
ELPHEXA	716			4. ***	3 .7777 .6296	4 .2691	-1.1498	0.5958	1.1858	0.3718	0.3153
ELPHEXA	717			4.	4 .6339 .5145	2 .4797	-0.5331	-0.3868	0.6399	1.2044	
ELPHEXA	718			4.	4 .8929 .6768	2 .1003	-1.7245	1.80505	1.6001	0.8676	1.1879
ELPHEXA	719			4.	4 .8528 .7323	2 .2440	0.4972	0.5078	0.914	2.0458	
ELPHEXA	720			4. ***	2 .3760 .5000	4 .4934	-1.5502	1.1900	0.7768	0.363	0.4351
ELPHEXA	721			4.	4 .6198 .6336	2 .1923	1.2267	0.8191	0.585	0.7804	
							-1.2522	-0.3893	-0.3371	1.6334	0.0444
							0.4494	-0.1860	1.5007	0.2019	1.1819
							-1.3615	-1.1509	0.6028	-0.7348	
							2.1447	-1.4866	1.7429	-1.6034	0.1866
							5.9047	0.2049	-0.0362	2.3980	
							-2.0205	1.9844	1.2421	-0.8409	0.0103
							-1.8022	1.6144	-0.7687	0.9815	0.8329
							-1.1332	-0.5235	1.6555	-0.4054	
							1.9404	-1.227	0.3073	-0.267	0.0343
							-1.2128	0.9879	2.3558	0.287	-0.1578
							-1.7659	0.2101	1.2192	0.0267	
							-1.1951	-0.7513	1.5779	1.10776	1.0598
							1.4791	0.0955	0.2036	1.7495	
							-1.7329	-1.033533	-1.7584	-1.0194	1.0339
							-1.6186	0.0813	1.8193	-0.4600	0.6603
							-0.5461	-0.0809	0.0451	1.6109	
							1.02499	-0.8679	1.6575	1.3219	1.1102
							1.4213	0.5707	0.7250	1.9861	
							-0.7370	-1.3502	1.3173	0.9153	0.0515
							1.0340	-1.1315	0.3334	1.8797	

CLASSIFICATION RESULTS =

ACTUAL GROUP	NO. OF CASES	PREDICTED GROUP MEMBERSHIP									
		1	2	3	4	5	6	7	8	9	
GROUP 1	163	156 95.7	1 .6	0	1 .6	1 .6	0	0	0	4 2.5	0
GROUP 2	61	7 11.5	43 70.5	4 6.6	3 4.9	0	1 1.6	0	0	1 1.6	2 3.3
GROUP 3	51	1 2.0	3 5.9	30 70.6	1 2.0	4 7.8	0	4 7.8	0	0	0
GROUP 4	134	22 16.4	21 15.7	5 3.7	83 61.9	1 .7	0	0	0	0	0
GROUP 5	46	1 2.2	0	0	0	43 93.5	0	2 4.3	0	0	0
GROUP 6	69	0	2 2.9	1 1.4	0	0	63 91.3	0	0	1 1.4	0
GROUP 7	20	0	0	0	0	2 10.0	0	18 90.0	0	0	0
GROUP 8	67	2 3.0	0	0	0	0	0	0	64 95.5	1 1.5	0
GROUP 9	100	0	0	1 1.0	0	0	0	0	0	3 3.0	96 96.0
GROUP 10	10	0	0	0	0	0	0	0	0	0	0

ACTUAL GROUP		NO. OF CASES	PREDICTED GROUP MEMBERSHIP
			10
GROUP	1	163	0
GROUP	2	61	0
GROUP	3	51	3.9
GROUP	4	134	1.5
GROUP	5	46	0
GROUP	6	69	2.9
GROUP	7	20	0
GROUP	8	67	0
GROUP	9	100	0
GROUP	10	10	100.0

PERCENT OF GROUPED CASES CORRECTLY CLASSIFIED - 84.80

APPENDIX E: Specimen depositories.

Representative collections have been deposited with the following persons / departments: Ann Miller, Department of Geology, Dalhousie; Dr. S. W. Snyder, Department of Geology, East Carolina University, Greenville, N. Carolina, 27834; Dr. G. Vilks/Dr. C. Schafer, Atlantic Geoscience Centre, Bedford Institute of Oceanography, Dartmouth, N.S. B2Y 4A2; Dr. M. A. Buzas, Department of Paleobiology, U. S. N. M. Nat. History, Smithsonian Inst., Wash., D. C. 20560; Dr. C. W. Poag, U. S. G. S., Woods Hole, Mass., 02543; Prof. R. Feyling-Hanssen, Department of Micropaleotology, University of Århus, Universitetsparken, DR-8000, Århus C. Denmark; Department Invert. Palaeo., R. O. M., 100 Queen's Park Blvd., Toronto, Ont., M5S 2C6; Dr. L. Osterman, INSTAAR, University of Colorado, R. B. #1, Boulder, Co., 80302; Miss Ruth Todd, P. O. Box 4687, Vineyard Haven, Mass., 02568; Dr. K. McDougall, U. S. G. S., 345 Middlefield Rd., Menlo Park, Ca., 94025; Dr. D.H. McNeil, Inst. of Sedimentary and Petroleum Geology, Geological Survey of Canada, Calgary, Alberta T2L 2A7; Prof. J.R. Haynes, Dept. of Geology, University College of Wales, Aberysthwyth, Cards., SY23 3DB, U.K.; Prof. J.W. Murray, Dept. of Geology, University of Exeter, Exeter, BX4 4QE, U.K.; and Dept. of Invertebrate Palaeontology, British Museum of Natural History, Cromwell Road, London, SW7 5BD, U.K.

Elphidium excavatum (Terquem)
forma *tumida* Natland

Elphidium (*Cellanthus*) *tumidum*
Natland
PARKER *et al.*, 1953; TODD and BRÖNNIMANN, 1957; LEHMANN, 1957; PHLEGER, 1960b; SCOTT, 1976; SCOTT *et al.*, 1976.
(II)

Criboelphidium (*Elphidium*) *trinitatense*
Cushman and Brönnimann, 1948; TODD and BRÖNNIMANN, 1957.
(S)

Elphidium tumidum
Natland, 1938
(P)

Elphidium excavatum (Terquem)
forma *gunteri* Cole

Elphidium oceanense (d'Orbigny)
MURRAY, 1971, 1979; BOLTOVSKOY, 1977; APHORPE, 1980
(VV)

Elphidium gunteri Cole
CUSHMAN, 1939; PARKER *et al.*, 1953; PHLEGER, 1954, 1960a, 1960b; BANDY, 1956; LEHMANN, 1957; VAN VOORTHUYSEN, 1957, 1960; LANKFORD, 1959; HAAKE, 1962; RICHTER, 1964a; LÉVY, 1966; LÉVY *et al.*, 1969
(DD)

Elphidium littorale
Le Calvez and Le Calvez, 1951
(X)

Elphidium oceanense (d'Orbigny)
CUSHMAN, 1939
(FF)

Polystomella oceanensis
Fornasini, 1904, ab d'Orbigny, 1825
Nomen dubium
(A)

Elphidium gunteri Cole, 1931
(L)

Elphidium gunteri Cole
forma *typicum*
Poag, 1978
(BBB)

Elphidium waddense
van Voorthuysen
HAYNES, 1973

Elphidium guntheri (sic) Cole
var. *waddensis*
VAN VOORTHUYSEN, 1951
(Y)

CUSHMAN, 1930; CUSHMAN and COLE, 1930; PHLEGER, 1951; PHLEGER and PARKER, 1951; TODD and BRÖNNIMANN, 1957

Elphidium gunteri Cole
var. *galvestonense*
Kornfeld, 1931
(M)

Elphidium excavatum (Terquem)
forma *galvestonense* Kornfeld

Elphidium galvestonense
Kornfeld forma *typicum*
Poag, 1978
(DDD)

Elphidium galvestonense
Kornfeld
PARKER *et al.*, 1953; PHLEGER, 1954, 1960a; LEHMANN, 1957; PARKER and ATHEARN, 1959; TODD and LOW, 1961
(EE)

Elphidium brooklynense
Shupack, 1934
(Q)

Elphidium (*Polystomella*) *excavatum* (Terquem)
HERON-ALLEN and EARLAND, 1932; CUSHMAN, 1944; PARKER, 1952a, 1952b
(J)

Elphidium selseyense (Heron-Allen and Earland)
CUSHMAN, 1939; PARKER, 1952b
(GG)

Polystomella stratopunctata (Fichtel and Moll)
var. *selseyensis*
HERON-ALLEN and EARLAND, 1909, 1911
(F)

Polystomella excavata
Terquem, 1875, 1876
(E)

Polystomella stratopunctata (Fichtel and Moll)
(K)

Elphidium excavatum (Terquem)
forma *selseyensis*
(Heron-Allen and Earland)
MILLER *et al.*, 1982

Elphidium selseyense (Heron-Allen and Earland)
HAYNES, 1973; BANNER and CULVER, 1978. (ZZ)

Criboelphidium limnosum
Cushman and Brönnimann, 1948
PHLEGER and WALTON, 1950; TODD and BRÖNNIMANN, 1957
(T)

Elphidium florentinae
Shupack, 1934
(R)

Elphidium selseyense (sis) (Heron-Allen and Earland)
VAN VOORTHUYSEN, 1957, 1960; BRIAND, 1941; RICHTER, 1961, 1964a
(LL)

Polystomella arctica
Parker and Jones
TERQUEM, 1876
(G)

Polystomella stratopunctata (Fichtel and Moll)
(K)

Elphidium excavatum (Terquem)
forma *lidoensis* Cushman
MILLER *et al.*, 1982

Elphidium ct. advenum
Sensu, Todd and Low

Elphidium excavatum (Terquem)
forma *selseyensis*
(Heron-Allen and Earland)
FEYLING-HANSSSEN, 1972

Cribranonion (*Elphidium*) *excavatum* (Terquem)
LUTZE, 1965, 1968; LÉVY, 1966; HAAKE, 1967; LÉVY *et al.*, 1969, 1975; MURRAY, 1971
(KK)

Elphidium selseyense (sis) (Heron-Allen and Earland)
VAN VOORTHUYSEN, 1957, 1960; BRIAND, 1941; RICHTER, 1961, 1964a
(LL)

Polystomella arctica
Parker and Jones
TERQUEM, 1876
(G)

Polystomella stratopunctata (Fichtel and Moll)
(K)

Elphidium excavatum (Terquem)
forma *excavata* Terquem
MILLER *et al.*, 1982

Elphidium excavatum (Terquem)
forma *excavata* Terquem
(Terquem)

Elphidium excavatum (Terquem)
forma *lidoensis* Cushman
FEYLING-HANSSSEN, 1972

Criboelphidium vadesens
Cushman and Brönnimann, 1948
LEHMANN, 1957 (U)

Cribranonion (*Elphidium*) *lidoense*
ACCORDI and SOCINI, 1951; LÉVY, 1966; CITA and PREMOLI-SILVA, 1967; LÉVY *et al.*, 1969
(MM)

Elphidium selseyense (sis) (Heron-Allen and Earland)
VAN VOORTHUYSEN, 1957, 1960; BRIAND, 1941; RICHTER, 1961, 1964a
(LL)

Polystomella arctica
Parker and Jones
TERQUEM, 1876
(G)

Polystomella stratopunctata (Fichtel and Moll)
(K)

Elphidium excavatum (Terquem)
forma *clavata* Cushman
MILLER *et al.*, 1982

Elphidium excavatum (Terquem)
forma *clavata* Cushman
(Cushman)

Elphidium excavatum (Terquem)
forma *clavata* Cushman
FEYLING-HANSSSEN, 1972

Criboelphidium salsum
Cushman and Brönnimann, 1948
(V)

Elphidium clavatum
Cushman
HANSEN, 1965; KNUDSEN, 1971a, 1971b
(NN)

Elphidium clavatum Cushman
LOEBLICH and TAPPAN, 1953; BARTLETT, 1963, 1964; BUZAS, 1965a, 1965b; TODD and LOW, 1961, 1967. (AA)

Elphidium incertum
var. *clavatum*
Cushman, 1930, 1939, 1948; PARKER, 1952a, 1952b; TODD and LOW, 1961; BARTLETT, 1965b
(H)

Polystomella umbilicatala
var. *incerta*
Williamson, 1858
(D)

Elphidium excavatum (Terquem)
forma *magna*
Miller *et al.*, 1982

Elphidium excavatum (Terquem)
forma *magna*
Miller *et al.*, 1982

Elphidium excavatum (Terquem)
forma *alba* Feyling-Hanssen, 1972

Elphidium clavatum
Cushman
WILKINSON, 1979 (8 subspecies); RODRIGUES and HOOPER, 1982.
(AAA)

Elphidium incertum clavatum "complex"
Cushman
BARTLETT, 1965b, 1966; GREGORY, 1970; WAGNER, 1970; SCHAFFER and COLE, 1978.
(☆)

Elphidium clavatum Cushman
LOEBLICH and TAPPAN, 1953; BARTLETT, 1963, 1964; BUZAS, 1965a, 1965b; TODD and LOW, 1961, 1967. (AA)

Elphidium incertum
var. *clavatum*
Cushman, 1930, 1939, 1948; PARKER, 1952a, 1952b; TODD and LOW, 1961; BARTLETT, 1965b
(H)

Polystomella umbilicatala
Williamson, 1858
(D)

Elphidium excavatum (Terquem)
forma *williamsoni* Haynes

Elphidium williamsoni
Haynes

Elphidium excavatum (Terquem)
forma *williamsoni* Haynes

Elphidium translucens
Natland
BANDY, 1953; PARKER *et al.*, 1953; PHLEGER, 1954, 1964; TODD and BRÖNNIMANN, 1957; TODD and LOW, 1961. (HH)

Elphidium williamsoni
Haynes, 1973, ab Williamson, 1858
HANSEN and LYKKE-ANDERSEN, 1976; CULVER and BANNER, 1978; KNUDSEN, 1979; SCOTT and MEDIOLI, 1980; SCOTT *et al.*, 1980.
(YY)

Elphidium umbilicatala (Williamson)
LÉVY *et al.*, 1969; KNUDSEN, 1971b
(XX)

Elphidium excavatum
CUSHMAN, 1930, 1939, 1949; VAN VOORTHUYSEN, 1957, 1960; TODD and LOW, 1961; RICHTER, 1961, 1964a, 1967; HAAKE, 1962, 1967; FEYLING-HANSSSEN, 1964; BRODNIWICZ, 1965; ADAMS and FRAMPTON, 1965; MURRAY, 1965a, 1968, 1970; SCOTT, 1977; SCOTT *et al.*, 1977.
(I)

Polystomella umbilicatala
Williamson, 1858
(D)

Elphidium excavatum (Terquem)
forma *williamsoni* Haynes

Elphidium translucens
Natland
VON DANIELS, 1970; HANSEN and LYKKE-ANDERSEN, 1976.

Elphidium translucens
Natland
VON DANIELS, 1970; HANSEN and LYKKE-ANDERSEN, 1976.

Elphidium translucens
Natland
BANDY, 1953; PARKER *et al.*, 1953; PHLEGER, 1954, 1964; TODD and BRÖNNIMANN, 1957; TODD and LOW, 1961. (HH)

Elphidium williamsoni
Haynes, 1973, ab Williamson, 1858
HANSEN and LYKKE-ANDERSEN, 1976; CULVER and BANNER, 1978; KNUDSEN, 1979; SCOTT and MEDIOLI, 1980; SCOTT *et al.*, 1980.
(YY)

Elphidium umbilicatala (Williamson)
LÉVY *et al.*, 1969; KNUDSEN, 1971b
(XX)

Elphidium excavatum
CUSHMAN, 1930, 1939, 1949; VAN VOORTHUYSEN, 1957, 1960; TODD and LOW, 1961; RICHTER, 1961, 1964a, 1967; HAAKE, 1962, 1967; FEYLING-HANSSSEN, 1964; BRODNIWICZ, 1965; ADAMS and FRAMPTON, 1965; MURRAY, 1965a, 1968, 1970; SCOTT, 1977; SCOTT *et al.*, 1977.
(I)

Polystomella umbilicatala
Williamson, 1858
(D)

Elphidium excavatum (Terquem)
forma *cuvillieri* Lévy

Elphidium williamsoni
Haynes

Elphidium translucens
Natland
VON DANIELS, 1970; HANSEN and LYKKE-ANDERSEN, 1976.

Elphidium translucens
Natland
BANDY, 1953; PARKER *et al.*, 1953; PHLEGER, 1954, 1964; TODD and BRÖNNIMANN, 1957; TODD and LOW, 1961. (HH)

Elphidium williamsoni
Haynes, 1973, ab Williamson, 1858
HANSEN and LYKKE-ANDERSEN, 1976; CULVER and BANNER, 1978; KNUDSEN, 1979; SCOTT and MEDIOLI, 1980; SCOTT *et al.*, 1980.
(YY)

Elphidium umbilicatala (Williamson)
LÉVY *et al.*, 1969; KNUDSEN, 1971b
(XX)

Elphidium excavatum
CUSHMAN, 1930, 1939, 1949; VAN VOORTHUYSEN, 1957, 1960; TODD and LOW, 1961; RICHTER, 1961, 1964a, 1967; HAAKE, 1962, 1967; FEYLING-HANSSSEN, 1964; BRODNIWICZ, 1965; ADAMS and FRAMPTON, 1965; MURRAY, 1965a, 1968, 1970; SCOTT, 1977; SCOTT *et al.*, 1977.
(I)

Polystomella umbilicatala
Williamson, 1858
(D)

Elphidium excavatum (Terquem)
forma *cuvillieri* Lévy

Elphidium williamsoni
Haynes

Elphidium translucens
Natland
VON DANIELS, 1970; HANSEN and LYKKE-ANDERSEN, 1976.

Elphidium translucens
Natland
BANDY, 1953; PARKER *et al.*, 1953; PHLEGER, 1954, 1964; TODD and BRÖNNIMANN, 1957; TODD and LOW, 1961. (HH)

Elphidium williamsoni
Haynes, 1973, ab Williamson, 1858
HANSEN and LYKKE-ANDERSEN, 1976; CULVER and BANNER, 1978; KNUDSEN, 1979; SCOTT and MEDIOLI, 1980; SCOTT *et al.*, 1980.
(YY)

Elphidium umbilicatala (Williamson)
LÉVY *et al.*, 1969; KNUDSEN, 1971b
(XX)

Elphidium excavatum
CUSHMAN, 1930, 1939, 1949; VAN VOORTHUYSEN, 1957, 1960; TODD and LOW, 1961; RICHTER, 1961, 1964a, 1967; HAAKE, 1962, 1967; FEYLING-HANSSSEN, 1964; BRODNIWICZ, 1965; ADAMS and FRAMPTON, 1965; MURRAY, 1965a, 1968, 1970; SCOTT, 1977; SCOTT *et al.*, 1977.
(I)

Polystomella umbilicatala
Williamson, 1858
(D)

LEGEND
— in synonymy with
- - - uncertain affinity
- - - errors
☆ a name erected by Bartlett, 1965b though never published by him subsequently published by others.