

**THE FATTY ACID-BINDING PROTEIN (*fabp*) GENES OF SPOTTED GREEN
PUFFERFISH (*TETRAODON NIGROVIRIDIS*) - COMPARATIVE
STRUCTURAL GENOMICS AND TISSUE-SPECIFIC DISTRIBUTION OF
THEIR TRANSCRIPTS**

by

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Dedicated to my family

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ABSTRACT

The fatty acid-binding protein (*fabp*) genes belong to the multigene family of intracellular lipid-binding proteins (iLBP). To date, 12 different FABPs have been identified in various vertebrate genomes. Owing to the fish-specific whole genome duplication (FSGD) event, many fishes have duplicated copies of the *fabp* genes. Here, I identified and characterized the *fabp* genes of spotted green pufferfish (*Tetraodon nigroviridis*). Initially, a BLAST search was performed and ten *fabp* genes were identified, out of which, three were retained in the pufferfish genome as duplicated copies. The putative pufferfish Fabp proteins shared greatest sequence identity and similarity with their teleost and tetrapod orthologs. Conserved gene synteny was evident between the pufferfish *fabp* genes and human, zebrafish, three-spined stickleback and medaka *FABP/fabp* genes, providing evidence that the duplicated copies of pufferfish *fabp* genes most likely arose as a result of the FSGD. The differential tissue-specific distribution of pufferfish *fabp* transcripts suggests divergent spatial regulation of duplicated pairs of *fabp* genes.

LIST OF ABBREVIATIONS USED

BLAST	Basic local alignment search tool
bp	Base pair(s)
cDNA	Complementary deoxyribonucleic acid
Fabp	Fish fatty acid-binding protein
<i>fabp</i>	Fish fatty acid-binding protein gene
FABP	Human fatty acid-binding protein
<i>FABP</i>	Human fatty acid-binding protein gene
FSGD	Fish-specific genome duplication
iLBP	Intracellular lipid-binding protein
kb	Kilobase
kDa	Kilodalton
LCN1	Human lipocalin 1 protein
mRNA	Messenger ribonucleic acid
NCBI	National Center for Biotechnology Information
PCR	Polymerase chain reaction
<i>rpl13α</i>	Ribosomal protein large subunit 13 alpha gene
<i>rpl7</i>	Ribosomal protein large subunit 7
RT-PCR	Reverse transcription-polymerase chain reaction
RT-qPCR	Reverse transcription-quantitative polymerase chain reaction
SEM	Standard error of mean

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CHAPTER 1

Introduction

1.1 Fatty acid-binding proteins (FABPs)

The members of the family of intracellular lipid-binding protein (iLBP) are known to play vital roles in the transport of hydrophobic ligands such as long-chain fatty acids and retinoid to fuel various metabolic requirements and to regulate gene expression by activating specific transcription factors (Bernlohr *et al.*, 1997). iLBPs are compact, low molecular mass proteins of 14-16 kDa, having a highly conserved gene structure and protein conformation. ILBP gene structure consists of four exons separated by three introns, a common characteristic of the iLBP gene family (Bernlohr *et al.*, 1997). The protein structure is composed of ten anti-parallel β -sheets and two short α -helices. The β -sheets together form a barrel like structure with a hollow central cavity, in which the ligand binds and the two α -helices at the N-terminal end act like a lid to the β -barrel structure (Veerkamp and Maatman, 1995). This conformation forms a characteristic three-dimensional clamshell-shaped structure (Schaap *et al.*, 2002). Based on ligand affinity, and specificity for different ligands, iLBPs can be divided into three subfamilies, the retinol-binding proteins (RBPs), the cellular retinoic acid-binding proteins (CRABPs) and the fatty acid-binding proteins (FABPs/Fabps) (Hanhoff *et al.*, 2002). The multigene family of iLBPs arose as a result of various duplications and evolutionary diversification of an ancestral iLBP gene (Schleicher *et al.*, 1995). So far, 18 different iLBPs including 12 FABPs, 4 RBPs and 2 CRABPs, have been identified in vertebrates. Schaap *et al.*, (2002) reported that due to the absence of iLBPs in plants and fungi, the ancestral

iLBP gene arose after the divergence of animals from plants and fungi approximately 1000-1200 million years ago.

The FABPs have a broad specificity in binding to various ligands such as long-chain fatty acids, bile salts and eicosanoids (Smathers and Petersen, 2011). Although the exact function(s) of FABPs is poorly defined, experimental evidence suggests their role in trafficking fatty acids for lipid storage, cellular signaling, peroxisomal or mitochondrial oxidation and enzymatic interactions (Furuhashi and Hotamisligil, 2008). They are also reported to be involved in the regulation of receptor-mediated gene expression by directly binding to fatty acids and other hypolipidemic drugs (Storch and Corsico, 2008).

When FABPs were first identified, they were named based on the initial tissue of isolation, e.g., intestinal FABP (I-FABP), liver-type FABP (L-FABP). However, due to overlapping patterns of tissue-specific distribution of transcripts and their proteins i.e. different *fabps* expressed in the same tissue, here I use the nomenclature proposed by Hertzel and Bernlohr (2000), where they used Arabic numerals in the chronological order based on the initial site of isolation, i.e., FABP1 (liver), FABP2 (intestine), FABP3 (heart), FABP4 (adipocyte), FABP5 (epidermal), FABP6 (ileal), FABP7 (brain), FABP8 (myelin), FABP9 (Testis), FABP10 (liver-basic).

1.2 Teleost Fabp/fabp

Although mammalian FABPs have been well studied for four decades (Ockner *et al.*, 1972), less is known about the teleost Fabps. The teleost Fabp was initially identified from the heart tissue of ocean pout, *Macrozoarces americanus* and sea raven, *Hemitripterus americanus* (Stewart and Driedzic, 1988). Subsequently, teleost

Fabp/fabps were identified in the nurse shark (*Ginglymostoma cirratum*) liver (Medzihradszky *et al.*, 1992), aerobic muscle of striped bass, *Morone saxatilis* (Londraville and Sidell, 1996), H-FABP of rainbow trout, *Oncorhynchus mykiss* (Ando *et al.*, 1998) and Antarctic teleost fishes, *Chaenocephalus aceratus*, *Cryodraco antarcticus*, *Gobionotothen gibberifrons* and *Notothenia coriiceps* (Vayda *et al.*, 1998). More recently, *fabp* genes have been studied extensively in zebrafish, *Danio rerio* (Alves-Costa *et al.*, 2008; Denovan-Wright *et al.*, 2000a, 2000b; Karanth *et al.*, 2008; Karanth *et al.*, 2009; Liu *et al.*, 2003a, 2003b, 2004, 2007; Sharma *et al.*, 2004, 2006; Venkatachalam *et al.*, 2009), medaka, *Oryzias latipes* (Parmar *et al.*, 2012a, 2012b; Parmar and Wright, 2013a, 2013b), three-spined stickle back, *Gasterosteus aculeatus* (Parmar *et al.*, 2012a, 2012b; Parmar and Wright, 2013a, 2013b) and Atlantic salmon, *Salmo salar* (Lai *et al.*, 2009; Lai *et al.*, 2012).

Studies on zebrafish *fabp* started with the discovery of intestinal fatty acid-binding protein gene (*fabp2*) (Pierce *et al.*, 2000), followed by the identification and tissue-specific expression patterns of other paralogous zebrafish *fabps* (Alves-Costa *et al.*, 2008; Denovan-Wright *et al.*, 2000a, 2000b; Karanth *et al.*, 2008; Karanth *et al.*, 2009; Liu *et al.*, 2003a, 2003b, 2004, 2007; Sharma *et al.*, 2004, 2006; Venkatachalam *et al.*, 2009). To date, twelve *fabps* have been reported in zebrafish, where eight genes (four pairs) are retained in the genome as duplicated copies, owing to the teleost fish-specific whole genome duplication (FSGD) event. The *fabp1b.2* gene however, was identified as a tandem duplicate of *fabp1b*, presumably due to unequal crossing over during meiosis, an event that most likely arose subsequent to the FSGD (Karanth *et al.*, 2009).

Identification and structural characterization of medaka and three-spined stickleback *fabp*

genes have shown that ten *fabp* genes (five pairs) were retained in the medaka and three-spined stickleback genomes as duplicated copies (Parmar *et al.*, 2012a, 2012b; Parmar and Wright, 2013a, 2013b).

1.3 *Tetraodon nigroviridis* (Marion de Procé, 1822)

Tetraodon nigroviridis, commonly called the spotted green pufferfish (hereafter referred to simply as pufferfish), lives in fresh/brackish waters of Southeast Asia. *Tetraodon* (meaning “four teeth” in Latin) has two fused upper and lower teeth, forming a beak like structure. Interest in *Tetraodon* as model organism for genomics arose after Hinegardner in 1968, discovered its remarkably small genome of about 380 million base pairs compared to the 1.6 billion base pairs of the zebrafish genome. The compact pufferfish genome is mostly due to the reduction in the size of intronic and intergenic regions, while the structure of coding regions is well conserved with that of other teleost fishes and vertebrates in general. It is also important to note that transposable elements which constitute about 45% of the human genome represent only 3.8% of the pufferfish genome and 2.7% of *Takifugu* sp. (Crollius *et al.*, 2000; Dasilva *et al.*, 2002). Another factor that may, in part, explain the compact pufferfish genome is the resistance of the genome to retrotransposon-induced insertions, as proposed after a comparative study of diodonts and tetraodontid pufferfish (Neafsey and Palumbi, 2003).

A comparative analysis based on the distribution of duplicated genes in the pufferfish genome with their human orthologs provides evidence for a FSGD (Jaillon *et al.*, 2004). Based on these findings, Jaillon *et al.*, (2004), proposed an ancient karyotype of the teleost fish, consisting of 12 chromosomes, where they confirmed the occurrence of a possible FSGD event after their divergence from the ancestral tetrapod lineage.

These characteristics make the pufferfish a suitable reference genome for the study of comparative and evolutionary genomics. A complete dataset of pufferfish genome is available at the National Center for Biotechnology Research (NCBI) database. To date, no studies have described members of the iLBP multigene family in the pufferfish genome, with the exception of a recent report by Parmar and Wright (*in press*) on *fabp7a*, *fabp7b* and *fabp10* of pufferfish. Here, I report all the other uncharacterized *fabp* genes of pufferfish.

1.4 Objectives of the study

The focus of this study was to answer the following questions:

1. Are the putative *fabps* identified in the pufferfish genome orthologs of their mammalian and teleost counterparts?
2. Based on their tissue distribution patterns, could I provide insight into the evolutionary processes that lead to the retention or loss of the duplicated pufferfish *fabp* genes?
3. How are the duplicated *fabp* genes retained in the pufferfish genome compared to duplicated *fabps* in other teleost genomes?

CHAPTER 2

Materials and Methods

2.1 Bioinformatics and computational study

2.1.1 Identification and structural elucidation of pufferfish *fabp* genes

The putative gene sequence of each pufferfish *fabp* gene was retrieved by performing separate BLAST (<http://useast.ensembl.org/Multi/blastview>) searches with corresponding orthologous Fabp protein sequence of zebrafish or medaka as a query (Table 1) to the Ensembl genome sequence database (www.ensembl.org). To determine the gene structure the nucleotide sequence database of NCBI was searched for the longest cDNA sequence encoded by each pufferfish *fabp* gene by performing a BLAST using the Ensembl-derived genomic sequence as a query. The location of a putative TATA box, exons and introns splice junctions, presence of polyadenylation signals were determined for all the *fabp* genes of pufferfish based on their location on other orthologous *fabp* genes in zebrafish (Alves-Costa *et al.*, 2008; Denovan-Wright *et al.*, 2000a, 2000b; Karanth *et al.*, 2008; Karanth *et al.*, 2009; Liu *et al.*, 2003a, 2003b, 2004, 2007; Sharma *et al.*, 2004, 2006; Venkatachalam *et al.*, 2009), medaka and three-spined stickleback (Parmar *et al.*, 2012a, 2012b; Parmar and Wright, 2013a, 2013b).

2.1.2 Multiple sequence alignment

Sequence alignment was performed using CLUSTALW (Thompson *et al.*, 1994) application available at BioEdit version 7.2.0 software (<http://www.mbio.ncsu.edu/bioedit/page2.html>). The protein sequence of each pufferfish Fabp retrieved from Ensembl database was aligned with Fabp protein

Table 1: Fatty acid-binding protein NCBI query sequence IDs of zebrafish used for the identification of pufferfish *fabp* genes

Pufferfish <i>fabps</i>	Query protein	Query sequence ID
<i>fabp1</i>	Fabp1	NP_001019822
<i>fabp2</i>	Fabp2	NP_571506
<i>fabp3</i>	Fabp3	NP_694493
<i>fabp6a</i>	Fabp6	NP_001002076
<i>fabp6b</i>	Fabp6	NP_001002076
<i>fabp7a</i>	Fabp7a	NP_571680
<i>fabp7b</i>	Fabp7b	NP_999979
<i>fabp10</i>	Fabp10a	NP_694492
<i>fabp11a</i>	Fabp11a	NP_001004682
<i>fabp11b</i>	Fabp11b	NP_001018394

sequences from human, rat, mouse, zebrafish, medaka and three-spined stickleback. The percentage of sequence similarity and identity was determined using the pairwise alignment of BLOSUM62 matrix (Gromiha, 2010).

2.1.3 Phylogenetic analysis and conserved gene synteny

The phylogenetic relationship of the pufferfish Fabp protein sequences were determined using MEGA5 (Tamura *et al.*, 2011), a computational genetic analysis tool. A bootstrapped neighbor-joining was constructed with Fabp/FABP sequences of human, rat, mouse, zebrafish, medaka and three-spined stickleback using the Poisson correction distance model based on amino acid substitution per site. A maximum-likelihood tree was constructed using WAG substitution model based on gamma distribution for rate heterogeneity.

Conserved gene synteny was determined based on some regions of the ancestral chromosome that remain unaltered in the course of evolution (Woods *et al.*, 2005). Genes that are present along with the specific pufferfish *fabp* genes in specific chromosome were identified in medaka, zebrafish, three-spined stickleback and human genome and the conserved gene synteny was determined manually using the region overview option of Ensembl database to locate the co-localized genes.

2.2 Husbandry of fish (*Tetraodon nigroviridis*)

Adult pufferfish were brought from a commercial supplier, Pets Unlimited, Halifax, Nova Scotia, Canada. All the animal care conditions and experimental protocols were reviewed by the Animal Care Committee of Dalhousie University, in accordance with the recommendations of Canadian Council on Animal Care.

2.3 Tissue dissection and RNA isolation

Fish were anesthetized using 0.2% (w/v) Tricaine methanesulfonate (FinquelMS-222, Argent Chemical Laboratories, Redmond, WA, USA). Liver, intestine, muscle, brain, heart, eye, swim bladder, gills, kidney, skin, ovary and testis were dissected from three male and three female fish. Tissues were immediately homogenized in TRIzol reagent (Invitrogen, Carlsbad, California, USA) and total RNA was isolated from each tissue based on the protocol suggested by the supplier. The concentration of the isolated RNA was measured using a spectrophotometer. The quality of the RNA was determined by calculating the ratio of absorbance at 260 and 280 nm (A_{260}/A_{280}), where the ratio of pure RNA is 2. About 2 μ g of total RNA with A_{260}/A_{280} ratio of greater than 1.8 was used for conversion of mRNA to cDNA.

2.4 Reverse transcription (RT), primer optimization and polymerase chain reaction (PCR)

Complimentary DNA (cDNA) was synthesized using the Omniscript Reverse Transcription Kit (Qiagen, Mississauga, Canada). For primer optimization, cDNA derived from a whole pufferfish total RNA was used. A pufferfish was anesthetized, frozen in liquid nitrogen, homogenized and RNA was isolated as described above. All the primers used in the study (Table 2) were designed using NCBI primer-BLAST (<http://www.ncbi.nlm.nih.gov/tools/primer-blast/>). To avoid amplification of products from contaminating genomic DNA or heterogeneous nuclear RNA, either the forward or the reverse primer was designed to span an exon-exon junction. Primers were purchased from Integrated DNA Technologies Inc. (Coralville, Iowa, USA) and were prepared according to the manufacturer's instructions to a final stock concentration of 100 μ M.

Table 2: Primers used for quantitative PCR (qPCR) amplification of pufferfish *fabp* genes and normalization genes, and primer sets used for sequencing cDNA.

Gene	Forward primer	Reverse primer	Tm (°C)*
RT-qPCR primers			
<i>fabp1</i>	AGGCCATTGGTGTTCCTG	TGTCCTCCAGACGAAAGACC	56.2
<i>fabp2</i>	CAACATGGTCAAGAGGAAGCTGG	GCTCCACGAACCCGATAGCTC	59.5
<i>fabp3</i>	GGAACCTCGTCAAAGCGA	CGGTAACAACGGACTTAACAT	54.7
<i>fabp6a</i>	TACGACGACTTCTGCAAAC TGC	CATGAAGACGGTGGCCTGAAAC	60.3
<i>fabp6b</i>	TCGTGATGGACGATGGGAGG	GAAGGTCACACCCTCTCTCC	56.8
<i>fabp11a</i>	TGAAAGCAATTGGTGTGGGT	CGAACGGCTCGTTGAGTTG	56.2
<i>fabp11b</i>	AAGACGATCGGTGTGAAC TTTGC	GTCATCCGACCATCTGCTGT	57.4
<i>rpl13a</i>	CCCTACGACAAGAGGAAGCG	GGTACTTCCAGCCAACCTCA	58.6
<i>rpl7</i>	CGAGAAAAAGGCCGCAAGG	GCTTGACGAACACTCCGTTG	61.2
Sequencing primers			
<i>fabp1 (genomic)</i>	GCTTGCAACGCCAGAGTG	GGCCAGTAGTGGTGGTGATT	57.0
<i>fabp2</i>	CAGCACTCGGGAGATGAGAT	TTGTAGCTCTGGACGAGTTC	55.2
<i>fabp3</i>	GGAACCTCGTCAAAGCGA	GAGCGTGAGCGTCAGAGTGA	57.3
<i>fabp6a</i>	GAGACCCAGGAGGGTTACGAC	GTTCTCTTCAGCACCACTGAGC	58.5
<i>fabp6b</i>	GAGTTCTGGATGCCATCG	GTGCAATT CATGATCAGCTG	54.0
<i>fabp11a</i>	TGAAAGCAATTGGTGTGGGT	GAGGAGACCTCCTCACACCC	57.3
<i>fabp11b</i>	AGGAAGTCTGCGTGTGAG	TGGCATGATGGAAGACTGGA	56.6

*Annealing temperature

For primer optimization, PCR was performed with cDNA prepared from RNA of a whole fish using the Platinum PCR Super mix (Invitrogen, Carlsbad, California, USA). RT-PCR reactions contained 0.4 μ M forward and reverse primers and 100 ng cDNA template in a final volume of 25 μ l. Optimization of annealing temperature was performed by using six different temperatures for each of *fabp* gene primers (data not shown). The RT-PCR conditions consisted of an initial denaturation at 94.0°C for 2 min, followed by denaturation at 94.0 °C for 30 sec and 35 cycles, annealing of primers at an optimum temperature (differs for each *fabp* gene) for 30 sec, extension at 72.0 °C for 1 min and a final extension at 72.0 °C for 5 min. The amplified PCR products were visualized on a 1% agarose gel electrophoresis to confirm the expected product size. All the RT-PCR products were sequenced by Macrogen corp., (Rockville, MD, USA) to confirm the product size and sequence. The gene structure of each pufferfish *fabps* were confirmed by comparing the sequence of the PCR product with the gene sequence data available at Ensembl and NCBI.

2.5 Quantitative PCR (RT-qPCR)

To determine the tissue distribution and steady-state mRNA levels of pufferfish *fabp* genes, quantitative PCR was performed. The PCR product mentioned above was subjected to a copy number calculation, where the spectrophotometric measurement of copy number of the double stranded DNA was determined in each tissue as outlined by Bustin *et al.*, 2005. A rotor-gene (RG-3000) thermal cycler system (Corbett Research, Sydney, NSW, Australia) and the QuantiTech SYBR Green PCR kit (Qiagen, Mississauga, Canada) were used for the qPCR analysis. The qPCR was run at a final volume of 20 μ l with 100 μ M each of forward primer, reverse primer and 100 ng cDNA

template. The qPCR conditions consisted of an initial denaturation for 15 min at 95.0 °C, followed by 40 cycles of denaturation at 95.0 °C for 20 sec, primer annealing at optimum temperature specific for each *fabp* gene (Table 2) for 30 sec and elongation at 72.0 °C for 30 sec.

Initially, standards were prepared by serial dilution of PCR product amplified from cDNA of a whole pufferfish. A qPCR was later performed for each *fabp* gene of pufferfish with the standards prepared as template. An appropriate standard curve was constructed with amplification efficiencies ranging from 90 to 100 %. To perform an absolute quantification analysis of a particular *fabp* gene transcript, the selected standards were amplified along with the unknown cDNA of the above mentioned tissue samples for each *fabp* gene. Also, every qPCR run included a negative control, in which the cDNA template was omitted. The steady-state levels of two reference genes, *rpl13α* and *rpl7*, which are constitutively expressed in all the tissues, were also determined. The ratio of copy number of the each pufferfish *fabp* transcript and copy number of the each reference gene transcript was calculated to determine the mean normalized relative copy number for each pufferfish *fabp* gene.

CHAPTER 3

Results and Discussion

3.1 Identification of pufferfish *fabp* genes and their gene structures

By searching the *Tetraodon nigroviridis* genome sequence in the Ensembl database using the orthologous Fabp sequences of zebrafish, medaka and three-spined stickleback as BLAST queries, the *fabp* genes of pufferfish were identified. A BLAST search of the NCBI database was performed with the same query sequences to identify *fabp* cDNA nucleotide sequences (Table 1) of pufferfish. By this approach, I identified *fabp1*, *fabp2*, *fabp3*, *fabp6a*, *fabp6b*, *fabp7a*, *fabp7b*, *fabp10*, *fabp11a* and *fabp11b* (Figure 1) in the genome of the pufferfish. The organization of introns and exons was determined in each *fabp* gene based on sequence alignments with the longest cDNA sequence identified in the NCBI database. All the pufferfish *fabp* genes have a conserved gene structure of four exons separated by three introns, and each gene codes for a polypeptide of 125-135 amino acid residues (Table 3). The alignment between the genomic sequence of the pufferfish *fabp1* and a cDNA sequence revealed many undetermined nucleotides in the genomic and cDNA sequence of pufferfish *fabp1*, therefore, a DNA fragment containing exon 1 and parts of intron 1 was PCR amplified and sequenced to clarify the sequence and exon-intron organization of pufferfish *fabp1* (supplementary material). Hence the putative transcription start site could not be located for *fabp1*. A putative TATA box, 3' untranslated region, transcription initiation site, polyadenylation signal and the exon-intron junction, which conforms to the GT/AG rule (Breathnach and Chambon, 1981), were identified in each pufferfish *fabp* gene, except for *fabp6b*, where a putative TATA box could not be found.

Figure 1: Nucleotide sequence of pufferfish *fabp* genes.

A) *fabp1*, **B)** *fabp2*, **C)** *fabp3*, **D)** *fabp6a*, **E)** *fabp6b*, **F)** *fabp7a* [reproduced from Parmar and Wright, *in press*], **G)** *fabp7b* [reproduced from Parmar and Wright, *in press*] **H)** *fabp10* [reproduced from Parmar and Wright, *in press*], **I)** *fabp11a*, **J)** *fabp 11b* were identified. Exons are shown in capital letters, where the coding region is in bold font. The corresponding amino acid sequences are indicated below the exons. Introns are represented in lowercase letters. The number scale on the right indicates the position of the nucleotide and +1 indicates the putative transcription initiation site. A putative TATA box in the 5' upstream region of each gene is indicated by bold letter and underlining. A putative polyadenylation signal is shown in bold letter and underlining in the 3' untranslated region of each gene. Primers used for RT-qPCR analysis are highlighted in green and yellow corresponding to the forward and the reverse primers, respectively. Primers used to confirm the sequence structure for *fabp1* is highlighted in blue and pink corresponding to the forward and reverse primers, respectively.

Figure1A

fabp1

Figure 1B

fabp2

5' ccgatcctcgccggcgacccttggagattgagaggtctgataggcttctaaag	-74
gccacatcagcactcgggagatgagataaagaggtggctgagc <u>TTTAAA</u> GCAGCCGCG	-16
+1	
gcgggagagggagacACTCCTCCCTGCAGTCGTCCCGTCGGCCCTCTCGGAACC CATGACT	45
M T	
TTCAACGGAACCTGGAAAGTTGATCGCAATGACAACATATGAGAAGTTCATGGAACAGATG	105
F N G T W K V D R N D N Y E K F M E Q M	
Ggtgagtgactggacggctgcagacggcgaggcccattgcagcagctgcattgcagcg	165
tgaaggctctcagccgtctcctggagctccgcggacactgcatttccttcacccatcagttccctct	225
ttccttctctgtcag <u>GAATCAACATGGTCAAGAGGAAGCTGGCTGCTCACGACAAC</u>	285
G I N M V K R K L A A H D N	
CTGAAAATAACCATCCAACAGGATGGAAACACACCTTCCACATCAAGGAGAGCAGCAATTTC	345
L K I T I Q Q D G N T F H I K E S S N F	
CGAACCTGGAAATGGACTTCACCCCTGGCGTCACCTTCGACTACAGCCTGGCAGACGGA	405
R N L E M D F T L G V T F D Y S L A D G	
ACAGAGCTATCG gtgagtaaacgtggccggaaacctgaaaccgcctgtggctcgagtc	465
T E L S	
ggaagcggttaccaccatgagcaagcgctcgcttgccgtcgac <u>GGTCATGGAGCAGTC</u>	525
G S W S M	
AGGGGGAGACCATGAAGGGGGTTCGTCCGGAAAGGACAACGGAAAACAAC TGACAACCA	585
E G E T M K G V F V R K D N G K Q L T T	
TCAGAACTCTCGAAGGCGCGA ACTCGTCCAG gtaaacgcacagtcatatgcagg	645
I R T L E G G E L V Q	
aatccggctgtgaacgtgcacgttttctactaaaatacagaaactgcagcgtgaggc	705
gcggatgtacacacaccactctggccttctgtcctcag AGCTACAAC TACGAAGGCGTGG	765
S Y N Y E G V D	
CGCTAAGAGGATTTCAAGAGGAGCTAGAGCCAAGCGGAGCCTGGAGCGCCCCCCTGGC	825
A K R I F K R S *	
CCCTCGTTACTGCCTGTACAGTCAGCACACCCCTTTTCATAAAGC <u>ATAAAA</u> GCCCTTG	885
TCAAAGCGTG 3'	895

Figure 1C

fabp3

5' ccgatcctcgccccggacccttggagattgagaggtctgataggcttctaaag	-74
gccacatcagcactcgggagatgagataaagaggtggctgagc	-16
+1	
gcgggagagggagacACTCCTCCCTGCAGTCGTCCGTCGGCCCTCTCGGAACC	45
ATGACT	
M T	
TTCAACGGAACCTGGAAAGTTGATCGCAATGACAACATATGAGAAGTTCATGGAACAGATG	105
F N G T W K V D R N D N Y E K F M E Q M	
Ggtgagtgactgggacggctgcagacgggcgaggccatgcagcagctgcac	165
tgcgttcagccgtctcggagctccgcggacacctgcac	225
ttccttctctgtcag	285
GAATCAACATGGTCAAGAGGAAGCTGGCTGCTCACGACAAC	
G I N M V K R K L A A H D N	
CTGAAAATAACCATCCAACAGGATGGAAACACCTTCCACATCAAGGAGAGCAGCAATT	345
L K I T I Q Q D G N T F H I K E S S N F	
CGCAACCTGGAAATGGACTTCACCCCTGGCGTCACCTTCGACTACAGCCTGGCAGACGGA	405
R N L E M D F T L G V T F D Y S L A D G	
ACAGAGCTATCG	465
gtgagtaaacgtggccggaaacctgaaaccgcctgtggcagtc	
T E L S	
ggaagcgttaccaccatgagcaagcgtcgcttgtggcgctgcag	525
GGTTCATGGAGCATGG	
G S W S M	
AGGGGGAGACCATGAAGGGGTGTTCGTCCGGAAGGACAACGGAAAACAAC	585
T G C A A C T G A A G G C G G C G A A C T C G T C A G	
I R T L E G G E L V Q	645
aatccggctgtgaacgtgcacgttttcatataaaaatacagaaactgcagcgtgagg	705
gcggatgttaaacacaccactctggccttctgtcctcag	765
AGCTACAAC	
CGCTAACAGAGGATTTCAAGAGGAGCTAGAGCCAAGCGGAGCCTGGAGCGCCGCCTGGC	825
A K R I F K R S *	
CCCTCGTTACTGCCTGTACAGTCAGCACACCCCTTTCATAAAGC	885
ATAAAA	
TCAAAGCGTG 3'	895

Figure 1D

fabp6a

+1				
5' g	TATATAA	gccaggcgagcagcgatctcca	CCTGCCTCAGCATCCCTGCTCTCGC	26
		ACCACCACCACCAACCACCACCAACCACC	ATGGCCTTCGCTGGAAGATGGCTAACTGAG	86
		M A F A G R W L T E		
	ACCCAGGAGGGT	TACGACGACTCGGCAAAC	TGCTTTgtgagaaggcacagcccgtcctc	146
	T Q E G Y D D F G K L L	ttcctcacatatgccgaacagccggggcttctccgagcgtgcttcctgc	206	
		cgcagGGATCCCCGCTGACATCATCGAGAAAGGCCGACTACAAGATGATCACGGAGGT	266	
	W I P A D I I E K G R D Y K M I T E V	GACCCAGGACGGAGACACCTTCTCCTGGACCCAGGTCTACCCCACAGACGCCAAGGTCAC	326	
	T Q D G D T F S W T Q V Y P T D A K V T	CAACACCTTCACCGTCGGCAAGGAAAGCGACATGGAGACCATCGGAGGGAAAGAAGTTCAA	386	
	N T F T V G K E S D M E T I G G K K F K	Ggtgagcggcagagtggagccgcctgtgagtcggtgacggaggccaggacgccagcagag	446	
	taacagccgtgtttcagGCCACCGTCTTCATGGAAAGGAGGCAGCTGAGCGTGACCTTC		506	
	A T V F M E G G K L S V T F			
	CCCAACTACCACACACCTCTGAGATCAGCGGAGGC	AAACTCATCGAGgtaccaggcag	566	
	P N Y H H T S E I S G G K L I E	agccggaggtcatcgccccgcagatctgatcctgcatcagctttcaagcagaaaaatg	626	
	tgattttgaatgattccgtttctggtgtctccgcagACGTCTAAAGCCGGCTCAGT		686	
	T S K A G S G			
	GGTGCTGAAGAGAACACAAGCAAGAAGCTGTAA	ATGCTGGATGCCCTTGACCCCTGCACAA	746	
	V L K R T S K K L *			
	CCCAAGCTGCAGACATAAATGACAAGAACACGAA	3'	782	

Figure 1E

fabp6b

5' ttccagggtgagagcgagcaaacctggaaacagcagccggcggtcgatgcggaca	-99
tcgacgcctcattctgcgcaacagaaggcagcggatgtggcgctcagcctgagcca	-51
+1	
ctccttcttcttgcaaaaagtgcgttcctgtctcgttctctGTTCTTCCACC ATGA	10
M	
CTTCACTGGAAATACGAGCTGGAGACTCAGGAAA A CTACTGTGAGTTCTGGATGCCA	70
T F T G K Y E L E T Q E N Y C E F L D A	
TCGgtacacacacacttcttctcatacgatgtcacaaaaggatggatgtgatttcct	130
I	
ttcgctgcgcag GACTTCTCAGCGCCAAGACGGACCACAAAGTGATAACGGAGGTGAGC	190
G L L S A K T D H K V I T E V S	
CAGGACGGAAACGACTTCACCTGGACTCAAAGCATTCCCCACTGGACGTGGACAAACACC	250
Q D G N D F T W T Q S I P H W T W T N T	
TTCAGCGTGGTCGGGAATGTGAGCTGGTGACCATGAAGGGCACCAAGTTCAAGgtgagc	310
F S V G R E C E L V T M K G T K F K	
aggcggaaagaccagccagcatccctgatgtctggaggcttccatccctttccgtgcag	370
tcgggggtgaacgtgtctctgacccgcag GCGCTGTCGTGATGGACGATGGGAGGAT	430
A P V V M D D G R I	
CTCAGTCCAGTTCCCCAGTACCATTACAGCCGAGGTCAAGCAGGGACAAGCTGATCAT	490
S V Q F P Q Y H F T A E V S G D K L I M	
G taaaagaaccagtggagcgtggatttagtgggtaacgtcacgtaaaa	550
gctcccctttctacttcgaaccctccag AATTGCACAATTCCAGGAGAGAAGGGTGTGA	610
N C T I P G E K G V	
CCTCAGGAGAACCA CAGCAGGAGGGTCTGAAGGCCAAGACTCAGCATGTCCTCATGGAAC	670
T F R R T S R R V *	
GCCGGGACATTGGGAGAGTAGAAAAGTTCGCCTTCGTTGGTTGTGATAGTTAATA	730
AAGATTAAAAGACCAACTGTGACCTTGAGCGGTTGTGA <u>AATAAA</u> ACGTCACCTGTGAC	790
AACTAGGAAA 3'	800

Figure 1F

fabp7a

5' gccaagaataacatgattaaatgttacgattaagagcaatgacaacacgtacgctgat	121
aagcaaacggcgaccactcgaacccaggaatggcgtaacggtcacccgtctccggc	61
cgaggcgctagccccgcccccgctggctg TATTA tctggggatggagacgcgggttcc	1
+1	
ACTGCGGGGAGACCCGAGGCTGCGCAGGGCTGCTGACCCGGAGCCGAAGCTGCTGCAGA	60
CATGGTCGACGCTTCTGCCACATGGAAGCTGGTCGACAGCCAGAACTTGATGACTA	120
M V D A F C A T W K L V D S Q N F D D Y	
CATGAAGGGCTAG gtgagcaagttcacctcttgaattatgttgtttagtcaatt	180
M K A L	
Gcatttttatggctgaaaatttagggtgacaaatttagtattgttctttgtttgtttg	240
ttttcttatagttagatttaatgttcatatgcataatggtaactgacactctg	300
ttttattttag GTGTTGGCTTGCAACAAGGCAAGTCGGCAATGTCACCAAACCAACTGT	360
G V G F A T R Q V G N V T K P T V	
AGCCATTAGCAAGGATGGAGACAAAGTGGTATAAAGACCATGAGCACCTCAGGAACAC	420
A I S K D G D K V V I K T M S T F R N T	
GGAGATCTCCGCCAACGCTGGGAGAGGAGGTTGATGAGACCACGCCCTGATGATCGACATGT	480
E I S A K L G E E F D E T T P D D R H V	
CAAA gtgagctgaataatgttagaaataaaatataactctgacattgtccatccgc	540
K	
ttaaaatggcgtgaaattctcggtgtgttcttgcag TCAACTTCTCTATGGAGGGAG	600
S T F S M E G	
ACAAGCTGATGCAGGTGCAGAAGTGGAACCGCAAGGAGACCAAATTGTCAGAGAAATCA	660
D K L M Q V Q K W N G K E T K F V R E I	
AGGATGGAAAGATGGTATG gtaagaagtgcaaaaaatggtgacagttgaagg	720
K D G K M V M	
atgctgttcaaaccatattatctcactatattacagtgtgcagtaacttatattccaa	780
atgaaaacaatctgtttgtgcacatgtacaacatttatcgccccctcttttg	840
acag ACTTTGACTTCAAGGGTCACGGCGTTCGCACGTATGAGAAAGCCTAA TCTCA	900
T L T F E G V T A V R T Y E K A *	
GCAAGTCATCCAGCACTCAACAATATCTCAACGTTGTTACAATATGCAGTGGATC	960
TTTTGTCAGATCTTTGAATTAAACGCTAAAGTGTGACTATGTGTGAACAAAATTGT	1020
TACAAAAA AATAAA GTGCTTCAGTAATGTTAAATGTG 3'	1058

Figure 1G

fabp7b

5'	aacgagctcatctagccgtctcccagccctctgcgggtcagtaggaccatgacccaat	121
	ttaggatccaataaaaacctgaggaaaaatctccctgagccaatgggacctatggctcg	61
	cccacccacctgcatactaattgaggaggaga <u>TAATAA</u> cacactggggggaaacttgagg	1
+1		
	GATCGCAGCTCGTCCAGCGTGCTCTCCAGATTACTCTGCTTCTAATCAACCGTCACC ATG	60
	M	
	GTCGACGCCTCTGTGCAACCTGGAAACTGGTGGAGAGCGAGAACCTTGATGATTACATG	120
V D A F C A T W K L V E S E N F D D Y M		
AAAGCGCTCG gtcagtgtgcttatgttgcgtttgatctgttgcgttttagtccga	180	
K A L		
gtgctgaaatgtgcttatttgcctatgtctccag	GCGTGGGTTTCGCCACCCGACAGGT	240
G V G F A T R Q V		
CGGTAACGTGACCAAACCGACTGTGATCATCAGCCAGGAGGGCGACAAGGTGGCGATCCG	300	
G N V T K P T V I I S Q E G D K V A I R		
CACACAGAGCACCTTCAAAAACACAGAGATCTCCTCAAGCTGGAGAGGGAGTTCGATGA	360	
T Q S T F K N T E I S F K L G E E F D E		
AACCACCGCCGACGACAGGA^{ACTGTAAA}gtatgacgaaacgtctcgatgttcccactaa	420	
T T A D D R N C K		
accctaattggaaatctgtttgttcaggatgaatttagtcggtagttcgctaaaggtcacc	480	
atgtgtcttgtctctcag	TCCACAGTGACTCTGGACGGAGACAAGTTGGTCCACGTCCAG	540
S T V T L D G D K L V H V Q		
AAGTGGGATGGCAAAGAGACCAAGTTGTCAAGAGAGATCAAGGATGGCAAACACTGGTCATG	600	
K W D G K E T K F V R E I K D G K L V M		
gtaggattcaattaaaaaggatctaaattatgacactgataaaattcggaaagcattaa	660	
aaatggaaatcttggaaatctggctgttttaaagtgcagattctataactctaataatgaagag	720	
atcattacctgattattatgatggggactccgctgtaaacaatgagctctaggtgtgatt	780	
tctaggcagcagaaaaatcaacaatggctctttcactctccctttcttcgt	840	
caacacacccctcagatattgtgtctgtccatagtaaccttgcgtctccctcgt	900	
cag	GACCTGACCTTGAAGACGTCCACGCAGTGCACCTACGAGAAGGCATGAAGGGCC	960
D L T F E D V H A V R T Y E K A *		
CGACCTTGACTCCAGGAGGTTCTGCAGGCTGCACGTTTGTA TTTATACAAACTGTTCTGTATGATGCATCGTCTTGCAGCCCCGCGTGAACG ATTAAA	1020	
AAAGTGAACAGGCTAA 3'	1080	
	1097	

Figure 1H

fabp10

5' tgacaaatatcaccttacacttggtcgaaggcaccattggccaaacgggagttacgtg	121
aacctgaggtgttcaaacaacagtgattacctcactggatctattactcattagacag	61
agagctggggccccgacctgtcctc TATAAGAA ccaggaccctctgatcacatcctc	1
+1	
ACATTGTGTTGAGCTTCACACAGCTGTACAGCCTCCAGCCCATCTGGTTAAGGAGAAG	60
CCAGATCTAAGAAC ATGGACTTCAACGGGACATGGCAGGTTATGCCAGGAGAACTAC	120
M D F N G T W Q V Y A Q E N Y	
GAGGAGTTCCCTCAAGGCCATGGgtgagaagtaacaccaataccaggctgtcctatTTTA	180
E E F L K A M	
gttttctcaattaaataaacgtgaacatgtttcttgcatttaccag AACTGCCCGAGGATGTC	240
E L P E D V	
ATCAAGATGCCAAAGACATTAAGCCGATTACTGAGATCAAGCAAATGGCAAAGACTTT	300
I K M A K D I K P I T E I K Q N G K D F	
GTTGTTACCTCCAAAACGCCCGGGAAAGACTTGTACCAACTCCTTACCATGGCAAAGAG	360
V V T S K T P G K T L T N S F T I G K E	
GCTGATATAACCACCATGGACGGCAAGAAGCTCAAGgttcgctggcccccctccgggtgat	420
A D I T T M D G K K L K	
tctgcttggtcacgttctcatTTTgatacgatcccttgcgttcatTTTcacaacac	480
acag TGCACGTCACTATGGAGGGAGCAAACGGTCTGCAAACCGGAGAAGTTCTGTCA	540
C T V T M E G S K L V C K T E K F C H	
CACACAGGAGCTCAAAGGGGAGAGATGATTGAGgtacggatTTTatTTTaa	600
T Q E L K G G E M I E	
gatactggttcggtgaaatgttttggaaagaccacacagacggacgctgacaatcgactg	660
attcattttcag ACTTTGACCAAGGGCTCAACAACCTTGTCAAGAAAGAGTAAGAAGATG	720
T L T K G S T T F V R K S K K M	
AACTAA GACAAAGGAAAACAAGGTTGAATAAAATAAAAGTGTTCGGACAGGCG	780
N *	
TGGTTGAGTTAATTCTTACAGTTCACCTATCGGTCTGGCTAAAGGTCAAAGCTGCGA	840
GGACGAGGCCGTGCCGATGCTATTGCCTAAGGCTATCAATAAAATGATGAGCTGTAATT	900
TCAACTGAGGAAAAAAACACAGACAAGAAGTGGGGAAAGGGAGTTATTTT	960
CTTTTTTTAAATTAGACATGAATTAGAGTCATTGCCTTTTTT AATAAA AAC	1020
ACCAATTAAACAAATCTTGAAA 3'	1043

Figure 1I

fabp11a

+1		
5' tgggcgcgaggcccgccgtt	TAAAA ggcagcaggttcccacagcacagAGACCGCGTC	10
CACGCACTCCCCACGGGCTGACCAGGGTCACGCCGGCTGCAAAC	ATGGTTGAGAAG	70
M V E K		
TTTGTGGGACGTGGAAAATGATTCCAGCGAGAACCTTGACGATTACT	TGAAAGCAATT	130
F V G T W K M I S S E N F D D Y L K A I		
Gtacgagagcaaaaagagggataacgggatgagtagtatcactgaggcaacagtgattata		190
gtaccacagtagaaaaacgcatttagaaacatttctgcttcctgtatgattgttattag		250
atttatccatgcttaccttattctgcag	GTGTTGGTTCCGCACCCGGCAGGTGGGCA	310
G V G F A T R Q V G		
ATCGGACCAAGCCAATCTGGTGGTACCGTGGACGACCAGGGACGGTGTGCATGAGGA		370
N R T K P N L V F T V D D Q G T V C M R		
CTCAGAGCACCTTCAAAAACACCGAGATCAAGTT	CAAACCTCAACGAGCCGTTCGACGAGA	430
T Q S T F K N T E I K F K L N E P F D E		
CCACCGCCGACGACAGGAAGACGAGG	gttagtgccccccgtgaacgcatcacggccg	490
T T A D D R K T R	cgcggcggttggcgccggctctcacggctttctgtcccccg	550
	cag	
	ACCGTGGTGAC	
	T V V T	
TCTGGAGAACGGCAAGCTTGTGCAGAACAGAACTGGGACGGCAAAGAAACCTGCATCGA		610
L E N G K L V Q K Q N W D G K E T C I E		
AAGGGAGATCTGGACGGGAAGTTGATAGCG	gtgagacgttccctgtttccactttta	670
R E I S D G K L I A		
ttgagtctagttgtcgccgggttttagtggctttaactcattaatgcagtctgcga		730
atgttttgacagaaaaacatgaaactgaacccctaaatgctaattaacagcagc		790
agcagttattaccgcttacgggtgaaatccgcataatttgcattgtattgatcgatca		850
gtctgttttatctgttttcttcccccccg	cagAAATGCATCATGGGAGATGTGGTT	910
K C I M G D V V		
GCTGTCAGGACATATGTGAGAGAGGCATGA	GGAGGGTGTGAGGAAGGTCTCCTCTGGAA	970
A V R T Y V R E A *		
CAGGCTCAGTTCAATGAGCGTCGGATACCAAACACTGTTCTTCTGTCTCCTCTTTCTT	1030	
TACTGTGTTCTACAGCTGCAGCCTGTACTCCAGACCCCACACCAGCAACCTCTCCTT	1090	
TTTTTGATGATTGG AATAAAA ATGAATTATCCACATGG 3'		1130

Figure 1J

fabp11b

Table 3: Fatty acid-binding protein genes of pufferfish with their corresponding Ensembl gene ID, GenBank accession numbers of their cDNA sequences, location and number of amino acid.

Gene	Ensembl ID	GenBank ID	Location	Number of amino acids
<i>fabp1</i>	ENSTNIG00000016680	CR720131.2	Chromosome 12: 9,154,751-9,155,415	127
<i>fabp2</i>	ENSTNIG00000012360	CR734529.2	unassigned_random: 14,883,076-14,884,115	132
<i>fabp3</i>	ENSTNIG00000016979	CR637284.2	Chromosome 21: 1,218,101-1,220,308	133
<i>fabp6a</i>	ENSTNIG00000017612	CR733593.2	Unassigned_random: 20,198,312-20,199,077	127
<i>fabp6b</i>	ENSTNIG00000004652	CR699217.2	Chromosome 1: 1,118,490-1,119,289	125
<i>fabp7a</i>	ENSTNIG00000017217	CR705358.2	Chromosome 10: 8,694,895-8,695,952	132
<i>fabp7b</i>	ENSTNIG00000003393	CR683239.2	Chromosome 14: 9,433,632-9,434,728	132
<i>fabp10</i>	ENSTNIG00000002424	CR639897.2	Unassigned_random: 52,169,316-52,170,358	127
<i>fabp11a</i>	ENSTNIG00000017781	CR733066.2	Chromosome 21: 1,566,398-1,567,527	134
<i>fabp11b</i>	ENSTNIG00000018400	CR671793.2	Chromosome 8: 7,093,697-7,094,465	134

However, all *Fabp* transcripts were detected in the RT-PCR assay, using primers specific for each pufferfish *fabp* gene indicating that all the *fabp* genes identified are functional and the presence of a possible TATA-less promoter in the *fabp6b*.

3.2 The sequence identities and similarities of pufferfish *fabp* genes with teleost fish and human *fabp/FABP* genes

The sequence identity and similarity showed that pufferfish Fabps and other Fabps from other vertebrates shared 45-95% sequence identity (Figure 2). A default cut-off <40% identity was set while aligning orthologous sequences (Thompson *et al.*, 1994). Meanwhile, the percentage identity decreased significantly (15%-40%) when aligned with other paralogs of pufferfish Fabps. However, the pufferfish fish Fabps shared 50-95% sequence identity with other teleost fish Fabps, suggesting that the gene coding for these proteins are orthologous and most likely arose from a common ancestral gene.

To infer the evolutionary relationship between the pufferfish Fabps and Fabps from other vertebrates, neighbor-joining and maximum-likelihood phylogenetic trees were constructed using MEGA5 software (Figure 3). Human lipocalin 1 (LCN1) protein was used as a designated outgroup. Both trees generated identical topologies indicating similar phylogenetic relationship, where each of the ten Fabps of pufferfish clustered in distinct clades with their orthologs of other teleost and tetrapod Fabps. Based on the robust bootstrap values >80%, the *fabp* genes of pufferfish appear to be orthologs of other teleost and tetrapod *fabp* genes.

Figure 2: Amino acid sequence alignments of various orthologous and paralogous sequences showing percentage identity and similarity to pufferfish Fabp.

Amino acid sequence of pufferfish (Pf) **A**) Fabp1, **B**) Fabp2, **C**) Fabp3, **D**) Fabp6a, **E**) Fabp6b, **F**) Fabp11a and **G**) Fabp11b aligned with Fabp/FABP of zebrafish (Zf), Medaka (Me), three-spined stickleback (St), fugu (Fu), human (Hu), mouse (Mo) and rat (Rt). The percentage of sequence identity and similarity are show at the end of each sequence. The dots in the alignment represent sequence identity, whereas the dashes represent missing amino acids.

Figure 2A

Fabp1

	10	20	30	40	50	60
Pf Fabp1	M--SLSGKYQQVSQENFEPFMKAIGVPDEIIQQIKDIMSSTEIVQ-NGNDFKITTTTGPK					
St Fabp1	.---FT.....LET.....A.....V.L..DL..KG...K.IS..EE-..DN..V.V...S.					
Me Fabp1	.---FT.....LE.S..Y.A....V.IS..N..LT.GLK.I..MEE-..DN.RV.V...SH					
Hu FABP1	.---F.....LQ.....A.....L.E.L..KG...KGVS....-..KH..F.I.A.S.					
Mo FABP1	.---NF.....LQ.....L.EDL..KG...KGVS...H-E.KKI.L.I.Y...					
Rt FABP1	.---NF.....VQ.....M.L.EDL..KG...KGVS...H-E.KKV.L.I.Y.S.					
Zf Fabp1b.1	.---FT.....LE...G.VE....V.L..DM.EKG...K.VS..EE-...Q..V.V...S.					
Zf Fabp1a	.---AFT.....LE.H....A....V....DEVEKG...K.IS..H.-D.K...V.V.A.T.					
Zf Fabp1b.2	--PF...FELEYK.GW.E.....-HQ.LLKKAN.GKTQF..QE-....YCL..RS.G.					
Pf Fabp11a	.ADAFI.TWNLKESK..DEY..G...GFATR.VANMTKPT.I.SK-E.DV.TLK.VSTF.					
Pf Fabp3	.VDKFV.TWKMTTSD..DEY.....GFATR.VGNRTKPNLVVCVDEQGLICMKSQSTF.					
Zf Fabp2	.--TFN.TWKVDRN..Y.K..EQM..NMVKRKLAHDNLKITLE.-T.DK.NVKEVSTFR					
	70	80	90	100	110	120
Pf Fabp1	VTVNQGTIGKETEMDAISGEKIKTVFRLLED-NKLKVSLKN----IESVTELVDPNTLVAV					
St Fabp1	.L..TFK..Q.A.IET.T...V.A.VKRDG-..T.C..G---M.....L.K..I.NT					
Me Fabp1	.II.SF...Q....LSVT.....VKK.G-T..MM...R---ADI.....G....TT					
Hu FABP1	.IQ.EF.V.E.C.LETMT...V...VQ..GD...VTTF.....K.....-NGDIITNT					
Mo FABP1	.VR.EF.L.E.C.LETMT...V.A.VK..GD..MVTTF.G----K.....-NGD.ITNT					
Rt FABP1	.IH.EF.L.E.C.LETMT...V.A.VKM.GD..MVTTF.G----K.....F-NGD.ITNT					
Zf Fabp1b.1	.LT.SF...Q.ADIETLT...V...VHKDG-Q...AA.G----.TLTW..L.E...LIT					
Zf Fabp1a	.ILYSF.V.E.C.LETFT.DRA...VQMDG-...TAFV.G----...GD.ISNT					
Zf Fabp1b.2	.LN.SF...QD..IQML..D.V...VHKDG-Q...AA.G----.TLTW..L.E...LIT					
Pf Fabp11a	S.EINFKL.E.FDETTADDR.V.S.IT.DGGKL.H.QKWD--GKETTLRE.SD.N.TLT					
Pf Fabp3	T.EIKFKLNEPF.ETTADDR.TT..MTI.NGKLVQKQTWD--GKESTIERE.SDGK.I.K					
Zf Fabp2	TLEINF.L.VTFDYSLAD.TELTGSWVI.GDTLKGTFRKDNGKVLTVRTIVNGE..QS					
	130	Identity (%)		Similarity (%)		
Pf Fabp1	MTLGD-IVYKSTS KRV-	100		100		
St Fabp1	I...S-....R....M-	60		77		
Me Fabp1-....T.Y..I-	58		74		
Hu FABP1-..F.RI...I-	58		74		
Mo FABP1-....RV...I-	55		73		
Rt FABP1-....RV...I-	51		72		
Zf Fabp1b.1	LSTA.N.A..RF....V	51		70		
Zf Fabp1a	LSFNG-....RI.R.IS	48		70		
Zf Fabp1b.2	LSTA.N.A..RF....V	48		63		
Pf Fabp11a	L.....STRHYVKAE	23		35		
Pf Fabp3	CKM..-V.AVR.YVKEA	22		38		
Zf Fabp2	YSYDG-VEA.RIF..A-	16		35		

Figure 2B

Fabp2

	10	20	30	40	50	60
Pf Fabp2	M--TFNGTWKVRNDNYEKFMEQMGINMVKRKLAHDNLKITIQQDGNTFHKESSN-FR					
St Fabp2a	.---D....I.....D.....				N..K.AV.....-	
Me Fabp2a	.----...SE.....V.V.....				E.T.DK..V.....	
Zf Fabp2	.---.....E.....V.....				LE.T.DK.NV..V.T..	
St Fabp2b	.---.....D.....V.IM....E.....V..E.T.DQ.Q.....T..					
Me Fabp2b	.--A.....A..SE..D..L.RI.V.VM.....E.....E.T.DN..V.....T..					
Hu FABP2	.--A.DS.....SE..D....K..V.I.....L..T.E..K.TV.....A..					
Mo FABP2	.--A.D.....E.....K...VM....G.....L..T...K.TV.....-					
Rt FABP2	.--A.D.....E.....K...V.....G.....L..T.E..K.TV.....-					
Zf Fabp6	.--A...K.ETESQEG..P.CKLI..PDDVIAKGRDFK.VTE.V.N.DD.TWTQYYPNHH					
Pf Fabp10	--D....Q.YAQE...E.LKA.ELPEDVI.M.KDIKPITE.K.N.KD.VVTSKTP-GK					
Pf Fabp11a	.VEK.V....MISSE.FDDY.KAI.VGFAT.QVGNRTKPNLVFTV.DQGTVCMMRTQST.K					
	70	80	90	100	110	120
Pf Fabp2	NLEMDFTLGVTFDYSLADGTELGSWSMGESES MKGVFVRKDNGKQLTTIRTLEGELVQS					
St Fabp2a	...IN....D..E.....N...DML..I.N.....T.IVQRD..I..					
Me Fabp2a	.I.I.....E.....A.AL..DVL..T.T.....A...T.VIQ.D.MI..					
Zf Fabp2	T..IN.....T...VI..DTL..T.T.....V...V..IVN.....					
St Fabp2b	TKDI.....P.....V..T.E...DLL..K.T....S.V...T.A.V.....					
Me Fabp2b	TK.F.....K.....V..T.E...DML..L.N....N.L...T.SIVN.....T					
Hu FABP2	.I.VV.E.....N.N.....R.T..L..NKLI.K.K.T...NE.N.V.EII.D....T					
Mo FABP2	.IDVV.E....N.P.....T.A.TI..NKLI.K.T.V....E.IAV.EVS.N..I.T					
Rt FABP2	.IDVV.E....D.A.I.GNCCHFT.T.T...NKLV.K.K.V....E.IAV.EIS.N..I.T					
Zf Fabp6	VVTNK.IV.KES.METVG.KKFK.IV....GKLTIS.PK-----YQQTTEIS..K..ET					
Pf Fabp10	T.TNS..I.KEA.ITTM..KK.KCTVT...SKLVCKTEK-----FCHTQE.K...MIET					
Pf Fabp11a	.T.IK.K.NEP..ETT..DRKTRTVVTL.NGKLVQKQNWD--..ETCIE.EISD.K.IAK					
	130	140	Identity (%)	Similarity (%)		
Pf Fabp2	YNYEGVDAKRIFKRS----		100	100		
St Fabp2aKC----		80	92		
Me Fabp2a	...D.....G----		77	92		
Zf Fabp2	.S.D..E.....A----		76	91		
St Fabp2bKQ----		75	86		
Me Fabp2bKH----		69	85		
Hu FABP2	.V....E.....KD----		66	83		
Mo FABP2	.T....E....F..KE----		66	82		
Rt FABP2	.T....E.....KE----		62	78		
Zf Fabp6	STAS.AQGTAVLV.TSKKV-		22	36		
Pf Fabp10	LTK----GSTT.V.KSKKMN		22	43		
Pf Fabp11a	CIMGD.V.V.TYV.EA----		21	40		

Figure 2C

Fabp3

	10	20	30	40	50	60
Pf Fabp3	MAEAFA	GATWN	LVKSEKFDEYM	KELGVGLAMR	KMGNLAKPTLSI	TIEGDKVTLKNSSTF
Fu Fabp3V.IS.D.V.L.	..I.GM.	...STI.S.D.
Me Fabp3V.MKEA.F.T.V.T.TI.SMD.R.V.TQ.I.N
St Fabp3	KVN..V.	K.E.KE.V.	D...K...	F.T.Q...	T...TI.SV...	V..V.TQ.SL.N
Zf Fabp3	..D..I...	KE.KN...	GI...	F.T.QVA.MT...	TI.SK...	VF...TV....S
Rt FABP3	..D..V...	K..D.KN..D...	S...	F.T.QVASMT...	TI.EKN..TI.I.TH...	N
Hu FABP3	.VD..L...	K..D.KN..D...	S...	F.T.QVASMT...	TI.EKN..IL...	TH....N
Mo FABP3	..D..V...	K..D.KN..D...	S...	F.T.QVASMT...	TI.EKN..TI.I.TQ...	N
Zf Fabp2	--T.N...	KVDRN.NYEKF.EQM..NMVK..	LAAHDNLKITLEQT...	FNV.EV...R.		
Pf Fabp10	--D.N...	QVYAQ.NYE.FL.AMELPEDVI..	AKDI..ITE.KQN.KDFVVTSKTPG..			
Pf Fabp1	--SLS.KYQQ.	SQ.N.EPF..AI..	PDEIIQQIKDIMSFT	E.VQN.NDFKITTTGP.V		
	70	80	90	100	110	120
Pf Fabp3	TEVSFKLGE	EFDESTADGRNVKS	VVTVED--GKLHVQ	KWDGKETSLV	REVEG-NNLT	LT
Fu Fabp3	..I.....T.NIV.	---M..A....D...	T.....ND-KS...		
Me Fabp3	..LT.....	T...D.K...I..L...	-----			N.-.....
St Fabp3	..L....D...	T...E.K...F.K.S.D-	-----			S....-A....
Zf Fabp3	..IN.....	T...D.K...I.LDG--	...L.....	T.L...SD-		
Rt FABP3	..I..Q..V...	V...D.K.....LDG--Q..T.T..LSD-GK.I..			
Hu FABP3	..I.....V...	D.K...I..LDG--L....Q..T....LID-GK.I..			
Mo FABP3	..IN.Q..I....V...	D.K...L..LDG--	...I....N.Q..T.T..LVD-GK.I..			
Zf Fabp2	L.IN.T..VT..Y.L...	TELTGWSVI.GDTL.GTFTR.DN..VLTT..TIVN-GE.VQS				
Pf Fabp10	LTN..TI.K.A.IT.M..KKL.CT..M.G----	SKLVCKTEKFCHTQ.LK.-GEMIE.				
Pf Fabp1	.VNQGTI.K.TEMDAIS.EKI.T.FRL.....	NKL.VSL.NIES.T.LVDP.T.VAV				
	130	Identity (%)	Similarity (%)			
Pf Fabp3	LTLGNVVCTRHYEKVQ	100	100			
Fu Fabp3K.....AE	75	86			
Me Fabp3	..M.D.....R...AE	75	87			
St Fabp3	..MDD..S.....V.AE	67	78			
Zf Fabp3DI.S.....V.AE	66	82			
Rt FABP3	..H.....S...T...EA	62	74			
Hu FABP3	..H.TA.....T...EA	61	74			
Mo FABP3	..H.S...S...T...EA	58	74			
Zf Fabp2	YSYDG.EAK.IFKRA-	28	50			
Pf Fabp10	..K.STTFV.KSK.MN	26	45			
Pf Fabp1	M...DI.YKSTS	22	41			

Figure 1D

Fabp6a

	10	20	30	40	50	60
Pf Fabp6a	M--AFAGRWLTTETQEGYDDFCKLLGIPADIIEKGRDYKMITTEVTQDGDTFSWTQVYPTDA					
Fu Fabp6a	.----T...E..N.....E.....D.....N..L.....E.....I....N.					
St Fabp6a	.--...K.E..S.....A.....N.V.....L.....N...C.I...N.					
Me Fabp6a	.--T....EI.S.....E...VV.....V.....AV.....N..E.T.....-I					
Zf Fabp6	.--..N.K.E..S....EP....I...D.V.A....F.LV..IV.N..D.T...Y..NNH					
Hu FABP6	.--..T.KFEM.SEKN..E.M.....SS.V...A.NF.IV...Q...QD.T.S.H.SGGH					
Mo FABP6	.--..S.KYEF.SEKN..E.M.R..L.G.V..R..NF.I....Q...QD.T.S.S.SGGN					
Rt FABP6	.--..T.KYEF.SEKN..E.M.R..L.E.V..R..NF.I....Q...EN.T.S.S.SGGN					
Pf Fabp6b	.--..T.T.KYEL....N.EE.LDAI.TA.GLLSAKT.H.V....S...ND.T...SI.H-W					
Pf Fabp7a	.--..T.CAT.KLVDSQNF..YM.A...VGFATRQV.NVT.PTVAISK...KVVIKT-MS.FR					
Pf Fabp2	.--..T.N.T.KVDRNDN.EK.MEQM..NMVKRKLAAHDNLKITIQ...N..HIKE-SSNFR					
	70	80	90	100	110	120
Pf Fabp6a	KVTNTFTVGKESDMETIGGKKFKATVFMEGGKLSVTFPNYHHTSEIS-----GGKLIET					
Fu Fabp6aI...C.....HL.....-----					
St Fabp6a	R...N..I...C.....H.....TTA....Q..T.....					
Me Fabp6a	...K..I....E..SL.....NL....MT.N....TQ.N.....					
Zf Fabp6	V...K.I.....V.....GI.S.....TIS..K.QQ.T.....V..					
Hu FABP6	TM..K.....NIQ.M...T.....Q.....V.N.....Q....V-----D..V.V					
Mo FABP6	IMS.K..I...CE.Q.M.....K.....VVAE.....Q...VV-----D..V.I					
Rt FABP6	IMS.K..I...CE.Q.M.....K.....VVAD.....Q...VV-----D..V.I					
Pf Fabp6b	TW....S..R.CELV.MK.T....P.V.DD.RI...Q..Q..F.A.V.-----D...MN					
Pf Fabp7a	NTEISAKL.E.F.ET.PDDRHV.S.FS...D..MQVQKWNGKETKFVR--EIKD..MVM.					
Pf Fabp2	NLEMD..L.VTF.YSLAD.TELSGSWS...ESMKGV.VRKDNGKQLTTIRTLE..E.VQS					
	130	140	Identity (%)	Similarity (%)		
Pf Fabp6a	SKA---GSVVLKRTSKKL-	100	100			
Fu Fabp6aR..-	88	95			
St Fabp6a	.T.----A.....I-	81	89			
Me Fabp6a	C.-----RKV...I-	74	88			
Zf Fabp6	.T.SGAQ.TA..V.....V-	64	80			
Hu FABP6	.TI----.G.TYE.V..R.A	57	75			
Mo FABP6	.TI----.D.TYE.V..R.A	54	75			
Rt FABP6	.TI----.D.TYE.V..RVA	53	75			
Pf Fabp6b	CTIPG-EKG.TFR...RRV-	44	64			
Pf Fabp7a	LTF----EG.TAV..YE.A-	25	37			
Pf Fabp2	YNY----EG.DA..IF.RS-	21	37			

Figure 2E

Fabp6b

	10	20	30	40	50	60	
Pf Fabp6b	M--TFTGKYELETQENYEEFLDAIGT-AAGLLSAKTDHKVITEVSQDGNDFTWTQSIP-H						
Fu Fabp6b	.--.....E....QIT...G.....V.N.....T..-N						
St Fabp6b	.--A.A.....S.....A.-----.N.....L...SG.....-N						
Me Fabp6b	.--K.A.....S.S.....E.----D.VN...E...V...V.....-N						
Rt FABP6	.--A.....F.SEK..D..MKRL.-LPEDVIERGRNF.I....Q...EN...S..YSGG						
Mo FABP6	.--A.S....F.SEK..D..MKRL.-LPGDVIERGRNF.I....Q...Q...S..YSGG						
Pf Fabp6a	.--A.A.RWLT....G.DD.CKLL.-IP.DIIEKGR.Y.M....T...DT.S...VY.TD						
Hu FABP6	.--A....F.M.SEK..D..MKLL.-ISSDVIEKARNF.IV....Q...Q...S.HYSGG						
Zf Fabp6	.--A.N..W.T.S..G..P.CKL..-IPDDVIAKGR.F.LV..IV.N.D.....YY.NN						
Pf Fabp2	.---N.TWKVDRND...K.MEQM.-INMVKRKLAAHDLKITIQ....T.HIKE.SN-F						
Pf Fabp7b	.VDA.CATWK.VES..FDDYMK.L.-VGFATRQVGNVT.PTVII..E.DKVAIRTQST-F						
		70	80	90	100	110	120
Pf Fabp6b	WTWTNTFSVGRECELVTMKGTKFKA	PVVMDGRISVQFPQYHFTAEVSGDKLIMNCTIPG					
Fu Fabp6b	...S.....Q.....IQ.....R.EN.K.....N.....T..						
St Fabp6b	...S.K.T..QD...T....V..T.A.T.EG.K..IP.....LIE...V.I.LT..						
Me Fabp6b	.S...K.T..Q....L...NI....L.TLEN.KL.IP....L....IID...V.T.VT..						
Rt FABP6	NIMS.K.TI.K....MQ..G.K....T.K.EG.KVVAD..N..Q.S..V....VEIS...--						
Mo FABP6	NIMS.K.TI.K....MQ..G.K....T.K.EG.KVVAE..N..Q.S..V....VEIS...--						
Pf Fabp6a	AKV....T..K.SDME.IG.K....T.F.EG.KL..T..N..H.S.I..G...ETSKA--						
Hu FABP6	H.M..K.T..K.SNIQ..G.KT....T.Q.EG.KLV.N..N..Q.S.IV....VEVS...--						
Zf Fabp6	HVV..K.I..K.SDME.VG.K...GI.S.EG.KLTIS..K.QQ.T.I..G..VETS.AS.						
Pf Fabp2	RNLEMD.TL.VTFDYSRAD..ELSGSWS.EGESMKGV.VRKDNGKQLTTIRTLEGELVQ						
Pf Fabp7b	KNTEIS.KL.E.FDET.ADDRNC.ST.TL.GD--KLVHV.KWDGK.TKFVRE.KDGKLM						
		130	Identity (%)	Similarity (%)			
Pf Fabp6b	---EKGVTFRRRTSRRV-	100	100				
Fu Fabp6b	---.....K.I...L-	83	93				
St Fabp6b	---.S.....I-	71	82				
Me Fabp6b	---.....VNK.I-	65	82				
Rt FABP6	---GD..YE.V.K..A	46	64				
Mo FABP6	---GD..YE.V.K.LA	45	65				
Pf Fabp6a	----GS.VLK...KKL-	44	64				
Hu FABP6	----G...YE.V.K.LA	44	65				
Zf Fabp6	A--QGTAVLV...KK.-	41	61				
Pf Fabp2	SYNYE..DAK.IFK.S-	19	38				
Pf Fabp7b	DLTFED.HAV..YEKA-	18	36				

Figure 2F

Fabp11a

	10	20	30	40	50	60
Pf Fabp11a	MVEKFVGTWKMISSENFDYMKAIGVGFATRQVGNRTKPNLVFTVDDQGTVCMRTQSTFK					
Fu Fabp11a	VS.....	KS.....
Me Fabp11a	.A.....	VSIE....	IS.KS.....
St Fabp11aA	IV..E.D.V..	LKS.....
Zf Fabp11a	..D.....TT.D...E	VC..E..LI..KS
Pf Fabp11b	..Q.....TLA.....E..T..N.....M..LV.....	S.AED.FISLKAE
Me Fabp11b	..Q.A...TLA.....E....V.M.....M..I.....	IQ.GAD.LIS.KAE
St Fabp11b	..DQ.....TL.A.....E.....L.....M..LA.....	IG..EA.LIS.KSET
Zf Fabp11b	..QY..K..T..D..E....V.AS..S..MA.LA..S.LIA..E..VIT.KAVT
Pf Fabp7b	..DA.CA...LVE.....L.....V....TVIISQEGD-K.AI
Pf Fabp7a	..DA.CA...LVD.Q.....L.....V....TVAISK.GD-K.VIK.M...R
Pf Fabp2	--T.N....VDRND.YEKF.EQM.INMVK.KLAHDNLKITIQQ.GN-.FHIKES.N.R
	70	80	90	100	110	120
Pf Fabp11a	NTEIKFKLNEPFDETTADDRKTRTV--VTLENGKLVQKQNWGDGKETCIEREISDGKLIAK					
Fu Fabp11aE.M.....
Me Fabp11a	T..V.....E.....T.--I..D.....S.....S.....
St Fabp11a	T...R....V..E.....T.--.S..G.....S.....N.....T.....
Zf Fabp11a	T.....E.....T.--M.I.....T.....ST.....V.....
Pf Fabp11b	T.....E.E....G.M.KADGRM.K.....H.K.....T.....Q....T..
Me Fabp11b	T.....R.G.E.....V.G.NVKST--F..D.....TSTL.....QE.....
St Fabp11b	T..F.....EC.....G.T.K.L--..KD.....Q.T.....T.TL.....QG.NMS..
Zf Fabp11b	TL.....D.E.....VK.T--MS.AD....I....T.E..T.I.....Q.T.M..
Pf Fabp7bS...G.E.....NCKST---DGD...HV.K.....KFV...K....VMD
Pf Fabp7aSA..G.E....P...HVKST--FSM.GD..M.V.K.N....KFV...K...MVMT
Pf Fabp2	..L.MD.T.GVT..YSL..GTELGSWSMEG.SM.G.FVRKDN..QLTTI.TLEG.E.VQS
	130	Identity (%)	Similarity (%)			
Pf Fabp11a	CIMGDVVAVRTYVREA	100	100			
Fu Fabp11a	..I.....K..	94	98			
Me Fabp11aK..	86	95			
St Fabp11aI.....K..	84	94			
Zf Fabp11a	.K.....K..	82	93			
Pf Fabp11b	..AD....L...E.V-	67	77			
Me Fabp11b	...D.....EKAT	66	79			
St Fabp11b	.V.D.....AKK.	65	82			
Zf Fabp11b	.T.D....I...EKD.	61	79			
Pf Fabp7b	LTFE..H.....EKA-	61	71			
Pf Fabp7a	LTFEG.T.....EKA-	53	67			
Pf Fabp2	YNYEG.D.K.IFK.S-	24	44			

Figure 2G

Fabp11b

	10	20	30	40	50	60
Pf Fabp11b	MVEQFVGTWTLASSENFDEYMKTIGVNFA	TROMGNLVKPNI	VFSVAEDGFISLKAESTFK			
Fu Fabp11bN..V..D..D.....		M.....MGD.....	M.S.....		
Me Fabp11bA.....	AV.MG.....	IT.....IQ.GA..L..M.....			
St Fabp11b	..D.....IA.....	A..GL.....	A.....IG.D.A.L..M.S.T..			
Fu Fabp11a	..K.....KMI.....	D..A..G.....	V..RT.....V..DDQ.TVCM.SQ.....			
Pf Fabp11a	..K.....KMI.....	D..A..G.....	V..RT.....T..DDQ.TVCMRTQ.....			
Me Fabp11a	.A.K.....KMI.....	D..A..G.....	V..RT.....V.IEDQ.T..M.S.Q.....			
Zf Fabp11a	..DK.....KMTT.D.....	A..G.....	V..RT.....VC.D.Q.L.CM.SQ.....			
St Fabp11a	...K.....KMIA.....	D..A..G.....	V..RT.....IVT.ED..VVC..SQ.....			
Zf Fabp11bY..K.KMT..D.....	AV.AS..S...A..A..S.LIA.D.Q.V.TM..VT...				
Pf Fabp6a	--A.A.R.LTETQ.GY.DFC.LL.IPADIIIEK.RDY.MITEVTQDG.T.SWTQVYP.DA					
Pf Fabp10	--D.N...QVYAQ..YE.FL.AMELPEDVIK.AKDI..ITEIKQNGKD.VVTSKTP-G.					
Pf Fabp6b	--T.T.KYE.ETQ..YE.FLDA..TAAGLLSAKTDH.VITEV.QDGND.TWTQSIP-HW					
	70	80	90	100	110	120
Pf Fabp11b	TTEIKFKLN	EEFEETTADGRMTKADGRMTKENG	KLVQHQ	KWDGKETTIEREI	QDGKL	TAK
Fu Fabp11bT.....	N..T--VV.F.....				
Me Fabp11bR.G..D..V..NV.S--TF.LD.....	K.N.....TS.L.....E..I..				
St Fabp11b	...F.....CD.....	T..T--LV.LKD.....	Q.T.....T..L.....G.NMS..			
Fu Fabp11a	N.....P...M..D.K.RT--VV.L.....	K.N.....C.....S.....I..				
Pf Fabp11a	N.....P.D.....D.K.RT--VV.L.....	K.N.....C.....S.....I..				
Me Fabp11a	...V.....P.....D.K.TT--VI.LD.....	K.S.....S.....S.....I..				
Zf Fabp11aP.....D.K.TT--V..I.....	K.T.....S.....VS.....I..				
St Fabp11aR....VP.....D.K.TT--VVSL.G.....	K.S.....N.....T.....I..				
Zf Fabp11b	.L.....D...D.....D.KV.T--T.SLAD...I.K.T.E..T.I.....	T.I.....T.MI..				
Pf Fabp6a	KVTNT.TVGK.SDME.IG.KKF.--TVFM.G...SVTF---	PNYHHTS..SG...IET				
Pf Fabp10	.LTNS.TIGK.ADI..M..KKL.C--TV.M.GS...CKT---	EKFCHTQ.LKG.EMIET				
Pf Fabp6b	.WTNT.SVGR.C.LV.MK.TKF...-PVVMDD.RISVQF---	PQYHFTA.VSGD..IMN				
	130	Identity (%)	Similarity (%)			
Pf Fabp11b	CIAD---DVVALRTYERV-	100	100			
Fu Fabp11b---	86	92			
Me Fabp11b	..M.----V....KAT	72	81			
St Fabp11b	.VM.----V...AKKA	69	80			
Fu Fabp11a	.IG----V...VKEA	67	77			
Pf Fabp11a	.MG----V...V.EA	67	77			
Me Fabp11a	.MG----V...VKEA	67	78			
Zf Fabp11a	.KMG----V...VKEA	67	77			
St Fabp11a	.MG----I.V...VKEA	64	77			
Zf Fabp11b	.TM.----I....KDA	61	76			
Pf Fabp6a	SK.G---S..LK..SKKL-	23	36			
Pf Fabp10	LTKG---STTFV.KSKKMN	21	41			
Pf Fabp6b	.TIPGEKG.TFR..SR..-	21	33			

Figure 3: Phylogenetic analysis of pufferfish Fabp with other vertebrates.

A) A neighbour-joining tree showing the phylogenetic relationship between Fabp/FABPs of pufferfish (Pf), zebrafish (Zf), medaka (Me), three-spined stickleback (St), tilapia (Ti), human (Hu) and chicken (Ch). The bootstrap values are placed at the nodes and indicates the percentage number of times, out of 1000 bootstrap replications. The polypeptide sequence used in the phylogenetic analysis and their corresponding Ensembl/NCBI accession numbers are Pf Fabp1 (ENSTNIG00000016680), Me Fabp1 (ENSORLG00000003820), St Fabp1 (ENSGACG00000006574), Hu FABP1 (NP_001434), Pf Fabp 2 (ENSTNIG00000012360), Zf Fabp2 (NP_571506), St Fabp2a (ENSGACG00000017790), St Fabp2b (ENSGACG00000019474), Me Fabp2a (ENSORLG00000019478), Me Fabp2b (ENSORLG00000004146), Hu FABP2 (NP_000125), Pf Fabp3 (ENSTNIG00000016979), Me Fabp3 (ENSORLG00000011573), Ti Fabp3 (ENSONIG00000002309), Hu FABP3 (NP_004093), Hu FABP4 (NP_001433), Hu FABP5 (NP_001435), Pf Fabp6a (ENSTNIG00000017612), Pf Fabp6b (ENSTNIG00000004652), Me Fabp6a (ENSORLG00000005456), Me Fabp6b (ENSORLG000000012622), St Fabp6a (ENSGACG00000014354), St Fabp6b (ENSGACG00000018140), Hu FABP6 (NP_001436), Pf Fabp7a (ENSTNIG00000017217), Pf Fabp7b (ENSTNIG00000003393), Me Fabp7a (ENSORLG00000013475), Me Fabp7b (ENSORLG00000010773), St Fabp7a (ENSGACG00000010166), St Fabp7b (ENSGACT00000016034), Hu FABP7 (NP_001437), Ch FABP7 (NP_990639), Hu FABP8 (NP_002668), Hu FABP9 (NP_001073995), Pf Fabp10 (ENSTNIG00000002424), Me Fabp10a (ENSORLG00000014794), Me Fabp10b (ENSORLG00000007702), St Fabp10a (ENSGACG00000013639), St Fabp10b (ENSGACG00000002234), Ch FABP10 (P80226), Pf Fabp11a (ENSTNIG00000017781), Pf Fabp11b (ENSTNIG00000018400), Me Fabp11a (ENSORLG00000010830), Me Fabp11b (ENSORLG00000008282), St Fabp11a (ENSGACG00000003457), St Fabp11b (ENSGACG00000008725), Zf Fabp 11a (NP_001004682), Zf Fabp11b (NP_001018394) and Hu FABP12 (NP_001098751). The human lipocalin 1 protein (Hu LCN1, NP_002288) was used as outgroup. The scale bar indicates 0.2 substitutions per site. **B)** A maximum-likelihood tree showing the

evolutionary relationship of vertebrate Fabps, using the same amino acid sequence as mentioned above. The bootstrap values shown at the nodes indicate percentage based on 1000 replicates. The scale bar indicates 0.2 substitutions per site.

Figure 3A

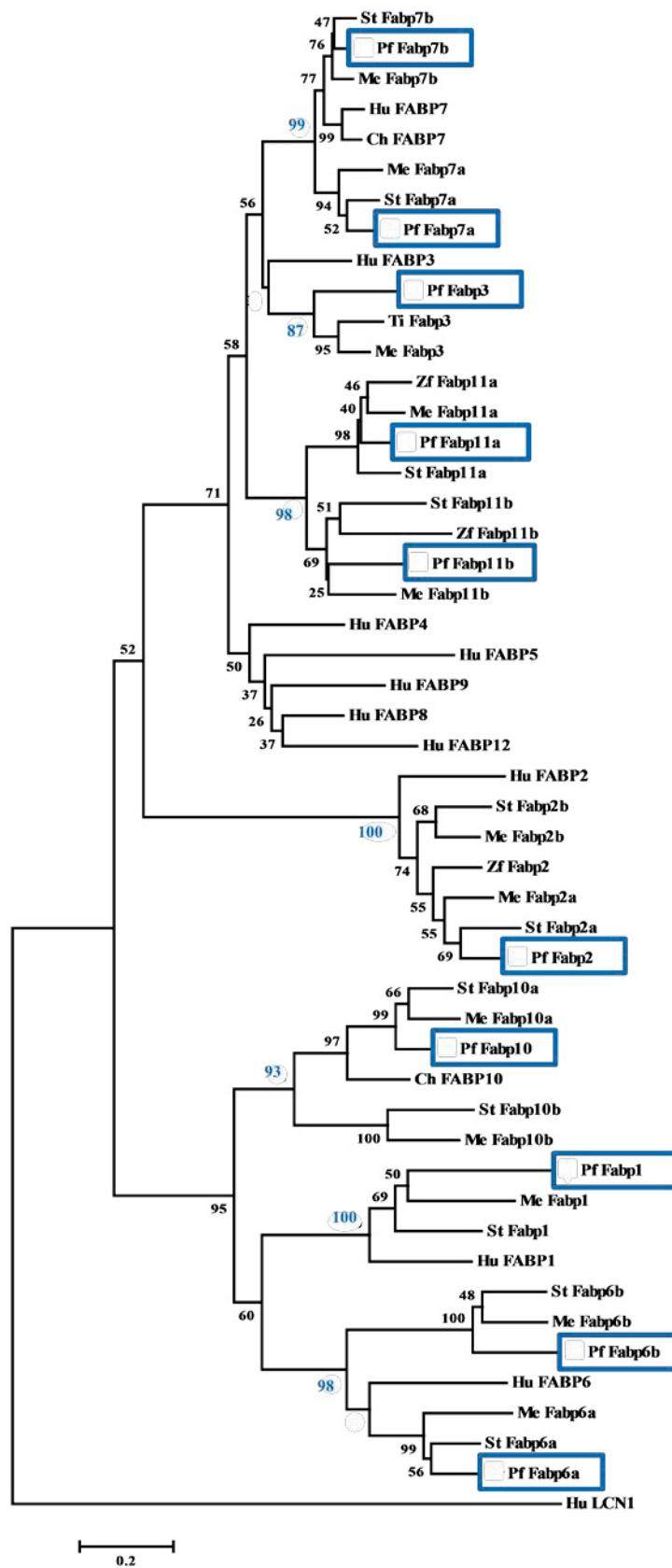
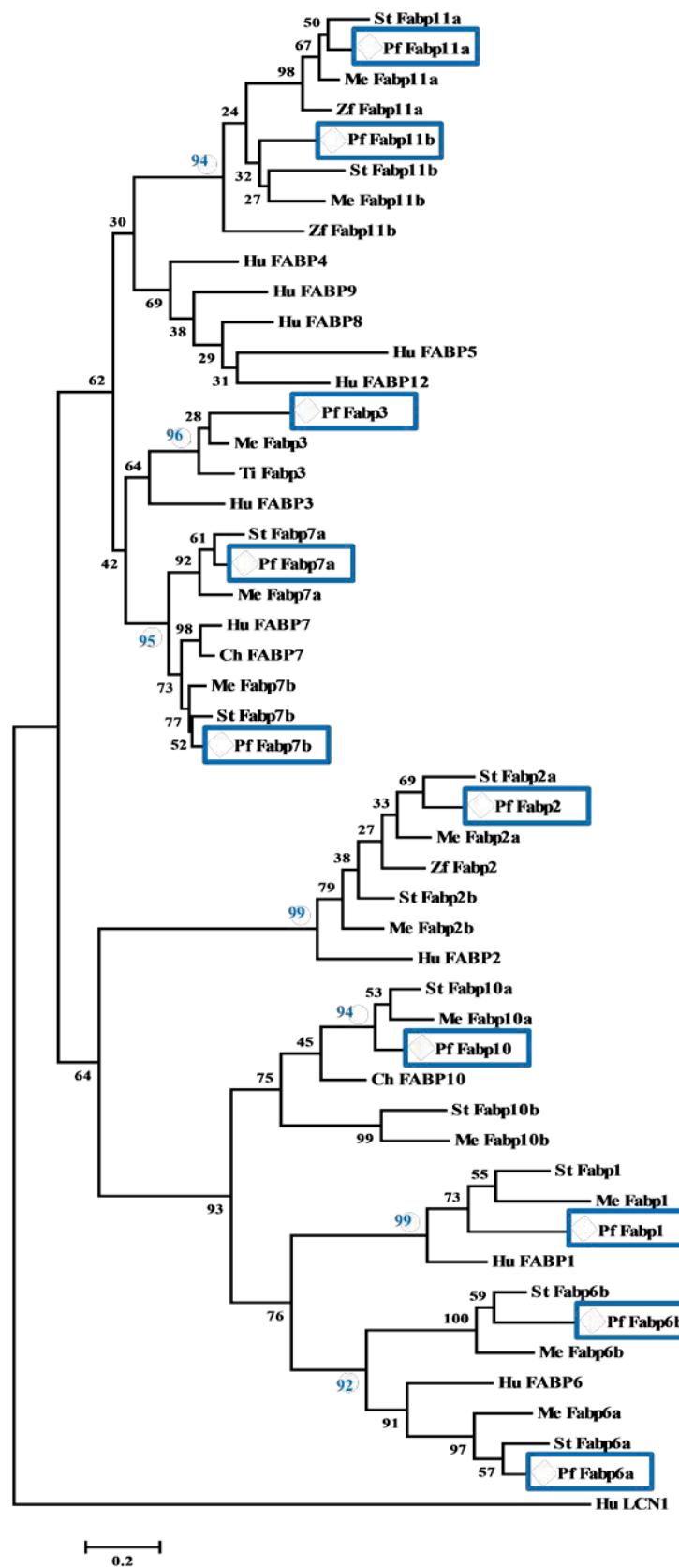


Figure 3B



3.3 Conserved gene synteny

Conserved gene synteny was evident between *fabp* genes of pufferfish and the orthologous *Fabp/fabp* genes of human and other teleost fish (Figure 4). Conserved gene synteny of human *FABP1* on chromosome 2 with zebrafish *fabp1a* located on chromosome 5 and *fabp1b* located on zebrafish chromosome 8, suggests that the duplicated copies arose owing to the FSGD event (Sharma *et al.*, 2006), followed by a tandem-duplication event, which lead to the retention of *fabp1b.1* and *fabp1b.2* on zebrafish chromosome 8 (Karanth *et al.*, 2009). The syntenic genes from pufferfish chromosome 12 (location of *fabp1*) and 4, medaka chromosome 9 (location of *fabp1*) and 12, three-spined stickleback chromosome XII (location of *fabp1*) and XIV exhibit conserved gene synteny with the syntenic genes of human chromosome 2, suggesting a possible loss of duplicated *fabp1* gene from chromosome 4 of pufferfish, chromosome 12 of medaka and chromosome XIV of three spined stickleback (Figure 4A). In the Ensembl database location of pufferfish *fabp2* was designated ‘un_random’, indicating an unassigned chromosome. The synteny of genes on human chromosome 4(location of *FABP2*) is conserved with the order of genes in medaka scaffold 461(location of *fabp2a*) and chromosome 18 (location of *fabp2b*), three spined stickleback chromosome IX (location of *fabp2a*) and VII (location of *fabp2b*), suggest that these orthologous genes arose from the same ancestral genes as a result of the FSGD event. The conserved gene synteny of zebrafish chromosome 1 (location of *fabp2*) and chromosome 7, and pufferfish unassigned chromosome (location of *fabp2*) and chromosome 18 with human chromosome 4 suggest a possible loss of duplicated *fabp2* gene from zebrafish chromosome 7 and pufferfish chromosome 18 (Figure 4B). Figure 4C displays the

Figure 4: **(A)** Conserved gene synteny of the pufferfish, zebrafish, medaka and three-spined stickleback *fabp1* with human *FABP1*. The syntenic genes of human *FABP1* located chromosome (ch) 2, exhibit conserved gene synteny with pufferfish ch 12 and 4, three-spined stickleback ch XIII and XIV, medaka ch 9 and 12, and zebrafish ch 5 and 8. **(B)** The syntenic genes of human *FABP2* located on ch 4, exhibit conserved gene synteny with pufferfish unassigned chromosome and ch 18, three-spined stickleback ch IX and VII, medaka scaffold 461 and ch 18, and zebrafish ch 1 and ch 7. **(C)** The syntenic genes of human *FABP3* located on ch 1, exhibit conserved gene synteny with pufferfish ch 21, three-spined stickleback ch X and medaka ch 11. **(D)** The syntenic genes of human *FABP6* located on ch 5, exhibit conserved gene synteny with pufferfish unassigned chromosome and ch 1, three-spined stickleback ch III and IV, medaka ch 17 and ch 10, and zebrafish ch 21 and ch 2. **(E)** The syntenic genes of zebrafish duplicated *fabp11a/fabp11b* located on ch 19 and ch 16, respectively, exhibit conserved gene synteny with pufferfish ch 21 and ch 8, three-spined stickleback ch X and XX, and medaka ch 11 and ch 18.

Figure 4A

fabp1

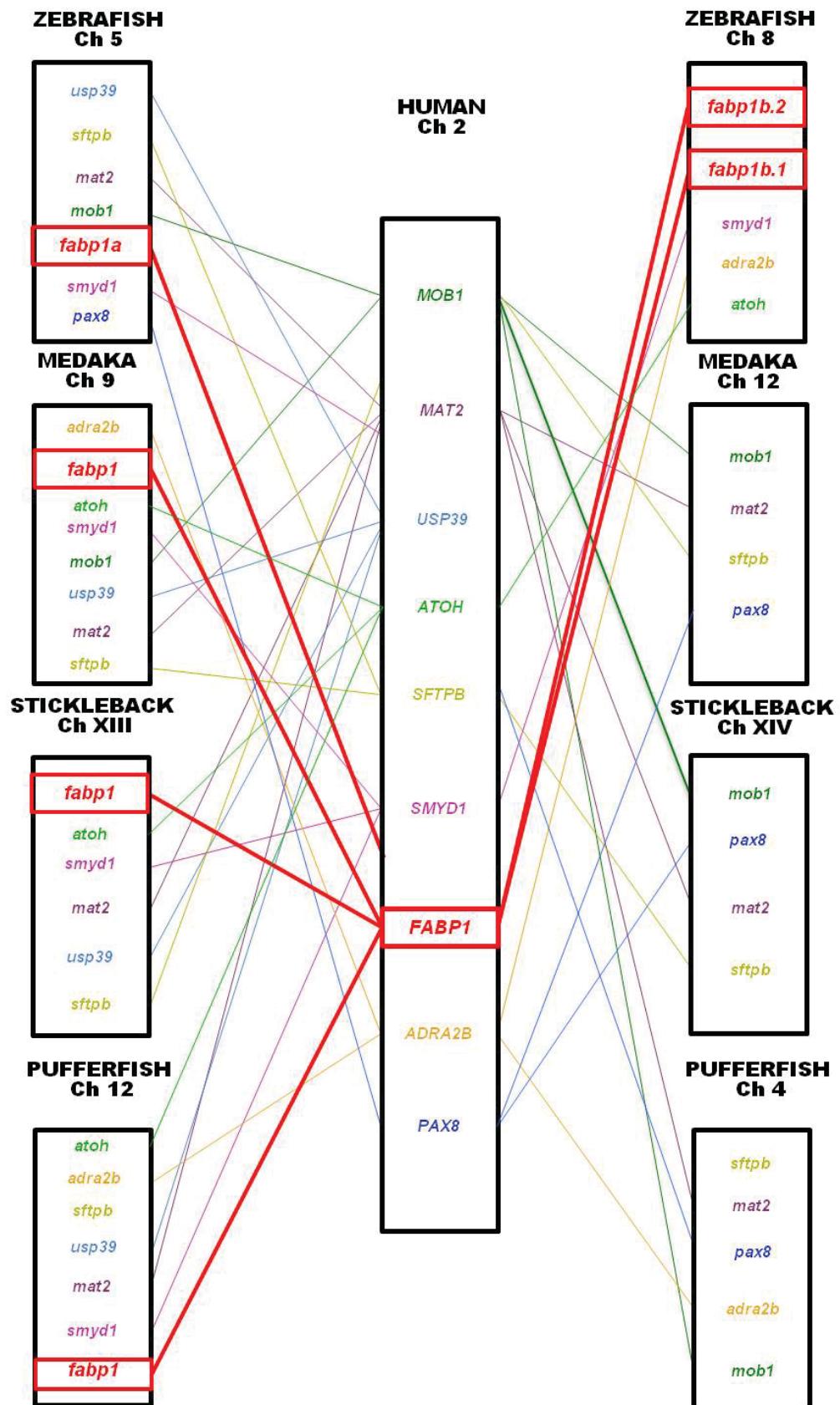


Figure 4B

fabp2

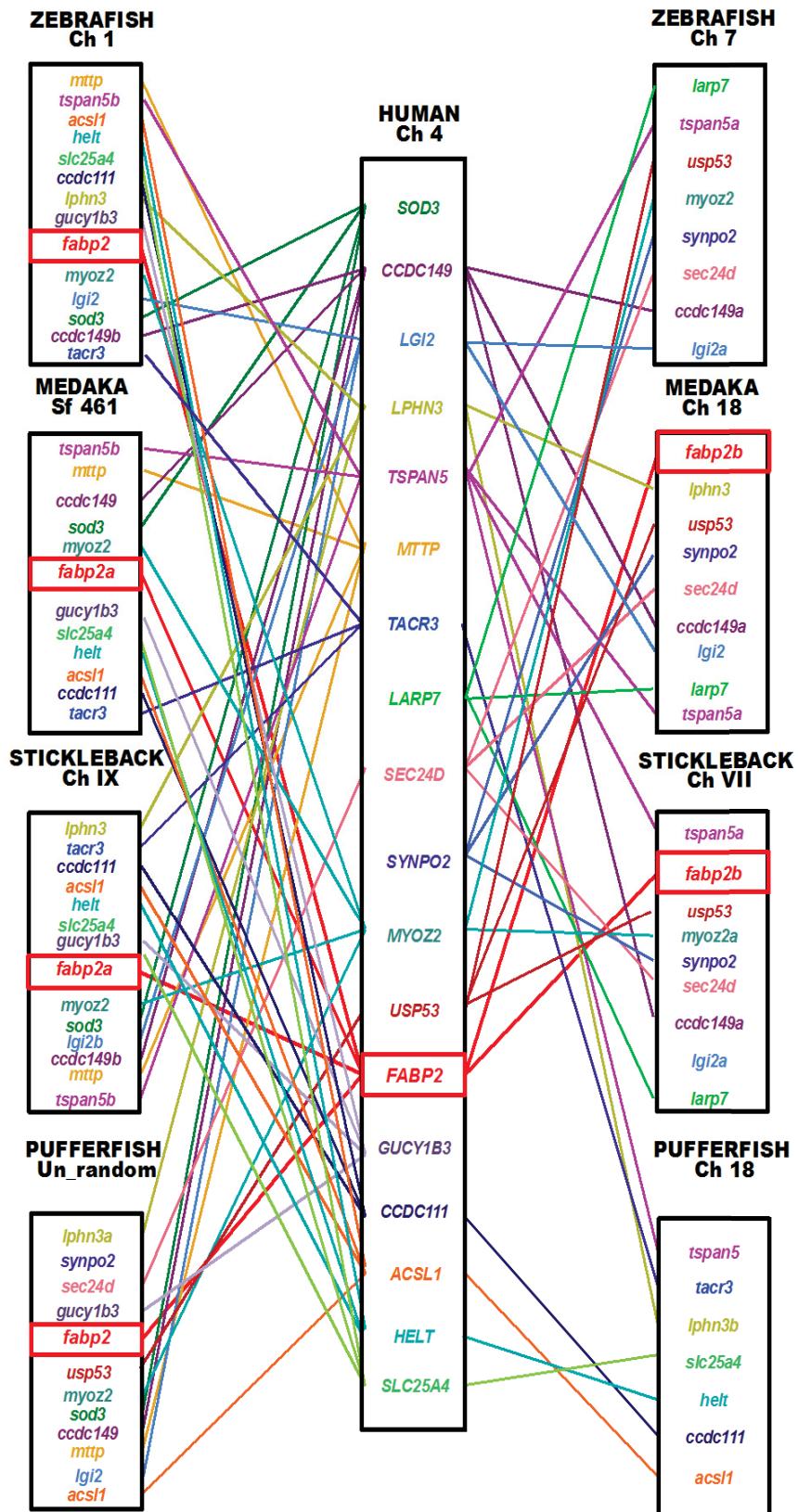


Figure 4C

fabp3

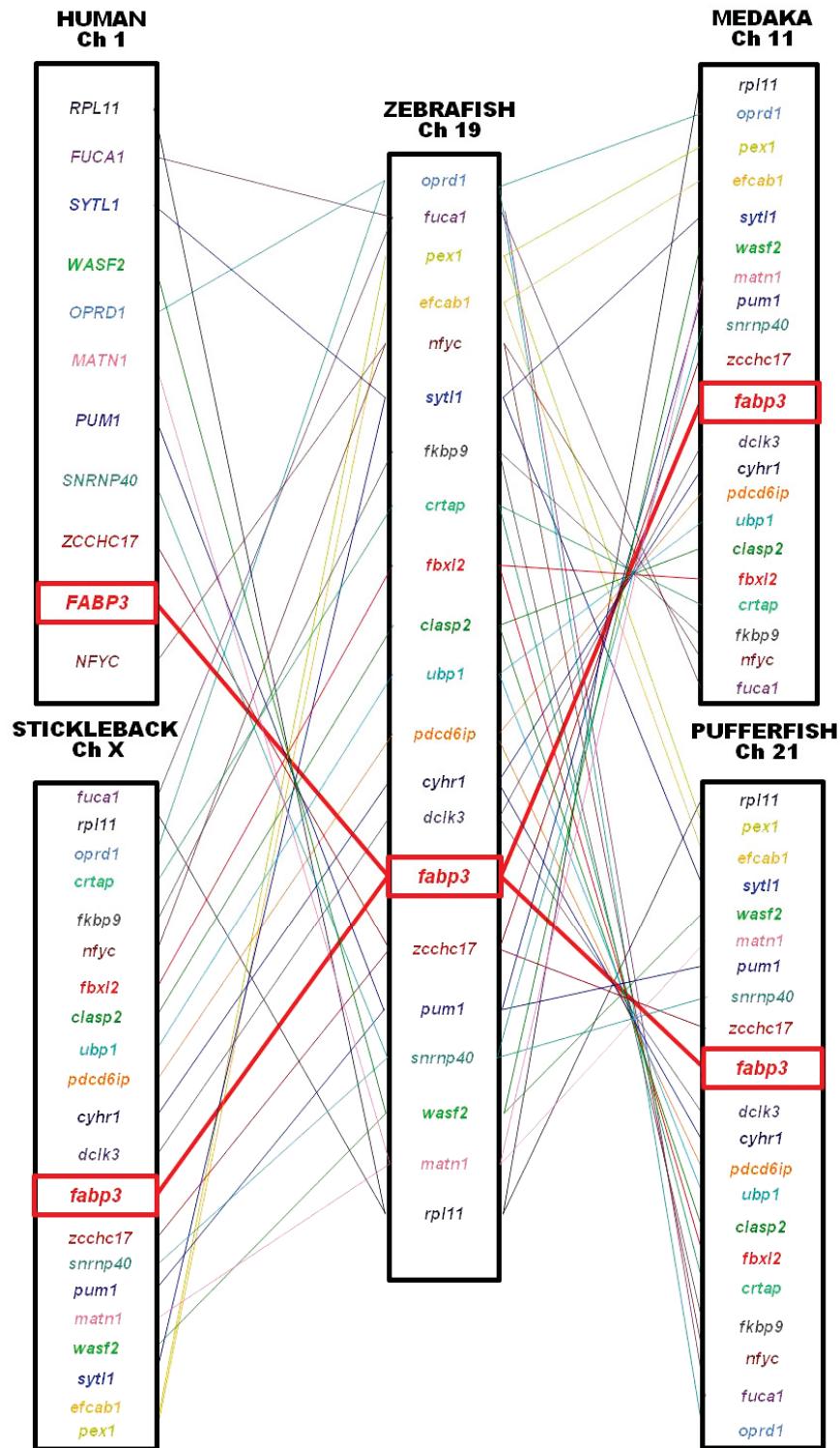


Figure 4D

fabp6

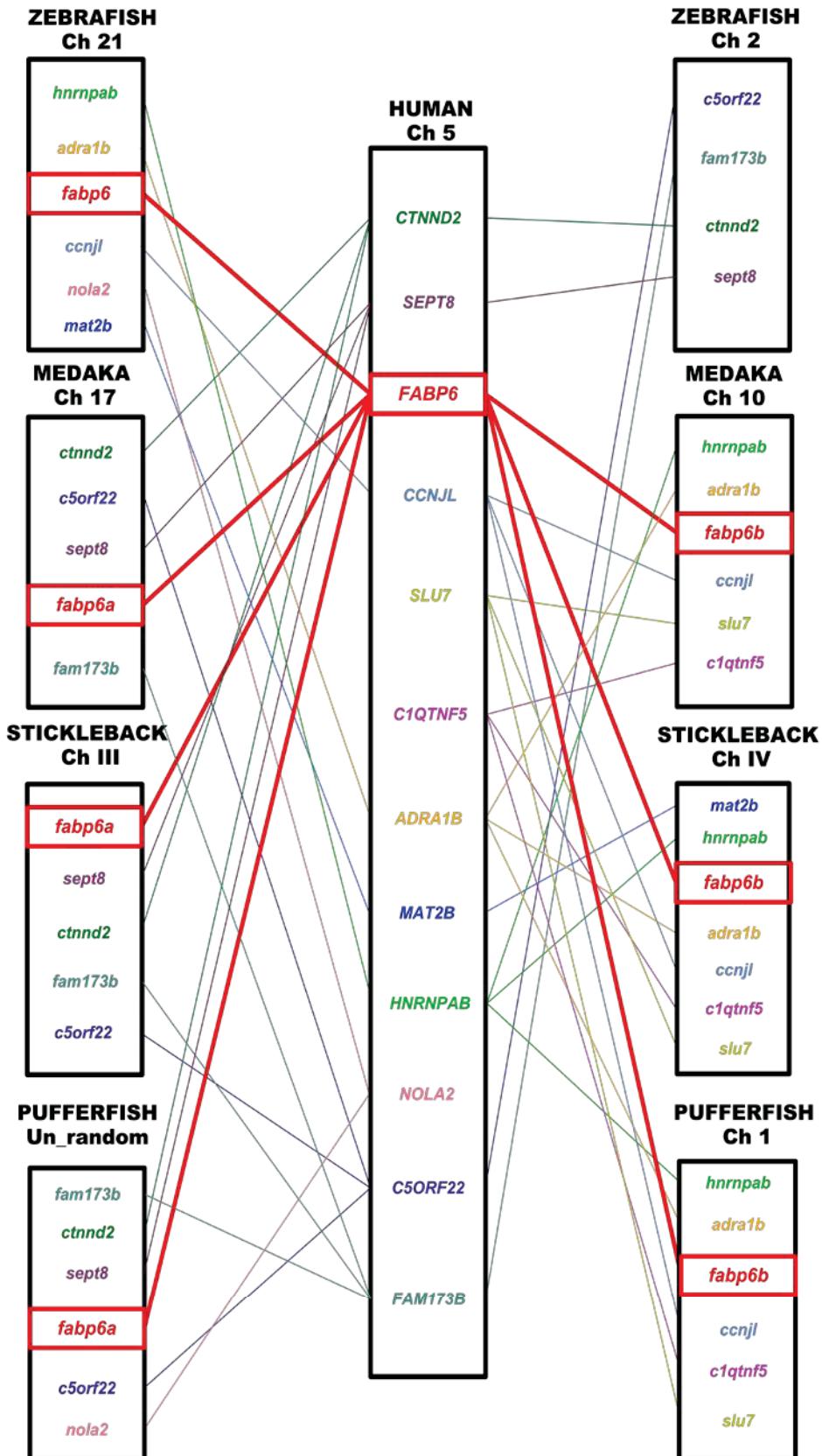
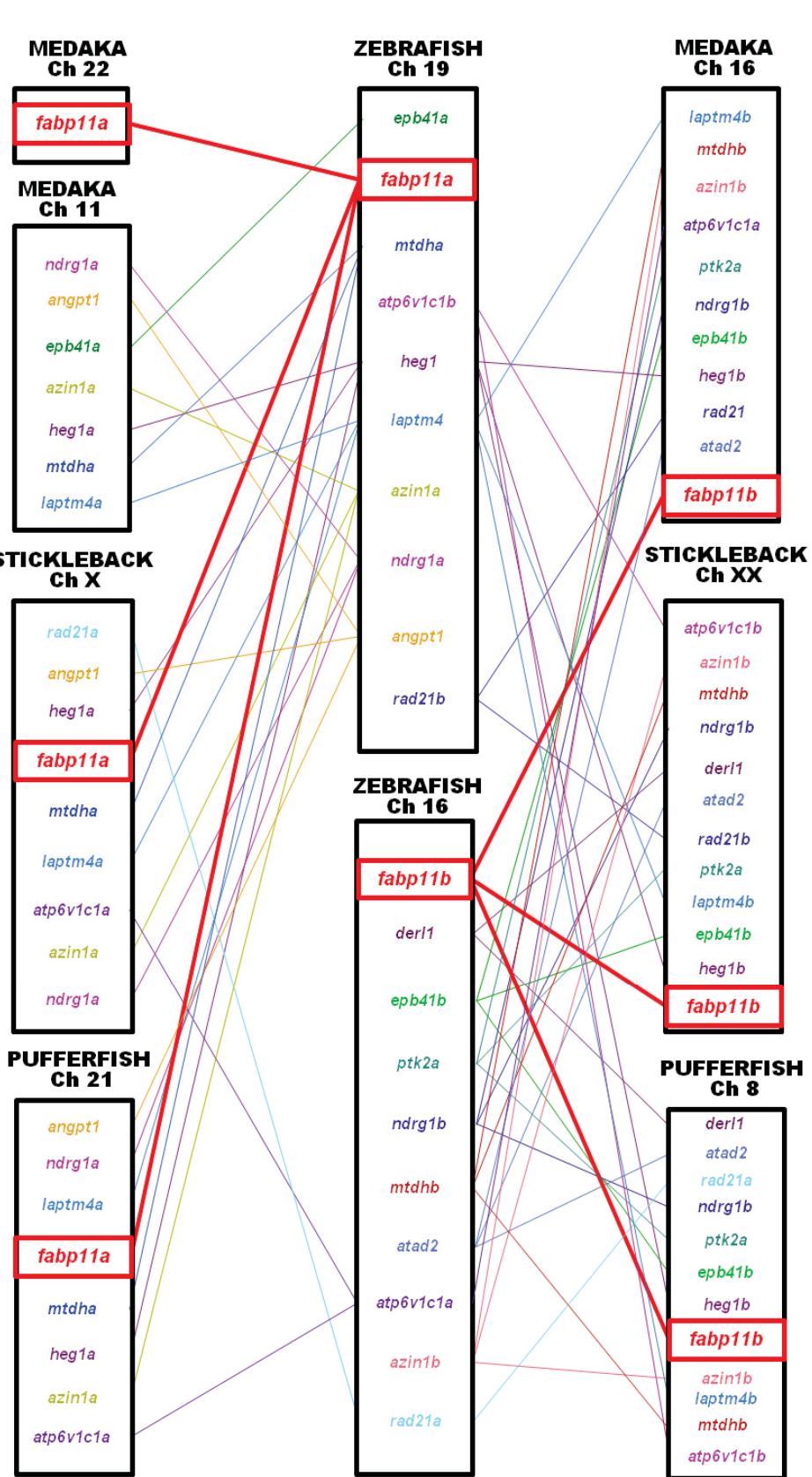


Figure 4E



conserved gene synteny of pufferfish chromosome 21 (location of *fabp3*), with human chromosome 1, zebrafish chromosome 19, medaka chromosome 11 and stickleback chromosome X, however, no duplicated *fabp3* copy were identified in any other teleost fish, suggesting a possible loss of the duplicated genes. Similarly, syntenic genes of human *FABP6* on chromosome 5 are conserved with the syntenic genes of *fabp6a* and *fabp6b* from pufferfish on unassigned chromosome and on chromosome 1, three-spined stickleback chromosome III and IV, and medaka chromosome 17 and 10 respectively (Figure 4D), whereas conserved gene synteny on chromosome 21 (location of *fabp6*) and chromosome 2 with human chromosome 5, indicate a possible loss of duplicated *fabp6* copy from chromosome 2. The gene synteny of pufferfish *fabp11* on chromosome 21 (location of *fabp11a*) and 8 (location of *fabp11b*) was found to be conserved with chromosome X (location of *fabp11a*) and XX (location of *fabp11b*) of three-spined stickleback, and chromosome 22 (location of *fabp11a*) and 16 (location of *fabp11b*) of medaka (Figure 4E). There was no synteny observed between chromosome 22 of medaka and other teleost *fabp11a* containing chromosomes, however, a much evident synteny of conserved genes were observed in chromosome 11 of medaka.

3.4 Tissue-specific distribution and steady-state levels of pufferfish *fabp* gene transcripts

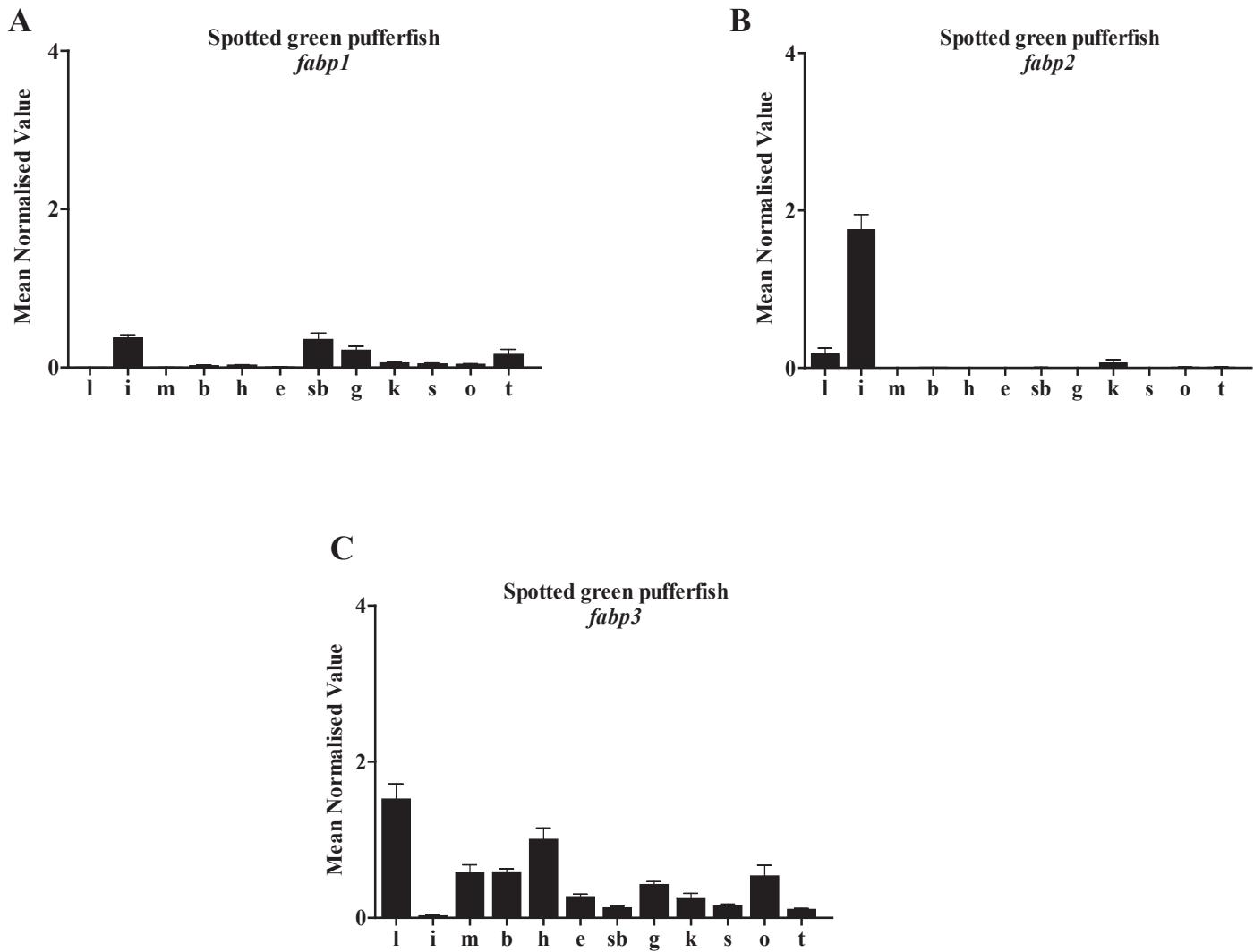
To further investigate the reason behind the retention of pufferfish *fabp* duplicates in their genome, the tissue-specific distribution of *fabp* mRNA transcripts in various tissues was determined. The transcripts of pufferfish *fabp1* were detected in all the tissues assayed except for the liver, muscle and eye (Figure 5A). Although Fabp1 was initially called the liver-type fatty acid binding protein, its mRNA transcripts were not detected in

the liver of pufferfish nor zebrafish (Sharma *et al.*, 2006; Karanth *et al.*, 2009), but high transcript levels were evident in the intestine, when compared to other tissues. Unlike pufferfish, duplicated copies of *fabp1* exist in the zebrafish genome owing to the FSGD event, followed by a tandem duplication event, presumably due to unequal crossing-over during meiosis (Sharma *et al.*, 2006 and Karanth *et al.*, 2009). Previous studies have also revealed that the zebrafish genome has undergone extensive translocation leading to considerable inter-chromosomal rearrangements (Woods *et al.*, 2005). The pufferfish genome, however, contains fewer inter-chromosomal rearrangements when compared to other teleost genome, which lead Jaillon *et al.*, to deduce the proto-karyotype of the ancestral bony fish vertebrate consisting of 12 chromosomes, after the divergence from mammals about 450 million years ago. Force *et al.*, (1999) proposed that one of the duplicated gene copies most often assumes redundancy in gene function(s), thereby resulting in functional loss of a gene owing to accumulation of deleterious mutations (nonfunctionalization). This process may account for the loss of one of the duplicated *fabp1* following the FSGD in the pufferfish genome. When nonfunctionalization does not occur, as proposed for zebrafish *fabp1a* and *fabp1b* (Sharma *et al.*, 2006), owing to positive selection and mutation, neofunctionalization may occur, where one copy acquires a new function, while the other duplicated gene copy maintains the function(s) of the ancestral gene. The tissue-specific distribution of pufferfish *fabp1*, when compared with the duplicated copies of zebrafish *fabp1* shows redundancy in function (Figure 5A). The expression levels of pufferfish *fabp1* transcripts were found in the intestine, brain, heart, swim bladder, gills, kidney, skin, ovaries and testis, whereas in the case of

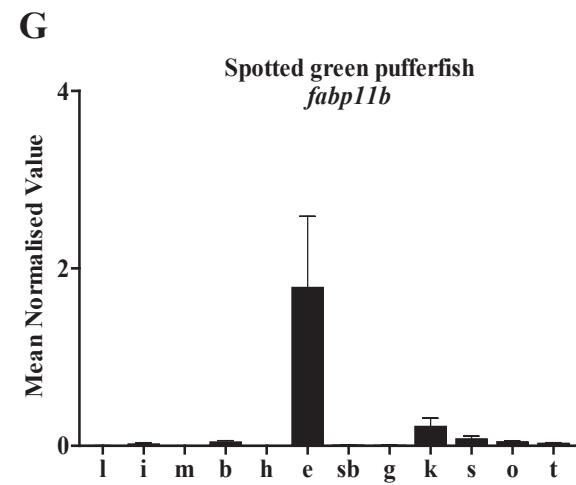
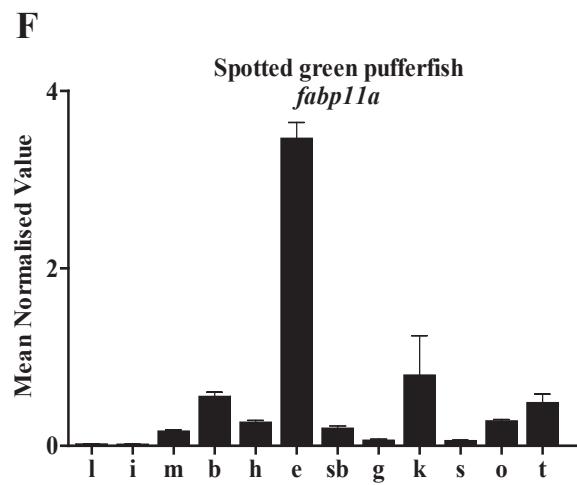
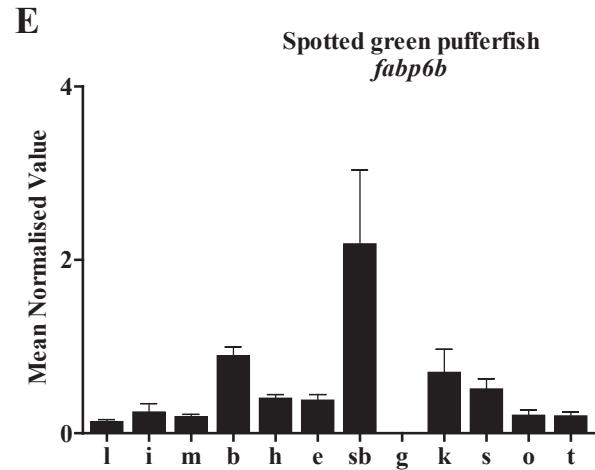
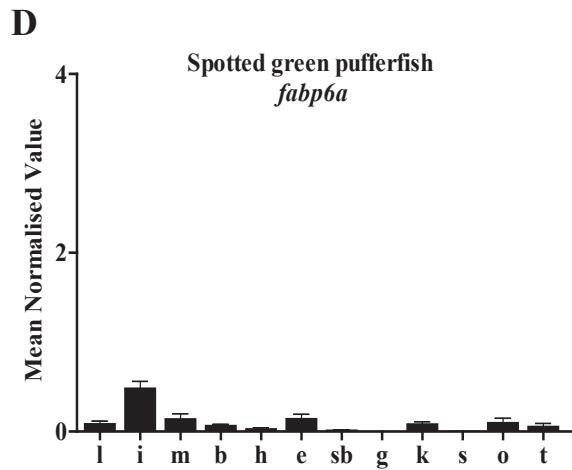
Figure 5: Tissue-specific distribution and steady-state levels of pufferfish *fabp* transcripts in various tissues.

The steady state levels of pufferfish *fabp* transcripts in liver (l), intestine (i), muscle (m), brain (b), heart (h), eye (e), swim bladder (sb), gills (g), kidney (k), skin (s), ovary (o), testis (t). The pufferfish *fabp* transcripts investigated for tissue-specific distribution are **A)** *fabp1*, **B)** *fabp2*, **C)** *fabp3*, **D)** *fabp6a*, **E)** *fabp6b*, **F)** *fabp11a* and **G)** *fabp11b*. The bar graph depicted shows a mean value of three male and three female fishes combined. No significant statistical difference was evident between the tissue-specific steady-state levels of male and female fish. Single black line on each bar represents the standard error of mean.

Figure 5



OS



zebrafish, tissue-specific distribution of *fabp1b.1* transcripts were found in the intestine, liver, heart, ovary and testis, while the tissue-specific distribution of zebrafish *fabp1b.2* transcripts were found in intestine, heart, brain, eye, skin and ovary. This pattern indicates a functional divergence in the tissue-specific distribution, possibly owing to subfunctionalization (Karanth *et al.*, 2009), where the combined functions of both gene copies partition the functions of the ancestral gene. Transcripts of pufferfish *fabp2* were more abundant in the intestine than in the liver and kidney, and absent in muscle, brain, heart, eye, swim bladder, gills, skin, ovaries and testis (Figure 5B). While duplicated copies of medaka and three-spined stickleback *fabp2* are retained in the genome (Parmar *et al.*, 2012), the duplicated copy of zebrafish and pufferfish *fabp2* has been lost from the respective genomes. The tissue distribution of pufferfish *fabp2* transcripts was similar to the distribution of *fabp2* in zebrafish (Parmar *et al.*, 2013a). The steady-state level of pufferfish *fabp2* transcripts were low in kidney, compared to the levels of medaka *fabp2a* (Parmar *et al.*, 2012). The tissue-specific distribution of pufferfish *fabp3* mRNA transcripts were distributed in all tissue assayed, except intestine (Figure 5C), with levels most abundant in liver, heart and ovaries. *fabp3* exists as a single copy in all the teleost fishes studied to date (Liu *et al.*, 2007; BLAST search in Ensembl genome database), however, *fabp3* of Antarctic teleost fishes are found to be retained as duplicated copies (Vayda *et al.*, 1998). The pufferfish *fabp3* mRNA levels are high in heart and liver tissues, in contrast to lower levels of expression in all other tissues. These results correlate with the findings of Liu *et al.*, in 2003(a), where they first reported *fabp3* of zebrafish. The steady-state levels of pufferfish *fabp6a* mRNA transcripts were higher in intestine, when compared to liver, muscle, brain, heart, eye, kidney, ovaries and testis (Figure 5D),

whereas, *fabp6b*, was abundant in swim bladder, brain and kidney. Swim bladder, brain, heart, kidney and skin showed high steady-state levels of *fabp6b* transcripts when compared to *fabp6a*. This tissue-specific pattern of mRNA distribution may contribute to the retention of duplicated copies in the pufferfish genome after the FSGD event. Parmar and Wright (*in press*) showed that *fabp7* exists as duplicated copy and *fabp10* as single copy in the pufferfish genome. They identified and discussed the various tissue-specific distribution patterns of the orthologous *fabp7* and *fabp10* transcripts in medaka, zebrafish, three spined stickleback and pufferfish. The *fabp7a* mRNA transcripts are found in all the tissues, except for swim bladder and ovaries, whereas *fabp7b* transcripts were found only in intestine, heart and kidney. The tissue-specific distribution of *fabp7b* mRNA transcripts was found only in liver, intestine, brain, gills and kidney (Parmar and Wright, *in press*). The steady-state level of pufferfish *fabp11a* transcripts was abundant in brain, kidney, eye, ovary and testis, when compared to low levels of *fabp11a* mRNA transcripts in muscle, heart, swim bladder, gills and skin. Similar to their orthologs in medaka and zebrafish (Parmar *et al.*, 2012b), both the duplicated copies of pufferfish *fabp11* were dominant in eye. The tissue-specific distribution of *fabp11a* was found to be extensively distributed in most of the tissue samples assayed, when compared to *fabp11b*. This pattern of tissue-specific distribution suggests either a subfunctionalization, where the functions of an ancestral gene is divided between the duplicates or neofunctionalization, where one copy of the sister duplicate acquires a novel function.

Susumu Ohno in 1970 proposed that at least two rounds of whole genome duplication in early vertebrate lineage contributed to the increased complexity and genome size of vertebrates. Three whole genome duplication events have so far been

proposed in the evolutionary history of vertebrates. However, studies on timing of the first two-rounds of genome duplication in the vertebrate lineage remains debatable (Dehal and Boore, 2005). The third-round or the FSGD is clearly evident in the evolution of vertebrate lineage (Jaillon *et al.*, 2004; Woods *et al.*, 2005). The comparative analysis of about 6000 pufferfish genes to the human genome showed patterns of ‘double conserved synteny’, where one chromosomal region in mammal matches with two in the pufferfish genome (Jaillon *et al.*, 2004). The study described here was performed to identify and structurally characterize the *fabp* genes in the pufferfish genome, to determine if the duplicated copies of pufferfish *fabp* genes arose as a consequence of FSGD, approximately 230-400 million years ago in the ray-finned fish lineage and to provide evidence regarding the evolutionary processes that lead to the loss or retention of these duplicated copies after the FSGD. I identified ten different paralogs of pufferfish *fabp* genes, three of which were found to be retained as duplicated copies (*fabp6*, *fabp7* (Parmar and Wright, *in press*) and *fabp11*). Multiple sequence alignment and phylogenetic analysis showed that the pufferfish Fabps are orthologous to vertebrate Fabps/FABPs. Conserved gene syntenies for each pufferfish *fabp* gene suggest that the duplicated genes arose as a result of the FSGD event. A comparative study on the pattern of tissue-specific distribution of duplicated pufferfish *fabp* mRNA transcripts (Table 4) with other teleost orthologs, concluded that, following the FSGD event, a considerable loss of duplicated genes from the pufferfish genome were evident in pufferfish *fabp1*, *fabp2*, *fabp3* and *fabp7* transcripts. Also evident is the differential tissue-specific distribution patterns, which suggests considerable divergence in the *cis*-regulatory elements of pufferfish *fabp6*, *fabp10* and *fabp11* since their duplication.

Table 4: The tissue-specific distribution of the steady-state levels for pufferfish *fabp* transcripts. + / - indicates presence/ absence of transcripts, respectively.

Tissues	Gene transcripts						
	<i>fabp1</i>	<i>fabp2</i>	<i>fabp3</i>	<i>fabp6a</i>	<i>fabp6b</i>	<i>fabp11a</i>	<i>fabp11b</i>
Liver	-	+	+	+	+	-	-
Intestine	+	+	+	+	+	-	-
Muscle	-	-	+	+	+	+	-
Brain	+	-	+	+	+	+	+
Heart	+	-	+	-	+	+	-
Eye	+	-	+	+	+	+	+
Swim bladder	+	-	+	-	+	+	-
Gills	+	-	+	-	-	+	-
Kidney	+	+	+	+	+	+	+
Skin	+	-	+	-	+	+	+
Ovary	+	-	+	+	+	+	+
Testis	+	-	+	+	+	+	-

Further insight into the retention or loss of *fabp* genes in teleost fishes may be gained from the study of tissue-specific distribution of *fabp* gene transcripts in a basal extant species of ray-finned fishes (e.g. spotted gar and bichir), which diverged from the teleost fish lineage before the FSGD event or the ancient fish, coelacanth, a member of the lobe-finned fishes. This type of study might shed some light on the spatial distribution of an ancient *fabp* gene transcript, thereby providing insight into the evolutionary processes that lead to the retention or loss of duplicated *fabps* in the teleost fish genome. Pre Ensembl, a preliminary database, contains partial datasets for spotted gar and coelacanth, while the fully annotated genomic dataset can soon be expected in the Ensembl database.

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Appendix

Sequence alignment of Ensembl-derived genomic DNA, NCBI-derived cDNA and PCR amplified genomic of pufferfish *fabp1*. The amino acid sequences are shown below in bold. The dots represent identity and the dashes represent missing nucleotides. The star symbol indicates the stop codon.

Genomic	1	TTTGGGTGTATAAAACGGGCAGGTGAGCAGGTGTTCCCTCTACTCTTCGTCACTCACTGT
cDNA	1	-----
<i>fabp1</i>	1 M
Genomic	61	CTCTCTCATCTGGAAAATACCAGCAGGTGTCATTCCAGGTAGAACCTTGAGCCTTTATG
cDNA	1	-----
<i>fabp1</i>	55 S L S G K Y Q Q V S Q E N F E P F M
Genomic	121	AAGGCCATTGTGAAGAGTCCAGCTTCGTCTTAACAGCCCTGAGTTCTCATTGATGATA
cDNA	1	-----CTC...CTGG.AAATACCAGCAGGTG.C.C.G..-G
<i>fabp1</i>	108 K A I
Genomic	181	AACATTGTTGCTTGTCTGCCTGTTCTTTAGGTGTTCTGATGAAATCATCCAGCAG
cDNA	37	...T...A...C.T..A..AAG.CCA.T----- K A I G V P D E I I Q Q
<i>fabp1</i>	168 G V P D E I I Q Q
Genomic	241	ATCAAAGACATCATGAGCTTCACTGAGATTGTGCAGAATGGCAATGATTTCAAAATCACC
cDNA	91 I K D I M S F T E I V Q N G N D F K I T
<i>fabp1</i>	228 I K D I M S F T E I V Q N G N D F K I T
Genomic	301	ACCACTACTGGCCCAAAGGTAACGGTCAACCAATTACCATGGAAAGGAGACAGAGATG
cDNA	151 T T T G P K V T V N Q F T I G K E T E M
<i>fabp1</i>	288 T T T G P K V T V N Q F T I G K E T E M

Genomic	361	GACGCCATCAGCGGAGAAAAGATAAAGGTGAGGCAAGACAAGAAAGTCTGTTGCTTGAGA
cDNA	211-----
		D A I S G E K I K
<i>fabp1</i>	348-----
		D A I S G E K I K
Genomic	421	AAAGTCTGAGCGCGGCCATGGTAACCTCATTCTGCTTCATGTCGGAAGACGGTGTTC
cDNA	237	-----.....-----
		T V F
<i>fabp1</i>	408-----
		T V F
Genomic	481	GTCTGGAGGACAACAAACTGAAAGTCTCCCTGAAAAATATAGAGTCGGTCACAGAACTGG
cDNA	248-----
		R L E D N K L K V S L K N I E S V T E L
<i>fabp1</i>	468-----
		R L E D N K L K V S L K N I E S V T E L
Genomic	541	TGGATCAAACACGCTCGTTGCCGTAAGTAGTTGA-----ATGCTGAGTGACTCCAAA
cDNA	308-----
		V D P N T L V A
<i>fabp1</i>	528-----
		V D P N T L V A ..N..
Genomic	595	-TCTA---GAATAAATCTCTAAATTCTGCCTATCTATCTGCAGGTGATGACTTGGCGA
cDNA	368	-----.....-----A.
		V M T L G N
<i>fabp1</i>	583	A.....-----T..A..T..T.....
		V M T L G D
Genomic	650	CATCGTGTACAAGTCAACAAAGTAAACGCGTGTAAAGAACCGAGAACTTGTGCTTACACCAA
cDNA	428-----
		I V Y K S T S K R V *
<i>fabp1</i>	643-----
		I V Y K S T S K R V *