

# Fusobacterial Infections

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Anaerobic, Gram-negative bacilli of the genus *Fusobacterium* have been implicated in the etiology, pathophysiology, and complications of several diseases, including periodontal diseases, Lemierre's syndrome, tropical skin ulcers, and intraamniotic infections (IAI). As part of the normal flora of the oral cavity, female genital tract, and gastrointestinal tract, fusobacteria have a number of natural entry points to cause disease. *F. nucleatum* plays a critical role in the development of periodontal diseases by acting as a microbial bridge between early and late (pathogenic) colonizers of the oral tissues. *F. necrophorum* is the causative agent of Lemierre's syndrome, a rare infection that can have devastating effects on the joints, lungs, and central nervous system. A variety of fusobacteria have been implicated in the development of tropical skin ulcers, which continue to cause significant debility in regions of the tropics. Fusobacteria have been associated with a significant proportion of preterm low birth weight infants due to IAI. Morbidity and mortality may result from IAI, and the incidence of IAI has not decreased in recent years. Typically, antimicrobial drugs provide effective treatment of fusobacterial infections, which can affect people of all age groups.

## INTRODUCTION

Fusobacteria, which form part of the family *Bacteroidaceae*, are asaccharolytic obligately anaerobic, non-spore forming, Gram-negative bacilli (1,2). Historically, the production of butyric acid, rather than isobutyric or isovaleric acids, has been used to differentiate fusobacteria from other members of the family *Bacteroidaceae*. More recently, fusobacteria have been subdivided into species and subspecies by the comparative analysis of cellular fatty acid patterns and small-subunit rRNA gene sequences (1,3,4).

Fusobacteria, either alone or in combination with other anaerobes and aerobes, have been isolated from a wide variety of clinically significant anaerobic infections. However, positive identifications cannot always be made because these bacteria require specific growth conditions, appear with varying cell morphologies, and give primarily negative responses in routine biochemical tests (5). On the other hand, the occurrence of fusobacteria as part of the normal flora of the oral cavity,

female genital tract, and gastrointestinal tract (5-7) provides many opportunities for these bacteria to initiate infections.

## PERIODONTAL DISEASES

Severe destructive periodontal diseases affect 5-20% of the population, at considerable social and economic costs (8,9). These diseases are characterized by local tissue inflammation, tissue destruction, and bone loss (8). Periodontal diseases, which eventually result in the loss of teeth, can also have a variety of systemic effects (8,10). It is well known that transient bacteremias can occur as a result of normal oral hygiene practices, such as the brushing of teeth (10,11). Such transient bacteremias can cause complications in susceptible patients. For example, patients with damaged heart valves or prosthetic devices are susceptible to infective endocarditis or infection of the prosthesis (10,12,13), and precautions are taken to avoid complications due to transient bacteremias arising from professional dental care (12). Infection by *Chlamydia pneumoniae* and other microbial factors have been implicated as risk factors for atherogenesis (14-16), and many epidemiologic studies have indicated that patients suffering from periodontitis are also at an increased risk of developing coronary ar-

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tery disease (CAD) (17-19). A cross-sectional study by Mattila *et al.* (18) has suggested that dental infections, particularly gingivitis and periodontal diseases, are as important as the classical risk factors (age, smoking, diabetes, hypertension, and elevated serum triglycerides) in the pathogenesis of CAD. DeStefano *et al.* carried out a prospective cohort study (9760 subjects followed for a median of 14 years) in which it was determined that men under 50 with periodontitis had a stronger risk for CAD, but overall, they found that periodontal disease was associated with a small increased risk (19). Diabetic patients may have a reduced need for insulin following treatment for periodontitis (10,20,21), and it has been noted that potential respiratory pathogens may become established in the oral flora of patients with periodontal disease (10). A number of studies which examined the relationship between oral health and the sense of well-being, especially in the elderly, have concluded that eating difficulties lead to social withdrawal, especially in the elderly (10,22,23).

Infectious periodontal diseases, including gingivitis and periodontitis, are complex, multifactorial diseases, primarily due to the interaction between organisms and the immune system (1). Many reviews have described the pathogenesis in detail (1,2,24-29); a simplified summary follows. The initial event in periodontal diseases is the growth of predominantly aerobic, saccharolytic bacteria along a clean gingival margin (30,31). These early colonizers are primarily Gram-positive streptococci and Gram-positive rods which are capable of adhering to human tissues, such as tooth enamel and gingiva. The anaerobic, asaccharolytic bacteria (1,2) responsible for periodontal diseases do not adhere directly to human tissue, and buildup of these species occurs by coaggregation with other bacteria, a phenomenon particularly associated with oral bacteria (1,30). The interactions among the various genera and species colonizing the oral mucosa (1,30,32-38) are cell specific; the late colonizers found in periodontal diseases do not coaggregate with the early colonizers. The ability of *Fusobacterium nucleatum* to coaggregate with most early and late colonizers (1,30) suggests that it acts as a microbial bridge between the early and late stages of infection (1,30,36,38). Because of this role, relatively large quantities of *F. nucleatum* are often found in subgingival pockets affected by periodontal disease (1,39-41). While *F. nucleatum* coaggregates with the three microorganisms that are normally implicated in the etiology of periodontal diseases (1,30) - *Actinobacillus actinomycetemcomitans*, *Porphyromonas gingivalis*, and *Bacteroides forsythus* - the relationship between *F. nucleatum* and *P. gingivalis* appears particularly strong (1,35,36).

*F. nucleatum*, being part of the normal oral flora, can be isolated from the healthy gingiva of most adults (6,42). This is not to say, however, that the organism is benign; the many virulence factors that enable it to be pathogenic in periodontal diseases are likely to be quite important in systemic diseases as well. The lipopolysaccharide (LPS) in the cell wall of *F. nucleatum* is structurally related to the LPS of other Gram-negative bacteria, and has a biological activity similar to that of *Escherichia coli* (1,43-45). In addition to its toxic properties, the LPS of *F. nucleatum* activates complement,

provoking an inflammatory response, resulting in further tissue destruction (1,2,46). The production of butyric acid by fusobacteria inhibits the proliferation of gingival fibroblasts which normally compromises the rapid healing of wounds (1,47,48).

The outer membrane proteins (OMPs) of *F. nucleatum* have a role in bacterial nutrition (1) and may play a role in the pathogenesis of adult periodontitis (49). OMPs in *F. nucleatum* display bioactivities similar to those of LPS, but are present in greater quantities (49). In other Gram-negative bacteria, OMPs provide a route for the uptake of antibiotics, and the OMPs in *F. nucleatum* may display similar activities (1,50). Evidence for the production of extracellular proteolytic enzymes by fusobacteria is weak (51,52), although its occurrence cannot be discounted (53).

## LEMIERRE'S SYNDROME

Lemierre's syndrome (synonyms: postanginal sepsis, necrobacillosis) classically presents with a severe sore throat, followed by fever, rigors, and painful cervical lymphadenopathy in a previously healthy child or young adult (6,54-58). Jaundice may also be present (57). Both exudative and non-exudative tonsillar and peritonsillar abscesses, and/or lesions in the mouth and jaw may be present (56). Lemierre reported that the syndrome, which he referred to as "post-anginal septicemia", may result following otitis media, mastoiditis, appendicitis, urinary tract infection, or "purulent" endometritis following parturition (56). Of these alternative presentations, otitis media is most frequently encountered, but overall, Lemierre's syndrome has not been very common in the antibiotic era (6,57-59). Recently, there has been some disagreement as to whether Epstein-Barr virus (EBV) may predispose patients to this condition (6,55).

Infections of *Fusobacterium necrophorum* are responsible for Lemierre's syndrome. Following oropharyngeal infection, the jugular vein becomes palpable as bacteria begin to colonize it (as well as other local veins) (6,54,60). The suppurative internal jugular vein may be mistaken for lymphadenopathy (6,54). Septic emboli from the jugular vein allow distant metastatic spread of *F. necrophorum* to the lungs, as well as to the joints and central nervous system (CNS) (6, 57,58). As infection spreads to the lungs, multiple infiltrates quickly cavitate and can result in pleural effusion, empyema, and/or pneumothorax (54,57). Septic arthritis usually affects one or more large joints, such as the hip or knee (6). *F. necrophorum* meningitis can follow pharyngeal infection and may also be otogenic (59). Cranial palsies and brain abscesses leading to infarction are possible CNS sequelae (6,61).

Lemierre's syndrome is relatively easy to diagnose clinically and timely antibiotic therapy can prevent complications or death (54). However, rates of morbidity and mortality (mortality: 4-18%) remain high, partly because unfamiliarity with the disease leads to delays in diagnosis, underdiagnosis, and delays in the choice of a proper antimicrobial agent. Unfortunately, identification of the pathogen by culture is often the first indication of the disease (6), but

growth on solid media in a laboratory takes at least 48 hours. This emphasizes the need for a prompt clinical diagnosis (6,55,58,62). Computed tomography (CT) or ultrasound of the neck may be used to confirm involvement of the internal jugular vein (6). X-rays may be used to localize some cranial and pulmonary lesions, and to follow the progress of treatment. The fairly recent reviews by Eyken and Sinave *et al.* should be helpful in correctly diagnosing this syndrome (57,58). It is ironic that Lemierre himself stated: "The appearance and repetition several days after the onset of a sore-throat (and particularly of a tonsillar abscess) of severe pyrexial attacks with an initial rigor, or still more certainly the occurrence of pulmonary infarcts and arthritic manifestations, constitute a syndrome so characteristic that mistake is almost impossible" (56).

## TROPICAL SKIN ULCERS

Tropical skin ulcers (synonyms: Naga sore, tropical septic ulcer, *ulcus tropicum*, tropical phagedenic ulcer, tropical sloughing phagedena) (63) are common among children and young adults in the tropics, but are not confined to these areas (64,65). Patients are predominantly between 5 and 15 years old, with those over 35 years of age being rarely affected (63). Although tropical skin ulcers are relatively common and are the leading cause of morbidity in parts of the tropics (65), they remain understudied. This is mainly because they usually occur in rural areas, away from large research centers (66). Much of what is known about the etiology and pathogenesis of tropical skin ulcers is due to epidemiologic studies by Adriaans, and an experimental study by McAdam, who induced these ulcers in 20 volunteers by bathing intact skin with ulcer pus for 6-10 days (63,67,68).

The major etiological factors of tropical skin ulcers have been identified as trauma and secondary infection. The trauma may be extremely minor, such as leg contact with previously infected shrubbery and plants (68). For this reason, the lesions usually occur on exposed skin (68,69). A small localized inflammatory reaction develops into a pustule about 1 cm in diameter after 5 or 6 days (68). Once the pustule ruptures, a foul-smelling blood-stained pus is discharged, and the round ulcer is raised above the surrounding edematous skin (68). The ulcer, which is usually solitary, involves the skin and subcutaneous tissues, but if the deep fascia is penetrated, the ulcer can destroy tendons, muscles, joints and bone (68,70). During its acute stage, the ulcer is extremely painful, leading to difficulties in sleeping and ambulating. An ulcer can bleed as much as 90 mL in 15 minutes (68). The margins of the ulcers often have what is described as pseudo-epitheliomatous hyperplasia (67); squamous carcinoma, perhaps due to the hyperplasia, occurs in about 2-15% of ulcers of more than 3 years duration (63). The nutritional status of the patient does not appear to be particularly relevant to the development of ulcers, as was once thought (63). Moisture is apparently necessary to induce infection and subsequent ulceration, as there is an increased incidence in the wet season (63,68). Patients often do not develop immunity to the infec-

tious agents; it has been observed that recurrence of the ulcers is possible if patients are re-exposed to the causative organisms (63).

Fusobacteria are the most frequently isolated bacteria from early tropical ulcers, having been implicated in about 35% of all cases (67). *F. nucleatum*, *F. necrophorum*, and *F. ulcerans* are the species usually associated with this disease, and the rapid tissue destruction involved has led to the inference of bacterial toxin production (5,64,65).

## INTRAAMNIOTIC INFECTION

Premature infants with low birth weights (<2500 g) are a major social and economic public health problem. A more intensive hospital-based management of low birth weight infants, and not a decline in incidence, has resulted in the most recent reductions of infant mortality in more developed nations (71-73). Although the exact pathogenesis of intraamniotic infection (IAI) (synonyms: clinical chorioamnionitis, amnionitis, amniotic fluid infection, intrapartum infection) is unknown, infection has been established as a major etiological factor in premature rupture of chorioamniotic membranes and as a cause of prematurity (74-79). The incidence of IAI has been reported to be 1-4% by Gibbs and Duff, although incidence rates up to 10.5% have also been described (74,75,80).

The hallmark of IAI is maternal fever, although uterine tenderness, foul-smelling amniotic fluid, maternal tachycardia, and fetal tachycardia have also been noted (74-76). Common maternal complications associated with IAI have been dysfunctional labor and postpartum infections (74,79,81). Neonatal complications are usually related to premature birth; the newborn may also be born with an infection or sepsis (78).

The bacteria most related to prematurity have been reported to be fusobacteria and group B streptococci (82). According to Altshuler and Hyde, fusobacterial infections have been associated with 18% of IAI cases that result in prematurity, but figures as high as 30% have been reported (77,83-88). Although the mechanism by which bacteria precipitate labor is unclear, translocation of endotoxins and maternally-produced prostaglandins in response to bacterial phospholipase A<sub>2</sub> have been implicated (71,77,89).

## OTHER INFECTIONS

Case reports have documented the involvement of fusobacteria in osteomyelitis (90,91), urinary tract infections (92), pulmonary nodules (93), infective endocarditis (94), pericarditis (95), septic arthritis of the sternoclavicular joint (96), liver and splenic abscesses (97-100), fatal pneumonia (101), and disseminated intravascular coagulation (102), among others. In most of these infections, the most probable route of entry for fusobacteria is the oropharynx. Fusobacterial sternoclavicular infections are now exceedingly rare, but in the preantibiotic era, *F. necrophorum* was the most important anaerobe in these infections (96). However, splenic abscesses

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may be more common in specific subgroups within the population, particularly intravenous drug users who have the habit of licking needles before injection to ease passage of the needle, to check the potency of the drug, and to ensure that the bevel is sharp (99,100).

Although *F. nucleatum* and *F. necrophorum* are considered the most pathogenic in this genus, other fusobacteria do occasionally cause infection. *F. russi* has been associated with animal bite infections, and *F. varium* with conjunctivitis and intra-ocular infections (5). *F. mortiferum* sepsis has been documented as well (103).

## THE ROLE OF ANTIMICROBIAL THERAPY IN FUSOBACTERIAL INFECTIONS

In general, fusobacteria display variable resistance to vancomycin, erythromycin, amoxicillin, ampicillin, and aminoglycosides (e.g. neomycin) (1,101,104-109). There are no general guidelines for choosing an antimicrobial agent, but antibiotic susceptibility testing is advised if the case is not urgent. Otherwise, empiric use of broad spectrum antibiotics with activity against anaerobes will suffice (110). Some commonly used antibiotics for anaerobic coverage include metronidazole, imipenem, and penicillins (110).

In periodontal diseases, antibiotic therapy is usually directed against *P. gingivalis*, *B. forsythus*, and *A. actinomycetemcomitans*, and typically involves a combination of a common tetracycline or penicillin antibiotic with metronidazole or ciprofloxacin (29). Antibiotics can be administered systemically or locally in the periodontal pocket. *F. nucleatum* often displays resistance to tetracyclines, and beta-lactamase producing strains are becoming more common (1,106,108,111). It has been estimated that 40-60% of clinically isolated fusobacteria strains are beta-lactamase producing, but the clinical significance of this finding is not yet clear (110). Treatment of periodontal diseases is not usually directed toward elimination of *F. nucleatum*, although certainly many antibiotic therapies may act against this organism (29).

Despite *in vitro* susceptibility to a number of antibiotics, numerous case reports have documented the ineffectiveness of many antibiotics in treating *F. necrophorum* infections (Lemierre's syndrome). As a result, the drug of choice is a combination of metronidazole usually with a penicillin for aerobic coverage, although certainly other drugs may also be effective (1,6,24,53-55,59,60,62,90,101,102,112,113). Surgical interventions may be necessary in some cases (54). If treatment is delivered effectively, full recovery without sequelae is the rule unless there is cerebral involvement or osteomyelitis. Antibiotic treatment should be at least 6 weeks in duration for Lemierre's syndrome (58).

Antibiotics are effective in the early stages of tropical skin ulcers (64,68), but they should be administered systemically, as local application often causes sensitization (63,114). The epithelium usually begins healing around the margin of the ulcer within 24 hours of administration (68). For more severe ulcers, skin grafts may be necessary (68).

There is no broad agreement on the selection of antibiotics for IAI, but there is accordance that both antibiotic therapy and delivery are essential to cure this condition (75). Gibbs and Duff report that many retrospective and prospective studies have evaluated the use of a penicillin with an aminoglycoside (75).

## CONCLUSION

Fusobacteria are capable of producing infections that result in significant morbidity and mortality. The presence of *F. nucleatum* in the mouth is critical to the development of periodontal diseases, which affect a significant proportion of elderly patients. *F. necrophorum* causes Lemierre's syndrome, an entity which is less common in the antibiotic era, but which can have potentially devastating consequences if unrecognized. Lemierre's syndrome usually affects young adults, and often begins as a pharyngeal infection. Tropical skin ulcers have been attributed to a number of different species of fusobacteria; although the incidence of these ulcers seems to be declining with better living standards, they remain a significant cause of morbidity in parts of the tropics. Fusobacteria, although not the most common cause of IAI, have been associated with a significant proportion of premature births. The incidence of prematurity does not appear to have declined. Fusobacterial infections can affect a wide range of age groups throughout the world. The presence of fusobacteria as part of the normal flora of the oropharynx, female genital tract, and gastrointestinal system appears to be critical to the pathogenesis of a number of these infections.

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## REFERENCES

1. Bolstad AI, Jensen HB, Bakken V. Taxonomy, biology, and periodontal aspects of *Fusobacterium nucleatum*. *Clin Microbiol Rev* 1996;9:55-71.
2. Smalley JW. Pathogenic mechanisms in periodontal disease. *Adv Dent Res* 1994;8:320-328.
3. Tuner K, Baron EJ, Summanen P, Finegold SM. Cellular fatty acids in *Fusobacterium* species as a tool for identification. *J Clin Microbiol* 1992;30:3225-3229.
4. Lawson PA, Gharbia SE, Shah HN, Clark DR. Recognition of *Fusobacterium nucleatum* subgroups Fn-1, Fn-2, and Fn-3 by ribosomal RNA gene restriction patterns. *FEMS Microbiol Lett* 1989;65:41-46.
5. Bennett KW, Eley A. Fusobacteria: new taxonomy and related diseases. *J Med Microbiol* 1993;39:246-254.
6. Burden P. *Fusobacterium necrophorum* and Lemierre's syndrome. *J Infect* 1991;23:227-231.
7. Falkow S. *Bacteroides* and *Fusobacterium*. In: Davis BD, Dulbecco R, Eisen HN, Ginsburg HS, eds. *Microbiology*. 4<sup>th</sup> ed. Philadelphia: J.B. Lippincott, 1990:589-593.

8. Wactawski-Wende J, Grossi SG, Trevisan M, Genco RJ, Tezal M, Dunford RG, Ho AW, Hausmann E, Hreshchyshyn MM. The role of osteopenia in oral bone loss and periodontal disease. *J Periodontol* 1996;67:1076-1084.
9. Genco RJ. Current view of risk factors for periodontal diseases. *J Periodontol* 1996;67:1041-1049.
10. Committee on Research, Science and Therapy. Periodontal disease as a potential risk factor for systemic diseases. *J Periodontol* 1998;69:841-850.
11. Silver JG, Martin AW, McBride BC. Experimental transient bacteremias in human subjects with varying degrees of plaque accumulation and gingival inflammation. *J Clin Periodontol* 1977;4:92-99.
12. Kaye D. Infective endocarditis. In: Fauci AS, Braunwald E, Isselbacher KJ, Wilson JD, Martin JB, Kasper DL, Hauser SL, Longo DL, eds. *Harrison's Principles of Internal Medicine* 14<sup>th</sup> ed. New York: McGraw-Hill, 1998:785-791.
13. Gristina AG. Biomaterial-centered infection: microbial adhesion versus tissue integration. *Science* 1987;237:1588-1595.
14. Thom DH, Grayston JT, Siscovick DS, Wang S, Weiss NS, Daling JR. Association of prior infection with *Chlamydia pneumoniae* and angiographically demonstrated coronary artery disease. *JAMA* 1992;268:68-72.
15. Saikku P, Leinonen M, Tenkanen L, Linnanmaki E, Ekman M, Manninen V. Chronic *Chlamydia pneumoniae* infection as a risk factor for coronary heart disease in the Helsinki heart study. *Ann Intern Med* 1992;116:273-278.
16. Lopes-Virella MF, Virella G. Immunological and microbiological factors in the pathogenesis of atherosclerosis. *Clin Immunol Immunopathol* 1985;37:377-386.
17. Beck J, Garcia R, Heiss G, Vokonas PS, Offenbacher S. Periodontal disease and cardiovascular disease. *J Periodontol* 1996;67:1123-1137.
18. Mattila KJ, Valle MS, Nieminen MS, Valtonen VV, Hietaniemi KL. Dental infections and coronary atherosclerosis. *Atherosclerosis* 1993;103:205-211.
19. DeStefano F, Anda RF, Kahn HS, Williamson DF, Russell CM. Dental disease and risk of coronary heart disease and mortality. *BMJ* 1993;306:688-691.
20. Williams RC, Mahan CJ. Periodontal disease and diabetes in young adults. *JAMA* 1960;172:776-778.
21. Miller LS, Manwell MA, Newbold D, Reding ME, Rasheed A, Blodgett J, Kornman KS. The relationship between reduction in periodontal inflammation and diabetes control: a report of 9 cases. *J Periodontol* 1992;63:843-848.
22. Ettinger RL. Oral disease and its effect on the quality of life. *Gerodontology* 1987;3:103-106.
23. Smith JM, Sheiham A. How dental conditions handicap the elderly. *Commun Dent Oral Epid* 1979;7:305-310.
24. Dahlen G. Role of suspected periodontopathogens in microbiological monitoring of periodontitis. *Adv Dent Res* 1993;7:163-174.
25. Liebana J, Castillo A. Physiopathology of primary periodontitis associated with plaque. Microbial and host factors. A review. Part 1. *Aust Dent J* 1994;39:228-232.
26. Liebana J, Castillo A. Physiopathology of primary periodontitis associated with plaque. Microbial and host factors. A review. Part 2. *Aust Dent J* 1994;39:310-315.
27. Enwonwu CO. Interface of malnutrition and periodontal diseases. *Am J Clin Nutr* 1995;61(Suppl):430S-436S.
28. Dibart S. Children, adolescents and periodontal diseases. *J Dent* 1997;25:79-89.
29. Section 11 Consensus Report from the 1996 World Workshop in Periodontics. Periodontal diseases: pathogenesis and microbial factors. *J Am Dent Assoc* 1998;129(Suppl):58S-62S.
30. Kolenbrander PE, London J. Adhere today, here tomorrow: oral bacterial adherence. *J Bacteriol* 1993;175:3247-3252.
31. Nyvad B, Kilian M. Microbiology of the early colonization of human enamel and root surfaces in vivo. *Scand J Dent Res* 1987;95:369-380.
32. Kolenbrander PE. Intrageneric coaggregation among human oral bacteria and ecology of dental plaque. *Annu Rev Microbiol* 1988;42:627-656.
33. Kolenbrander PE. Surface recognition among human oral bacteria: multigeneric coaggregations and their mediators. *Crit Rev Microbiol* 1989;17:137-159.
34. Kolenbrander PE, Andersen RN. Multigeneric aggregations among oral bacteria: a network of independent cell-to-cell interactions. *J Bacteriol* 1986;168:851-859.
35. Kolenbrander PE, Andersen RN, Holdeman LV. Coaggregation of oral *Bacteroides* species with other bacteria: central role in coaggregation bridges and competitions. *Infect Immun* 1985;48:741-746.
36. Kolenbrander PE, Andersen RN, Moore LVH. Coaggregation of *Fusobacterium nucleatum*, *Selenomonas flueggei*, *Selenomonas infelix*, *Selenomonas noxia*, and *Selenomonas sputigena* with strains from 11 genera of oral bacteria. *Infect Immun* 1989;57:3194-3203.
37. Kolenbrander PE, Andersen RN, Moore LVH. Intrageneric coaggregation among strains of human oral bacteria: potential role in primary colonization of the tooth surface. *Appl Environ Microbiol* 1990;56:3890-3894.
38. Kolenbrander PE, London J. Ecological significance of coaggregation among oral bacteria. *Adv Microb Ecol* 1992;12:183-217.
39. Dzink JL, Socransky SS, Haffajee AD. The predominant cultivable microbiota of active and inactive lesions of destructive periodontal diseases. *J Clin Periodontol* 1988;15:316-323.
40. Dzink JL, Tanner ACR, Haffajee AD, Socransky SS. Gram-negative species associated with active destructive periodontal lesions. *J Clin Periodontol* 1985;12:648-659.
41. Tanner A, Bouldin H. The microbiota of early periodontitis lesions in adults. *J Clin Periodontol* 1989;16:467-471.
42. Duerden BI. *Bacteroides*, *Fusobacterium*, and *Leptotrichia*. In: Parker MT, Collier LH, eds. *Topley and Wilson's principles of bacteriology, virology, and immunity*. 8<sup>th</sup> ed. Vol. 2 Philadelphia: B.C. Decker, 1990:552-575.
43. Hamada S, Koga T, Nishihara T, Fujiwara T, Okahashi N. Characterization and immunobiologic activities of lipopolysaccharides from periodontal bacteria. *Adv Dent Res* 1988;2:284-291.
44. Hase S, Hofstad T, Rietschel ET. Chemical structure of the lipid A component of lipopolysaccharides from *Fusobacterium nucleatum*. *J Bacteriol* 1977;129:9-14.
45. Hofstad T, Skaug N, Sveen K. Stimulation of B lymphocytes by lipopolysaccharides from anaerobic bacteria. *Clin Infect Dis* 1993;16(Suppl 4):200-202.
46. Horiba N, Maekawa Y, Yamauchi Y. Complement activation by lipopolysaccharides purified from gram-negative bacteria isolated from infected root canals. *Oral Surg Oral Med Oral Pathol* 1992;74:648-651.
47. Bartold PM, Gully NJ, Zilm PS, Rogers AH. Identification of components in *Fusobacterium nucleatum* chemostat-culture supernatants that are potent inhibitors of human gingival fibroblast proliferation. *J Periodont Res* 1991;26:314-322.
48. Singer RE, Buckner BA. Butyrate and propionate: important components of toxic dental plaque extracts. *Infect Immun* 1981;32:458-463.
49. Takada H, Ogawa T, Yoshimura F, Otsuka K, Koikeguchi S, Kato K, Umemoto T, Kotani S. Immunobiological activities of a porin fraction isolated from *Fusobacterium nucleatum* ATCC 10953. *Infect Immun* 1988;56:855-863.
50. Harder KJ, Nikaido H, Matsushashi M. Mutants of *Escherichia coli* that are resistant to certain beta-lactam compounds lack the *ompF* porin. *Antimicrob Agents Chemother* 1981;20:549-552.
51. Brokstad KA, Jensen HB. Purification and characterization of a 65-kDa diisopropylfluorophosphate-binding protein in the outer membrane of *Fusobacterium nucleatum* Fev1. *Scand J Dent Res* 1991;99:20-29.
52. Bakken V, Hogh BT, Jensen HB. Utilization of amino acids and peptides by *Fusobacterium nucleatum*. *Scand J Dent Res* 1989;97:43-53.
53. Brook I. Fusobacterial infections in children. *J Infect* 1994;28:155-165.
54. Moreno S, Altozano JG, Pinilla B, Lopez JC, de Quiros B, Ortega

- A, Bouza E. Lemierre's disease: postanginal bacteremia and pulmonary involvement caused by *Fusobacterium necrophorum*. *Rev Infect Dis* 1989;11:319-324.
55. Martin MJ, Wright ED. A case of *Fusobacterium necrophorum* sepsis. *J Infect* 1995;31:151-152.
  56. Lemierre A. On certain septicaemias due to anaerobic organisms. *Lancet* 1936;1:701-703.
  57. Sinave CP, Hardy GJ, Fardy PW. The Lemierre syndrome: suppurative thrombophlebitis of the internal jugular vein secondary to oropharyngeal infection. *Medicine* 1989;68:85-94.
  58. Eykyn SJ. Necrobacillosis. *Scand J Infect Dis* 1989;62(Suppl):41-46.
  59. Pace-Balzan A, Keith AO, Curley JWA, Ramsden RT, Lewis H. Otogenic *Fusobacterium necrophorum* meningitis. *J Laryng Otol* 1991;105:119-120.
  60. Jones TH, Bergvall V, Bradshaw, JPP. Carotid artery stenoses and thrombosis secondary to cavernous sinus thromboses in *Fusobacterium necrophorum* meningitis. *Postgrad Med J* 1990;66:747-750.
  61. Spencer CH, Slusher CW, Sanders CV, Aldridge KE. *Fusobacterium necrophorum* sepsis with cerebral infarction. *South Med J* 1989;82:1040-1043.
  62. Wolf RFE, Konings JG, Prins TR, Weits J. *Fusobacterium* pyomyositis of the shoulder after tonsillitis - report of a case of Lemierre's syndrome. *Acta Orthop Scand* 1991;62:595-596.
  63. Adriaans B. Tropical ulcer - a reappraisal based on recent work. *Trans Royal Soc Trop Med Hyg* 1988;82:185-189.
  64. Adriaans B, Garelick H. Cytotoxicity of *Fusobacterium ulcerans*. *J Med Microbiol* 1989;29:177-180.
  65. Falkler WA Jr, Montgomery J, Nauman RK, Alpers M. Isolation of *Fusobacterium nucleatum* and electron microscopic observations of spirochetes from tropical skin ulcers in Papua New Guinea. *Am J Trop Med Hyg* 1989;40:390-398.
  66. Tropical ulcers [editorial]. *Lancet* 1987;2:835.
  67. Adriaans B, Hay R, Drasar B, Robinson D. The infectious aetiology of tropical ulcer - a study of the role of anaerobic bacteria. *Br J Derm* 1987;116:31-37.
  68. McAdam I. Tropical phagedenic ulcers in Uganda: report of an investigation. *J Roy Coll Surg Edin* 1966;11:196-205.
  69. Maegrath B. Tropical ulcer. In: *Adams and Maegrath: clinical tropical diseases*. 9<sup>th</sup> ed. London: Blackwell, 1989:364-368.
  70. Adriaans B, Hay RJ, Drasar BS, Robinson DCA. Anaerobic bacteria in tropical ulcer - the application of a new transport system for their isolation. *Trans Royal Soc Trop Med Hyg* 1986;80:793-794.
  71. Offenbacher S, Katz V, Fertik G, Collins J, Boyd D, Maynor G, McKaig R, Beck J. Periodontal infection as a possible risk factor for preterm low birth weight. *J Periodontol* 1996;67:1103-1113.
  72. McCormick MC. The contribution of low birth weight to infant mortality and childhood mortality. *N Engl J Med* 1985;312:82-90.
  73. Williams RL, Chen PM. Identifying the sources of the recent decline in perinatal mortality in California. *N Engl J Med* 1982;306:207-214.
  74. Casey BM, Cox SM. Chorioamnionitis and endometritis. *Infect Dis Clin North Am* 1997;11:203-222.
  75. Gibbs RS, Duff P. Progress in pathogenesis and management of clinical intramniotic infection. *Am J Obstet Gynecol* 1991;164:1317-1326.
  76. Gibbs RS, Blanco JD, St. Clair PJ, Castaneda YS. Quantitative bacteriology of amniotic fluid from patients with clinical intraamniotic infection at term. *J Infect Dis* 1982;145:1-8.
  77. Altshuler G, Hyde S. *Fusobacteria*: an important cause of chorioamnionitis. *Arch Pathol Lab Med* 1985;109:739-743.
  78. Sperling RS, Newton E, Gibbs RS. Intraamniotic infection in low-birth-weight infants. *J Infect Dis* 1988;157:113-117.
  79. Daikoku NH, Kaltreider DF, Johnson TB Jr, Johnson JC, Simmons MA. Premature rupture of membranes and preterm labor: neonatal infection and perinatal mortality risks. *Obstet Gynecol* 1981;58:417-425.
  80. Soper DE, Mayhall CG, Dalton HP. Risk factors for intraamniotic infection: a prospective epidemiologic study. *Am J Obstet Gynecol* 1989;161:562-568.
  81. Siegel JD, McCracken GH Jr. Sepsis neonatorum. *N Engl J Med* 1981;304:642-647.
  82. Hillier SL, Krohn MA, Kiviat NB, Watts DH, Eschenbach DA. Microbiological causes and neonatal outcomes associated with chorioamnion infection. *Am J Obstet Gynecol* 1991;165:955-961.
  83. Altshuler G, Hyde S. Clinicopathologic considerations of *Fusobacteria* chorioamnionitis. *Acta Obstet Gynecol Scand* 1988;67:513-517.
  84. Easterling TR, Garite TJ. *Fusobacterium*: anaerobic occult amnionitis and premature labor. *Obstet Gynecol* 1985;66:825-828.
  85. Wallace RL, Herrick CN. Amniocentesis in the evaluation of premature labor. *Obstet Gynecol* 1981;57:483-486.
  86. Bobitt JR, Hayslip CC, Damato JD. Amniotic fluid infection as determined by transabdominal amniocentesis in patients with intact membranes in premature labor. *Am J Obstet Gynecol* 1981;140:947-952.
  87. Wahbeh CJ, Hill GB, Eden RD, Gall SA. Intra-amniotic bacterial colonization in premature labor. *Am J Obstet Gynecol* 1984;148:739-743.
  88. Hameed C, Tejani N, Verma UL, Archibald F. Silent chorioamnionitis as a cause of preterm labor refractory to tocolytic therapy. *Am J Obstet Gynecol* 1984;149:726-730.
  89. Bejar R, Curbelo V, Davis C, Gluck L. Premature labor. II. Bacterial sources of phospholipase. *Obstet Gynecol* 1981;57:479-482.
  90. Foulkes GD, Johnson CE, Katner HP. *Fusobacterium osteomyelitis* associated with intraosseous gas. *Clin Orthoped Rel Res* 1990;251:246-248.
  91. Beauchamp RD, Cimolai, N. Osteomyelitis of the pelvis due to *Fusobacterium nucleatum*. *Can J Surg* 1991;34:618-620.
  92. Ribot S, Gal K, Goldblat MV, Eslami HH. The role of anaerobic bacteria in the pathogenesis of urinary tract infections. *J Urol* 1981;126:852-853.
  93. Hsu CY, Luh KT. Cytology of pulmonary *Fusobacterium nucleatum* infection. *Acta Cytol* 1995;39:114-117.
  94. Shammam NW, Murphy GW, Eichelberger J, Klee D, Schwartz R, Bachman W. Infective endocarditis due to *Fusobacterium nucleatum*: case report and review of the literature. *Clin Cardiol* 1993;16:72-75.
  95. Truant AL, Menge S, Milliorn K, Lairscey R, Kelly MT. *Fusobacterium nucleatum* pericarditis. *J Clin Microbiol* 1983;17:349-351.
  96. Lau ES, Shuckett R. *Fusobacterium* septic arthritis of the sternoclavicular joint. *J Rheumatol* 1993;20:1979-1981.
  97. Crippin JS, Wang KK. An unrecognized etiology for pyogenic hepatic abscesses in normal hosts: dental disease. *Am J Gastroenterol* 1992;7:1740-1743.
  98. Scoular A, Corcoran GD, Malin A, Evans BA, Davies A, Miller RF. *Fusobacterium nucleatum* bacteraemia with multiple liver abscesses in an HIV-1 antibody positive man with IgG<sub>2</sub> deficiency. *J Infect* 1992;24:321-325.
  99. Sastre J, Casas E, Sierra J, Puig JG, Gil A. Splenic abscess due to *Fusobacterium necrophorum*. *Rev Infect Dis* 1991;13:1249-1250.
  100. Haber SW, Perlino CA. Splenic abscess from *Fusobacterium nucleatum*. *Ann Int Med* 1989;110:948.
  101. Paaske PB, Rasmussen BM, Illum P. *Fusobacterium pneumonia* and death following uvulo-palato-pharyngoplasty. *Head & Neck* 1994;16:450-452.
  102. Page Y, Comtet C, Tardy B, Zeni F, Thevenet D, Lucht F, Bertrand JC. Disseminated intravascular coagulation in *Fusobacterium necrophorum* septicemia. *Scand J Infect Dis* 1990;22:743-747.
  103. Prout J, Glymph R. Anaerobic septicemia secondary to *Fusobacterium mortiferum*. *J Nat Med Assoc* 1986;78:334-335.
  104. Abu Fanas SH, Drucker DB, Hull PS, Reeder JC, Ganguli LA. Identification, and susceptibility to seven antimicrobial agents, of 61 Gram-negative anaerobic rods from periodontal pockets. *J Dent* 1991;19:46-50.
  105. Appelbaum PC, Spangler SK, Jacobs MR. Susceptibility of 539 Gram-positive and Gram-negative anaerobes to new agents, including RP59500, biapenem, trospectomycin, and piperacillin/tazobactam. *Antimicrob Chemother* 1993;32:223-231.

106. Hedberg M, Lindqvist L, Tuner K, Nord CE. Effect of clavulanic acid, sulbactam and tazobactam on three different b-lactamases from *Bacteroides uniformis*, *Clostridium butyricum* and *Fusobacterium nucleatum*. *J Antimicrob Chemother* 1992;30:17-25.
107. Rowland MD, Del Bene VE, Lewis JW. Factors affecting antimicrobial susceptibility of *Fusobacterium* species. *J Clin Microbiol* 1987;25:476-479.
108. Tuner K, Lindqvist L, Nord CE. Characterization of a new b-lactamase from *Fusobacterium nucleatum* by surface profiles and chromatofocusing patterns. *J Antimicrob Chemother* 1985;16:23-30.
109. Williams JD, Maskell JP, Shain H, Chrysos G, Sefton AM, Fraser HY, Hardie JM. Comparative in-vitro activity to azithromycin, macrolides (erythromycin, clarithromycin and spiramycin) and streptogramin RP59500 against oral organisms. *J Antimicrob Chemother* 1992;30:27-37.
110. Kasper DL. Infections due to mixed anaerobic organisms. In: Fauci AS, Braunwald E, Isselbacher KJ, Wilson JD, Martin JB, Kasper DL, Hauser SL, Longo DL, eds. *Harrison's Principles of Internal Medicine* 14<sup>th</sup> ed. New York: McGraw-Hill, 1998:991-997.
111. Roberts MC, Moncla BJ. Tetracycline resistance and TetM in oral anaerobic bacteria and *Neisseria perflava-N. sicca*. *Antimicrob Agents Chemother* 1990;32:1271-1273.
112. Landsaat PM, van der Lelie H, Bongaerts G, Kuijper EJ. *Fusobacterium nucleatum*, a new invasive pathogen in neutropenic patients? *Scand J Infect Dis* 1995;27:83-84.
113. Ray MS, Feldman S. Mixed *Fusobacterium* and *Actinomyces* pulmonary infection. *Clin Ped* 1989;28:426-428.
114. Angelini G, Rantuccio F. Contact dermatitis in patients with leg ulcers. *Cont Derm* 1975;1:81-87.



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